

Review

# Gut Microbial-Derived Short Chain Fatty Acids: Impact on Adipose Tissue Physiology

Karolline S. May<sup>1,2</sup> and Laura J. den Hartigh<sup>1,2,\*</sup> 

<sup>1</sup> Department of Medicine, Division of Metabolism, Endocrinology, and Nutrition, University of Washington, Seattle, WA 98109, USA

<sup>2</sup> UW Medicine Diabetes Institute, 750 Republican Street, Box 358062, Seattle, WA 98109, USA

\* Correspondence: lauradh@u.washington.edu

**Abstract:** Obesity is a global public health issue and major risk factor for pathological conditions, including type 2 diabetes, dyslipidemia, coronary artery disease, hepatic steatosis, and certain types of cancer. These metabolic complications result from a combination of genetics and environmental influences, thus contributing to impact whole-body homeostasis. Mechanistic animal and human studies have indicated that an altered gut microbiota can mediate the development of obesity, leading to inflammation beyond the intestine. Moreover, prior research suggests an interaction between gut microbiota and peripheral organs such as adipose tissue via different signaling pathways; yet, to what degree and in exactly what ways this inter-organ crosstalk modulates obesity remains elusive. This review emphasizes the influence of circulating gut-derived short chain fatty acids (SCFAs) i.e., acetate, propionate, and butyrate, on adipose tissue metabolism in the scope of obesity, with an emphasis on adipocyte physiology in vitro and in vivo. Furthermore, we discuss some of the well-established mechanisms via which microbial SCFAs exert a role as a prominent host energy source, hence regulating overall energy balance and health. Collectively, exploring the mechanisms via which SCFAs impact adipose tissue metabolism appears to be a promising avenue to improve metabolic conditions related to obesity.

**Keywords:** gut microbiome; short-chain fatty acids; adipose tissue; obesity; inflammation; acetate; propionate; butyrate; metabolism



**Citation:** May, K.S.; den Hartigh, L.J. Gut Microbial-Derived Short Chain Fatty Acids: Impact on Adipose Tissue Physiology. *Nutrients* **2023**, *15*, 272. <https://doi.org/10.3390/nu15020272>

Academic Editor: Jonathan Jacobs

Received: 15 December 2022

Revised: 28 December 2022

Accepted: 3 January 2023

Published: 5 January 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

### 1.1. Pathophysiology of Obesity

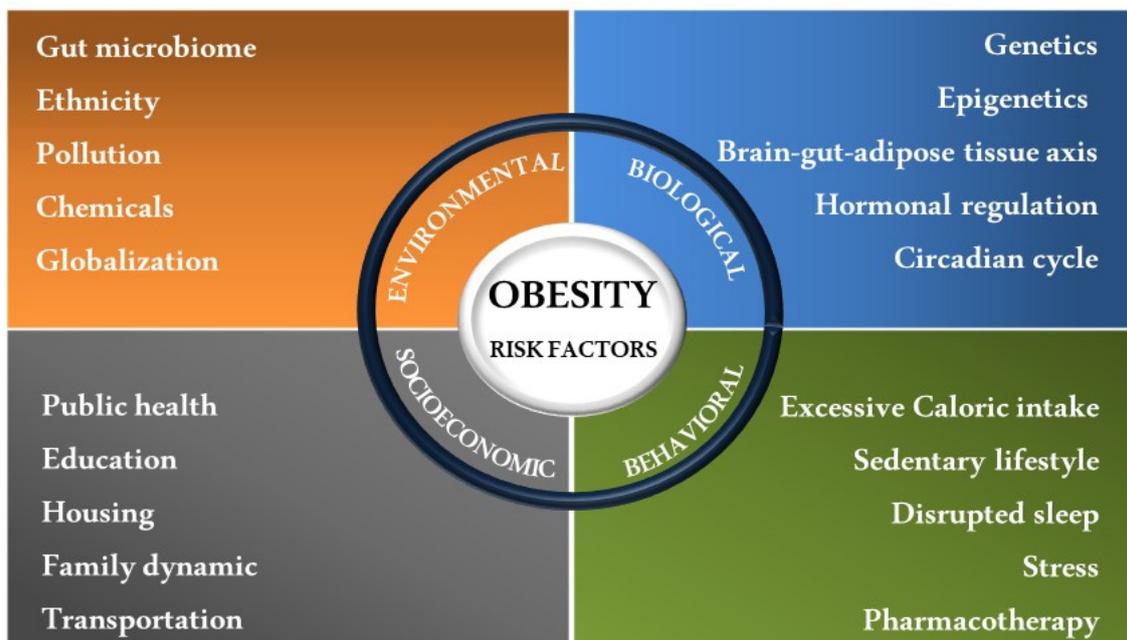
The prevalence of obesity has escalated in recent years, thus representing a serious impediment to public health worldwide [1]. Recent trends in the United States estimate that over 40% of the US population has either overweight or obesity [2], with this rate likely to be even higher among adults by 2030 [3,4]. Additionally, epidemiologic studies indicate that obesity-related pathological conditions, including type 2 diabetes (T2D) [5], dyslipidemia [6], coronary artery disease (CAD) [7,8], hepatic steatosis [9], and certain cancers [10–12], will continue to increase, potentially reducing life expectancy and increasing healthcare costs [13,14]. In this regard, there is an urgent need to identify biological mechanisms and effective approaches to manage obesity and its risk factors.

Obesity is a multifactorial disorder characterized by a sustained energy imbalance (energy intake > energy expenditure) [15], leading to adverse physiological consequences across individuals [1,16]. In obesity, adipose tissue (AT) metabolism plays an important role in driving the negative effects of increased adiposity [16,17]. As such, marked nuances in AT distribution throughout the body as well as AT phenotypes (white, brown, and beige) [18] play a key role in various metabolic complications [16]. Moreover, due to the energy imbalance that characterizes obesity, white adipose tissue (WAT) undergoes expansion and remodeling coupled with changes in AT-resident immune cells [19–21].

Failure to modulate WAT mass via hypertrophy in adipocytes, hyperplasia of resident pre-adipocytes, or combined mechanisms [18] drives excessive ectopic fat deposition in peripheral tissues (i.e., heart, liver, skeletal muscles, pancreas), imbalanced energy storage, increased reactive oxygen species (ROS) production, inadequate vascularization, hypoxia, hormonal dysregulation, and low-grade systemic inflammation, with macrophage infiltration and polarization [17,21–25]. Hence, through interactions with distinct organs and a multitude of signaling pathways, WAT contributes to cardiometabolic health, ultimately impacting whole-body energy homeostasis [17,21,26]. A more detailed description of the metabolic changes in AT elicited by obesity will be further discussed herein. Given the overall health burden, various hypotheses have emerged to explain the underlying causes of obesity that are summarized in [27–29], but will not be the main focus of this review.

### 1.2. Obesity and the Gut Microbiota

Evidence has suggested that the etiology of obesity is quite complex and depends on several elements [1,29,30]. A combination of individual behaviors and societal, biological, and environmental factors may affect body weight and adiposity [1,29,30] and should therefore be utilized to inform therapeutic approaches to obesity management and treatment. Among the environment-attributable factors related to obesity, research in animal models [31–35] and humans [36–38] has shown that the gut microbiota is a new keystone linked to obesity-related metabolic complications [39,40]. Previous research highlights the relevant contribution of the intestinal microbial ecosystem to energy harvest from the diet as well as its impact on genetic and diet-induced adiposity [41–45], suggesting that targeting the gut microbiota may be an avenue to treat environment-related obesity and other related metabolic diseases. Figure 1 depicts the wide spectrum of factors that influence obesity and related comorbidities.



**Figure 1. Obesity-attributable risk factors.** Targetable individual behavioral, societal, biological, and environmental factors that contribute to the complex etiology of obesity and obesity-related metabolic complications in humans.

The gut microbiota harbors a wide variety of bacterial genera, the majority of which are represented by the phyla *Firmicutes* and *Bacteroidetes*. These can be further distributed across many diverse bacterial species [46–48], thereby conferring the host with an individualized microbiota composition [46,49,50]. Interestingly, differences in the composition and distribution of the microbiota have been correlated to adiposity, as revealed by mecha-

nistic studies [35,37]. The composition of the diet, how nutrients are ingested, intestinal transit time, and the duration of fasting throughout the day can greatly affect microbial populations, which could modulate energy balance and body weight [51,52]. Initially, Ley et al. showed that obesity correlates with a decreased abundance of *Bacteroidetes* with a higher proportion of *Firmicutes*, whereas lean individuals harbored a higher proportion of *Bacteroidetes* by a corresponding degree [35]. Such profound changes in the bacterial community structure, associated with an alteration in body weight, were observed regardless of food consumption in genetically obese mice [35]. In parallel, reduced levels of *Bacteroidetes* were found in cecal samples from leptin-deficient (*ob/ob*) obese mice relative to their lean counterparts [32], an effect also observed in obese humans [37]. However, an accumulation of subsequent metagenomics data revealed that the *Firmicutes* to *Bacteroidetes* relative ratio is not a consistent indicator of obesity status, suggesting far more complexity regarding the imbalance of gut microbial phyla related to metabolic disease status [53–56]. The notion that alterations in microbial ecology can modulate obesity, or at least can contribute to a host-mediated adaptive response to energy imbalance, has led to many studies in germ-free (GF) and conventionalized mice [31–34,57]. Proof-of-concept experiments showed that an “obese microbiome” was capable of driving host adiposity through increased adipocyte lipoprotein lipase (LPL) activity, adipocyte hypertrophy [31], reduced proportions of *Bacteroidetes*, and an enrichment in genes related to energy metabolism pathways in adult GF mice [32,34]. These findings suggest that the gut microbiota may efficiently harvest energy from a given diet and act as a crucial environmental factor in regulating energy harvesting, storage, and expenditure [31,32,34]. Other mechanisms that explain how gut dysbiosis mediates obesity pathophysiology include increased intestinal permeability accompanied with metabolic endotoxemia, dysregulated gut immunity, contribution to low-grade systemic inflammation, and hormonal imbalance in the gut–brain–adipose axis [58–62]. Furthermore, particular gut microbial metabolites such as short-chain fatty acids (SCFAs), the majority of which include acetate, propionate, and butyrate, could also contribute to host energy homeostasis and have been extensively investigated in this context [63–71].

While gut microbiota are intimately involved in host energy metabolism, their contribution to weight loss, and potential direct signaling to adipose tissue, is less well-defined. Herein, the focus of this review will be on the direct influence of circulating gut-derived SCFAs on adipocyte metabolism *in vitro*. We also will discuss potential mechanisms whereby microbial SCFAs impact adipose tissue and adipocyte metabolism *in vivo*, hence regulating overall energy balance and health. Literature that contributed to this review was found using Pubmed, with search terms that included combinations of the terms SCFAs, adipose tissue, and obesity *in vitro* and *in vivo*. Collectively, exploring SCFAs as potential targets that impact the gut–adipose tissue axis may be a promising avenue to improve metabolic conditions related to obesity.

## 2. Gut Microbial-Derived Short Chain Fatty Acids (SCFAs) and Obesity

The composition and functional properties of the gut microbiota affect whole-body metabolism, thus playing a role in obesity and its complications [72,73]. As such, evidence indicates that nutritional strategies, including consumption of a fiber-rich or plant-based diet, seem to improve host bacterial richness and overall colonic health, with the potential to mitigate the metabolic complications of obesity [74–77]. Particular microbial metabolites, including SCFAs derived from the fermentation of indigestible dietary fibers [78,79], exert beneficial effects on both the host and its microbiome. SCFAs serve as (1) an energy source for both microbiota and the host to maximize gut health and (2) circulating factors facilitating the crosstalk between the host and gut microbiota [72,74]. Indeed, adipose tissue boasts a relatively high expression of SCFA receptors relative to other tissues (described in more detail in Section 3). In this context, precise roles of SCFAs and their direct impact on host adipose tissue metabolism have been underexplored [74,78,79].

SCFAs refer to organic monocarboxylic acids with fewer than six carbons, encompassing primarily acetate (C2), propionate (C3), and butyrate (C4) at an approximate molar

ratio of 3:1:1, respectively, in the intestinal lumen [78,80]. These small molecules account for approximately 95% of all SCFAs and are produced by specific bacterial strains depending on the type and amount of fiber consumed, the extent of substrate degradation, and colonic transit time [81–83]. Acetate, the most abundant systemic SCFA, and propionate are primarily produced by particular strains of *Bacteroidetes* (i.e., *Bacteroides* spp., *Bifidobacterium* spp., *Lactobacillus* spp.), whereas butyrate-producing bacteria are primarily of the phylum *Firmicutes* (i.e., *Enterococcus* spp., *Eubacterium* spp., *Roseburia* spp.) [84–86]. SCFAs circulate at high levels in rodents and humans, typically in the range of several hundred micromolar. In contrast to what is observed in the intestinal lumen, the majority of circulating SCFAs in mice and humans are represented by acetate, with a smaller proportion represented by propionate and butyrate [87–89], suggesting a potentially important systemic function of these SCFAs.

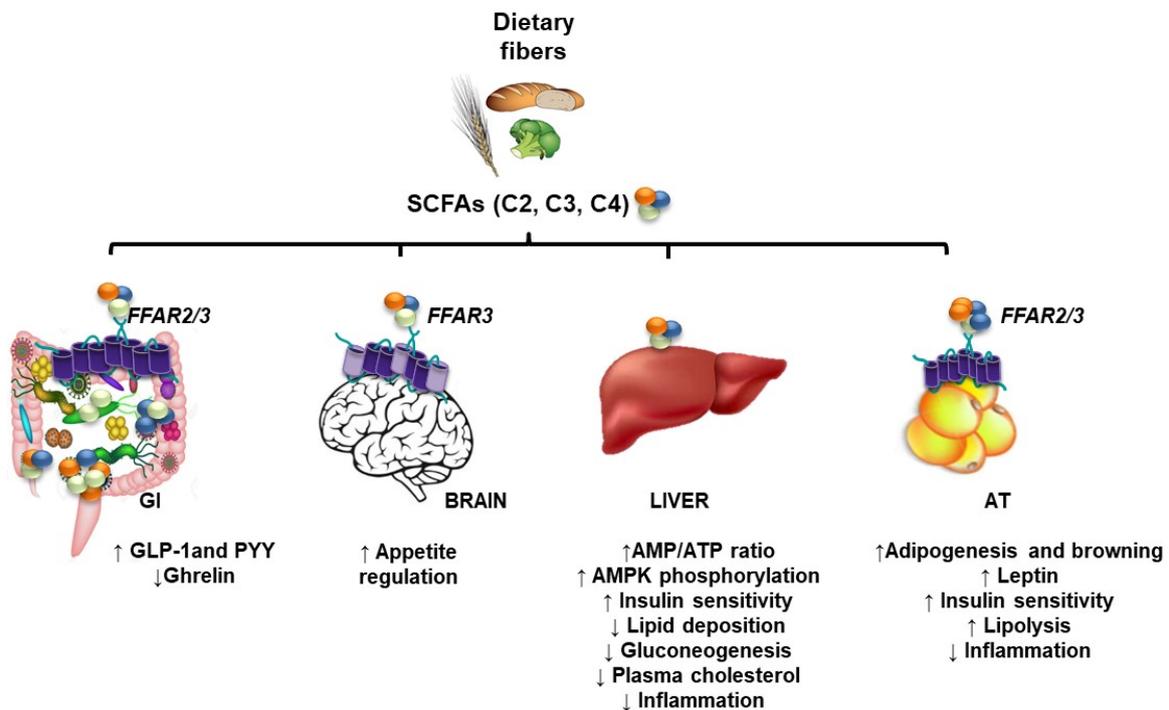
Dietary substrates are fermented to SCFAs by gut microbes, then efficiently absorbed via either simple passive diffusion or anion sodium/potassium ( $\text{Na}^+/\text{K}^+$ ) exchange in the cecum and proximal colon [80,90]. Once absorbed by colonocytes, SCFAs are highly metabolized in the colonic epithelium, which preferentially uses butyrate, followed by acetate, as the main energy sources under physiological conditions [91,92]. Alternatively, residual butyrate and propionate as well as up to 70% of acetate are metabolized by hepatocytes for gluconeogenesis. Finally, the remaining acetate is oxidized by muscle cells for energy production [93,94]. A study conducted by Cummings et al. demonstrated that SCFAs are present in the portal vein and the liver at high concentrations, then progressively decline in the peripheral systemic circulation, but remain in the range of several hundred micromolar systemically [78]. In addition to stool, some excretion of SCFAs from the body may occur in urine and breath (~5 to 20%) [95]. While SCFA have a clear positive impact on the intestinal environment via reduction of colonic pH [96] and modulation of microbial and colonic energy homeostasis [74,97], SCFAs counteract obesity-induced inflammation [98] and regulate glucose [99] and lipid metabolism [100] (Figure 2). To what extent these effects directly or indirectly control the physiological function of host adipose tissue will be discussed in further sections.

### 2.1. Gut Microbial-Derived SCFAs in Mice with Obesity

Several studies in mice point towards a correlative link between body weight and gut-derived SCFA levels. In a genetic mouse model of spontaneous obesity and diabetes, analysis of the gut microbiome and SCFA profiles showed that the abundance of *Bacteroidetes* decreased in parallel with plasma acetate levels [101]. In the same mice, the relative abundance of *Firmicutes* and fecal butyrate were higher relative to their littermate controls, suggesting that shifts in gut microbiota may lead to the loss of type and quantity of SCFAs [101]. In contrast to genetically-induced obesity, nutritionally obese rodents fed a hypercaloric diet had a lower abundance of *Bacteroidetes* as well as a reduction in fecal acetate and propionate content over time [102], an effect that was reversed by the addition of dietary fiber [103]. Another study also showed that shifting the gut microbial composition towards higher SCFA production using dietary fiber rescued the obesity-inducing effects of a high fat diet in mice [104], suggesting that dietary strategies to increase endogenous SCFA production could mitigate obesity.

Our recent findings indicate that an obesogenic diet followed by distinct weight loss interventions alter gut microbial structure and composition, which also impacted SCFA production in mice [88]. In this study, low-density lipoprotein receptor (LDLR) knockout mice administered a high fat high sucrose diet (HFHS) supplemented with *trans-10,cis-12* conjugated linoleic acid (*t10,c12* CLA, a dietary compound that promotes weight loss) presented an enrichment in butyrate-producing bacterial species as well as an elevated fecal butyrate content in conjunction with weight loss and decreased adiposity [88]. Such weight loss and gut microbiota changes were also associated with increased systemic acetate. Collectively, previous work suggests that changes in body weight are coincident with changes in fecal and systemic SCFAs in mice, but whether such changes are the direct result

or causal of weight loss remains unknown. Additional studies on the associations between fecal SCFA and obesity in animal models have been reviewed by us previously [105].



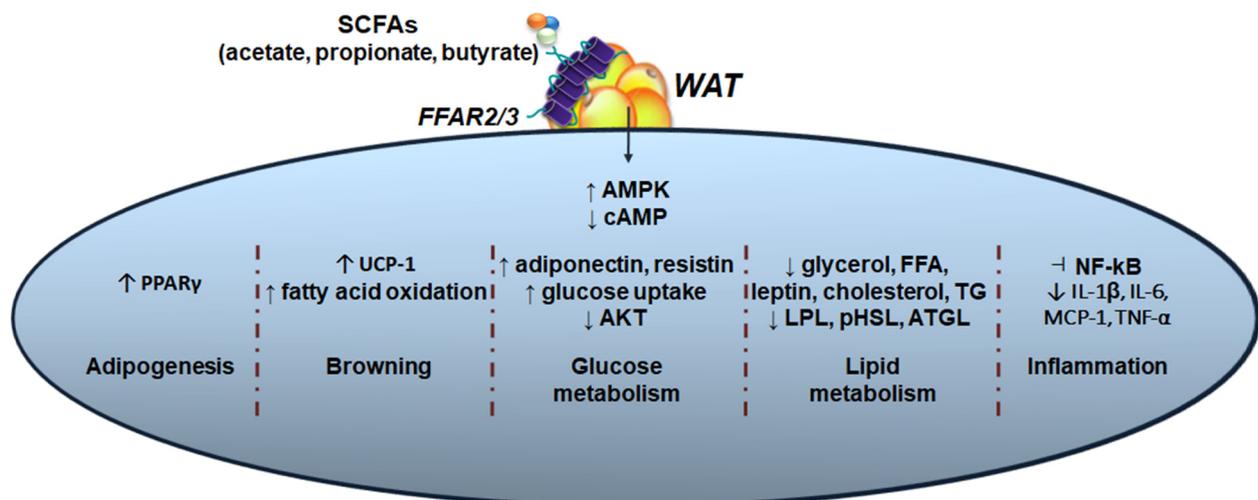
**Figure 2. Overview of host responses to short chain fatty acids (SCFAs).** SCFAs acetate (C2), propionate (C3), and butyrate (C4) derive from the microbial fermentation of non-digestible dietary fibers. These metabolites are utilized by the host, thus promoting a range of beneficial metabolic responses in peripheral tissues beyond the gut. After being absorbed through simple diffusion or signaling-mediated free fatty acid receptors (FFAR2 or FFAR3), SCFAs stimulate hormonal release (GLP-1, PYY, Ghrelin) in the gastrointestinal tract (GI), which directly signal the brain to monitor energy intake and appetite regulation. At micromolar concentrations, SCFAs enter the circulation and reach the liver via the portal vein to attenuate glucose production and lipid deposition and inflammation, possibly by activating AMPK. Additionally, SCFAs may directly or indirectly affect the brain and adipose tissue (AT) and, therefore, modulate lipid buffering capacity, adipokine secretion, insulin sensitivity, and inflammation.

## 2.2. Gut Microbial-Derived SCFAs in Humans with Obesity

In contrast with animal models, associations between body weight and SCFAs in humans are less clear. While the administration of probiotics has been shown to associate with reductions in body weight and adiposity in a recent meta-analysis in humans [106], whether these beneficial effects of probiotics involved SCFAs was not assessed. To complicate the matter, fecal SCFA levels have been reported to be lower in children with obesity [107], higher in pediatric subjects with obesity [108], elevated in adults with obesity relative to lean healthy control subjects [109,110], and to decrease with age [111], observations that are quite contradictory. A recent meta-analysis has shown that among seven studies evaluated, people with obesity had a significantly higher fecal SCFA content compared to lean controls [112]. Another study similarly showed that among 490 Chinese adults aged 30–68, plasma SCFA levels positively associated with body mass index (BMI) [113]. An investigation led by Schwieritz et al. observed that the ratio of *Firmicutes* to *Bacteroidetes* shifted in favor of *Bacteroidetes* in overweight and obese humans, with a significant increase in fecal propionate [55]. Other studies suggest that fecal SCFAs are negatively associated with BMI, waist circumference, and visceral adiposity in humans [114–116]. Thus, whether SCFAs are involved in energy balance in humans is unclear and requires further study.

### 3. SCFA Receptor Signaling

Microbial-derived SCFAs are absorbed into the bloodstream and impact whole-body physiology through mechanisms that may involve G protein-coupled receptors (GPCRs), which are also termed free fatty acid receptors (FFARs) [117,118]. FFAR2 (GPR43) and FFAR3 (GPR41) are selectively activated by SCFAs, thus activating metabolic pathways in the setting of obesity and obesity-related diseases [119,120]. Acetate preferably binds to FFAR2, whereas propionate and butyrate show high affinity to FFAR3 as revealed in HEK293 cells [121]. FFAR2/3 have been extensively described as ~40% homologous to each other in humans and mice, with proximity on chromosomes 1 (humans) and 7 (mice) [117,121]. In terms of specificity, FFAR2 displays a dual coupling through  $G_{i/o}$  and pertussis toxin-insensitive  $G_q$  protein, further influencing cyclic adenosine monophosphate (cAMP), the extracellular signal-regulated protein kinase 1/2 (ERK1/2) cascade, and calcium channels, whereas FFAR3 couples exclusively with the  $G_{i/o}$  protein family [120,121]. Both cell surface receptors are detected in a variety of organs beyond the intestine [122–125], with expression described in immune cells [126–128], liver [129,130], and adipose tissue depots [131–133]. The importance of the hydroxycarboxylic acid receptor HCA2 (GPR109A) and olfactory receptor 78 (OLFR78) as potential SCFA receptors has been less extensively explored [133,134]. In this section, we will discuss the putative effects of SCFAs on host adipose tissue that likely occur via activation of FFAR2/3 (Figure 3) and whether these receptors play a role as therapeutic targets for metabolic conditions.



**Figure 3.** SCFA-FFAR2/3 signaling pathways in adipose tissue. Gut bacterial SCFA-activated free fatty acid receptors 2 and 3 (FFAR2/3) modulate host adipose tissue physiology, thus regulating body weight and whole-body metabolic homeostasis. In white adipose tissue (WAT), studies indicate that SCFA-FFAR2/3 putatively impact nutritional status and energy balance through signaling via various molecular pathways including adipogenesis [135–141], browning [133,135,142], glucose [132,142,143] and lipid metabolism [131–133,138,139,142,144,145], and inflammation [132,133,135,137,140,142,146–148].

#### 3.1. FFAR2 and FFAR3 Signaling in Adipose Tissue of Rodents

FFAR2 and FFAR3 are expressed in nearly all metabolically active tissues, including adipose tissue [117,120,149], thus promoting a potential association between the gut microbiota, SCFAs, and adipose tissue. Notably, given that FFAR2/3 share an overlapping range of ligands [149], it is possible that their metabolic roles in adipose tissue are partially redundant, leading to the potential for conflicting data in this area. The majority of studies on the function of FFAR2/3 activation in cultured adipocytes, rodent models, and human studies demonstrate that SCFA-activated FFAR2/3 impact body weight and fat storage capacity in adipocytes [133,135], as well as other adipose tissue functions such as glucose

and lipid homeostasis [132,137], adipogenesis [136,137], lipolysis [138–140], adipokine release [131,144,150], and low-grade inflammation [146–148].

Several studies have reported that SCFAs impart metabolic benefits on host energy homeostasis, which is likely mediated by FFAR2 and/or FFAR3 [132,133,135,142]. A study conducted by Lu et al. showed that a 5% (wt/wt) mixture of SCFAs incorporated into a high fat diet (HFD) reduced body weight, total cholesterol, and inflammatory mediators such as IL-1 $\beta$ , IL-6, and monocyte chemoattractant protein (MCP-1), potentially via increased expression of FFAR2/3 in the epididymal adipose tissue in mice [133]. SCFA-mediated increases in FFAR2/3 mRNA levels were also accompanied by elevated expression of the insulin-sensitizing adipokine adiponectin, increased expression of fat oxidation genes, and decreased markers of lipolysis [133], suggesting beneficial metabolic effects via adipose tissue. To a similar extent, FFAR2/3 activation positively correlated with adipogenesis and thermogenesis [133], reinforcing the beneficial roles of these receptors in energy expenditure and obesity. In addition, a study in which mice were fed a HFD with and without the addition of individual SCFA for 5 weeks showed that SCFAs reduced weight gain and visceral fat mass, at least partly by reducing energy intake, and contributed to changes in plasma cholesterol, free fatty acids (FFA), and glucose [135]. In addition to potentially modulating lipid and glucose homeostasis via FFAR2/3 mRNA upregulation, these data revealed that SCFAs attenuate the inflammatory markers IL-1 $\beta$  and IL-6 while improving insulin sensitivity, hydrolysis of triglycerides, fatty acid oxidation, WAT browning, adipogenesis, and mitochondrial function in WAT [135].

SCFA-mediated FFAR2/3 signaling plays crucial roles in glucose and lipid metabolic pathways [142], and a few studies showcasing perturbed FFAR2 expression in mice underscore the important association between SCFA and cardiometabolic risk in models of obesity. Bjursell et al. generated the first mice globally deficient in FFAR2 (FFAR2-KO). When fed a HFD for 35 weeks, FFAR2-KO mice exhibited lower body fat mass and retroperitoneal WAT despite increased food intake, which was likely offset by increased energy expenditure, elevated core body temperature, and a reduced respiratory exchange ratio (RER), indicating increased fat oxidation [142]. Adipose tissue homeostasis was also improved, as mirrored by reduced macrophage infiltration and elevated adiponectin production. Importantly, FFAR3 expression was increased in WAT in FFAR2-KO mice fed a chow diet, a potential compensation for the loss of FFAR2. Moreover, HFD-fed FFAR2-KO mice had higher brown adipose tissue (BAT) activity, without significant alterations in genes involved in thermogenesis [142]. These findings suggest that this first FFAR2-deficient model improved obesity phenotypes [142]. In stark contrast, a second FFAR2-KO mouse model later reported by Kimura et al. showed that mice globally deficient in FFAR2 fed either normal chow or a HFD exhibited increased body weight and adiposity due to decreased energy expenditure, a phenotype that was dependent on an intact microbiota [132]. Further, transgenic mice with overexpression of FFAR2 specifically from adipocytes were protected against HFD-induced obesity and insulin resistance [132]. Collectively, the two mouse models reporting genetic perturbations in FFAR2 have yielded completely opposing phenotypes, requiring further study. Additional studies on FFAR2/3-mediated responses to SCFAs should be further explored in animal models and in human adipose tissue, which could support the design of dietary interventions and particular strategies to increase systemic availability of SCFAs. Table 1 lists studies describing mechanistic changes in FFAR2/3 expression levels in adipose tissue in rodents, highlighting the potential for FFAR isoforms as possible targets to improve diet-induced obesity, long-term energy balance, and weight management.

Several groups have also examined phenotypes related to body weight and/or obesity in FFAR3-KO mice. Jeffrey Gordon's group described the first global FFAR3-KO mice in 2008 [151]. FFAR3-KO mice were leaner than their WT counterparts in the presence of an intact microbiota on a chow diet [151]. By contrast, when fed a high fat diet, FFAR3-KO mice exhibited increased body weight and adiposity with reduced energy expenditure [152,153], suggesting that HFD feeding is required for FFAR3-mediated effects on body weight. However, an additional FFAR3-KO mouse model showed that HFD-induced obesity was

reduced compared to WT controls, potentially due to increased energy expenditure [142]. Thus, again, there is little consensus regarding the impact of FFAR2/3 on body weight and adiposity.

Beyond the scope of the current review, SCFA-activated FFAR2/3 can modulate energy homeostasis and indirectly affect host adipose tissue by increasing gut hormonal release and triggering inflammatory responses [125,151,154,155]. Global FFAR3-KO mice fed a carbohydrate-rich diet exhibited a decrease in leptin secretion as well as the enteroendocrine cell-derived peptide YY (PYY), a hormone that selectively reduces gut motility and impacts energy harvesting from the diet, thus implying that FFAR3 may be required for PYY-mediated effects [151]. These results were confirmed by Tolhurst et al., in which mice deficient in FFAR2 or FFAR3 exhibited lower secretion of PYY and the incretin glucagon-like peptide 1 (GLP-1) followed by a disruption of glucose tolerance [154]. In parallel, SCFA-mediated gut hormone secretion was also impaired in FFAR2/3<sup>-/-</sup> colonic cultures in vitro [154]. Conversely, a mechanistic study showed that butyrate and propionate protect against diet-induced obesity and regulate glucose-dependent insulinotropic peptide (GIP) and GLP-1 secretion independently of FFAR3, indicating a minor role of this FFA receptor in host metabolism [155]. In this regard, inflammation by SCFA-activated FFAR2/3 was assessed in mice fed SCFAs or given injections of antibodies that delay the immune response. The data showed that mice lacking FFAR2/3 did not exhibit major histological changes or evidence of altered inflammatory mediators in the intestine, which can be related to defective SCFA signaling. In sum, the authors emphasize the importance of FFAR2/3 to mitigate further whole-body inflammation and associated complications [155].

**Table 1.** Short chain fatty acid receptor signaling in host adipose tissue metabolism.

Model	Experimental Design	FFAR Expression	Metabolic Response	Reference
C57BL6/J mice	HFD-SCFA mixture: 5% (wt/wt <sup>-1</sup> ) acetate, propionate, and butyrate for 16 weeks	↑ FFAR2/3	↓ body weight, total cholesterol  ↓ IL-1β, IL-6, MCP-1  ↑ adiponectin, resistin ↑ lipolysis and FFA oxidation ↑ adipogenesis and mitochondrial biogenesis	Lu et al., 2016 [133]
C57BL6/J mice	HFD-SCFA individualized: 5% (wt/wt <sup>-1</sup> ) acetate, butyrate, or butyrate for 5 weeks	↑ FFAR2/3	↓ body weight, total cholesterol, FFA and glucose. ↓ energy intake ↓ IL-1β, IL-6 ↑ insulin sensitivity ↑ lipolysis and FFA oxidation ↑ browning, adipogenesis and mitochondrial function	Jiao et al., 2021 [135]
FFAR2-KO mice	HFD for 35 weeks	↔ FFAR2 ↑ FFAR3	↓ body weight, retroperitoneal WAT ↑ energy intake/expenditure, core body temperature, RER ↓ inflammation ↑ adiponectin ↑ BAT activity ↑ insulin sensitivity	Bjursel et al., 2011 [142]

Table 1. Cont.

Model	Experimental Design	FFAR Expression	Metabolic Response	Reference
FFAR2-KO			FFAR2-KO mice: ↑ body weight ↓ insulin sensitivity ↑ inflammation, TNF- $\alpha$	
aP2-FFAR2tg	HFD for 12 weeks	↑ FFAR2	aP2-FFAR2tg: ↓ body weight ↑ insulin sensitivity ↓ inflammation, TNF- $\alpha$ ↓ LPL activity	Kimura et al., 2013 [132]

### 3.2. FFAR2 and FFAR3 Signaling in Adipocytes In Vitro

Several in vitro studies have contributed to what we know regarding the influence of SCFA-activated FFAR2/3 on adipocyte metabolic regulation, informing the roles of SCFAs in the onset and progression of obesity [136–140,143,145]. Hong et al. demonstrated in 3T3-L1 adipocytes that microbial-derived acetate and propionate promote adipogenesis by increasing peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) expression and lipid accumulation and blunt lipolytic activity, mainly through FFAR2 [139]. Similar findings were confirmed years later by another group that showed that 3T3-L1 adipocytes exposed to acetate and propionate for 4 h exhibited increased FFAR2 expression with decreased evidence of lipolysis, namely, reduced free fatty acids and glycerol release [138]. In line with this, supra-physiological concentrations of acetate target FFAR2 via G<sub>i/o</sub>, thus suppressing insulin-induced protein kinase B (AKT) phosphorylation and LPL activity as well as fatty acid uptake [132]. Quantitative and qualitative assays determined that FFAR2, but not FFAR3, is expressed at late stages of adipocyte differentiation and impacts intracellular lipid content [144]. At micromolar doses, propionate did not influence lipid accumulation or leptin secretion in 3T3-L1 adipocytes, indicating that earlier expression of FFAR2 might be required to modulate such adipogenesis markers [144]. Additional studies have yielded mixed results [131,143]. First, in fully differentiated 3T3-L1 adipocytes, an exposure to distinct concentrations of propionate significantly increased insulin-stimulated glucose uptake, at least in part, through FFAR3. Moreover, silencing FFAR3 using small interfering RNA (siRNA) blunted approximately 30% of the SCFA-induced glucose effects in vitro, implying a potential role for FFAR3 to modulate glucose uptake in adipocytes [143]. Second, Xiong et al. examined whether SCFAs are required to stimulate leptin release via FFAR3 in mouse primary adipocytes. The results indicated that SCFA-upregulated FFAR3 led to leptin secretion, an effect that was abolished via pertussis toxin treatment. In addition, data showed that a marked decrease in cAMP levels supports the exclusive G<sub>i</sub>-coupled pathway mediated by FFAR3 [131]. The roles of FFAR3 in leptin stimulation of adipocytes in response to SCFAs was then explored in adipocytes from FFAR3-KO mice [144]. Interestingly, the authors concluded that FFAR3 was undetectable at a molecular level in mouse adipose tissue, in contrast to previous reports [131,143]. Additionally, reduced leptin production and lipolysis, in the presence of SCFAs, might be related to the downregulation of FFAR2 rather than other unknown mechanisms, suggesting a crucial interplay between the FFA receptors [144].

Despite accumulating evidence to support the influence of microbial-derived SCFA activation of FFAR2/3 in the regulation of host adipose tissue biology and physiology, studies in humans are needed to understand such ambiguous findings. Al-Lahham et al. demonstrated for the first time that propionate counteracts inflammation and increases leptin secretion via omental (OAT) and subcutaneous adipose tissue (SAT) explants from female subjects [150]. Such metabolic responses were attributed to high expression of FFAR2/3, confirmed by blocking the G<sub>i/o</sub> pathway with pertussis toxin [150]. Next, preadipocytes isolated from human OAT were incubated with acetate and propionate to evaluate the effects of FFAR2 agonists on adipogenesis [137]. Unlike what has been described in murine studies, results from Dewful et al. suggested that FFAR2 is not implicated in adipocyte dif-

differentiation, as exemplified by adipocyte fatty acid-binding protein 2 (aP2) expression, but might be associated with inflammatory outcomes in humans [137]. Moreover, propionate inhibited lipid accumulation and adipogenesis in human adipose-derived mesenchymal stem cells (MSCs) via FFAR2 [136]. Therefore, the role of FFAR2 protein should be further elucidated via the use of knockdown and overexpression in human adipocyte models.

To assess the differential effects of SCFAs on targeting FFA receptors, Jocken et al. proposed that individual SCFAs of a mixture could alter adipocyte lipolysis, thus impacting whole-body energy homeostasis [145]. In human multipotent adipose tissue-derived stem (hMADS) cells, which resemble human white adipocytes, high doses of acetate conferred an anti-lipolytic response followed by a reduced phosphorylation of hormone sensitive lipase (HSL) at serine 650 (pHSL<sub>SER650</sub>) in a FFAR2/3-dependent manner [145]. Collectively, the apparent discrepancies among these in vitro and ex vivo findings may stem from differences in SCFA concentrations and length of SCFA exposure, mouse strains with different backgrounds, age, sex, the use of human adipose tissue harvested from distinct sites, and varied metabolic conditions employed in each study protocol. In Table 2 we describe findings from experimental analyses in vitro and ex vivo, including conflicting metabolic outcomes.

**Table 2.** Short chain fatty acid receptor signaling in host adipose tissue metabolism in vitro and ex vivo.

Model	Experimental Design	FFAR Expression	Metabolic Response	Reference
3T3-L1 adipocytes	1 mM acetate and propionate for 7 days	↑ FFAR2 ↔ FFAR3	↑ adipogenesis, PPAR $\gamma$ ↑ lipid infiltration ↓ lipolysis	Hong et al., 2005 [139]
	0.1 to 0.3 mM acetate and propionate for 4 h	↑ FFAR2	↓ lipolysis (FFA, glycerol)	Ge et al., 2008 [138]
	10 mM acetate for 2 h	↑ FFAR2	↓ insulin-induced AKT, fatty acid uptake, LPL	Kimura et al., 2013 [132]
	Up to 0.01 mM acetate and propionate for 10 days	↑ FFAR2 ↔ FFAR3	↑ adipogenesis, lipid infiltration ↔ leptin	Frost et al., 2014 [141]
	Up to 1 mM propionate for 30 min	↑ FFAR3	↑ glucose uptake	Han et al., 2014 [143]
Primary murine adipocytes	1 to 3 mM acetate, propionate, and butyrate for 10 min	↑ FFAR3	↑ leptin ↓ cAMP	Xiong et al., 2004 [131]
FFAR3-KO adipocytes	Up to 03 mM acetate, propionate, and butyrate for 4h	↔ FFAR3	↓ leptin ↓ lipolysis	Zaibi et al., 2010 [144]
Human OAT and SAT	Up to 10 mM Propionate for 24 h	↑ FFAR2/3	↑ leptin ↔ adiponectin ↓ inflammation, resistin	Al-Lahham et al., 2010 [150]
Human OAT	0.01 mM acetate and propionate for 13 days	↔ FFAR2	↔ adipogenesis, aP2 ↓ inflammation	Dewufl et al., 2013 [137]
Human MSCs	3.3 mM propionate for 14 days	↑ FFAR2	↓ adipogenesis, PPAR $\gamma$ ↓ lipid accumulation	Iván et al., 2017 [136]
Human MADS	Up to 1 mM acetate, propionate, butyrate or mixed SCFAs for 6 h	↑ FFAR2/3	↓ lipolysis ↓ phospho-HSL <sub>ser650</sub>	Jocken et al., 2017 [145]

#### 4. SCFA Potential for Targeting Histone Deacetylases

The majority of findings to date indicate that activation of FFAR2/3 by SCFAs could be a promising anti-obesity strategy, with direct implications for adipose tissue function. Moreover, epigenetic mechanisms shed light on how SCFAs promote beneficial rather than detrimental metabolic responses, thus impacting whole-body energy homeostasis [156–158]. SCFAs, mainly butyrate, act as natural histone deacetylase (HDAC) inhibitors to epigenetically regulate chromatin acetylation and gene transcription involved in cell proliferation, differentiation, apoptosis, fatty acid oxidation, and inflammation [159–163]. Of note, SCFA-mediated HDAC inhibitory activity depends on the concentration of SCFAs and on which cell type or tissue the gut metabolites may target [159]. Li et al. showed that butyrate contributes to the differentiation of pre-adipocytes into mature adipocytes via increased histone acetylation at the promoters of PPAR $\gamma$  and CCAAT enhancer-binding protein alpha (CEBP $\alpha$ ), leading to upregulation of these adipogenic genes [164]. Moreover, additional work confirmed that butyrate and propionate, but not acetate, impact the rate of lipolysis via their shared activity as HDAC inhibitors in 3T3-L1 adipocytes, thus suggesting a possible molecular pathway to mitigate obesity-related outcomes [165]. As pointed out by the authors, *in vivo* analyses are required to confirm in detail whether such HDAC inhibitors can differentially impact whole-body metabolism [164,165]. In line with that, a study in type 2 diabetic rats fed a HFD revealed that butyrate treatment reduced body weight and adiposity while improving glucose homeostasis through hyperacetylation of histone H3, thus likely inhibiting HDAC activity as neither FFAR2 nor FFAR3 were modulated [162]. Similarly, butyrate activates the phosphorylation of the mitogen-activated protein kinase 3/p38/p38-regulated activated protein kinase (MKK3/p38/PRAK) signaling pathway and protects against HFD-induced cardiometabolic disruptions in response to HDAC inhibition in diabetic and obese mice [163]. Moreover, a short-term intervention with butyrate counteracted HFD-induced adiposity and insulin resistance, while promoting histone acetylation on the promoters of genes encoding adiponectin receptors and uncoupling proteins in C57BL6/J mice [161]. Lastly, adipose tissue levels of HDAC were significantly decreased in obese rats given acetate, with subsequent normalization of metabolic homeostasis via modulation of PPAR $\gamma$  and suppression of oxidative stress [166]. In summary, in addition to the possibility of therapeutically targeting HDAC, to what extent SCFAs selectively inhibit HDAC requires further investigation as another potential mechanism to manage obesity.

#### 5. Gut Microbial-Derived SCFAs Influence Host Adipose Tissue Inflammation

Adipose tissue is a dynamic tissue that contributes to local and systemic low-grade inflammation, thus orchestrating the progression of insulin resistance and pathophysiology of obesity [18,23]. Hence, chronic inflammation in adipose tissue promotes the recruitment and infiltration of various immune cells, including macrophages and T cells, which regulate the secretion of cytokines and activation of inflammatory signaling cascades [25,26]. While the contribution of adipose tissue inflammation to the pathology of obesity has been established, it is worth discussing this paradigm in the context of microbial-derived SCFAs *in vitro* and *in vivo* [140,147,167–171].

Effects of SCFA on inflammatory responses in monocytes and macrophages have yielded conflicting results. A study conducted by Al-Lahham et al. reported that propionate impacts human omental OAT and THP-1-derived macrophages, resulting in down-regulation of cytokines and chemokines, crucial mediators linked to ATM activation [147]. Indeed, the expression of specific macrophage markers was impacted by propionate, suggesting that macrophages could be targets for SCFAs within adipose tissue, either directly or indirectly [147]. In a different study, co-stimulation with SCFAs and the pro-inflammatory mediators tumor necrosis factor alpha (TNF- $\alpha$ ) and lipopolysaccharide (LPS) exerted opposing effects in monocytes and macrophages. Acetate, in combination with TNF- $\alpha$ , increased the expression of chemokines in THP-1 monocytic cells, but decreased MCP-1 in macrophages, implying a paradoxical SCFA-mediated inflammatory response that is dependent on cell type, culture conditions, and SCFAs concentration [171]. Immunomodulation

latory effects of SCFAs were further explored by Ohira et al. [140,167]. The authors showed that butyrate blunts inflammatory responses and activation of cell signaling pathways coordinated via the interaction of 3T3-L1 adipocytes and RAW264.7 macrophages [140,167]. In a similar co-culture experimental design, butyrate increased prostaglandin E2 (PGE2) production, which is correlated to adiposity, and signaling factors including cyclooxygenase-2 (COX2) and cytosolic phospholipase A2 (cPLA2), in a dose-dependent manner [167]. Thus, whether SCFAs promote or reduce inflammatory responses in monocytes and macrophages remains to be determined.

Importantly, ATM, upon stimulation with local environmental factors, may be further divided into “classically activated” (M1), “alternatively activated” (M2), or “metabolically activated” (MMe) macrophage phenotypes [25,172]. Bone marrow-derived macrophages exposed to IL-4 and different concentrations of butyrate activated the histone H3 lysine 9/signal transducer and activator of transcription 6 (H3K9/STAT6) signaling pathway and enhanced M2-related gene expression and chemokines, indicating a putative role of such SCFAs to facilitate M2 macrophage polarization [170]. The effects of butyrate inducing the M2 macrophage profile may contribute to adipose tissue homeostasis, thus alleviating obesity and related comorbidities. In a diet-induced obesity mouse model, butyrate increased M2 relative to M1 macrophages and regulatory T cells and associated genes, whereas pro-inflammatory markers were reduced in white adipose tissue [169]. Furthermore, analysis in 3T3-L1 adipocytes showed that butyrate markedly decreased the expression of endoplasmic reticulum (ER) stress proteins, suggesting a potential to mitigate inflammation and cell autophagy [169]. Recent work in peripheral blood monocytes and ATM from obese individuals provided evidence that SCFAs impact molecular pathways that reduce systemic inflammation, evidenced by decreased nuclear factor kappa B (NF- $\kappa$ B), mitogen-activated protein kinases (MAPK), and pro-inflammatory cytokines following weight loss [168]. A list of relevant studies describing the modulation of adipose tissue inflammation by SCFAs are listed in Table 3 and additionally reviewed in [105].

**Table 3.** Gut microbial-derived SCFAs influence adipose tissue inflammation.

Model	Experimental Design	Inflammatory Responses	Reference
Human OAT	3 mM propionate for 24 h	↓ IL-4, IL-10, G-CSF, IP-10, MIP-1 $\alpha/\beta$ , CCL5, TNF- $\alpha$	Al-Lahham et al., 2012 [147]
THP-1 M $\phi$	Up to 10 mM propionate for 2 h	↓ IL-10, G-CSF, MCP-1, CCL5, TNF- $\alpha$	Al-Lahham et al., 2012 [147]
THP-1 M $\phi$	100 mM acetate, 10 mM propionate, and 2 mM butyrate for 24 h	↑ MCP-1 ↓ NF- $\kappa$ B, MAPKs (p38, ERK1/2)	Al-Roub et al., 2021 [171]
3T3-L1 and RAW264.7 M $\phi$	Up to 1 mM butyrate for 24 h	↓ TNF- $\alpha$ , MCP-1, IL-6 ↓ NF- $\kappa$ B, MAPKs (p38, ERK1/2, JNK1/2)	Ohira et al., 2013 [140]
		↑ PGE2, COX2, cPLA2 ↓ PRKAR1A, cAMP	Ohira et al., 2016 [167]
M $\phi$ -BMD	Up to 1 mM butyrate between 1.5 to 24 h	↑ Arg1, Fizz1, Ym1 ↑ CCL2, CCL17, CCL22 ↑ H3K9, STAT6	Ji et al., 2016 [170]
C57BL6/J mice	5% <i>w/w</i> butyrate for 8 weeks	↑ M $\phi$ M2 (CD206+) ↑ Tregs (CD25+) ↑ IL-10, DioA2, Pgc-1, IL-4	Kushwaha et al., 2022 [169]

Table 3. Cont.

Model	Experimental Design	Inflammatory Responses	Reference
3T3-L1	1 mM butyrate for 24 h	↓ ER (pPERK, CHOP)	Kushwaha et al., 2022 [169]
Monocytes and ATM	300 mM acetate or 30 mM propionate, and butyrate for 15 h	↓ NF-κB, TNF-α, IL-6 ↓ MAPK1	Eslick et al., 2022 [168]

## 6. Administration of SCFAs to Impact Adipose Tissue

Proof-of-concept studies have investigated whether SCFAs influence adipose tissue and whether they ultimately impact energy balance and overall metabolism in rodent models and humans [68,71,133,173–188]. In this context, exogenous administration of SCFAs delivered via dietary supplementation, in the drinking water, oral gavage, via intraperitoneal (IP) injection, and other routes, has been done in rodents and humans [189,190]. Of note, orally delivered SCFAs reach the gastrointestinal (GI) tract, then signal local SCFA receptors, rather than being immediately absorbed by colonocytes. On the other hand, IP injection of SCFAs at supra-physiological concentrations rapidly targets organs beyond the gut, thus exerting an acute and potent response on host metabolism [189,190]. Because host metabolic outcomes can be heterogeneous depending on the method of SCFA delivery, interpretation of these study results should be done with care. Various studies that delivered SCFA endogenously to rodents and humans will be described below.

### 6.1. Effects of Exogenously-Administered SCFAs in Rodent Models

Several studies have examined obesity phenotypes in mice given various doses and combinations of SCFA admixed into a HFD. Gao et al. showed that adding butyrate (5% wt/wt) to a HFD increased energy expenditure, thermogenic function, and insulin sensitization and prevented body weight gain in mice [68]. A similar study, in which the HFD was also supplemented with 5% wt/wt SCFA for 12 weeks, showed that acetate, propionate, and butyrate all individually prevented diet-induced obesity and insulin resistance, with similar increases in fatty acid oxidation and energy expenditure [177]. In line with these studies, SCFAs incorporated into an obesogenic diet reduced body weight, mitigated systemic inflammation and other obesity-related markers in mice, with acetate exerting the major effects [133]. An experimental design by Arnoldussen et al. revealed that dietary delivery of butyrate (5%) ameliorated HFD-induced obesity and associated complications, contributing to weight loss and decreased adiposity in mid-adult LDLR KO mice [174]. Thus, there is broad consensus that dietary supplementation of SCFA promotes metabolic benefits that mitigate HFD-induced obesity [68,133,177].

Chronic dietary butyrate supplementation prevented HFD-induced body weight gain and adiposity, likely due to increased fat oxidation and thermogenesis, and improved overall lipid and glucose metabolism in apoE\*3-Leiden+CETP, a mouse model with a humanized lipid profile [187]. Additionally, acute oral delivery of butyrate was shown to decrease food intake and neuronal signals, suggesting a connection between the gut–brain axis and energy homeostasis [187]. Further, a single oral gavage of 1.5% acetate increased energy expenditure in parallel with suppression of body weight and fat mass in C57BL/6J mice [173], which was attributed to upregulation of PPARγ and fatty acid oxidation-related genes [175].

Interestingly, propionate added to drinking water or administered via IP injection conferred weight gain, severe hyperinsulinemia, and hyperglycemia as opposed to rectally administered propionate in mice [188]. Such metabolic responses appear to be partially mediated by the adipokine fatty acid-binding protein 4 (FABP4) [188]. Mice fed an obesogenic diet, then treated with IP injections of liposome-encapsulated acetate nanoparticles, showed reduced whole-body adiposity, insulin resistance, and lipolytic activity [181]. Herein, thermogenic function through browning of WAT was increased by these acetate nanoparticles with a coincident reduction in adiposity [181].

Administration of SCFAs similarly impacts lipid metabolism in rats [176,178]. First, intraperitoneal (IP) injections of acetate, given 5 times per week for 6 months, led to decreased body weight while increasing energy expenditure [176]. Acetate also increased lipolytic and lipogenic gene expression in this rat model of spontaneous diabetes [176]. In another study using rats, IP administration of a mixture of SCFAs at a 3:1:1 molar ratio for 7 days had no effects on body weight and abdominal adiposity [178], implying that a longer treatment period is required to exert a body weight phenotype. However, acetate, propionate, and butyrate impacted lipid and glucose metabolism as evidenced by reduced total cholesterol, triglycerides, and glucose levels in a sex-dependent manner [178]. Therefore, SCFAs may exert positive metabolic benefits in both sexes via distinct mechanisms when administered exogenously [178].

Collectively, these discrepant findings could be attributed to variable substrate metabolism or host microbiota signatures, thus leading to diverse physiological and metabolic outcomes in responses to exogenous SCFAs [189,190]. As such, the unique host-specific and site-specific mechanisms whereby SCFAs affect adipose tissue and energy balance regulation should be further investigated. The effects of SCFA administration via different routes on aspects of obesity and related complications in rodents are detailed in Table 4.

**Table 4.** Exogenous administration of SCFAs to impact adipose tissue metabolism in rodents.

Rodent Studies	Experimental Design	Metabolic Response	Reference
C57BL/6J mice	5% wt/wt butyrate via diet supplementation for 16 weeks	↓ body weight ↑ energy expenditure ↑ PGC-1 $\alpha$ , UCP-1 ↑ insulin sensitivity	Gao et al., 2009[68]
C57BL/6J mice	5% wt/wt acetate, propionate, butyrate via diet supplementation for 12 weeks	↓ body weight, WAT mass, adipocytes size ↑ energy expenditure, fat oxidation, ↑ insulin sensitivity	den Besten et al., 2015 [177]
C57BL/6J mice	5% wt/wt <sup>-1</sup> acetate, propionate, and butyrate or SCFAs mixture diet supplementation for 16 weeks	↔ food and energy intake ↓ body weight ↓ glucose, FFA, IL-1 $\beta$ , IL-6, MCP-1	Lu et al., 2016 [133]
LDLR KO-Leiden mice	5% wt/wt butyrate via diet supplementation for 15 weeks	↔ food and energy intake ↓ body weight, omental and inguinal fat ↓ cholesterol, triglycerides, insulin	Arnoldussen et al., 2017 [174]
ApoE*3-Leiden.CETP	5% wt/wt butyrate via diet supplementation for 9 weeks 6 M butyrate via oral gavage/15 or 150 mM IV injection	↓ food intake ↓ body weight, epididymal WAT ↓ plasma triglycerides, insulin ↑ fat oxidation and BAT activity	Li et al., 2018 [187]
C57BL/6J mice	1.5% acetate via oral gavage for 1 day	↑ energy expenditure ↓ body weight and fat mass	Hattori et al., 2010 [173]
C57BL/6J mice	(1) ~15 mg/kg propionate via drinking water for 6 weeks (2) I.P. at 0.5 to 2 g/kg of BW (3) 1g/kg of BW rectal infusion	↑ body weight ↑ glucose, insulin ↑ FABP4	Tiroshi et al., 2019 [188]
C57BL6/J mice	Liposome encapsulated acetate nanoparticle via I.P. at 1g/kg of BW for 6 weeks	↔ body weight, adipocytes ↓ whole-body adiposity, lipolysis, FFA ↑ insulin sensitivity ↑ browning (UCP1, PRDM16)	Sahuri-Arisoylu et al., 2016 [181]

Table 4. Cont.

Rodent Studies	Experimental Design	Metabolic Response	Reference
Otsuka Long-Evans Tokushima Fatty rats	acetate via I.P. at 5.2 mg/kg of BW for 6 months	↓ body weight, lipid droplets size ↑ energy balance ↑ LCAD, 3KACT, PPAR $\gamma$	Yamashita et al., 2009 [176]
Long-Evans rats	acetate, propionate, butyrate via I.P at 60:20:20 M of BW for 7 days	↔ body weight, abdominal fat and food intake, NEFA ↓ cholesterol, triglycerides, glucose	Shah et al., 2021 [178]

### 6.2. Effects of Exogenously Administered-SCFAs in Humans

Mechanistic evidence from rodent studies has shown that the various routes to deliver exogenous SCFAs affect host whole-body substrate and energy metabolism [173,174,176,178,181,187,188]. Yet, the previously described long-term strategies as well as metabolic effects may not all directly translate into humans to prevent and/or manage obesity complications (reviewed extensively in [191]), thus warranting additional investigation.

In one of the first human randomized controlled trials, dietary supplementation of inulin-propionate ester increased the secretion of gut hormones PYY and GLP-1, leading to a reduction in energy intake, body weight, intra-abdominal adipose tissue mass, and plasma cholesterol, HDL, and LDL [192]. Additional evidence demonstrated that orally-delivered propionate modulated energy balance via the promotion of whole-body lipid oxidation in healthy subjects [71], in line with a previous study conducted in mice [177]. Conversely, a small pilot investigating orally-delivered butyrate showed no improvements in glucose metabolism or thermogenic function in insulin-resistant individuals with obesity, reinforcing the concept that butyrate could have been quickly metabolized by colonocytes and may not have been readily available to extra-intestinal organs in a systemic manner [182]. Moreover, butyrate increased total cholesterol and LDL levels after 4 weeks of supplementation compared with healthy lean subjects [182]. In another study, Harstra et al. determined that oral butyrate had no effect on body weight, energy expenditure, and insulin sensitivity in people with MetS, but favored a decrease in metabolic parameters including HbA1c [186]. The lack of effects on body weight with oral butyrate supplementation may be due to lower doses of butyrate relative to body weight as well as a lack of daily dose adjustment, as suggested previously [68,177]. Acetate and acetic acid (vinegar) supplementation has been reported to counteract the deleterious effects of obesity and related complications, hence conferring healthy metabolic functions among host species [193]. Kondo et al. first revealed that continuous oral intake of vinegar lowered weight gain and visceral and subcutaneous fat mass as well as serum triglycerides levels, with no changes in energy balance in a Japanese obese cohort [175]. These results suggest that oral delivery of various SCFA preparations can ameliorate metabolic syndrome complications, thus having a clinical contribution in humans [175].

Notably, distinct routes of SCFA administration markedly impact the metabolic responses in obesity. Rectal or intravenous (IV) infusions with acetate increased gut-derived PYY and GLP-1 secretion, which was accompanied by decreased ghrelin in hyperinsulinemic, overweight woman [185]. In a situation of excessive caloric intake, microbial acetate production was increased, ultimately promoting insulin secretion and whole-body metabolism [193]. Colonic-delivered acetate markedly affected energy expenditure and fat oxidation, then increased circulating glucose, insulin, and PYY in overweight and obese men [183]. Further, supraphysiological rectal infusions of a SCFA mixture for four days modulated energy balance via enhancing fat oxidation and PYY, while attenuating lipolysis [184]. Various studies depicting SCFA administration in humans followed by their metabolic responses are listed in Table 5.

**Table 5.** Routes of administration of SCFAs to impact adipose tissue metabolism in humans.

Human Studies	Experimental Design	Metabolic Response	Reference
Healthy men/women	10 g/day inulin-propionate via dietary supplementation for 24 weeks	↓ food intake, body weight and intra-abdominal AT ↑ PYY, GLP-1 ↓ cholesterol, HDL, LDL	Chambers et al., 2015 [192]
	6.8 g propionate via oral administration for 2 h/2 days	↑ energy expenditure ↑ lipid oxidation	Chambers et al., 2018 [71]
Healthy and MetS male	4 g/day butyrate via oral administration for 4 weeks	↔ BMI, BAT activity ↔ energy expenditure, insulin sensitivity ↑ cholesterol, LDL	Bouter et al., 2018 [182]
MetS males/females	4 g/day via oral administration butyrate for 4 weeks	↔ body weight, ↔ energy expenditure, insulin sensitivity ↓ Cholesterol, triglycerides, HbA1c	Harstra et al., 2020 [186]
Men/women with obesity	Up to 1.5 g acetate via oral administration for 12 weeks	↔ energy expenditure ↓ body weight ↓ visceral, subcutaneous fat mass ↓ triglycerides	Kondo et al., 2009 [175]
Hyperinsulinemic and overweight women	60 mM/L acetate via rectal infusions and 20 mM/L via IV for up to 1 h	↔ glucose and insulin ↑ PYY, GLP-1 ↓ ghrelin	Freeland et al., 2010 [185]
Men with overweight and/or obesity	Up to 180 mM/L acetate via rectal infusions for 3 days	↑ energy expenditure, fat oxidation ↑ glucose, insulin, PYY	Van der Beek et al., 2016 [183]
	Up to 200 mM/L SCFAs mixture via rectal infusions for 4 days	↑ energy expenditure, fat oxidation ↑ PYY ↓ lipolysis	Canfora et al., 2017 [184]

## 7. Gut-Derived SCFAs to Modulate Adipose Tissue Lipolysis

The contributions of SCFAs to host adipose tissue and obesity are becoming increasingly recognized [194–198]. For instance, experimental designs have centered on the mechanisms whereby SCFAs differentially impact signal transduction pathways to regulate lipid and glucose metabolism in WAT, such as lipolysis [138,140,145,165,199–205]. Briefly, AT lipolysis is a catabolic process leading to the hydrolysis of triacylglycerols (TAGs) into glycerol and free fatty acids (FFA) [206], a process that is initiated when the mobilization of energy stored in adipocytes is required. Hence, targeting adipocyte-specific mechanisms to reduce lipolysis, and increase fat oxidation and energy expenditure, may be crucial for the management of metabolic diseases. In this section, we will summarize the effects of SCFAs on AT lipolysis *in vitro* and *in vivo*.

In primary adipocytes from mice and in 3T3-L1 adipocytes, exposure to acetate and propionate (0.1–0.3 mM) inhibited lipolysis and plasma FFA levels in a dose-dependent manner through activation of GPCR-FFAR2 [138]. Treatment with butyrate ( $\geq 0.2$  mM) and TNF- $\alpha$  attenuated lipolysis as well as glycerol and FFA release in co-cultured 3T3-L1 and RAW264.7 macrophages, which correlated with the decreased protein expression of adipose triglyceride lipase (ATGL), HSL, and phosphorylation of HSL<sub>(Ser660)</sub> [140]. Under supra-physiological concentrations, Aberdein et al. demonstrated that acetate (4 mM) reduced non-esterified fatty acid (NEFA) and phosphorylation of HSL<sub>(Ser563)</sub>, with no changes in glycerol release, likely via FFAR2 in mature 3T3-L1 adipocytes [202]. Similarly, propionate and butyrate (10 mM) inhibited lipolysis, followed by a reduction in isoproterenol-induced phosphorylation of HSL<sub>(Ser563)</sub> and HSL<sub>(Ser562)</sub> in rat and human adipocytes, respectively,

an effect that involved FFAR2 [203]. Further, research conducted by Rumberger et al. indicated that butyrate (5 mM) increases the rate of lipolysis in 3T3-L1 adipocytes, underscoring the effects of butyrate on HDAC inhibitory activity [165]. Findings to date showed that SCFA mixtures, in particular those that include acetate, impact lipolysis via attenuation of HSL<sub>(Ser650)</sub> phosphorylation in human adipocytes [145]. Collectively, these results in cultured cells suggest that SCFAs play an important role in the lipolytic pathway, with the potential to modulate WAT lipid buffering capacity, insulin sensitivity, and improve energy balance.

In *in vivo* settings, rodents orally given acetate exhibited an increase in the transcription of genes involved with energy metabolism and lipolysis, including long-chain acyl-CoA dehydrogenase (LCAD), 3-ketoacyl-CoA thiolase (3KACT), and PPAR $\gamma$  [176]. Moreover, acetate suppressed lipolysis in subcutaneous AT and decreased circulating FFA levels, mainly through the downregulation of mRNA expression of ATGL as well as target genes involved in fatty acid oxidation and energy expenditure, carnitine palmitoyltransferase I (CPT1) and acyl-CoA oxidase 1 (ACOX1), in HFD-fed mice [181]. Lu et al. subsequently found that long-term dietary supplementation with acetate, propionate, and butyrate promoted a decrease in adipose HSL expression, without affecting CPT1 and FFA levels [133]. In humans, an initial study showed no changes in plasma glycerol, NEFA release, or fat oxidation following the consumption of 2 mg acetate [204]. Conversely, in another study, acetate blunted whole-body lipolysis, as indicated by decreased plasma glycerol, in healthy men [205] and in men with overweight/obesity [184]. In healthy individuals, acute SCFA infusion reduced circulating FFA concentrations [93,199], indicating that lipolysis was reduced [200]. In addition, recent cross-sectional analyses have reported that circulating rather than fecal SCFAs are negatively correlated to systemic FFA and whole-body lipolysis, suggesting that SCFAs impact lipid metabolism, which can improve peripheral insulin sensitivity and ultimately benefit metabolic health in humans [201]. In addition, measuring systemic SCFA levels is more predictive of metabolic status than stool SCFA content [201].

## 8. Conclusions

Over the last several decades, a handful of studies have been performed to highlight gut microbi-derived SCFAs as novel therapeutic targets for managing the risk of obesity-related metabolic disorders. Herein, we have provided an update in this field to show that SCFAs influence adipose tissue biology and physiology and thus impact body weight control, likely via activation and/or inhibition of multiple signaling pathways associated with regulating feeding behavior, ameliorating inflammatory responses, and favorably impacting lipid metabolism. In this contribution, we have described how different host species with varied genetic backgrounds and unique gut microbiota patterns, dietary sources, and composition as well as different methods of exposure to SCFAs could explain the underlying mechanisms whereby SCFAs can directly impact adipose tissue and host metabolic health. Taken together, while the beneficial effects of SCFAs on some adipose tissue functions have been described, the conflicting findings across studies provide a rationale for additional long-term and well-controlled investigations. Furthermore, a complete regulatory system focused on the interplay among host adipose tissue, host genetics, and SCFAs should be explored to inform potential treatment strategies for obesity and chronic diseases.

**Author Contributions:** K.S.M. and L.J.d.H. wrote the manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** The authors are thankful for research support from the University of Washington's training program in Diabetes, Obesity and Metabolism, #T32 DK007247. This work was also supported by funding from the United States Department of Agriculture (USDA) National Institute of Food and Agriculture (NIFA) award #2020-67001-30716.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

10,12 CLA: trans-10,cis-12 conjugated linoleic acid; 3KACT: 3-ketoacyl-CoA thiolase; ACOX1: acyl-CoA oxidase 1; AKT: protein kinase B; AMPK: AMP-activated protein kinase; aP2: adipocyte fatty acid-binding protein 2; apoE: Apolipoprotein E; Arg1: arginase 1; AT: adipose tissue; ATGL: adipose triglyceride lipase; ATM: adipose tissue macrophage; BAT: brown adipose tissue; BMI: body mass index; C2: acetate; C3: propionate; C4: butyrate; CAD: coronary artery disease; cAMP: cyclic adenosine monophosphate; CCL17: CC motif chemokine ligand 17; CCL2: CC motif chemokine ligand 2; CCL22: CC motif chemokine ligand 22; CCL5: CC motif chemokine ligand 5; CEBP $\alpha$ : CCAAT/enhancer-binding protein alpha; CHOP: CEBP homologous protein; CETP: cholesteryl ester transfer protein; COX2: cyclooxygenase-2; cPLA2: cytosolic phospholipase A2; CPT1: carnitine palmitoyltransferase I; DioA2: iodothyronine deiodinase-2; ER: endoplasmic reticulum; ERK1/2: extracellular signal-regulated protein kinase 1/2; FABP4: fatty acid binding protein 4; FABP4: fatty acid-binding protein 4; FFA: free fatty acid; FFAR2: free fatty acid receptor 2 (GPR43); FFAR3: free fatty acid receptor 3 (GPR41); Fizz1: Found in inflammatory zone protein 1; GF: germ free; GI: gastrointestinal; GLP-1: glucagon-like peptide 1 (GLP-1); GPCR: G-protein coupled receptor; G-CSF: granulocyte colony-stimulating factor; H3K9: histone H3 lysine 9; HCA2: hydroxycarboxylic acid receptor; HDAC: histone deacetylase; HFD: high fat diet; HFHS: high fat high sucrose; hMADS: human multipotent adipose tissue-derived stem cells; HSL: hormone sensitive lipase; IL-1 $\beta$ : interleukin 1 $\beta$ ; IL-4: interleukin 4; IL-6: interleukin 6; IP: intraperitoneal; IP-10: interferon-gamma induced protein; IV: intravenous; JNK 1/2: c-Jun N-terminal kinase; LCAD: long-chain acyl-coenzyme A (acyl-CoA) dehydrogenase; LDLR: low-density lipoprotein receptor; LPL: lipoprotein lipase; LPS: lipopolysaccharide; M $\phi$ : macrophage; M1: classically activated macrophage; M2: alternatively activated macrophage; MAPK: mitogen-activated protein kinases; MCP-1: monocyte chemoattractant protein—1; MetS: metabolic syndrome; MIP-1 $\alpha/\beta$ : macrophage inflammatory proteins-1 alpha and beta; MKK3: mitogen-activated protein kinase 3; MMe: metabolically activated macrophage; mRNA: messenger RNA; MSC: mesenchymal stem cells; NEFA: non-esterified fatty acids; NF- $\kappa$ B: nuclear factor kappa B; OAT: omental adipose tissue; OLF78: olfactory receptor 78; Pgc-1: peroxisome proliferator-activated receptor gamma coactivator 1 alpha; PGE2: prostaglandin E2; PPAR $\gamma$ : peroxisome proliferator-activated receptor gamma; pPERK: phospho-protein kinase-like ER kinase; PRAK: p38-regulated activated protein kinase; PRDM16: PR-domain containing 16; PRKAR1A: c-AMP-dependent protein kinase type 1-alpha; PYY: peptide YY; RER: respiratory exchange ratio; RNA: ribonucleic acid; ROS: reactive oxygen species; SAT: subcutaneous adipose tissue; SCFA: short chain fatty acid; siRNA: small interfering RNA; STAT6: signal transducer and activator of transcription 6; T2D: type 2 diabetes; TNF- $\alpha$ : tumor necrosis factor alpha; UCP1: uncoupling protein 1; WAT: white adipose tissue; Ym1: chitinase-like protein.

## References

1. Blüher, M. Obesity: Global epidemiology and pathogenesis. *Nat. Rev. Endocrinol.* **2019**, *15*, 288–298. [[CrossRef](#)] [[PubMed](#)]
2. Hales, C.M.; Carroll, M.D.; Fryar, C.D.; Ogden, C.L. Prevalence of Obesity and Severe Obesity Among Adults: United States. *2017–2018 NCHS Data Brief.* **2020**, *360*, 1–8.
3. Kelly, T.; Yang, W.; Chen, C.S.; Reynolds, K.; He, J. Global burden of obesity in 2005 and projections to 2030. *Int. J. Obes.* **2008**, *32*, 1431–1437. [[CrossRef](#)] [[PubMed](#)]
4. Voelker, R. Escalating obesity rates pose health, budget threats. *JAMA* **2012**, *308*, 1514. [[CrossRef](#)] [[PubMed](#)]
5. Pagidipati, N.J.; Zheng, Y.; Green, J.B.; McGuire, D.K.; Mentz, R.J.; Shah, S.; Aschner, P.; Delibasi, T.; Rodbard, H.W.; Westerhout, C.M.; et al. Association of obesity with cardiovascular outcomes in patients with type 2 diabetes and cardiovascular disease: Insights from TECOS. *Am. Heart J.* **2020**, *219*, 47–57. [[CrossRef](#)]
6. Saraswathi, V.; Heineman, R.; Alnouti, Y.; Shivaswamy, V.; Desouza, C.V. A combination of Omega-3 PUFAs and COX inhibitors: A novel strategy to manage obesity-linked dyslipidemia and adipose tissue inflammation. *J. Diabetes Complicat.* **2020**, *34*, 107494. [[CrossRef](#)]

7. Raffield, L.M.; Howard, A.G.; Graff, M.; Lin, D.Y.; Cheng, S.; Demerath, E.; Ndumele, C.; Palta, P.; Rebholz, C.M.; Seidemann, S.; et al. Obesity Duration, Severity, and Distribution Trajectories and Cardiovascular Disease Risk in the Atherosclerosis Risk in Communities Study. *J. Am. Heart Assoc.* **2021**, *10*, e019946. [\[CrossRef\]](#)
8. Lau, E.S.; Paniagua, S.M.; Zarbafian, S.; Hoffman, U.; Long, M.T.; Hwang, S.J.; Courchesne, P.; Yao, C.; Ma, J.; Larson, M.G.; et al. Cardiovascular Biomarkers of Obesity and Overlap With Cardiometabolic Dysfunction. *J. Am. Heart Assoc.* **2021**, *10*, e020215. [\[CrossRef\]](#)
9. Marin-Alejandro, B.A.; Abete, I.; Cantero, I.; Monreal, J.I.; Elorz, M.; Herrero, J.I.; Benito-Boillos, A.; Quiroga, J.; Martinez-Echeverria, A.; Uriz-Otano, J.I.; et al. The Metabolic and Hepatic Impact of Two Personalized Dietary Strategies in Subjects with Obesity and Nonalcoholic Fatty Liver Disease: The Fatty Liver in Obesity (FLiO) Randomized Controlled Trial. *Nutrients* **2019**, *11*, 2543. [\[CrossRef\]](#)
10. Kabat, G.C.; Kim, M.Y.; Lee, J.S.; Ho, G.Y.; Going, S.B.; Beebe-Dimmer, J.; Manson, J.E.; Chlebowski, R.T.; Rohan, T.E. Metabolic Obesity Phenotypes and Risk of Breast Cancer in Postmenopausal Women. *Cancer Epidemiol. Biomark. Prev.* **2017**, *26*, 1730–1735. [\[CrossRef\]](#)
11. Sanfilippo, K.M.; McTigue, K.M.; Fidler, C.J.; Neaton, J.D.; Chang, Y.; Fried, L.F.; Liu, S.; Kuller, L.H. Hypertension and obesity and the risk of kidney cancer in 2 large cohorts of US men and women. *Hypertension* **2014**, *63*, 934–941. [\[CrossRef\]](#)
12. Liu, P.H.; Wu, K.; Ng, K.; Zauber, A.G.; Nguyen, L.H.; Song, M.; He, X.; Fuchs, C.S.; Ogino, S.; Willett, W.C.; et al. Association of Obesity With Risk of Early-Onset Colorectal Cancer Among Women. *JAMA Oncol.* **2019**, *5*, 37–44. [\[CrossRef\]](#)
13. Cao, B. Future healthy life expectancy among older adults in the US: A forecast based on cohort smoking and obesity history. *Popul. Health Metr.* **2016**, *14*, 23. [\[CrossRef\]](#)
14. Cawley, J.; Biener, A.; Meyerhoefer, C.; Ding, Y.; Zvenyach, T.; Smolarz, B.G.; Ramasamy, A. Direct medical costs of obesity in the United States and the most populous states. *J. Manag. Care Spec. Pharm.* **2021**, *27*, 354–366. [\[CrossRef\]](#)
15. Schwartz, M.W.; Seeley, R.J.; Zeltser, L.M.; Drewnowski, A.; Ravussin, E.; Redman, L.M.; Leibel, R.L. Obesity Pathogenesis: An Endocrine Society Scientific Statement. *Endocr. Rev.* **2017**, *38*, 267–296. [\[CrossRef\]](#)
16. Saxton, S.N.; Clark, B.J.; Withers, S.B.; Eringa, E.C.; Heagerty, A.M. Mechanistic Links Between Obesity, Diabetes, and Blood Pressure: Role of Perivascular Adipose Tissue. *Physiol. Rev.* **2019**, *99*, 1701–1763. [\[CrossRef\]](#)
17. Koenen, M.; Hill, M.A.; Cohen, P.; Sowers, J.R. Obesity, Adipose Tissue and Vascular Dysfunction. *Circ. Res.* **2021**, *128*, 951–968. [\[CrossRef\]](#)
18. Chait, A.; den Hartigh, L.J. Adipose Tissue Distribution, Inflammation and Its Metabolic Consequences, Including Diabetes and Cardiovascular Disease. *Front Cardiovasc. Med.* **2020**, *7*, 22. [\[CrossRef\]](#)
19. Choe, S.S.; Huh, J.Y.; Hwang, I.J.; Kim, J.I.; Kim, J.B. Adipose Tissue Remodeling: Its Role in Energy Metabolism and Metabolic Disorders. *Front Endocrinol.* **2016**, *7*, 30. [\[CrossRef\]](#)
20. Vishvanath, L.; Gupta, R.K. Contribution of adipogenesis to healthy adipose tissue expansion in obesity. *J. Clin. Investig* **2019**, *129*, 4022–4031. [\[CrossRef\]](#)
21. Sakers, A.; De Siqueira, M.K.; Seale, P.; Villanueva, C.J. Adipose-tissue plasticity in health and disease. *Cell* **2022**, *185*, 419–446. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Regnier, S.M.; Sargis, R.M. Adipocytes under assault: Environmental disruption of adipose physiology. *Biochim. Biophys. Acta* **2014**, *1842*, 520–533. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Lee, Y.S.; Olefsky, J. Chronic tissue inflammation and metabolic disease. *Genes. Dev.* **2021**, *35*, 307–328. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Bays, H.E. Adiposopathy is “sick fat” a cardiovascular disease? *J. Am. Coll. Cardiol.* **2011**, *57*, 2461–2473. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Lumeng, C.N.; Bodzin, J.L.; Saltiel, A.R. Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J. Clin. Investig.* **2007**, *117*, 175–184. [\[CrossRef\]](#)
26. Longo, M.; Zatterale, F.; Naderi, J.; Parrillo, L.; Formisano, P.; Raciti, G.A.; Beguinot, F.; Miele, C. Adipose Tissue Dysfunction as Determinant of Obesity-Associated Metabolic Complications. *Int. J. Mol. Sci.* **2019**, *20*, 2358. [\[CrossRef\]](#)
27. Genné-Bacon, E.A. Thinking evolutionarily about obesity. *Yale J. Biol. Med.* **2014**, *87*, 99–112.
28. Heymsfield, S.B.; Wadden, T.A. Mechanisms, Pathophysiology, and Management of Obesity. *N. Engl. J. Med.* **2017**, *376*, 254–266. [\[CrossRef\]](#)
29. Nicolaidis, S. Environment and obesity. *Metabolism* **2019**, *100S*, 153942. [\[CrossRef\]](#)
30. Qasim, A.; Turcotte, M.; de Souza, R.J.; Samaan, M.C.; Champredon, D.; Dushoff, J.; Speakman, J.R.; Meyre, D. On the origin of obesity: Identifying the biological, environmental and cultural drivers of genetic risk among human populations. *Obes. Rev.* **2018**, *19*, 121–149. [\[CrossRef\]](#)
31. Bäckhed, F.; Ding, H.; Wang, T.; Hooper, L.V.; Koh, G.Y.; Nagy, A.; Semenkovich, C.F.; Gordon, J.I. The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 15718–15723. [\[CrossRef\]](#)
32. Turnbaugh, P.J.; Ley, R.E.; Mahowald, M.A.; Magrini, V.; Mardis, E.R.; Gordon, J.I. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* **2006**, *444*, 1027–1031. [\[CrossRef\]](#)
33. Bäckhed, F.; Manchester, J.K.; Semenkovich, C.F.; Gordon, J.I. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 979–984. [\[CrossRef\]](#)
34. Turnbaugh, P.J.; Bäckhed, F.; Fulton, L.; Gordon, J.I. Diet-Induced Obesity Is Linked to Marked but Reversible Alterations in the Mouse Distal Gut Microbiome. *Cell Host Microbe* **2008**, *3*, 213–223. [\[CrossRef\]](#)

35. Ley, R.E.; Bäckhed, F.; Turnbaugh, P.; Lozupone, C.A.; Knight, R.D.; Gordon, J.I. Obesity alters gut microbial ecology. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 11070–11075. [[CrossRef](#)]
36. Bjørneklett, A.; Viddal, K.O.; Midtvedt, T.; Nygaard, K. Intestinal and gastric bypass. Changes in intestinal microecology after surgical treatment of morbid obesity in man. *Scand. J. Gastroenterol.* **1981**, *16*, 681–687. [[CrossRef](#)]
37. Ley, R.E.; Turnbaugh, P.J.; Klein, S.; Gordon, J.I. Microbial ecology: Human gut microbes associated with obesity. *Nature* **2006**, *444*, 1022–1023. [[CrossRef](#)]
38. Turnbaugh, P.J.; Hamady, M.; Yatsunenkov, T.; Cantarel, B.L.; Duncan, A.; Ley, R.E.; Sogin, M.L.; Jones, W.J.; Roe, B.A.; Affourtit, J.P.; et al. A core gut microbiome in obese and lean twins. *Nature* **2009**, *457*, 480–484. [[CrossRef](#)]
39. Arora, T.; Bäckhed, F. The gut microbiota and metabolic disease: Current understanding and future perspectives. *J. Intern. Med.* **2016**, *280*, 339–349. [[CrossRef](#)]
40. Wu, H.; Tremaroli, V.; Bäckhed, F. Linking Microbiota to Human Diseases: A Systems Biology Perspective. *Trends Endocrinol. Metab.* **2015**, *26*, 758–770. [[CrossRef](#)]
41. de Vos, W.M.; Tilg, H.; Van Hul, M.; Cani, P.D. Gut microbiome and health: Mechanistic insights. *Gut* **2022**, *71*, 1020–1032. [[CrossRef](#)] [[PubMed](#)]
42. Duranti, S.; Ferrario, C.; van Sinderen, D.; Ventura, M.; Turrone, F. Obesity and microbiota: An example of an intricate relationship. *Genes. Nutr.* **2017**, *12*, 18. [[CrossRef](#)] [[PubMed](#)]
43. Cani, P.D.; Delzenne, N.M.; Amar, J.; Burcelin, R. Role of gut microflora in the development of obesity and insulin resistance following high-fat diet feeding. *Pathol. Biol.* **2008**, *56*, 305–309. [[CrossRef](#)] [[PubMed](#)]
44. Cani, P.D.; Delzenne, N.M. Gut microflora as a target for energy and metabolic homeostasis. *Curr. Opin. Clin. Nutr. Metab. Care* **2007**, *10*, 729–734. [[CrossRef](#)] [[PubMed](#)]
45. Delzenne, N.M.; Cani, P.D. Gut microflora is a key player in host energy homeostasis. *Med. Sci.* **2008**, *24*, 505–510. [[CrossRef](#)]
46. Eckburg, P.B.; Bik, E.M.; Bernstein, C.N.; Purdom, E.; Dethlefsen, L.; Sargent, M.; Gill, S.R.; Nelson, K.E.; Relman, D.A. Diversity of the human intestinal microbial flora. *Science* **2005**, *308*, 1635–1638. [[CrossRef](#)]
47. Lloyd-Price, J.; Mahurkar, A.; Rahnnavard, G.; Crabtree, J.; Orvis, J.; Hall, A.B.; Brady, A.; Creasy, H.H.; McCracken, C.; Giglio, M.G.; et al. Strains, functions and dynamics in the expanded Human Microbiome Project. *Nature* **2017**, *550*, 61–66. [[CrossRef](#)]
48. Lloyd-Price, J.; Mahurkar, A.; Rahnnavard, G.; Crabtree, J.; Orvis, J.; Hall, A.B.; Brady, A.; Creasy, H.H.; McCracken, C.; Giglio, M.G.; et al. Erratum: Strains, functions and dynamics in the expanded Human Microbiome Project. *Nature* **2017**, *551*, 256. [[CrossRef](#)]
49. Cresci, G.A.; Bawden, E. Gut Microbiome: What We Do and Don't Know. *Nutr. Clin. Pract.* **2015**, *30*, 734–746. [[CrossRef](#)]
50. Turnbaugh, P.J.; Quince, C.; Faith, J.J.; McHardy, A.C.; Yatsunenkov, T.; Niazi, F.; Affourtit, J.; Egholm, M.; Henrissat, B.; Knight, R.; et al. Organismal, genetic, and transcriptional variation in the deeply sequenced gut microbiomes of identical twins. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 7503–7508. [[CrossRef](#)]
51. Scott, K.P.; Gratz, S.W.; Sheridan, P.O.; Flint, H.J.; Duncan, S.H. The influence of diet on the gut microbiota. *Pharmacol. Res.* **2013**, *69*, 52–60. [[CrossRef](#)]
52. Delzenne, N.M.; Rodriguez, J. Nutrition and Microbiome. *Handb. Exp. Pharmacol.* **2022**, *274*, 57–73. [[CrossRef](#)]
53. Walters, W.A.; Xu, Z.; Knight, R. Meta-analyses of human gut microbes associated with obesity and IBD. *FEBS Lett.* **2014**, *588*, 4223–4233. [[CrossRef](#)]
54. Sze, M.A.; Schloss, P.D. Looking for a Signal in the Noise: Revisiting Obesity and the Microbiome. *mBio* **2016**, *7*, e01018-16. [[CrossRef](#)]
55. Schwiertz, A.; Taras, D.; Schäfer, K.; Beijer, S.; Bos, N.A.; Donus, C.; Hardt, P.D. Microbiota and SCFA in lean and overweight healthy subjects. *Obesity* **2010**, *18*, 190–195. [[CrossRef](#)]
56. Duncan, S.H.; Lobeley, G.E.; Holtrop, G.; Ince, J.; Johnstone, A.M.; Louis, P.; Flint, H.J. Human colonic microbiota associated with diet, obesity and weight loss. *Int. J. Obes.* **2008**, *32*, 1720–1724. [[CrossRef](#)]
57. Rabot, S.; Membrez, M.; Bruneau, A.; Gérard, P.; Harach, T.; Moser, M.; Raymond, F.; Mansourian, R.; Chou, C.J. Germ-free C57BL/6J mice are resistant to high-fat-diet-induced insulin resistance and have altered cholesterol metabolism. *FASEB J.* **2010**, *24*, 4948–4959. [[CrossRef](#)]
58. Yi, C.X.; Tschöp, M.H. Brain-gut-adipose-tissue communication pathways at a glance. *Dis. Model Mech.* **2012**, *5*, 583–587. [[CrossRef](#)]
59. Cani, P.D.; Possemiers, S.; Van de Wiele, T.; Guiot, Y.; Everard, A.; Rottier, O.; Geurts, L.; Naslain, D.; Neyrinck, A.; Lambert, D.M.; et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. *Gut* **2009**, *58*, 1091–1103. [[CrossRef](#)]
60. Cani, P.D.; Amar, J.; Iglesias, M.A.; Poggi, M.; Knauf, C.; Bastelica, D.; Neyrinck, A.M.; Fava, F.; Tuohy, K.M.; Chabo, C.; et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* **2007**, *56*, 1761–1772. [[CrossRef](#)]
61. Zheng, D.; Liwinski, T.; Elinav, E. Interaction between microbiota and immunity in health and disease. *Cell Res.* **2020**, *30*, 492–506. [[CrossRef](#)] [[PubMed](#)]
62. Cani, P.D.; Bibiloni, R.; Knauf, C.; Waget, A.; Neyrinck, A.M.; Delzenne, N.M.; Burcelin, R. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes* **2008**, *57*, 1470–1481. [[CrossRef](#)] [[PubMed](#)]

63. Cunningham, A.L.; Stephens, J.W.; Harris, D.A. Intestinal microbiota and their metabolic contribution to type 2 diabetes and obesity. *J. Diabetes. Metab. Disord.* **2021**, *20*, 1855–1870. [[CrossRef](#)] [[PubMed](#)]
64. Peterson, C.T.; Perez Santiago, J.; Iablokov, S.N.; Chopra, D.; Rodionov, D.A.; Peterson, S.N. Short-Chain Fatty Acids Modulate Healthy Gut Microbiota Composition and Functional Potential. *Curr. Microbiol.* **2022**, *79*, 128. [[CrossRef](#)] [[PubMed](#)]
65. Byrne, C.S.; Chambers, E.S.; Morrison, D.J.; Frost, G. The role of short chain fatty acids in appetite regulation and energy homeostasis. *Int. J. Obes.* **2015**, *39*, 1331–1338. [[CrossRef](#)] [[PubMed](#)]
66. Blaut, M. Gut microbiota and energy balance: Role in obesity. *Proc. Nutr. Soc.* **2015**, *74*, 227–234. [[CrossRef](#)]
67. Hu, J.; Lin, S.; Zheng, B.; Cheung, P.C.K. Short-chain fatty acids in control of energy metabolism. *Crit. Rev. Food Sci. Nutr.* **2018**, *58*, 1243–1249. [[CrossRef](#)]
68. Gao, Z.; Yin, J.; Zhang, J.; Ward, R.E.; Martin, R.J.; Lefevre, M.; Cefalu, W.T.; Ye, J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* **2009**, *58*, 1509–1517. [[CrossRef](#)]
69. Canfora, E.E.; Blaak, E.E. Acetate: A diet-derived key metabolite in energy metabolism: Good or bad in context of obesity and glucose homeostasis? *Curr. Opin. Clin. Nutr. Metab. Care* **2017**, *20*, 477–483. [[CrossRef](#)]
70. Zhang, L.; Liu, C.; Jiang, Q.; Yin, Y. Butyrate in Energy Metabolism: There Is Still More to Learn. *Trends Endocrinol. Metab.* **2021**, *32*, 159–169. [[CrossRef](#)]
71. Chambers, E.S.; Byrne, C.S.; Aspey, K.; Chen, Y.; Khan, S.; Morrison, D.J.; Frost, G. Acute oral sodium propionate supplementation raises resting energy expenditure and lipid oxidation in fasted humans. *Diabetes Obes. Metab.* **2018**, *20*, 1034–1039. [[CrossRef](#)]
72. Hill, J.H.; Round, J.L. Snapshot: Microbiota effects on host physiology. *Cell* **2021**, *184*, 2796. [[CrossRef](#)]
73. Sanna, S.; van Zuydam, N.R.; Mahajan, A.; Kurilshikov, A.; Vich Vila, A.; Vösa, U.; Mujagic, Z.; Masclee, A.A.M.; Jonkers, D.M.A.E.; Oosting, M.; et al. Causal relationships among the gut microbiome, short-chain fatty acids and metabolic diseases. *Nat. Genet.* **2019**, *51*, 600–605. [[CrossRef](#)]
74. Koh, A.; De Vadder, F.; Kovatcheva-Datchary, P.; Bäckhed, F. From Dietary Fiber to Host Physiology: Short-Chain Fatty Acids as Key Bacterial Metabolites. *Cell* **2016**, *165*, 1332–1345. [[CrossRef](#)]
75. Cani, P.D. Targeting gut microbiota with a complex mix of dietary fibers improves metabolic diseases. *Kidney Int.* **2019**, *95*, 14–16. [[CrossRef](#)]
76. Beam, A.; Clinger, E.; Hao, L. Effect of Diet and Dietary Components on the Composition of the Gut Microbiota. *Nutrients* **2021**, *13*, 2795. [[CrossRef](#)]
77. Marques, F.Z.; Nelson, E.; Chu, P.Y.; Horlock, D.; Fiedler, A.; Ziemann, M.; Tan, J.K.; Kuruppu, S.; Rajapakse, N.W.; El-Osta, A.; et al. High-Fiber Diet and Acetate Supplementation Change the Gut Microbiota and Prevent the Development of Hypertension and Heart Failure in Hypertensive Mice. *Circulation* **2017**, *135*, 964–977. [[CrossRef](#)]
78. Cummings, J.H.; Pomare, E.W.; Branch, W.J.; Naylor, C.P.; Macfarlane, G.T. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut* **1987**, *28*, 1221–1227. [[CrossRef](#)]
79. Morrison, D.J.; Preston, T. Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes* **2016**, *7*, 189–200. [[CrossRef](#)]
80. Cook, S.I.; Sellin, J.H. Review article: Short chain fatty acids in health and disease. *Aliment. Pharm.* **1998**, *12*, 499–507. [[CrossRef](#)]
81. den Besten, G.; van Eunen, K.; Groen, A.K.; Venema, K.; Reijngoud, D.J.; Bakker, B.M. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J. Lipid. Res.* **2013**, *54*, 2325–2340. [[CrossRef](#)] [[PubMed](#)]
82. Bliss, D.Z.; Weimer, P.J.; Jung, H.J.; Savik, K. In vitro degradation and fermentation of three dietary fiber sources by human colonic bacteria. *J. Agric. Food Chem.* **2013**, *61*, 4614–4621. [[CrossRef](#)] [[PubMed](#)]
83. Cox, L.M.; Blaser, M.J. Pathways in microbe-induced obesity. *Cell Metab.* **2013**, *17*, 883–894. [[CrossRef](#)] [[PubMed](#)]
84. Louis, P.; Young, P.; Holtrop, G.; Flint, H.J. Diversity of human colonic butyrate-producing bacteria revealed by analysis of the butyryl-CoA:acetate CoA-transferase gene. *Environ. Microbiol.* **2010**, *12*, 304–314. [[CrossRef](#)] [[PubMed](#)]
85. Louis, P.; Flint, H.J. Formation of propionate and butyrate by the human colonic microbiota. *Environ. Microbiol.* **2017**, *19*, 29–41. [[CrossRef](#)]
86. Tsukuda, N.; Yahagi, K.; Hara, T.; Watanabe, Y.; Matsumoto, H.; Mori, H.; Higashi, K.; Tsuji, H.; Matsumoto, S.; Kurokawa, K.; et al. Key bacterial taxa and metabolic pathways affecting gut short-chain fatty acid profiles in early life. *ISME J.* **2021**, *15*, 2574–2590. [[CrossRef](#)]
87. Perry, R.J.; Peng, L.; Barry, N.A.; Cline, G.W.; Zhang, D.; Cardone, R.L.; Petersen, K.F.; Kibbey, R.G.; Goodman, A.L.; Shulman, G.I. Acetate mediates a microbiome-brain- $\beta$ -cell axis to promote metabolic syndrome. *Nature* **2016**, *534*, 213–217. [[CrossRef](#)]
88. den Hartigh, L.J.; Gao, Z.; Goodspeed, L.; Wang, S.; Das, A.K.; Burant, C.F.; Chait, A.; Blaser, M.J. Obese Mice Losing Weight Due to trans-10,cis-12 Conjugated Linoleic Acid Supplementation or Food Restriction Harbor Distinct Gut Microbiota. *J. Nutr.* **2018**, *148*, 562–572. [[CrossRef](#)]
89. Skutches, C.L.; Holroyde, C.P.; Myers, R.N.; Paul, P.; Reichard, G.A. Plasma acetate turnover and oxidation. *J. Clin. Investig.* **1979**, *64*, 708–713. [[CrossRef](#)]
90. Fleming, S.E.; Choi, S.Y.; Fitch, M.D. Absorption of short-chain fatty acids from the rat cecum in vivo. *J. Nutr.* **1991**, *121*, 1787–1797. [[CrossRef](#)]
91. Clausen, M.R.; Mortensen, P.B. Kinetic studies on colonocyte metabolism of short chain fatty acids and glucose in ulcerative colitis. *Gut* **1995**, *37*, 684–689. [[CrossRef](#)]

92. Jørgensen, J.R.; Clausen, M.R.; Mortensen, P.B. Oxidation of short and medium chain C2-C8 fatty acids in Sprague-Dawley rat colonocytes. *Gut* **1997**, *40*, 400–405. [[CrossRef](#)]
93. Wolever, T.M.; Spadafora, P.; Eshuis, H. Interaction between colonic acetate and propionate in humans. *Am. J. Clin. Nutr.* **1991**, *53*, 681–687. [[CrossRef](#)]
94. Wong, J.M.; de Souza, R.; Kendall, C.W.; Emam, A.; Jenkins, D.J. Colonic health: Fermentation and short chain fatty acids. *J. Clin. Gastroenterol.* **2006**, *40*, 235–243. [[CrossRef](#)]
95. Boets, E.; Gomand, S.V.; Deroover, L.; Preston, T.; Vermeulen, K.; De Preter, V.; Hamer, H.M.; Van den Mooter, G.; De Vuyst, L.; Courtin, C.M.; et al. Systemic availability and metabolism of colonic-derived short-chain fatty acids in healthy subjects: A stable isotope study. *J. Physiol.* **2017**, *595*, 541–555. [[CrossRef](#)]
96. Walker, A.W.; Duncan, S.H.; McWilliam Leitch, E.C.; Child, M.W.; Flint, H.J. pH and peptide supply can radically alter bacterial populations and short-chain fatty acid ratios within microbial communities from the human colon. *Appl. Environ. Microbiol.* **2005**, *71*, 3692–3700. [[CrossRef](#)]
97. Xiao, S.; Jiang, S.; Qian, D.; Duan, J. Modulation of microbially derived short-chain fatty acids on intestinal homeostasis, metabolism, and neuropsychiatric disorder. *Appl. Microbiol. Biotechnol.* **2020**, *104*, 589–601. [[CrossRef](#)]
98. Ohira, H.; Tsutsui, W.; Fujioka, Y. Are Short Chain Fatty Acids in Gut Microbiota Defensive Players for Inflammation and Atherosclerosis? *J. Atheroscler. Thromb.* **2017**, *24*, 660–672. [[CrossRef](#)]
99. Portincasa, P.; Bonfrate, L.; Vacca, M.; De Angelis, M.; Farella, I.; Lanza, E.; Khalil, M.; Wang, D.Q.; Sperandio, M.; Di Ciaula, A. Gut Microbiota and Short Chain Fatty Acids: Implications in Glucose Homeostasis. *Int. J. Mol. Sci.* **2022**, *23*, 1105. [[CrossRef](#)]
100. Popeijus, H.E.; Zwaan, W.; Tayyeb, J.Z.; Plat, J. Potential Contribution of Short Chain Fatty Acids to Hepatic Apolipoprotein A-I Production. *Int. J. Mol. Sci.* **2021**, *22*, 5986. [[CrossRef](#)]
101. Nishitsuji, K.; Xiao, J.; Nagatomo, R.; Umemoto, H.; Morimoto, Y.; Akatsu, H.; Inoue, K.; Tsuneyama, K. Analysis of the gut microbiome and plasma short-chain fatty acid profiles in a spontaneous mouse model of metabolic syndrome. *Sci. Rep.* **2017**, *7*, 1–10. [[CrossRef](#)] [[PubMed](#)]
102. Murphy, E.F.; Cotter, P.D.; Healy, S.; Marques, T.M.; O’Sullivan, O.; Fouhy, F.; Clarke, S.F.; O’Toole, P.W.; Quigley, E.M.; Stanton, C.; et al. Composition and energy harvesting capacity of the gut microbiota: Relationship to diet, obesity and time in mouse models. *Gut* **2010**, *59*, 1635–1642. [[CrossRef](#)] [[PubMed](#)]
103. Zhang, M.; Liu, J.; Li, C.; Gao, J.; Xu, C.; Wu, X.; Xu, T.; Cui, C.; Wei, H.; Peng, J.; et al. Functional Fiber Reduces Mice Obesity by Regulating Intestinal Microbiota. *Nutrients* **2022**, *14*, 2676. [[CrossRef](#)] [[PubMed](#)]
104. Arora, T.; Rudenko, O.; Egerod, K.L.; Husted, A.S.; Kovatcheva-Datchary, P.; Akrami, R.; Kristensen, M.; Schwartz, T.W.; Bäckhed, F. Microbial fermentation of flaxseed fibers modulates the transcriptome of GPR41-expressing enteroendocrine cells and protects mice against diet-induced obesity. *Am. J. Physiol. Endocrinol. Metab.* **2019**, *316*, E453–E463. [[CrossRef](#)] [[PubMed](#)]
105. May, K.S.; den Hartigh, L.J. Modulation of Adipocyte Metabolism by Microbial Short-Chain Fatty Acids. *Nutrients* **2021**, *13*, 3666. [[CrossRef](#)] [[PubMed](#)]
106. Borgeraas, H.; Johnson, L.K.; Skattebu, J.; Hertel, J.K.; Hjelmessaeth, J. Effects of probiotics on body weight, body mass index, fat mass and fat percentage in subjects with overweight or obesity: A systematic review and meta-analysis of randomized controlled trials. *Obes. Rev.* **2018**, *19*, 219–232. [[CrossRef](#)]
107. Barczyńska, R.; Litwin, M.; Sliżewska, K.; Szalecki, M.; Berdowska, A.; Bandurska, K.; Libudzisz, Z.; Kapuśniak, J. Bacterial Microbiota and Fatty Acids in the Faeces of Overweight and Obese Children. *Pol. J. Microbiol.* **2018**, *67*, 339–345. [[CrossRef](#)]
108. Riva, A.; Borgo, F.; Lassandro, C.; Verduci, E.; Morace, G.; Borghi, E.; Berry, D. Pediatric obesity is associated with an altered gut microbiota and discordant shifts in Firmicutes populations. *Environ. Microbiol.* **2017**, *19*, 95–105. [[CrossRef](#)]
109. Fernandes, J.; Su, W.; Rahat-Rozenbloom, S.; Wolever, T.M.; Comelli, E.M. Adiposity, gut microbiota and faecal short chain fatty acids are linked in adult humans. *Nutr. Diabetes* **2014**, *4*, e121. [[CrossRef](#)]
110. Rahat-Rozenbloom, S.; Fernandes, J.; Gloor, G.B.; Wolever, T.M. Evidence for greater production of colonic short-chain fatty acids in overweight than lean humans. *Int. J. Obes.* **2014**, *38*, 1525–1531. [[CrossRef](#)]
111. Rios-Covian, D.; González, S.; Nogacka, A.M.; Arboleya, S.; Salazar, N.; Gueimonde, M.; de Los Reyes-Gavilán, C.G. An Overview on Fecal Branched Short-Chain Fatty Acids Along Human Life and as Related With Body Mass Index: Associated Dietary and Anthropometric Factors. *Front. Microbiol.* **2020**, *11*, 973. [[CrossRef](#)]
112. Kim, K.N.; Yao, Y.; Ju, S.Y. Short Chain Fatty Acids and Fecal Microbiota Abundance in Humans with Obesity: A Systematic Review and Meta-Analysis. *Nutrients* **2019**, *11*, 2512. [[CrossRef](#)]
113. Wang, Y.; Wang, H.; Howard, A.G.; Meyer, K.A.; Tsilimigras, M.C.B.; Avery, C.L.; Sha, W.; Sun, S.; Zhang, J.; Su, C.; et al. Circulating Short-Chain Fatty Acids Are Positively Associated with Adiposity Measures in Chinese Adults. *Nutrients* **2020**, *12*, 2127. [[CrossRef](#)]
114. Yin, X.Q.; An, Y.X.; Yu, C.G.; Ke, J.; Zhao, D.; Yu, K. The Association Between Fecal Short-Chain Fatty Acids, Gut Microbiota, and Visceral Fat in Monozygotic Twin Pairs. *Diabetes Metab. Syndr. Obes.* **2022**, *15*, 359–368. [[CrossRef](#)]
115. Miranda, V.P.N.; Dos Santos Amorim, P.R.; Bastos, R.R.; de Faria, E.R.; de Castro Moreira, M.E.; do Carmo Castro Franceschini, S.; do Carmo Gouveia Peluzio, M.; de Lucis Fortes Ferreira, C.L.; Priore, S.E. Abundance of Gut Microbiota, Concentration of Short-Chain Fatty Acids, and Inflammatory Markers Associated with Elevated Body Fat, Overweight, and Obesity in Female Adolescents. *Mediat. Inflamm.* **2019**, *2019*, 7346863. [[CrossRef](#)]

116. Murugesan, S.; Ulloa-Martínez, M.; Martínez-Rojano, H.; Galván-Rodríguez, F.M.; Miranda-Brito, C.; Romano, M.C.; Piña-Escobedo, A.; Pizano-Zárate, M.L.; Hoyo-Vadillo, C.; García-Mena, J. Study of the diversity and short-chain fatty acids production by the bacterial community in overweight and obese Mexican children. *Eur. J. Clin. Microbiol. Infect. Dis.* **2015**, *34*, 1337–1346. [[CrossRef](#)]
117. Mishra, S.P.; Karunakar, P.; Taraphder, S.; Yadav, H. Free Fatty Acid Receptors 2 and 3 as Microbial Metabolite Sensors to Shape Host Health: Pharmacophysiological View. *Biomedicines* **2020**, *8*, 154. [[CrossRef](#)]
118. Stoddart, L.A.; Smith, N.J.; Milligan, G. International Union of Pharmacology. LXXI. Free fatty acid receptors FFA1, -2, and -3: Pharmacology and pathophysiological functions. *Pharmacol. Rev.* **2008**, *60*, 405–417. [[CrossRef](#)]
119. Miyamoto, J.; Hasegawa, S.; Kasubuchi, M.; Ichimura, A.; Nakajima, A.; Kimura, I. Nutritional Signaling via Free Fatty Acid Receptors. *Int. J. Mol. Sci.* **2016**, *17*, 450. [[CrossRef](#)]
120. Kimura, I.; Ichimura, A.; Ohue-Kitano, R.; Igarashi, M. Free Fatty Acid Receptors in Health and Disease. *Physiol. Rev.* **2020**, *100*, 171–210. [[CrossRef](#)]
121. Brown, A.J.; Goldsworthy, S.M.; Barnes, A.A.; Eilert, M.M.; Tcheang, L.; Daniels, D.; Muir, A.I.; Wigglesworth, M.J.; Kinghorn, I.; Fraser, N.J.; et al. The orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *J. Biol. Chem.* **2003**, *278*, 11312–11319. [[CrossRef](#)] [[PubMed](#)]
122. Li, G.; Su, H.; Zhou, Z.; Yao, W. Identification of the porcine G protein-coupled receptor 41 and 43 genes and their expression pattern in different tissues and development stages. *PLoS ONE* **2014**, *9*, e97342. [[CrossRef](#)] [[PubMed](#)]
123. Li, G.; Su, H.; Zhou, Z.; Yao, W. Correction: Identification of the Porcine G Protein-Coupled Receptor 41 and 43 Genes and Their Expression Pattern in Different Tissues and Development Stages. *PLoS ONE* **2020**, *15*, e0239768. [[CrossRef](#)] [[PubMed](#)]
124. Wu, W.; Sun, M.; Chen, F.; Cao, A.T.; Liu, H.; Zhao, Y.; Huang, X.; Xiao, Y.; Yao, S.; Zhao, Q.; et al. Microbiota metabolite short-chain fatty acid acetate promotes intestinal IgA response to microbiota which is mediated by GPR43. *Mucosal. Immunol.* **2017**, *10*, 946–956. [[CrossRef](#)]
125. Kim, M.H.; Kang, S.G.; Park, J.H.; Yanagisawa, M.; Kim, C.H. Short-chain fatty acids activate GPR41 and GPR43 on intestinal epithelial cells to promote inflammatory responses in mice. *Gastroenterology* **2013**, *145*, 396–406. [[CrossRef](#)]
126. Nakajima, A.; Nakatani, A.; Hasegawa, S.; Irie, J.; Ozawa, K.; Tsujimoto, G.; Suganami, T.; Itoh, H.; Kimura, I. The short chain fatty acid receptor GPR43 regulates inflammatory signals in adipose tissue M2-type macrophages. *PLoS ONE* **2017**, *12*, e0179696. [[CrossRef](#)]
127. Trompette, A.; Gollwitzer, E.S.; Yadava, K.; Sichelstiel, A.K.; Sprenger, N.; Ngom-Bru, C.; Blanchard, C.; Junt, T.; Nicod, L.P.; Harris, N.L.; et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat. Med.* **2014**, *20*, 159–166. [[CrossRef](#)]
128. Xu, M.; Jiang, Z.; Wang, C.; Li, N.; Bo, L.; Zha, Y.; Bian, J.; Zhang, Y.; Deng, X. Acetate attenuates inflammasome activation through GPR43-mediated Ca. *Exp. Mol. Med.* **2019**, *51*, 1–13. [[CrossRef](#)]
129. Lin, M.; Jiang, M.; Yang, T.; Zhao, G.; Zhan, K. Overexpression of GPR41 attenuated glucose production in propionate-induced bovine hepatocytes. *Front. Vet. Sci.* **2022**, *9*, 981640. [[CrossRef](#)]
130. Luo, Q.J.; Sun, M.X.; Guo, Y.W.; Tan, S.W.; Wu, X.Y.; Abassa, K.K.; Lin, L.; Liu, H.L.; Jiang, J.; Wei, X.Q. Sodium butyrate protects against lipopolysaccharide-induced liver injury partially via the GPR43/ $\beta$ -arrestin-2/NF- $\kappa$ B network. *Gastroenterol. Rep.* **2021**, *9*, 154–165. [[CrossRef](#)]
131. Xiong, Y.; Miyamoto, N.; Shibata, K.; Valasek, M.A.; Motoike, T.; Kedzierski, R.M.; Yanagisawa, M. Short-chain fatty acids stimulate leptin production in adipocytes through the G protein-coupled receptor GPR41. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 1045–1050. [[CrossRef](#)]
132. Kimura, I.; Ozawa, K.; Inoue, D.; Imamura, T.; Kimura, K.; Maeda, T.; Terasawa, K.; Kashihara, D.; Hirano, K.; Tani, T.; et al. The gut microbiota suppresses insulin-mediated fat accumulation via the short-chain fatty acid receptor GPR43. *Nat. Commun.* **2013**, *4*, 1829. [[CrossRef](#)]
133. Lu, Y.; Fan, C.; Li, P.; Chang, X.; Qi, K. Short Chain Fatty Acids Prevent High-fat-diet-induced Obesity in Mice by Regulating G Protein-coupled Receptors and Gut Microbiota. *Sci. Rep.* **2016**, *6*, 37589. [[CrossRef](#)]
134. Nishida, A.; Miyamoto, J.; Shimizu, H.; Kimura, I. Gut microbial short-chain fatty acids-mediated olfactory receptor 78 stimulation promotes anorexigenic gut hormone peptide YY secretion in mice. *Biochem. Biophys. Res. Commun.* **2021**, *557*, 48–54. [[CrossRef](#)]
135. Jiao, A.; Yu, B.; He, J.; Yu, J.; Zheng, P.; Luo, Y.; Luo, J.; Yan, H.; Wang, Q.; Wang, H.; et al. Sodium acetate, propionate, and butyrate reduce fat accumulation in mice via modulating appetite and relevant genes. *Nutrition* **2021**, *87–88*, 111198. [[CrossRef](#)]
136. Iván, J.; Major, E.; Sipos, A.; Kovács, K.; Horváth, D.; Tamás, I.; Bay, P.; Dombrádi, V.; Lontay, B. The Short-Chain Fatty Acid Propionate Inhibits Adipogenic Differentiation of Human Chorion-Derived Mesenchymal Stem Cells Through the Free Fatty Acid Receptor 2. *Stem Cells Dev.* **2017**, *26*, 1724–1733. [[CrossRef](#)]
137. Dewulf, E.M.; Ge, Q.; Bindels, L.B.; Sohet, F.M.; Cani, P.D.; Brichard, S.M.; Delzenne, N.M. Evaluation of the relationship between GPR43 and adiposity in human. *Nutr. Metab.* **2013**, *10*, 11. [[CrossRef](#)]
138. Ge, H.; Li, X.; Weiszmann, J.; Wang, P.; Baribault, H.; Chen, J.L.; Tian, H.; Li, Y. Activation of G protein-coupled receptor 43 in adipocytes leads to inhibition of lipolysis and suppression of plasma free fatty acids. *Endocrinology* **2008**, *149*, 4519–4526. [[CrossRef](#)]
139. Hong, Y.H.; Nishimura, Y.; Hishikawa, D.; Tsuzuki, H.; Miyahara, H.; Gotoh, C.; Choi, K.C.; Feng, D.D.; Chen, C.; Lee, H.G.; et al. Acetate and propionate short chain fatty acids stimulate adipogenesis via GPCR43. *Endocrinology* **2005**, *146*, 5092–5099. [[CrossRef](#)]

140. Ohira, H.; Fujioka, Y.; Katagiri, C.; Mamoto, R.; Aoyama-Ishikawa, M.; Amako, K.; Izumi, Y.; Nishiumi, S.; Yoshida, M.; Usami, M.; et al. Butyrate attenuates inflammation and lipolysis generated by the interaction of adipocytes and macrophages. *J. Atheroscler. Thromb.* **2013**, *20*, 425–442. [[CrossRef](#)]
141. Frost, G.; Cai, Z.; Raven, M.; Otway, D.T.; Mushtaq, R.; Johnston, J.D. Effect of short chain fatty acids on the expression of free fatty acid receptor 2 (Ffar2), Ffar3 and early-stage adipogenesis. *Nutr. Diabetes* **2014**, *4*, e128. [[CrossRef](#)] [[PubMed](#)]
142. Bjursell, M.; Admyre, T.; Göransson, M.; Marley, A.E.; Smith, D.M.; Oscarsson, J.; Bohlooly, Y.M. Improved glucose control and reduced body fat mass in free fatty acid receptor 2-deficient mice fed a high-fat diet. *Am. J. Physiol. Endocrinol. Metab.* **2011**, *300*, E211–E220. [[CrossRef](#)] [[PubMed](#)]
143. Han, J.H.; Kim, I.S.; Jung, S.H.; Lee, S.G.; Son, H.Y.; Myung, C.S. The effects of propionate and valerate on insulin responsiveness for glucose uptake in 3T3-L1 adipocytes and C2C12 myotubes via G protein-coupled receptor 41. *PLoS ONE* **2014**, *9*, e95268. [[CrossRef](#)] [[PubMed](#)]
144. Zaibi, M.S.; Stocker, C.J.; O'Dowd, J.; Davies, A.; Bellahcene, M.; Cawthorne, M.A.; Brown, A.J.; Smith, D.M.; Arch, J.R. Roles of GPR41 and GPR43 in leptin secretory responses of murine adipocytes to short chain fatty acids. *FEBS Lett.* **2010**, *584*, 2381–2386. [[CrossRef](#)] [[PubMed](#)]
145. Jocken, J.W.E.; González Hernández, M.A.; Hoebbers, N.T.H.; van der Beek, C.M.; Essers, Y.P.G.; Blaak, E.E.; Canfora, E.E. Short-Chain Fatty Acids Differentially Affect Intracellular Lipolysis in a Human White Adipocyte Model. *Front Endocrinol.* **2017**, *8*, 372. [[CrossRef](#)]
146. Le Poul, E.; Loison, C.; Struyf, S.; Springael, J.Y.; Lannoy, V.; Decobecq, M.E.; Brezillon, S.; Dupriez, V.; Vassart, G.; Van Damme, J.; et al. Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. *J. Biol. Chem.* **2003**, *278*, 25481–25489. [[CrossRef](#)]
147. Al-Lahham, S.; Roelofsen, H.; Rezaee, F.; Weening, D.; Hoek, A.; Vonk, R.; Venema, K. Propionic acid affects immune status and metabolism in adipose tissue from overweight subjects. *Eur. J. Clin. Investig.* **2012**, *42*, 357–364. [[CrossRef](#)]
148. Vieira, A.T.; Macia, L.; Galvão, I.; Martins, F.S.; Canesso, M.C.; Amaral, F.A.; Garcia, C.C.; Maslowski, K.M.; De Leon, E.; Shim, D.; et al. A Role for Gut Microbiota and the Metabolite-Sensing Receptor GPR43 in a Murine Model of Gout. *Arthritis Rheumatol.* **2015**, *67*, 1646–1656. [[CrossRef](#)]
149. Al Mahri, S.; Malik, S.S.; Al Ibrahim, M.; Haji, E.; Dairi, G.; Mohammad, S. Free Fatty Acid Receptors (FFARs) in Adipose: Physiological Role and Therapeutic Outlook. *Cells* **2022**, *11*, 750. [[CrossRef](#)]
150. Al-Lahham, S.H.; Roelofsen, H.; Priebe, M.; Weening, D.; Dijkstra, M.; Hoek, A.; Rezaee, F.; Venema, K.; Vonk, R.J. Regulation of adipokine production in human adipose tissue by propionic acid. *Eur. J. Clin. Investig.* **2010**, *40*, 401–407. [[CrossRef](#)]
151. Samuel, B.S.; Shaito, A.; Motoike, T.; Rey, F.E.; Backhed, F.; Manchester, J.K.; Hammer, R.E.; Williams, S.C.; Crowley, J.; Yanagisawa, M.; et al. Effects of the gut microbiota on host adiposity are modulated by the short-chain fatty-acid binding G protein-coupled receptor, Gpr41. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 16767–16772. [[CrossRef](#)]
152. Priyadarshini, M.; Cole, C.; Oroskar, G.; Ludvik, A.E.; Wicksteed, B.; He, C.; Layden, B.T. Free fatty acid receptor 3 differentially contributes to  $\beta$ -cell compensation under high-fat diet and streptozotocin stress. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2020**, *318*, R691–R700. [[CrossRef](#)]
153. Bellahcene, M.; O'Dowd, J.F.; Wargent, E.T.; Zaibi, M.S.; Hislop, D.C.; Ngala, R.A.; Smith, D.M.; Cawthorne, M.A.; Stocker, C.J.; Arch, J.R. Male mice that lack the G-protein-coupled receptor GPR41 have low energy expenditure and increased body fat content. *Br. J. Nutr.* **2013**, *109*, 1755–1764. [[CrossRef](#)]
154. Tolhurst, G.; Heffron, H.; Lam, Y.S.; Parker, H.E.; Habib, A.M.; Diakogiannaki, E.; Cameron, J.; Grosse, J.; Reimann, F.; Gribble, F.M. Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein-coupled receptor FFAR2. *Diabetes* **2012**, *61*, 364–371. [[CrossRef](#)]
155. Lin, H.V.; Frassetto, A.; Kowalik, E.J., Jr.; Nawrocki, A.R.; Lu, M.M.; Kosinski, J.R.; Hubert, J.A.; Szeto, D.; Yao, X.; Forrest, G.; et al. Butyrate and propionate protect against diet-induced obesity and regulate gut hormones via free fatty acid receptor 3-independent mechanisms. *PLoS ONE* **2012**, *7*, e35240. [[CrossRef](#)]
156. He, J.; Zhang, P.; Shen, L.; Niu, L.; Tan, Y.; Chen, L.; Zhao, Y.; Bai, L.; Hao, X.; Li, X.; et al. Short-Chain Fatty Acids and Their Association with Signalling Pathways in Inflammation, Glucose and Lipid Metabolism. *Int. J. Mol. Sci.* **2020**, *21*, 6356. [[CrossRef](#)]
157. Coppola, S.; Avagliano, C.; Calignano, A.; Berni Canani, R. The Protective Role of Butyrate against Obesity and Obesity-Related Diseases. *Molecules* **2021**, *26*, 682. [[CrossRef](#)]
158. Berni Canani, R.; Di Costanzo, M.; Leone, L. The epigenetic effects of butyrate: Potential therapeutic implications for clinical practice. *Clin. Epigenetics* **2012**, *4*, 4. [[CrossRef](#)]
159. Liu, H.; Wang, J.; He, T.; Becker, S.; Zhang, G.; Li, D.; Ma, X. Butyrate: A Double-Edged Sword for Health? *Adv. Nutr.* **2018**, *9*, 21–29. [[CrossRef](#)]
160. Bultman, S.J. Molecular pathways: Gene-environment interactions regulating dietary fiber induction of proliferation and apoptosis via butyrate for cancer prevention. *Clin. Cancer Res.* **2014**, *20*, 799–803. [[CrossRef](#)]
161. Hong, J.; Jia, Y.; Pan, S.; Jia, L.; Li, H.; Han, Z.; Cai, D.; Zhao, R. Butyrate alleviates high fat diet-induced obesity through activation of adiponectin-mediated pathway and stimulation of mitochondrial function in the skeletal muscle of mice. *Oncotarget* **2016**, *7*, 56071–56082. [[CrossRef](#)] [[PubMed](#)]
162. Khan, S.; Jena, G. Sodium butyrate reduces insulin-resistance, fat accumulation and dyslipidemia in type-2 diabetic rat: A comparative study with metformin. *Chem. Biol. Interact.* **2016**, *254*, 124–134. [[CrossRef](#)] [[PubMed](#)]

163. Zhang, L.; Du, J.; Yano, N.; Wang, H.; Zhao, Y.T.; Dubielecka, P.M.; Zhuang, S.; Chin, Y.E.; Qin, G.; Zhao, T.C. Sodium Butyrate Protects -Against High Fat Diet-Induced Cardiac Dysfunction and Metabolic Disorders in Type II Diabetic Mice. *J. Cell Biochem.* **2017**, *118*, 2395–2408. [[CrossRef](#)] [[PubMed](#)]
164. Li, G.; Yao, W.; Jiang, H. Short-chain fatty acids enhance adipocyte differentiation in the stromal vascular fraction of porcine adipose tissue. *J. Nutr.* **2014**, *144*, 1887–1895. [[CrossRef](#)] [[PubMed](#)]
165. Rumberger, J.M.; Arch, J.R.; Green, A. Butyrate and other short-chain fatty acids increase the rate of lipolysis in 3T3-L1 adipocytes. *PeerJ* **2014**, *2*, e611. [[CrossRef](#)]
166. Olaniyi, K.S.; Owolabi, M.N.; Atuma, C.L.; Agunbiade, T.B.; Alabi, B.Y. Acetate rescues defective brain-adipose metabolic network in obese Wistar rats by modulation of peroxisome proliferator-activated receptor- $\gamma$ . *Sci. Rep.* **2021**, *11*, 18967. [[CrossRef](#)]
167. Ohira, H.; Tsutsui, W.; Mamoto, R.; Yamaguchi, S.; Nishida, M.; Ito, M.; Fujioka, Y. Butyrate attenuates lipolysis in adipocytes co-cultured with macrophages through non-prostaglandin E2-mediated and prostaglandin E2-mediated pathways. *Lipids Health Dis.* **2016**, *15*, 213. [[CrossRef](#)]
168. Eslick, S.; Williams, E.J.; Berthon, B.S.; Wright, T.; Karihaloo, C.; Gately, M.; Wood, L.G. Weight Loss and Short-Chain Fatty Acids Reduce Systemic Inflammation in Monocytes and Adipose Tissue Macrophages from Obese Subjects. *Nutrients* **2022**, *14*, 765. [[CrossRef](#)]
169. Kushwaha, V.; Rai, P.; Varshney, S.; Gupta, S.; Khandelwal, N.; Kumar, D.; Nilkanth Gaikwad, A. Sodium butyrate reduces endoplasmic reticulum stress by modulating CHOP and empowers favorable anti-inflammatory adipose tissue immune-metabolism in HFD fed mice model of obesity. *Food Chem.* **2022**, *4*, 100079. [[CrossRef](#)]
170. Ji, J.; Shu, D.; Zheng, M.; Wang, J.; Luo, C.; Wang, Y.; Guo, F.; Zou, X.; Lv, X.; Li, Y.; et al. Microbial metabolite butyrate facilitates M2 macrophage polarization and function. *Sci. Rep.* **2016**, *6*, 24838. [[CrossRef](#)]
171. Al-Roub, A.; Akhter, N.; Al-Sayyar, A.; Wilson, A.; Thomas, R.; Kochumon, S.; Al-Rashed, F.; Al-Mulla, F.; Sindhu, S.; Ahmad, R. Short Chain Fatty Acid Acetate Increases TNF $\alpha$ -Induced MCP-1 Production in Monocytic Cells via ACSL1/MAPK/NF- $\kappa$ B Axis. *Int. J. Mol. Sci.* **2021**, *22*, 7683. [[CrossRef](#)]
172. Coats, B.R.; Schoenfelt, K.Q.; Barbosa-Lorenzi, V.C.; Peris, E.; Cui, C.; Hoffman, A.; Zhou, G.; Fernandez, S.; Zhai, L.; Hall, B.A.; et al. Metabolically Activated Adipose Tissue Macrophages Perform Detrimental and Beneficial Functions during Diet-Induced Obesity. *Cell Rep.* **2017**, *20*, 3149–3161. [[CrossRef](#)]
173. Hattori, M.; Kondo, T.; Kishi, M.; Yamagami, K. A single oral administration of acetic acid increased energy expenditure in C57BL/6j mice. *Biosci. Biotechnol. Biochem.* **2010**, *74*, 2158–2159. [[CrossRef](#)]
174. Arnoldussen, I.A.C.; Wiesmann, M.; Pelgrim, C.E.; Wielemaker, E.M.; van Duyvenvoorde, W.; Amaral-Santos, P.L.; Verschuren, L.; Keijsers, B.J.F.; Heerschap, A.; Kleemann, R.; et al. Butyrate restores HFD-induced adaptations in brain function and metabolism in mid-adult obese mice. *Int. J. Obes.* **2017**, *41*, 935–944. [[CrossRef](#)]
175. Kondo, T.; Kishi, M.; Fushimi, T.; Ugajin, S.; Kaga, T. Vinegar intake reduces body weight, body fat mass, and serum triglyceride levels in obese Japanese subjects. *Biosci. Biotechnol. Biochem.* **2009**, *73*, 1837–1843. [[CrossRef](#)]
176. Yamashita, H.; Maruta, H.; Jozuka, M.; Kimura, R.; Iwabuchi, H.; Yamato, M.; Saito, T.; Fujisawa, K.; Takahashi, Y.; Kimoto, M.; et al. Effects of acetate on lipid metabolism in muscles and adipose tissues of type 2 diabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. *Biosci. Biotechnol. Biochem.* **2009**, *73*, 570–576. [[CrossRef](#)]
177. den Besten, G.; Bleeker, A.; Gerding, A.; van Eunen, K.; Havinga, R.; van Dijk, T.H.; Oosterveer, M.H.; Jonker, J.W.; Groen, A.K.; Reijngoud, D.J.; et al. Short-chain fatty acids protect against high-fat diet-induced obesity via a ppar $\gamma$ -dependent switch from lipogenesis to fat oxidation. *Diabetes* **2015**, *64*, 2398–2408. [[CrossRef](#)]
178. Shah, S.; Filliri, T.; Pham, T.H.; Thomas, R.; Cheema, S.K. Intraperitoneal Administration of Short-Chain Fatty Acids Improves Lipid Metabolism of Long-Evans Rats in a Sex-Specific Manner. *Nutrients* **2021**, *13*, 892. [[CrossRef](#)]
179. Lim, J.; Henry, C.J.; Haldar, S. Vinegar as a functional ingredient to improve postprandial glycemic control-human intervention findings and molecular mechanisms. *Mol. Nutr. Food Res.* **2016**, *60*, 1837–1849. [[CrossRef](#)]
180. Kimura, I.; Inoue, D.; Maeda, T.; Hara, T.; Ichimura, A.; Miyauchi, S.; Kobayashi, M.; Hirasawa, A.; Tsujimoto, G. Short-chain fatty acids and ketones directly regulate sympathetic nervous system via G protein-coupled receptor 41 (GPR41). *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 8030–8035. [[CrossRef](#)]
181. Sahuri-Arisoylu, M.; Brody, L.P.; Parkinson, J.R.; Parkes, H.; Navaratnam, N.; Miller, A.D.; Thomas, E.L.; Frost, G.; Bell, J.D. Reprogramming of hepatic fat accumulation and ‘browning’ of adipose tissue by the short-chain fatty acid acetate. *Int. J. Obes.* **2016**, *40*, 955–963. [[CrossRef](#)] [[PubMed](#)]
182. Bouter, K.; Bakker, G.J.; Levin, E.; Hartstra, A.V.; Kootte, R.S.; Udayappan, S.D.; Katiraei, S.; Bahler, L.; Gilijamse, P.W.; Tremaroli, V.; et al. Differential metabolic effects of oral butyrate treatment in lean versus metabolic syndrome subjects. *Clin. Transl. Gastroenterol.* **2018**, *9*, e155. [[CrossRef](#)] [[PubMed](#)]
183. van der Beek, C.M.; Canfora, E.E.; Lenaerts, K.; Troost, F.J.; Damink, S.W.M.O.; Holst, J.J.; Masclee, A.A.M.; Dejong, C.H.C.; Blaak, E.E. Distal, not proximal, colonic acetate infusions promote fat oxidation and improve metabolic markers in overweight/obese men. *Clin. Sci.* **2016**, *130*, 2073–2082. [[CrossRef](#)] [[PubMed](#)]
184. Canfora, E.E.; van der Beek, C.M.; Jocken, J.W.E.; Goossens, G.H.; Holst, J.J.; Olde Damink, S.W.M.; Lenaerts, K.; Dejong, C.H.C.; Blaak, E.E. Colonic infusions of short-chain fatty acid mixtures promote energy metabolism in overweight/obese men: A randomized crossover trial. *Sci. Rep.* **2017**, *7*, 1–12. [[CrossRef](#)] [[PubMed](#)]

185. Freeland, K.R.; Wolever, T.M. Acute effects of intravenous and rectal acetate on glucagon-like peptide-1, peptide YY, ghrelin, adiponectin and tumour necrosis factor- $\alpha$ . *Br. J. Nutr.* **2010**, *103*, 460–466. [[CrossRef](#)]
186. Hartstra, A.V.; Schüppel, V.; Imangaliyev, S.; Schranter, A.; Prodan, A.; Collard, D.; Levin, E.; Dallinga-Thie, G.; Ackermans, M.T.; Winkelmeijer, M.; et al. Infusion of donor feces affects the gut-brain axis in humans with metabolic syndrome. *Mol. Metab.* **2020**, *42*, 101076. [[CrossRef](#)]
187. Li, Z.; Yi, C.X.; Katiraei, S.; Kooijman, S.; Zhou, E.; Chung, C.K.; Gao, Y.; van den Heuvel, J.K.; Meijer, O.C.; Berbée, J.F.P.; et al. Butyrate reduces appetite and activates brown adipose tissue via the gut-brain neural circuit. *Gut* **2018**, *67*, 1269–1279. [[CrossRef](#)]
188. Tirosh, A.; Calay, E.S.; Tuncman, G.; Claiborn, K.C.; Inouye, K.E.; Eguchi, K.; Alcalá, M.; Rathaus, M.; Hollander, K.S.; Ron, I.; et al. The short-chain fatty acid propionate increases glucagon and FABP4 production, impairing insulin action in mice and humans. *Sci. Transl. Med.* **2019**, *11*, eaav0120. [[CrossRef](#)]
189. Kong, D.; Schipper, L.; van Dijk, G. Distinct Effects of Short Chain Fatty Acids on Host Energy Balance and Fuel Homeostasis With Focus on Route of Administration and Host Species. *Front. Neurosci.* **2021**, *15*, 755845. [[CrossRef](#)]
190. Shubitowski, T.B.; Poll, B.G.; Natarajan, N.; Pluznick, J.L. Short-chain fatty acid delivery: Assessing exogenous administration of the microbiome metabolite acetate in mice. *Physiol. Rep.* **2019**, *7*, e14005. [[CrossRef](#)]
191. Blaak, E.E.; Canfora, E.E.; Theis, S.; Frost, G.; Groen, A.K.; Mithieux, G.; Nauta, A.; Scott, K.; Stahl, B.; van Harsselaar, J.; et al. Short chain fatty acids in human gut and metabolic health. *Benef. Microbes* **2020**, *11*, 411–455. [[CrossRef](#)]
192. Chambers, E.S.; Viardot, A.; Psichas, A.; Morrison, D.J.; Murphy, K.G.; Zac-Varghese, S.E.; MacDougall, K.; Preston, T.; Tedford, C.; Finlayson, G.S.; et al. Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. *Gut* **2015**, *64*, 1744–1754. [[CrossRef](#)]
193. Moffett, J.R.; Puthillathu, N.; Vengilote, R.; Jaworski, D.M.; Namboodiri, A.M. Acetate Revisited: A Key Biomolecule at the Nexus of Metabolism, Epigenetics, and Oncogenesis—Part 2: Acetate and ACS2 in Health and Disease. *Front. Physiol.* **2020**, *11*, 580171. [[CrossRef](#)]
194. Xiao, H.; Kang, S. The Role of the Gut Microbiome in Energy Balance With a Focus on the Gut-Adipose Tissue Axis. *Front. Genet.* **2020**, *11*, 297. [[CrossRef](#)]
195. Bohan, R.; Tianyu, X.; Tiantian, Z.; Ruonan, F.; Hongtao, H.; Qiong, W.; Chao, S. Gut microbiota: A potential manipulator for host adipose tissue and energy metabolism. *J. Nutr. Biochem.* **2019**, *64*, 206–217. [[CrossRef](#)]
196. Nogal, A.; Valdes, A.M.; Menni, C. The role of short-chain fatty acids in the interplay between gut microbiota and diet in cardio-metabolic health. *Gut Microbes* **2021**, *13*, 1–24. [[CrossRef](#)]
197. Agus, A.; Clément, K.; Sokol, H. Gut microbiota-derived metabolites as central regulators in metabolic disorders. *Gut* **2021**, *70*, 1174–1182. [[CrossRef](#)]
198. Zhu, T.; Goodarzi, M.O. Metabolites Linking the Gut Microbiome with Risk for Type 2 Diabetes. *Curr. Nutr. Rep.* **2020**, *9*, 83–93. [[CrossRef](#)]
199. Wolever, T.M.; Brighenti, F.; Royall, D.; Jenkins, A.L.; Jenkins, D.J. Effect of rectal infusion of short chain fatty acids in human subjects. *Am. J. Gastroenterol.* **1989**, *84*, 1027–1033.
200. Suokas, A.; Kupari, M.; Heikkilä, J.; Lindros, K.; Ylikahri, R. Acute cardiovascular and metabolic effects of acetate in men. *Alcohol. Clin. Exp. Res.* **1988**, *12*, 52–58. [[CrossRef](#)]
201. Müller, M.; Hernández, M.A.G.; Goossens, G.H.; Reijnders, D.; Holst, J.J.; Jocken, J.W.E.; van Eijk, H.; Canfora, E.E.; Blaak, E.E. Circulating but not faecal short-chain fatty acids are related to insulin sensitivity, lipolysis and GLP-1 concentrations in humans. *Sci. Rep.* **2019**, *9*, 1–9. [[CrossRef](#)] [[PubMed](#)]
202. Aberdein, N.; Schweizer, M.; Ball, D. Sodium acetate decreases phosphorylation of hormone sensitive lipase in isoproterenol-stimulated 3T3-L1 mature adipocytes. *Adipocyte* **2014**, *3*, 121–125. [[CrossRef](#)] [[PubMed](#)]
203. Heimann, E.; Nyman, M.; Pålbrink, A.K.; Lindkvist-Petersson, K.; Degerman, E. Branched short-chain fatty acids modulate glucose and lipid metabolism in primary adipocytes. *Adipocyte* **2016**, *5*, 359–368. [[CrossRef](#)] [[PubMed](#)]
204. Smith, G.I.; Jeukendrup, A.E.; Ball, D. Sodium acetate induces a metabolic alkalosis but not the increase in fatty acid oxidation observed following bicarbonate ingestion in humans. *J. Nutr.* **2007**, *137*, 1750–1756. [[CrossRef](#)]
205. Crouse, J.R.; Gerson, C.D.; DeCarli, L.M.; Lieber, C.S. Role of acetate in the reduction of plasma free fatty acids produced by ethanol in man. *J. Lipid Res.* **1968**, *9*, 509–512. [[CrossRef](#)]
206. Morigny, P.; Boucher, J.; Arner, P.; Langin, D. Lipid and glucose metabolism in white adipocytes: Pathways, dysfunction and therapeutics. *Nat. Rev. Endocrinol.* **2021**, *17*, 276–295. [[CrossRef](#)]

**Disclaimer/Publisher’s Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.