

Supplementary Material

Defining and enhancing the biosynthesis of astaxanthin and Docosahexaenoic acid in *Aurantiochytrium* sp. SK4

Jingrun Ye^{a, b}, Mengmeng Liu^{a, b}, Mingxia He^a, Ying Ye^{a, b}, Junchao Huang^{a*}

^aKey Laboratory of Economic Plants and Biotechnology, Yunnan Key Laboratory for Wild Plant Resources, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201

^bUniversity of Chinese Academy of Sciences, Beijing 100049, People's Republic of China

***Corresponding author**

Email: huangjc@mail.kib.ac.cn.

Phone: +86-875-65228058

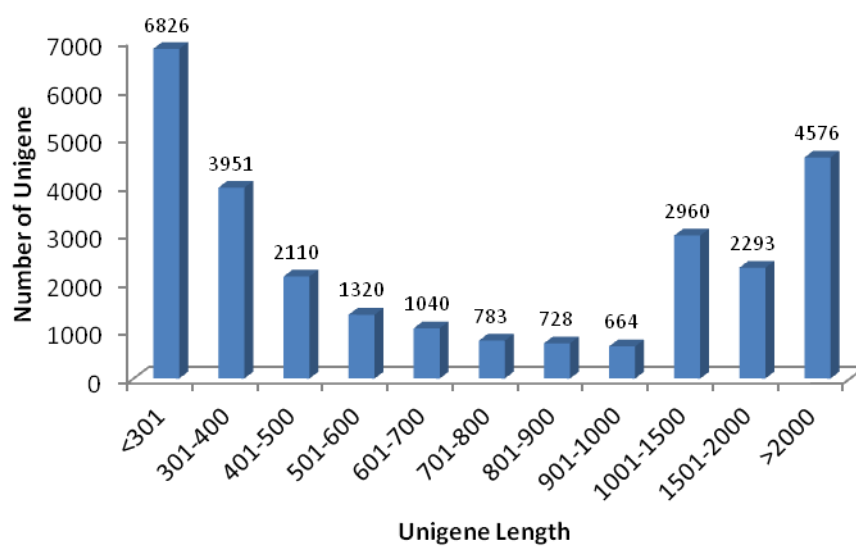


Figure S1. Uni-gene length distribution.

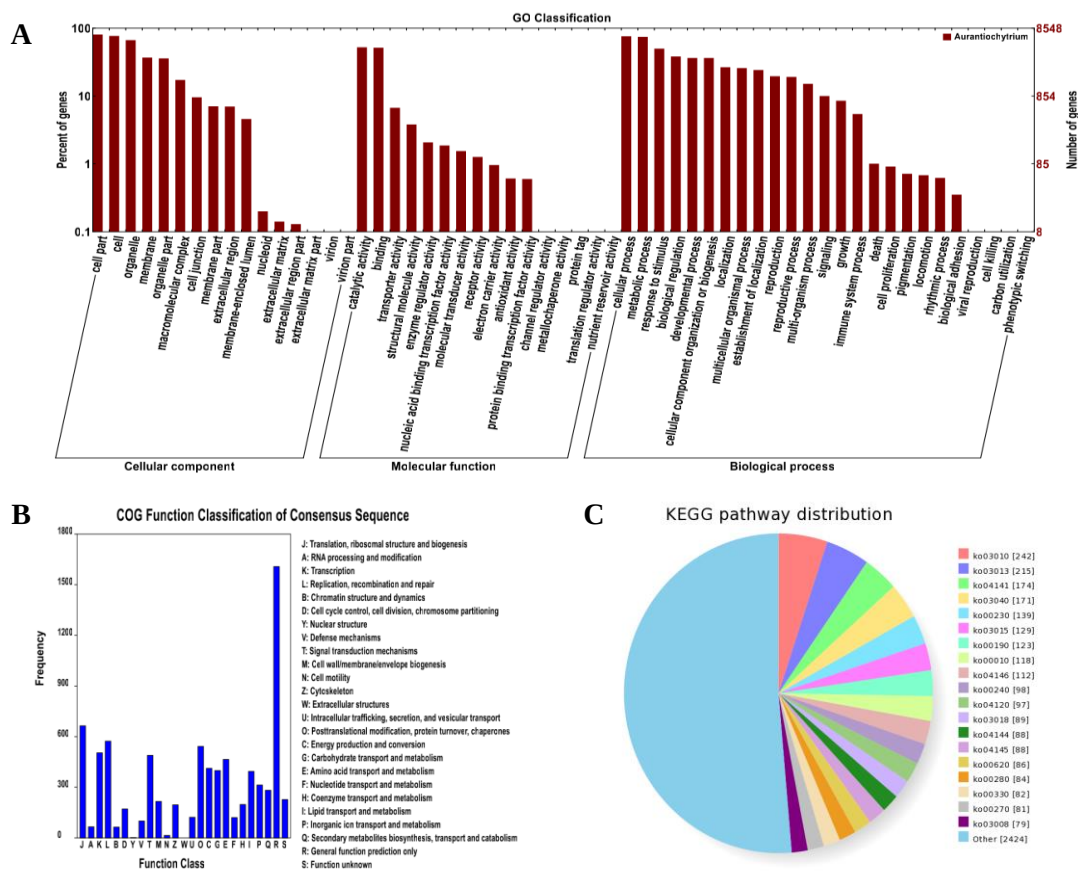


Figure S2. Functional annotation of assembled uni-genes in *Aurantiochytrium* sp. SK4. (A) GO classification, (B) COG classification, (C) KEGG pathway.

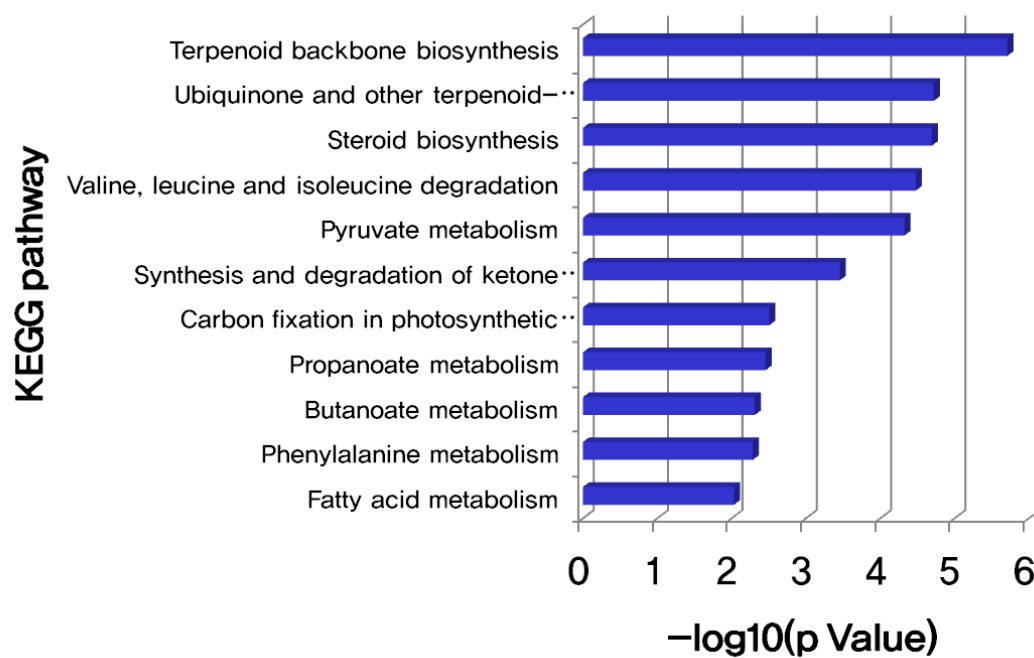


Figure S3. KEGG pathway enrichment analysis of differentially expressed genes (DEGs).

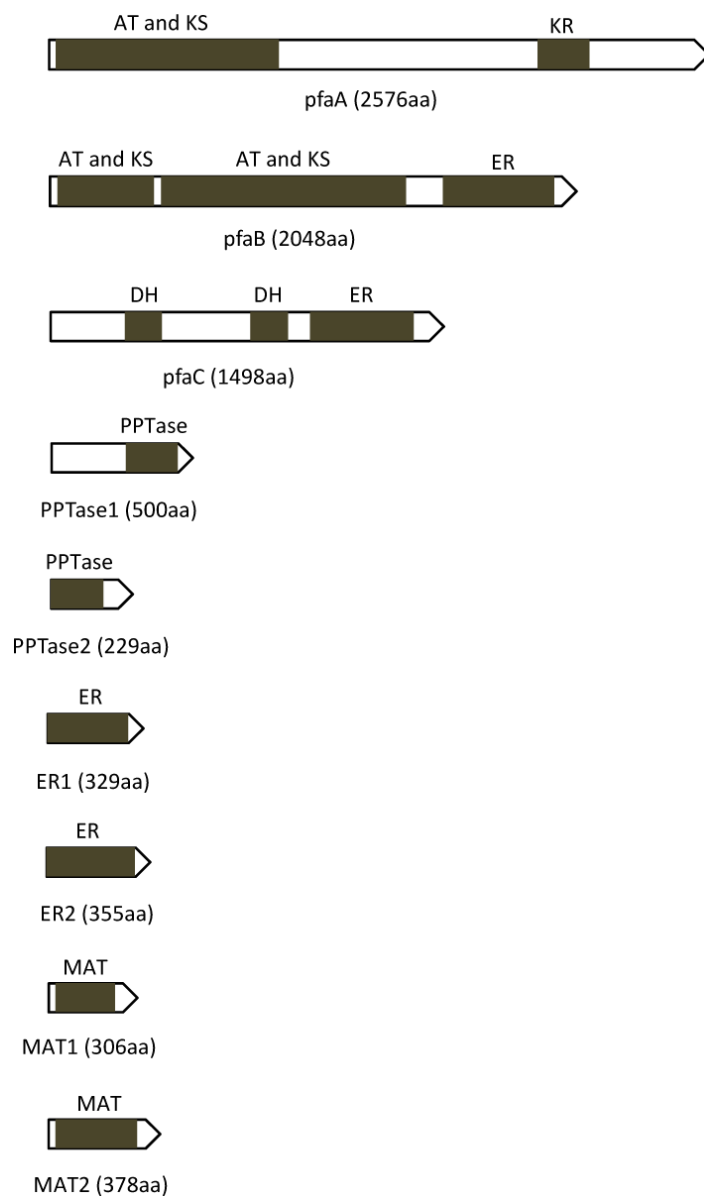


Figure S4. Genes encoding enzymes of the polyketide synthase (PKS pathway) in *Aurantiochytrium* sp.SK4. Dark gray areas indicate proposed enzymatic domains. KS, 3-ketoacyl synthase; KR, 3-ketoacyl-ACP reductase; ER, enoyl reductase; DH, dehydratase/isomerase; PPTase, phosphopantetheine transferase; MAT, malonyl-CoA:ACP acyltransferase. aa, amino acid.

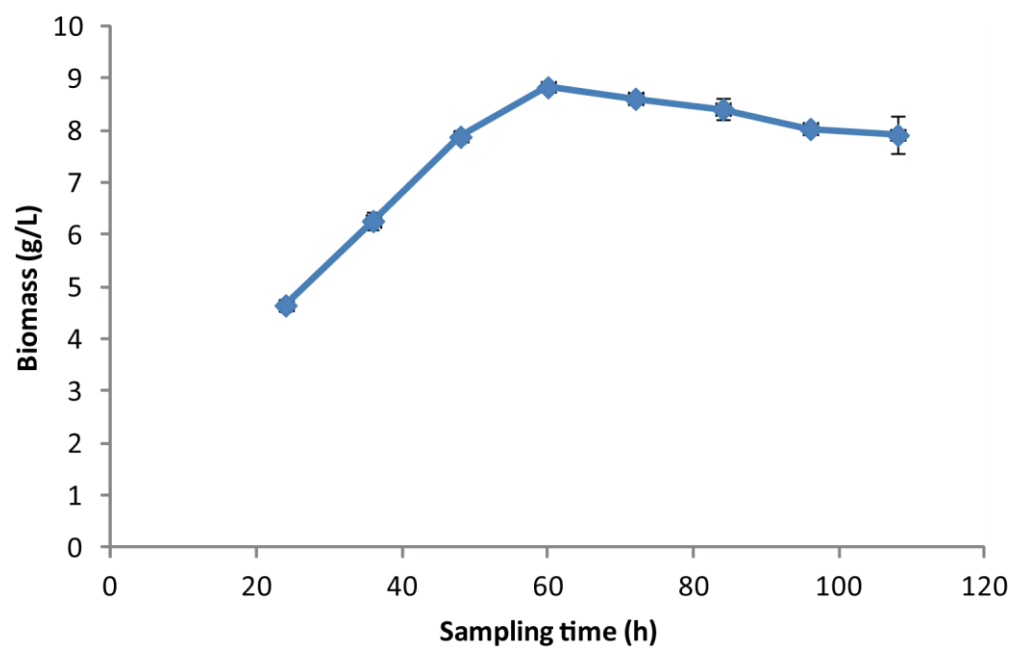


Figure S5. The growth curve of *Aurantiochytrium* sp.SK4 of Figure 1 and Figure 2.

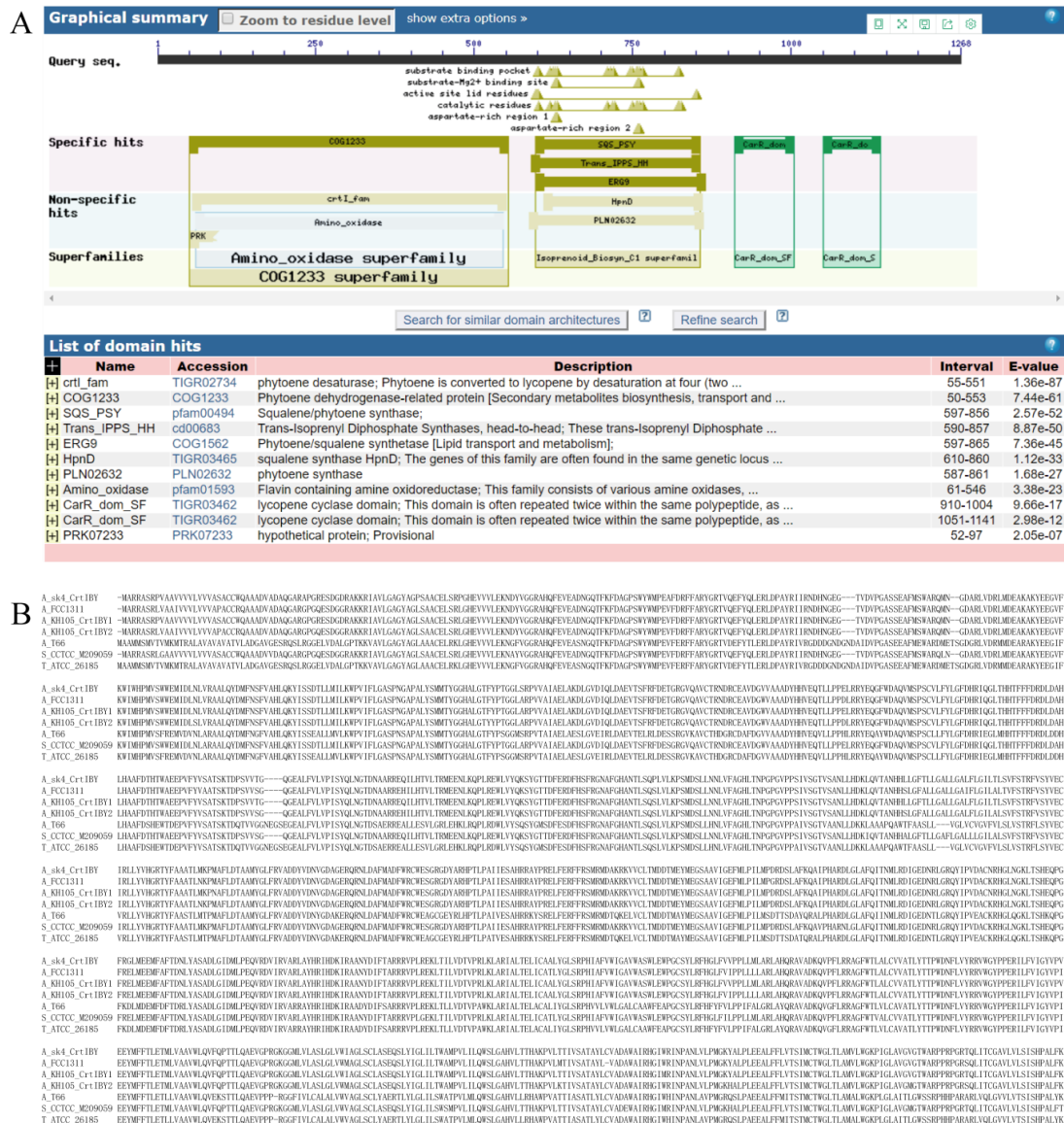


Figure S6. The conserved domains of CrtIBY and alignment of amino acid sequences of different CrtIBY. The conserved domains of CrtIBY from *Aurantiochytrium* sp.SK4 were analyzed by CDD/SPARCLE (A) and alignment of amino acid sequences of possible trifunctional β -carotene synthases, CrtIBY (B). Sequences of CrtIBY of *Aurantiochytrium* sp. SK4 are compared with those of *Aurantiochytrium* sp. FCC1311, *Aurantiochytrium* sp. KH105, *Schizochytrium* sp. CCTCC M209059, *Aurantiochytrium* sp. T66, and *Thraustochytrium* sp. ATCC 26185.

Table S1

Comparison of *Aurantiochytrium* sp. SK4 genome statistics to other five algae and the *Arabidopsis thaliana* genome.

Organism	<i>Aurantiochytrium</i> sp. SK4	<i>Arabidopsis</i> <i>thaliana</i>	<i>Chlamydomonas</i> <i>reinhardtii</i>	<i>Chlorella</i> sp. NC64A	<i>Chromocloris</i> <i>zofingiensis</i>	<i>Coccomyxa</i> <i>subellipsoides</i> C-169	<i>Monoraphidium</i> <i>neglectum</i>
Sequenced genome size	49Mbp	119Mbp	107Mbp	42Mbp	58Mbp	49Mbp	67Mbp
Percent G+C in sequenced genome	56.7%	36%	64%	67%	51%	53%	65%
Coding sequence in sequenced genome	63.0%	28%	37%	32%	39%	25%	26%
Percent G+C in coding sequence	56.9%	44%	70%	69%	53%	61%	70%
Average number of exons	2.4	5.2	8.5	8.3	5.0	8.1	5.0
Average exon length	903nt	237nt	261nt	166nt	291nt	159nt	207nt
Percentage transcript with at least one intron	55.5%	76%	92%	98%	82%	94%	82%

Table S2

The expression of genes associated with carotenoids and fatty acid biosynthesis in transcriptome.

Pathway	Gene	RPKM	
		24h	96h
Astaxanthin pathway	<i>HMGS</i>	293.68	2.67
	<i>HMGR</i>	141.10	1.42
	<i>MK</i>	194.72	10.42
	<i>PMK</i>	21.56	12.61
	<i>PPMD</i>	60.02	1.11
	<i>IDI</i>	29.57	1.29
	<i>CrtI</i> <i>BY</i>	9.10	19.95
	<i>CrtZ</i>	3.87	12.66
	<i>CrtO</i>	14.94	28.95
FAS pathway	<i>Type 1 fatty acid synthase</i>	613.72	247.47
	$\Delta 12$ desaturase	377.4	2.0
	$\Delta 5$ desaturase	18.2	29.1
	ω -3 desaturase	0	0.6
	$\Delta 4$ desaturase	114.4	19.7
	$\Delta 6$ desaturase	0.7	0
	$\Delta 9$ desaturase	1.08	0
PKS pathway	<i>PKS pfaA</i>	754.56	1.95
	<i>PKS pfaB</i>	193.79	9.46
	<i>PKS pfaC</i>	381.54	4.38

Table S3

Contents of squalene in wild-type SK4 and the transformant AT26 at different stages.

	Sampling time	SK4	AT26
Squalene content (mg × g ⁻¹ DW)	48h	10.98 ± 0.13	0.34 ± 0.02
	72h	21.08 ± 0.06	0.41 ± 0.06
	96h	13.18 ± 0.05	0.64 ± 0.07

Data are shown as mean ± SD, n = 3.

Table S4

Primers Used for qRT-PCR and the detection of the p--*VHb-ble-2A-IDI-2A-GPS* (VBIG).

Primer	Sequence
qRT-pcr-Actin-F	GAGGCCATGTTTCAGACCAT
qRT-pcr-Actin-R	ACGAGAGCCGTCATTTCTGT
qRT-pcr-HMGS-F	CGCCGGCGTCGACAGCAT
qRT-pcr-HMGS-R	GGGCACGGCGGGCAAGAC
qRT-pcr-HMGR-F	CCGGCGCAAAATGTCGAGTCT
qRT-pcr-HMGR-R	CCGCCGACAGTGCCAACCTC
qRT-pcr-MK-F	CCGCAACCACGAAATCCTCCAAAA
qRT-pcr-MK-R	CGAGAGCGCCGGCAGACTTG
qRT-pcr-PMK-F	GCCGTCTTTGCAGTTGTTGTTGATTG
qRT-pcr-PMK-R	GCCGCCGATCTTCACTCAGCAA
qRT-pcr-crtIBY-F	TGGTGACCTCGATCATGTGT
qRT-pcr-crtIBY-R	CGGCTCTACAGGTAATGAGT
qRT-pcr-FAS I -R	GAGAACGTCAGCACCTTTGC
qRT-pcr-FAS I -F	AGGCTCGAGAGAGCCTTGAC
qRT-pcr-pfaA-F	TGATCCCTTCGTGAATGACC
qRT-pcr-pfaA-R	GCTCGTTGTGGAAGTGAAGG
qRT-pcr-pfaB-F	GTCATTCTGCCCCTCATCATCAACC
qRT-pcr-pfaB-R	GACTGCTTGGCGACCTGGTTTAC
qRT-pcr-pfaC-F	CCGCCCCATCCACGTCATCCTC
qRT-pcr-pfaC-R	CCGGACTGCTTGGCGACCTGGTT
VBIGF	GGCTTTGGCGATGACGGTATTG
VBIGR	CCCCTCCTCATCTCGTCCCTGT