

Supplementary material for: Enzyme-linked phage receptor binding protein assay (ELPRA) enables identification of *Bacillus anthracis* colonies

Peter Braun ‡, Nadja Rupprich ‡, Diana Neif and Gregor Grass *

Bundeswehr Institute of Microbiology (IMB), Dept. of Bacteriology and Toxinology, 80937 Munich, Germany;
peter3braun@bundeswehr.org (P.B.); nadjarupprich@googlemail.com (N.R.); diananeif@bundeswehr.org (D.N.);
gregorgrass@bundeswehr.org (G.G.)

* Correspondence: gregorgrass@bundeswehr.org; Tel.: +49-99-2692-3981

‡ Both authors contributed equally to this work.

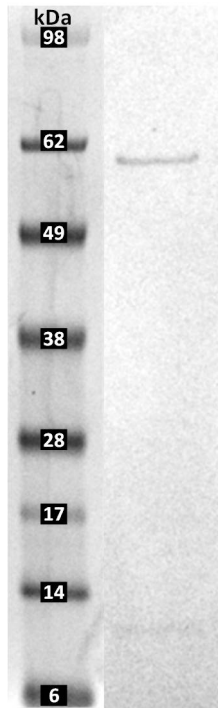
Supplementary Table S1: Strains, used in this work.

Species	Strain	Characteristics	Reference
<i>B. anthracis</i>	Sterne	pXO1 ⁺ , pXO2 ⁻ ; host of Gamma phage ⁻	[16]
<i>B. anthracis</i>	ATCC4229 (Pasteur)	pXO1 ⁻ , pXO2 ⁺ ; host of Gamma phage	American Type Culture Collection
<i>B. cereus</i>	ATCC10987	Not a host of Gamma phage	American Type Culture Collection
<i>B. cereus</i>	ATCC4342	Host of Gamma phage	American Type Culture Collection
<i>B. cereus</i>	CDC2000032805	Host of Gamma phage	Centers of Disease Control and Prevention
<i>B. cereus</i>	IMB-4-0-Rott	Not a host of Gamma phage; weak β -hemolysis on blood agar	This work; strain collection of the Bundeswehr Institute of Microbiology
<i>B. cereus</i> s.l.	IMB-2021-1	Host of Gamma phage, no β -hemolysis on blood agar	This work; strain collection of the Bundeswehr Institute of Microbiology

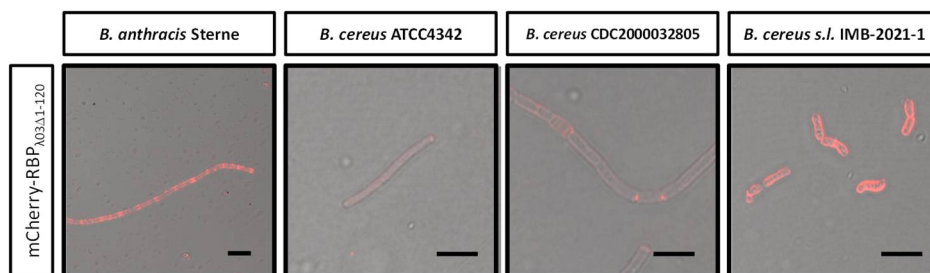
Supplementary Table S2: Oligonucleotide primers used for DNA sequencing and cloning in this work.

Oligonucleotide	Sequence (5'-3')	Reference
27f	AGAGTTTGATCMTGGCTCAG	[21]
341f	CCTACGGGAGGCAGCAG	[22]
534r	ATTACCGCGGCTGCTGG	[22]
1492r	TACGGYTACCTTGTTACGACTT	[21]
NanoLuc forward	AAAT <u>GTACAGTCTT</u> CACACTCGAAGATTTCGTT	This study
NanoLuc reverse	AAACTCGAGCGCCAGAATGCGTTCGCA	This study

M = A or C; Y = C or T. Restriction endonuclease recognition sites are underscored.



Supplementary Figure S1: SDS-PAGE of heterologously produced NanoLuc-RBP $\Delta 03\Delta 1-120$ -reporter protein. Affinity purified fusion protein was subjected to SDS-PAGE and NanoLuc-RBP $\Delta 03\Delta 1-120$ protein detected using ImperialTM protein stain (ThermoScientific, Dreieich, Germany). The expected size of the NanoLuc-RBP $\Delta 03\Delta 1-120$ protein was calculated as 66 kDa. Numbers indicate size-positions of the protein size marker (SeeBlue Plus2 prestained, ThermoFisher Scientific, Darmstadt, Germany).



Supplementary Figure S2: Binding of mCherry-RBP $\Delta 03\Delta 1-120$ -reporter to cells of cultures of *B. anthracis* and cross-reacting *B. cereus* cells, respectively. Cultures were grown overnight (16 h) at 37°C with shaking. Cells were labeled (red signals) by adding mCherry-RBP $\Delta 03\Delta 1-120$ protein, washed to remove unbound RBP and subjected to microscopy. Representative micrographs for labeling of the indicated, *B. cereus* cells are depicted as merged images from light- and fluorescence (excitation: 587 nm, emission: 610 nm) microscopy. Scale bar: 5 μ m.

References

16. Sterne M, Proom H Induction of motility and capsulation in *Bacillus anthracis*. J Bacteriol 1957, 74: 541-542.
21. Lane DJ, Pace B, Olsen GJ, Stahl DA, Sogin ML, Pace NR Rapid determination of 16S ribosomal RNA sequences for phylogenetic analyses. Proc Natl Acad Sci U S A 1985, 82: 6955-6959.
22. Watanabe K, Kodama Y, Harayama S Design and evaluation of PCR primers to amplify bacterial 16S ribosomal DNA fragments used for community fingerprinting. Journal of Microbiological Methods 2001, 44: 253-262.