

Review

Dietary Fibre for the Prevention of Post-Pancreatitis Diabetes Mellitus: A Review of the Literature and Future Research Directions

Xinye Li and Maxim S. Petrov * 

School of Medicine, University of Auckland, Auckland 1023, New Zealand

* Correspondence: m.petrov@auckland.ac.nz

Abstract: Post-pancreatitis diabetes mellitus—the most common sequela of pancreatitis—leads to poorer glycaemic control compared with type 2 diabetes. Because post-pancreatitis diabetes mellitus is an exemplar of secondary diabetes (with a clear underlying cause), much post-pancreatitis diabetes mellitus is preventable or treatable early. Earlier literature established the important role of dietary fibre in reducing plasma glucose in individuals with type 2 diabetes. The present review benchmarks available evidence on the role of habitual dietary fibre intake in pancreatitis and post-pancreatitis diabetes mellitus. It also paves the way for future research on the use of dietary fibre in the post-pancreatitis setting.

Keywords: dietary fibre; nutrition; acute pancreatitis; post-pancreatitis diabetes mellitus



Citation: Li, X.; Petrov, M.S. Dietary Fibre for the Prevention of Post-Pancreatitis Diabetes Mellitus: A Review of the Literature and Future Research Directions. *Nutrients* **2024**, *16*, 435. <https://doi.org/10.3390/nu16030435>

Academic Editor: Maria Teresa Guagnano

Received: 12 December 2023

Revised: 30 January 2024

Accepted: 30 January 2024

Published: 1 February 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Physicochemical Properties of Dietary Fibre

The concept of dietary fibre was first introduced in the 1950s when it was observed that some aspects of the diet that are related to “fibre” reduce the incidence of toxemia in pregnancy [1]. However, the definition of dietary fibre has been a subject of debate in the ensuing half-century, as it needs to encompass a wide range of dietary fibre types and related components. In 2010, the Codex Alimentarius Commission—an international food standards body established jointly by the World Health Organization and the Food and Agriculture Organization—published a definition of dietary fibre that met with broad international acceptance [2]. It characterises dietary fibre as carbohydrate polymers (that are not hydrolysed in the small intestine of humans) with at least 10 monomeric units [2]. Unlike many other nutrients, dietary fibre exerts its health effects mainly through its structural and chemical properties during digestion in the gastrointestinal tract.

1.1. Particle Size, Porosity, and Hydration Properties

Several physicochemical properties of dietary fibre (e.g., solubility, viscosity, fermentability) are closely linked to the role of dietary fibre in human physiology. The journey of dietary fibre in the human gastrointestinal tract involves several events during which its particle size changes. Before dietary fibre can reach the large intestine for fermentation, it is mechanically broken through chewing in the mouth and grinding in the stomach. In addition, food processing procedures (such as cooking and milling) result in alterations in the particle size and structure of dietary fibre, coupled with the mechanical breakdown in the mouth. This alters the digestibility of fibre and the degradation of other plant compounds [3]. For example, raw starch granules present in the plant cells can become soluble after cooking. Second, the level of porosity and available surface area of dietary fibre affect its ability to bind with other particles in the gut, such as enzymes and bacteria. It also affects the fermentability of dietary fibre by the gut microbiota. Third, modifications to the plant cell-wall structure through industrial processing (such as cooking, drying, heating, and

pH alteration) affect the hydration properties of dietary fibre, including swelling capacity, water-binding capacity, and water absorption [4].

1.2. Solubility and Viscosity

The solubility of dietary fibre is closely related to its role in physiology. In general, fibre that is relatively more soluble in water/solution tends to have a more branched structure (energetically unstable), which means it is less stable in its solid state. However, there are exceptions. For example, β -glucan with $\beta(1,4)$ linkages is insoluble, whereas $\beta(1,3)(1,4)$ linkages make β -glucan soluble [5,6]. The concept of viscosity is linked with the solubility of dietary fibre. Viscosity is defined as resistance to flow and is linked to the capacity of dietary fibre to form a viscous solution in a concentration-dependent manner when dissolved [7]. Solubility is generally positively associated with viscosity. Therefore, soluble dietary fibre is more likely to form a gel-like substance and increase the viscosity of the gut's contents compared with insoluble dietary fibre. The structure of soluble fibre determines how it forms a viscous solution. Soluble fibre with random-coil polysaccharides interacts with water molecules through entanglement, which then increases the viscosity. At the same time, soluble fibre with ordered-assembly polymers forms a gel network when divalent ions are present [8].

1.3. Interplay with Bile and Pancreatic Juice

Dietary fibre may interact with other chemical compounds in the gut, including bile acids and pancreatic enzymes. Primary bile acids are produced in the liver and secreted with bile, which is then converted into secondary bile acids upon bacterial fermentation. Bile acids are required for the digestion and absorption of lipids because they play a key role in the emulsification of lipids and the digestion and absorption of the lipophilic compounds [9]. When bile acids bind with dietary fibre in the gut, up to 95% of the bile acids (both primary and secondary bile acids) are no longer reabsorbed back through the enterohepatic circulation for the synthesis of bile acids. Instead, bile acids are excreted as bile salts in faeces [10]. The activity of pancreatic enzymes is known to be suppressed by several types of dietary fibre, including both soluble (e.g., pectin, guar gum) and insoluble (e.g., cellulose, wheat bran) ones. Pancreatic enzymes are secreted into the gut for the digestion and absorption of macronutrients and micronutrients. Dietary fibre alters the conformation of pancreatic enzymes and competes with nutrients for the binding site, which affects the catalytic activity performed by enzymes [11,12]. Pectin is known to suppress the activity of lipase [13,14]. Guar gum and cellulose are associated with the inactivation of α -amylase [15–17]. An in vitro study by Birkner and Kern found that adding dietary fibre (wheat bran, pectin, and guar gum) to human duodenal juice led to suppression of the activity of bile acids [18]. Similar findings were reported in other in vitro studies where slower nutrient digestion occurred when pancreatic enzymes were inhibited by both insoluble fibre (such as cellulose and rice bran) and soluble fibre (such as pectin) [17,19,20]. However, to date, no human in vivo study investigating the effects of dietary fibre on pancreatic enzymes has been conducted.

2. Physiological Effects of Dietary Fibre

2.1. Effects on Nutrients' Availability

Most nutrients do not exist on their own and are often combined with other nutrients or compounds in the human diet. Specifically, dietary fibre rarely appears on its own in natural food and is often present in the form of a plant cell wall with other nutrients (such as starch, vitamins, and minerals) [21,22]. The primary plant cell wall is the most common form of dietary fibre in the human diet, with a structured network of cellulose and hemicellulose embedded in a network of pectin, while the secondary plant cell wall is the less common form, which is made up of lignin and cellulose [23]. The cell-wall structure affects the bioavailability of other nutrients (such as resistant starch 1) present in plant-based foods (such as vegetables and fruit). When the cell-wall structure remains

intact, intracellular compounds (such as lipids and starch) are not available for absorption. During digestion, a series of events lead to structural changes in dietary fibre. This occurs through the intense mechanical breakdown of the plant cell wall, which allows the soluble components to dissolve or become hydrated. Evidence from studies of various plant-based foods (e.g., carrots, raw fruit, vegetables) showed thickened cell walls, reduced particle size, and solubilisation of pectin during digestion, which lead to a decrease in cell-wall integrity and an increase in cell-wall permeability [24,25]. As a result, nutrients within the plant cells become available for digestion. However, not all nutrients are absorbed before reaching the large intestine. Some of the nutrients may be released when dietary fibre is fermented by the gut microbiota, and these are still available for absorption through the colon's epithelium (e.g., polyphenols).

Dietary fibre can influence the bioavailability of micronutrients—both minerals and vitamins [26]. For example, dietary fibre (e.g., fructan) can enhance the absorption of calcium [27,28]. It is believed that an increase in dietary fibre intake results in a lower pH level in the gut, which increases the solubility of calcium ions [29,30]. The effects of dietary fibre on vitamins remain unclear due to mixed results from previous studies [31–33]. This gap in knowledge is of particular concern in the setting of chronic pancreatitis and its sequelae (such as new-onset diabetes mellitus) because of the accompanying malabsorption, which frequently leads to vitamin D deficiency. It is possible that, in individuals with exocrine pancreatic insufficiency and/or malnutrition, fibre acts as “anti-nutrients”—a concept derived from *in vitro* studies, suggesting that compounds present in plants may have untoward effects on certain biological systems. “Anti-nutrients” that act as pancreatic lipase inhibitors have been reviewed elsewhere [34].

2.2. Effect on Glucose Metabolism

Another physiological effect of dietary fibre is its ability to reduce blood glucose levels. Most studies on the health effects of dietary fibre have focused on its effects on gastric emptying and nutrient absorption [35]. It is believed that the glucose-lowering effect is achieved directly through the gel-forming ability of soluble dietary fibre, which results in the gut's contents becoming more viscous in the stomach and the intestine. In turn, increased viscosity of the gut's contents in the stomach and intestine leads to slower nutrient diffusion, slower gastric emptying, and delayed nutrient absorption in the small intestine [36,37]. As a result, carbohydrates are absorbed into the circulation at a reduced rate, and with less fluctuation in postprandial glycaemia [38]. Insoluble fibre has distinct characteristics, and its effects on glucose metabolism are different from those of soluble fibre. It was initially thought that insoluble fibre may reduce the risk of diabetes through the production of short-chain fatty acids in the colon and their effects on hepatic insulin sensitivity [39]. More recent thinking is that high insoluble fibre intake may improve insulin resistance independent of weight loss, by interfering with the absorption of dietary protein [40]. This is an ongoing area of active research, and future high-quality studies are expected to conclusively establish the mechanism by which insoluble fibre affects blood glucose control.

Studies of healthy individuals have found that intake of dietary fibre is associated with improved glycaemic control. Food and colleagues conducted a randomised controlled trial of 11 healthy individuals, where the participants were given either fibre-enriched bread (flax fibre) or control bread (white bread) to eat after overnight fasting. The authors found that the intake of flax-fibre-enriched bread was associated with a notable reduction in peak postprandial glucose levels when compared with the control group [41]. Similar findings were reported in a crossover randomised controlled trial of 16 healthy men: the intake of breakfast with a high amount of insoluble fibre (33 g) was associated with a significant reduction in glycaemic response after 75 min, compared with the intake of breakfast with a low amount of insoluble fibre (1 g) [42]. A randomised controlled trial by Weickert and colleagues in overweight and obese women showed that the consumption of insoluble-fibre-enriched white bread (compared with regular white bread) for 3 days

significantly increased whole-body insulin sensitivity in overweight and obese women [43]. Meta-analyses showed a significantly reduced risk of type 2 diabetes with higher fibre intake (in particular, cereal fibre intake) [44,45]. The role of dietary fibre in type 2 diabetes and prediabetes has been discussed comprehensively in previous studies and is beyond the scope of the present paper [46,47].

2.3. Effect on Lipid Metabolism

As far as the lipid-modulating effect of dietary fibre is concerned, evidence suggests that the interaction of dietary fibre with bile acids is the main contributor. A randomised controlled trial of a diet characterised by whole grains, legumes, and fruits and vegetables, compared with a diet high in refined grains, found that increased intake of dietary fibre leads to higher levels of circulating bile acids in healthy adults [48]. Since cholesterol is the precursor of bile acids, binding with dietary fibre results in increased excretion of bile acids in the form of bile salts [10,49]. As a result, the liver is required to synthesise bile acids using endogenous cholesterol, reducing serum cholesterol levels. Second, bile acids are essential for the emulsification of lipids and the digestion and absorption of lipophilic compounds. When dietary fibre is present, bile acids are no longer available for lipid emulsification. As a result, absorption of lipids and lipophilic compounds becomes reduced, which leads to reductions in serum triglycerides and total cholesterol [50]. In a 2023 large meta-analysis of randomised controlled trials, soluble fibre supplementation was shown to result in significant reductions in total cholesterol, LDL cholesterol, and apolipoprotein B [51]. The lipid-modulating effect of dietary fibre looks particularly promising in light of the PANDORA (PANcreatic Diseases Originating from intRa-pancreatic fAt) hypothesis, which postulates that excess fat in the pancreas is the main driver of all common diseases of both the endocrine (e.g., diabetes mellitus) and exocrine (e.g., pancreatitis) pancreas [52].

2.4. Effects on Gut Transit Time and Stool Mass

Dietary fibre is well known for its ability to influence colonic bulk (stool mass) and gut transit time. Dietary fibre, especially soluble fibre, can increase the water-binding activity in the colon and decrease the gut transit time. This is largely attributed to soluble fibre with ordered-assembly polymers (e.g., pectin), which allow the fibre to form gel networks in the colon when hydrated. A systematic review of healthy individuals showed that the gut transit time decreased with an increased intake of wheat fibre [53]. In addition, it was demonstrated that the bigger the particles of dietary fibre, the greater the stool mass when compared with refined dietary fibre particles [54]. Evidence suggests that both soluble and insoluble dietary fibre can influence stool mass. Soluble fibre increases the viscosity of the gut's contents and makes them viscoelastic, which is positively associated with increases in stool bulk and mass [55,56]. By contrast, insoluble fibre stimulates mucus secretion and colonic muscular contraction, which contribute to stool mass and stool consistency [57–59].

2.5. Interplay with Gut Microbiota

Over one thousand different bacterial species reside in the human intestine, especially in the colon [60]. There is an intricate relationship between bacterial species, the host, and the diet. Growing evidence suggests that the gut microbiota is vital for normal digestion. Undigested dietary fibre reaches the large intestine and is fermented extensively by the gut microbiota as an essential energy source for the bacterial species [61]. The gut microbiota is known to maintain the gut barrier by directly fighting against pathogenic bacteria or acting through a secondary product such as short-chain fatty acids (SCFAs), which are produced when dietary fibre is fermented by the gut microbiota [62,63]. Butyrate, propionate, and acetate account for 90–95% of all SCFAs that are produced in the gut, and their intraluminal fraction is 15%, 25%, and 60%, respectively [64]. Evidence suggests that dietary fibre is associated with an increased growth of the beneficial bacterial genera, such as *Bifidobacterium* and *Lactobacillus*, which have a common role in improving host immunity and enhancing gut development [65–67].

Earlier studies investigated the use of dietary fibre in the form of supplementation with either prebiotics or synbiotics. Prebiotics are defined as selective types of dietary fibre (e.g., inulin, fructooligosaccharide, and galactooligosaccharide) that provide health benefits to the host by stimulating the growth of bacterial species that help to maintain a healthy gut environment [68]. By contrast, synbiotics refer to mixtures of selective types of dietary fibre and bacterial species that provide health benefits to the host [69]. Similarly, the use of prebiotic or synbiotic supplementation has been shown to stimulate the growth of favourable bacteria such as *Bifidobacterium* and *Lactobacillus* [70]. In addition, a meta-analysis of randomised controlled trials in overweight or obese individuals found that the use of synbiotic supplementation was associated with significantly reduced levels of fasting plasma insulin, while the use of prebiotics was associated with reductions in plasma cholesterol and triglycerides [71]. Similar findings were also observed in another meta-analysis of randomised controlled trials of individuals with type 2 diabetes [72].

One of the potential mechanisms by which prebiotics or synbiotics improve glucose and lipid parameters is through the production of SCFAs. High intake of dietary fibre, especially dietary fibre with high fermentability (e.g., β -glucan, pectin, fructooligosaccharide, inulin), is associated with increased production of SCFAs by the gut microbiota [73–77]. SCFAs exert an important role in the gastrointestinal tract. First, high levels of SCFAs in the large intestine help maintain the gut environment by inhibiting the growth of pathogenic bacterial species (such as *Salmonella* spp. and *Escherichia coli*) by lowering the pH level [78,79]. Second, SCFAs reduce the gut transit time together with dietary fibre through stimulating colonic contractile activity [80]. Third, SCFAs may have an immunomodulatory effect through regulating both the size and the function of the regulatory T-cell pool, which is beneficial for the mucosal immune system [81]. Fourth, SCFAs are associated with better gut barrier integrity through strengthening the tight junctions between intestinal epithelial cells [82]. Lastly, dietary fibre also exerts its glucose-lowering effect through the SCFA pathway through metabolic activities in the liver, including decreasing gluconeogenesis [83,84]. While the above evidence is mainly related to soluble fibre, it is worth noting that insoluble fibre is also involved in the interplay with the gut microbiome [85]. First, insoluble fibre provides a substrate that supports the diversity of the microbiome (generally associated with favourable health outcomes). Second, the fermentation of insoluble fibre can contribute to immune modulation. Third, the fermentation products of insoluble fibre (e.g., butyrate) contribute to the maintenance of the gut barrier. Butyrate serves as the primary energy source for colonocytes and has anti-inflammatory properties [86,87]. It is worth noting that different types of insoluble fibre may have varying effects on the gut microbiome, and the overall impact depends on factors such as the existing composition of the gut microbiome, the individual's habitual diet, and the specific fibre consumed [88].

2.6. Interplay with Gut Hormones

Upon ingestion of food, the gastrointestinal tract secretes several different hormones, such as the incretin hormones and oxyntomodulin [89,90]. These gut hormones are secreted to regulate satiety, carbohydrate absorption, gastric emptying, and glucose metabolism [91,92]. Incretin hormones, such as glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide 1 (GLP-1), are responsible for augmenting the insulin-secretory response initiated by hyperglycaemia [93]. GIP was initially recognised as a gut hormone that inhibits gastric acid secretion (GIP was previously known as gastric-inhibitory polypeptide), but it was found that the main action of GIP is to stimulate insulin secretion [94]. GIP is produced from the enteroendocrine K cells located in the mucosa of the duodenum and upper jejunum, while GLP-1 is produced from the intestinal L cells located throughout the ileum and colon [95]. When nutrients, especially starch and sucrose, reach the gut, where the enteroendocrine K and intestinal L cells are located, GIP and GLP-1 are secreted and bind to their receptors in the cell membranes of pancreatic β -cells, which leads to an increase in insulin secretion [93,96]. The actions of incretin hormones go beyond glu-

cose homeostasis. For example, evidence has suggested their roles in promoting weight loss [97,98], enhancing lipoprotein lipase activity and promoting fat storage in subcutaneous adipose tissue [99,100], delayed gastric emptying (GLP-1 only) [101,102], delayed absorption of nutrients in the intestine [103], slower intestinal transit [104], and limiting bone resorption [105].

Dietary fibre intake has been associated with an early response of incretin hormones. A randomised controlled trial by Weickert and colleagues demonstrated that intake of cereal-fibre-enriched bread (matched portion) by healthy individuals resulted in earlier insulin responses and reduced postprandial glucose responses [106]. The study also found that an increase in dietary fibre intake was associated with an earlier response of GIP [106]. Similarly, a crossover randomised controlled trial by Ames and colleagues found that healthy individuals who consumed tortillas with high soluble β -glucan contents (11.6 g) for at least 1 week had a lower glycaemic response (no significant increase in postprandial blood glucose and less fluctuation in postprandial plasma insulin levels) compared with the group that consumed glucose drinks instead [107]. Furthermore, this study found that the other intervention group, which consumed high-insoluble-fibre tortillas (19.6 g), had an earlier GLP-1 response and higher plasma levels of GLP-1 [107]. These findings suggest that the intake of dietary fibre may exert a glucose-lowering effect through the GIP/GLP-1 pathway (along with other mechanisms).

Oxyntomodulin, another gut hormone, is secreted from the intestinal L cells along with GLP-1. It was first identified and named after its ability to regulate gastric acid secretion, and it was later proven to be involved in glucose metabolism through binding and activating GLP-1 and glucagon receptors [108,109]. Other actions of oxyntomodulin have also been proposed [110]. For example, a study in individuals with obesity showed that oxyntomodulin could lead to weight reduction through suppressing appetite and increasing energy expenditure [111]. While many studies have investigated the relationship between dietary fibre and incretin hormone activities, there is a paucity of studies investigating the relationship between oxyntomodulin and dietary fibre intake. However, a 2021 randomised controlled trial by Di Mauro and colleagues demonstrated that oxyntomodulin may be influenced by dietary fibre intake [112]. Findings from this study showed that, among obese/overweight individuals with type 2 diabetes, both a Mediterranean diet (4.8 g of fibre) and a high-fibre vegetarian diet (23 g of fibre) elicited significant increases in postprandial oxyntomodulin levels [112]. Therefore, it is possible that an increase in dietary fibre intake could stimulate the release of oxyntomodulin postprandially, which may, in turn, contribute to better glycaemic control. More detailed information on the impact of soluble fibre on incretins and other gastrointestinal hormones was comprehensively presented in a recent review [113].

3. Pancreatitis and Post-Pancreatitis Diabetes Mellitus

The pancreas is a dual-functional organ composed of the exocrine and endocrine glands. This gives rise to the key functions of the pancreas, which are the digestion of nutrients through secreting pancreatic enzymes and regulating glucose homeostasis through the actions of pancreatic hormones. Therefore, it is rather intuitive that both the endocrine and exocrine parts of the organ are reciprocally affected by pathological processes in the pancreas [114]. Pancreatic cancer, as the most ominous pancreatic disease, is one of the leading causes of cancer-specific mortality, with a dismal 5-year survival rate [115]. It has been shown that both acute pancreatitis (AP) and chronic pancreatitis (CP) are risk factors for developing pancreatic cancer. Studies have found that individuals with CP are at a 7.9-fold higher risk of developing pancreatic cancer within 5 years of the initial diagnosis of CP [116]. Regarding AP, studies from Denmark and Sweden have demonstrated that individuals with AP are at higher risk of pancreatic cancer [117,118]. A study from Denmark found that, compared with the general population, individuals with AP are at a 2-fold higher risk of developing pancreatic cancer after matching by age and sex [119]. Similarly, a study from Sweden found that individuals with AP have a 2.2-fold

higher risk of pancreatic cancer after adjustment for the recurrence of AP and CP after follow-up for 5 to 10 years [118].

Diabetes of the exocrine pancreas (DEP) is a common type of secondary diabetes [114,119,120]. DEP includes several nosologies, including post-pancreatitis diabetes mellitus (PPDM), pancreatic-cancer-related diabetes, and cystic-fibrosis-related diabetes [121]. PPDM is the most frequent sequela of AP and the most common subtype of DEP [119]. Since AP and CP are the two main types of pancreatitis, PPDM also has two subtypes: post-acute-pancreatitis diabetes mellitus (PPDM-A, the largest contributor to PPDM) and post-chronic-pancreatitis diabetes mellitus (PPDM-C) [120]. In the past, it was thought that the development of diabetes after pancreatitis only occurs after CP, or when pancreatitis results in pancreatic necrosis, which leads to a loss of pancreatic β -cell function [119]. However, it has recently been hypothesised that the natural course of PPDM represents a continuum, which is characterised by a gradual progression from insulin resistance after the first episode of non-necrotising AP to the loss of pancreatic β -cell function due to low-grade inflammation accompanied by end-stage CP [114,119]. It is important to acknowledge that most of the evidence to date has come from cross-sectional studies and, therefore, causality cannot be inferred. In particular, it is possible that individuals with pre-existing non-diabetic insulin resistance have a higher chance of acquiring AP (and maintaining insulin resistance when acquiring PPDM-A). While it remains to be established conclusively that sustained low-grade inflammation after AP induces novel insulin resistance, recent findings from the longitudinal LACERTA cohort study do indicate that fasting insulin is a significant risk factor for new-onset derangements in glucose metabolism following an attack of AP [122]. Other key factors involved in the pathogenesis of PPDM include intra-pancreatic fat deposition, altered lipid metabolism, and dysfunction of the pancreas–gut–brain axis [49,123,124]. Detailed recommendations on the differential diagnosis of PPDM and type 2 diabetes have been published elsewhere [115].

Several high-quality studies have established that the risk of developing PPDM is associated with a history of AP [125–128]. The 2020 prospective longitudinal LACERTA study by Bharmal and colleagues showed that individuals are at a higher risk of developing PPDM after an episode of AP [125]. The authors found that 40% of individuals developed new-onset prediabetes or diabetes within two years of an AP attack [125]. When looking at PPDM-A, a 2014 meta-analysis of 24 prospective clinical studies found that 23% (95% CI: 16 to 31%) of individuals developed PPDM-A after an attack of AP (after excluding individuals with CP, prior history of diabetes or prediabetes, and pancreatic resection) [126]. Similarly, two studies conducted in Taiwanese adults after an attack of AP who were previously normoglycaemic showed that the adjusted risks of new-onset diabetes after pancreatitis were 2.15 and 2.54, respectively, when compared with the general population with no history of diabetes or AP [127,128]. In comparison with individuals with type 2 diabetes, individuals with PPDM have poorer glycaemic control. A cohort study from the UK confirmed that individuals who developed PPDM-A had significantly higher HbA1c levels when compared with individuals with type 2 diabetes at the time of diagnosis (67 mmol/mol vs. 63 mmol/mol, $p = 0.002$), at 1 year (54 mmol/mol vs. 51 mmol/mol, $p < 0.001$), and at 5 years (60 mmol/mol vs. 55 mmol/mol, $p < 0.001$) [129]. The authors also found that, when compared with individuals with type 2 diabetes, individuals with PPDM-A were 6.4 times more likely to use insulin therapy 1 year after the diabetes diagnosis, and 5.2 times more likely 5 years after the diabetes diagnosis, after adjustment for common covariates (i.e., age, sex, ethnicity, alcohol consumption, smoking status, body mass index, and initial haemoglobin level at diagnosis) [129]. The above findings suggest that, even when individuals with PPDM-A started insulin therapy earlier, it did not translate into better glycaemic control when compared with individuals with type 2 diabetes.

The long-term use of metformin has been associated with a notable reduction in the risk of mortality (adjusted HR: 0.5; 95% CI: 0.36 to 0.70) for individuals with PPDM-A [130]. Notably, this beneficial effect of metformin was still 25% more pronounced when compared with individuals with type 2 diabetes (adjusted HR: 0.75; 95% CI: 0.72 to 0.77) [130].

However, long-term use of insulin therapy in PPDM was not as promising as metformin in reducing the mortality rate. Specifically, when compared with individuals with PPDM who had never used insulin, the long-term use of insulin was associated with an increased risk of progression from the first episode of AP to recurrent AP or CP among individuals with PPDM who were insulin-naïve (adjusted HR: 1.56; 95% CI: 1.15 to 2.11), after adjustment for pancreatic-related factors (i.e., aetiology, severity, and time since last AP attack) [131].

When looking at PPDM in the long term, individuals who developed PPDM had a higher risk of health complications when compared with individuals with type 2 diabetes [114,132]. First, individuals with PPDM (80.5 per 1000 person-years) showed a higher all-cause mortality rate, by an excess of 14.8 deaths per 1000 person-years, when compared with individuals with type 2 diabetes (65.5 per 1000 person-years), which was mostly attributed to cardiovascular mortality (mortality rate: 25.2 per 1000 person-years) [132]. Second, PPDM results in a significantly higher cancer mortality rate (excluding pancreatic cancer) than type 2 diabetes [132]. When looking at pancreatic cancer separately, a 2020 population-based study found that individuals with PPDM are at a 6.9-fold higher risk (adjusted HR: 6.94; 95% CI: 4.09 to 11.77) of developing primary pancreatic cancer than individuals with type 2 diabetes alone [133]. Moreover, the study found that individuals with PPDM showed a 2.3-fold higher risk of pancreatic cancer when compared with individuals with diabetes before an attack of pancreatitis, after adjustment for covariates [133]. This finding suggests that the increased risk of pancreatic cancer in individuals with PPDM is not merely due to the impact of pancreatitis as a comorbidity in individuals with type 2 diabetes, but rather that pancreatitis applies an impact beyond being a comorbidity in individuals with PPDM [119]. Overall, to improve the health outcomes of this sequela of AP, it is crucial to intervene at an early stage, with a view to preventing PPDM.

4. Dietary Fibre in Pancreatitis and Post-Pancreatitis Diabetes Mellitus

Studies of individuals with AP have shown that a high-fibre diet is associated with reduced length of hospital stay and decreased risk of complications [134,135]. Although investigations of the effects of dietary fibre in individuals with AP are scarce, evidence from the available studies suggests the potential effect of dietary fibre in improving health outcomes in individuals with AP. In 2007, Karakan and colleagues conducted a double-blind randomised controlled trial with 30 individuals with severe AP [135]. The intervention group that received fibre-enriched enteral nutrition (a total of 1.5 g/100 mL multi-fibre supplement as a prebiotic, including 0.7 g/100 mL soluble fibre and 0.8 g/100 mL insoluble fibre) showed significant reductions in hospital stays and overall complication rate compared with the control group who used the standard enteral nutrition formula. Another randomised controlled trial of 49 individuals with severe AP using fibre-enriched enteral feeding showed similar results [136]. The study found that the intervention group that received an extra 20 g of soluble fibre (polydextrose) in the enteral nutrition formula had significant reductions in feeding intolerance and plasma blood glucose when compared with the control group [136]. This reduction in blood glucose might be related to the improved intestinal gut function, absorption, and the direct glucose-lowering effect of soluble fibre [137]. Other notable studies that employed enteral feeding formulae were the two randomised controlled trials by Olah and colleagues [134,138]. The findings from these two trials showed significant reductions in pancreatic necrosis, organ failure, and systemic inflammatory response syndrome in the intervention groups (which received a fibre-enriched enteral tube feed and 10 g of oat fibre in the 2002 trial, and a 10 g mixture of four bioactive plant fibres— β -glucan, inulin, pectin, and resistant starch—in the 2007 trial). However, these two studies also used probiotics (along with fibre supplementation), making it impossible to determine whether it was dietary fibre that contributed to the improved clinical outcomes [139].

Clinical observations suggest that the use of dietary fibre in individuals with CP is associated with steatorrhoea, bloating, and abdominal distention [140]. It is believed that this is due to the maldigestion and malabsorption of fat in the gut, but also because

dietary fibre suppresses pancreatic enzymatic activity [141]. A rodent study showed that 20% wheat bran supplementation for 2 weeks resulted in significantly elevated levels of pancreatic enzymes (i.e., lipase, amylase, and trypsin) [142]. Unfortunately, to date, there have been no human trials to investigate the effects of dietary fibre on pancreatic enzyme activity in individuals with CP.

Nutritional management of PPDM has only recently started to attract research attention [143]. The first study of habitual dietary fibre intake in individuals after an episode of pancreatitis was conducted by Li and colleagues in 2021 as part of the ANDROMEDA project [144]. This was a cross-sectional study of 108 individuals following an episode of acute pancreatitis. Habitual dietary fibre intake was determined using the EPIC-Norfolk food frequency questionnaire. Multivariable regression analyses were conducted, adjusting for covariates such as age, sex, BMI, energy intake, use of antidiabetic medications, aetiology of acute pancreatitis, recurrence of acute pancreatitis, and the presence of pancreatic necrosis. The study showed that increased habitual intake of dietary fibre was significantly inversely associated with fasting plasma glucose in individuals with PPDM-A. This held true for total fibre, soluble fibre, and insoluble fibre. In particular, every 1% increase in the intake of total fibre, soluble fibre, and insoluble fibre was associated with a 0.15%, 0.13%, and 0.13% decrease in fasting plasma glucose, respectively. In the analysis of common sources of dietary fibre, the authors showed that increased intake of vegetables and nuts (but not of fruit and cereals) was significantly inversely associated with a reduction in fasting plasma glucose [144].

5. Directions for Further Research

Based on the findings presented above, several aspects require further investigations through purposely designed studies in individuals after an attack of AP. First, longitudinal studies and randomised controlled trials are warranted to investigate the causal relationships between dietary fibre and markers of glucose metabolism in the post-pancreatitis setting. Similar to studies in individuals with type 2 diabetes [106,145], randomised controlled trials may consider administering isocaloric bread or liquid with different amounts of dietary fibre to individuals with PPDM-A. In addition, future randomised controlled trials could look at introducing an equal number of different types of fibre (such as pectin versus β -glucan) in isocaloric meals or food to elucidate the effectiveness of each type of fibre in individuals after AP. Also, longitudinal studies could look at the relationship between dietary fibre and postprandial (as opposed to fasting) plasma glucose levels, for more comprehensive investigation in the context of post-pancreatitis glucose derangement.

Second, the use of prebiotics and synbiotics in individuals with metabolic diseases has been shown to be effective in improving hyperglycaemia and dyslipidaemia [146]. However, there are controversies around the dose of prebiotics (e.g., specific dietary fibres such as inulin, fructooligosaccharide, and galactooligosaccharide) when used in combination with bacterial species in synbiotic products. Many trials administered less than 1 g of dietary fibre as the prebiotic component, which is not in line with longitudinal and randomised controlled studies that investigated the sole use of dietary fibre in different disease settings [147–149]. Little is known about the effects of dietary fibre when used in combination with probiotics. Therefore, the effects of dietary fibre should be established by future randomised, double-blind, placebo-controlled trials to confirm the independent role of dietary fibre in regulating glucose homeostasis and other relevant parameters (e.g., intra-pancreatic fat deposition and lipid metabolism) in the post-pancreatitis setting. Future studies could also introduce synbiotics with different doses of dietary fibre to elucidate whether there are dose-dependent effects of synbiotics on the gut microbiota profile and glucose homeostasis.

Third, studies have suggested that dietary fibre is associated with the production of SCFAs, the alteration of the gut microbiota profile, the levels of circulating bile acids, and the earlier stimulation of incretins. However, the potential links between them and the underlying pathophysiology of how these mechanisms result in reductions in plasma

glucose are still poorly understood in the post-pancreatitis setting. Future randomised controlled trials are warranted to investigate the underlying pathophysiological events in post-pancreatitis individuals. For example, randomised controlled trials could administer different amounts of dietary fibre to individuals with PPDM-A and compare the circulating levels of bile acids and incretins in association with changes in fasting plasma glucose and insulin traits. In addition, future longitudinal studies could look at the effects of different levels of habitual dietary fibre intake on the gut microbiota profile (i.e., bacterial species and levels of luminal and circulating SCFAs).

Fourth, numerous studies have demonstrated that PPDM is different from type 2 diabetes in terms of pathology, glycaemic control, and risk of other complications [114,119,120]. It is conceivable that nutrition therapy may have different effectiveness in individuals with different types of diabetes [150–152]. Although previous studies showed that nutritional interventions had little effect in individuals with a longstanding history of type 2 diabetes [153–155], no study to date has investigated this in individuals with PPDM-A. Purposely designed interventional studies are now warranted to compare the effects of dietary fibre intake between these two types of diabetes. For example, randomised controlled trials could recruit individuals with newly diagnosed type 2 diabetes, PPDM-A, diagnosed with type 2 diabetes > 5 years, and diagnosed with PPDM > 5 years, and administer an equal amount of dietary fibre in their meals to investigate the effects on glucose markers and insulin traits.

Last, recent studies have shown that there could be potential interplay between dietary fibre and other nutrients or compounds (e.g., dietary iron, resistant starch, and β -hydroxybutyrate), which may lead to indirect metabolic pathways involved in glucose homeostasis. Indeed, earlier studies demonstrated that dietary fibre could interfere with mineral and metal ions in the gut through its porous surface and affect nutrient availability. Therefore, to comprehensively investigate the effect of dietary fibre in regulating plasma glucose, future studies could introduce nutrients that are known to interplay with dietary fibre to determine the independent effect of dietary fibre on glucose homeostasis in individuals after pancreatitis. Future randomised controlled trials in individuals with PPDM could compare the markers of glucose metabolism between individuals with or without exocrine pancreatic dysfunction when they have the same amount of dietary fibre in each meal.

6. Conclusions

The present review brings to the fore the role of fibre supplementation in pancreatitis and post-pancreatitis settings. The use of fibre-enriched enteral formulae helps preserving the gut barrier and, hence, patients with acute pancreatitis may potentially benefit from fibre supplementation alone (i.e., without concurrent use of probiotics) during hospitalisation. After hospital discharge, increased intake of dietary fibre (specifically, vegetables and nuts) may benefit individuals after an attack of acute pancreatitis, with a view to preventing PPDM. Individuals with advanced chronic pancreatitis, exocrine pancreatic insufficiency, and/or malnutrition may not benefit from fibre supplementation, because of exacerbated malabsorption and steatorrhoea following the use of fibre. As the above inferences are based on a rather limited body of knowledge about individuals with diseases of the pancreas, high-quality clinical research on the use of dietary fibre in pancreatitis and its sequelae is warranted.

Author Contributions: Conceptualisation, M.S.P.; writing—original draft preparation, X.L.; writing—review and editing, M.S.P.; supervision, M.S.P. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Hipsley, E.H. Dietary “fibre” and pregnancy toxemia. *Br. Med. J.* **1953**, *4833*, 420–422. [\[CrossRef\]](#)
2. Joint FAP/WHO Food Standards Programme, Secretariat of the CODEX Alimentarius Commission. In *Guidelines on Nutrition Labeling CAC/GL 2–1985 as Last Amended 2010*; FAO: Rome, Italy, 2010.
3. Carmody, R.N.; Bisanz, J.E.; Bowen, B.P.; Maurice, C.F.; Lyalina, S.; Louie, K.B.; Treen, D.; Chadaideh, K.S.; Maini Rekda, V.; Bess, E.N.; et al. Cooking shapes the structure and function of the gut microbiome. *Nat. Microbiol.* **2019**, *4*, 2052–2063. [\[CrossRef\]](#)
4. Capuano, E. The behavior of dietary fiber in the gastrointestinal tract determines its physiological effect. *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 3543–3564. [\[CrossRef\]](#)
5. Sikora, P.; Tosh, S.M.; Brummer, Y.; Olsson, O. Identification of high β -glucan oat lines and localization and chemical characterization of their seed kernel β -glucans. *Food Chem.* **2013**, *137*, 83–91. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Nasatto, P.L.; Pignon, F.; Silveira, J.L.; Duarte, M.E.; Nosedá, M.D.; Rinaudo, M. Methylcellulose, a cellulose derivative with original physical properties and extended applications. *Polymers* **2015**, *7*, 777–803. [\[CrossRef\]](#)
7. Dikeman, C.L.; Fahey, G.C. Viscosity as related to dietary fiber: A review. *Crit. Rev. Food Sci. Nutr.* **2006**, *46*, 649–663. [\[CrossRef\]](#) [\[PubMed\]](#)
8. Morris, E.R. Assembly and rheology of non-starch polysaccharides. In *Advanced Dietary Fibre Technology*; McCleary, B.V., Prosky, L., Eds.; Blackwell Science Ltd.: Cornwall, UK, 2001; pp. 30–41.
9. Al-Ani, Z.; Ko, J.; Petrov, M.S. Relationship of serum bile acids with fat deposition in the pancreas, liver, and skeletal muscle. *Clin. Exp. Gastroenterol.* **2023**, *16*, 137–146. [\[CrossRef\]](#) [\[PubMed\]](#)
10. Products E Panel on Dietetic, Nutrition and Allergies (NDA). Scientific opinion on the substantiation of a health claim related to barley beta-glucans and lowering of blood cholesterol and reduced risk of (coronary) heart disease pursuant to Article 14 of Regulation (EC) No 1924/2006. *EFSA J.* **2011**, *9*, 2470. [\[CrossRef\]](#)
11. Isaksson, G.; Lundquist, I.; Ihse, I. In vitro inhibition of pancreatic enzyme activities by dietary fiber. *Digestion* **1982**, *24*, 54–59. [\[CrossRef\]](#)
12. Kumar, A.; Chauhan, G.S. Extraction and characterization of pectin from apple pomace and its evaluation as lipase (steapsin) inhibitor. *Carbohydr. Polym.* **2010**, *82*, 454–459. [\[CrossRef\]](#)
13. Chater, P.I.; Wilcox, M.D.; Brownlee, I.A.; Pearson, J.P. Alginate as a protease inhibitor in vitro and in a model gut system; Selective inhibition of pepsin but not trypsin. *Carbohydr. Polym.* **2015**, *131*, 142–151. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Strugala, V.; Kennington, E.J.; Campbell, R.J.; Skjåk-Bræk, G.; Dettmar, P.W. Inhibition of pepsin activity by alginates in vitro and the effect of epimerization. *Int. J. Pharm.* **2005**, *304*, 40–50. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Slaughter, S.L.; Ellis, P.R.; Jackson, E.C.; Butterworth, P.J. The effect of guar galactomannan and water availability during hydrothermal processing on the hydrolysis of starch catalysed by pancreatic α -amylase. *Biochim. Biophys. Acta Gen. Subj.* **2002**, *1571*, 55–63. [\[CrossRef\]](#)
16. Hardacre, A.K.; Yap, S.Y.; Lentle, R.G.; Monro, J.A. The effect of fibre and gelatinised starch type on amylolysis and apparent viscosity during in vitro digestion at a physiological shear rate. *Carbohydr. Polym.* **2015**, *123*, 80–88. [\[CrossRef\]](#)
17. Dhital, S.; Gidley, M.J.; Warren, F.J. Inhibition of α -amylase activity by cellulose: Kinetic analysis and nutritional implications. *Carbohydr. Polym.* **2015**, *123*, 305–312. [\[CrossRef\]](#) [\[PubMed\]](#)
18. Birkner, H.; Kern, F. In vitro adsorption of bile salts to food residues, salicylazosulphapyridine and hemicellulose. *Gastroenterology* **1974**, *67*, 237–244. [\[CrossRef\]](#)
19. Qi, J.; Li, Y.; Yokoyama, W.; Majeed, H.; Masamba, K.G.; Zhong, F.; Ma, J. Cellulosic fraction of rice bran fibre alters the conformation and inhibits the activity of porcine pancreatic lipase. *J. Funct. Foods* **2015**, *19*, 39–48. [\[CrossRef\]](#)
20. Leng-Peschlow, E. Interference of dietary fibres with gastrointestinal enzymes in vitro. *Digestion* **1989**, *44*, 200–210. [\[CrossRef\]](#) [\[PubMed\]](#)
21. Norbitt, C.F.; Kimita, W.; Bharmal, S.H.; Ko, J.; Petrov, M.S. Relationship between habitual intake of vitamins and new-onset prediabetes/diabetes after acute pancreatitis. *Nutrients* **2022**, *14*, 1480. [\[CrossRef\]](#)
22. Norbitt, C.F.; Kimita, W.; Ko, J.; Bharmal, S.H.; Petrov, M.S. Associations of habitual mineral intake with new-onset prediabetes/diabetes after acute pancreatitis. *Nutrients* **2021**, *13*, 3978. [\[CrossRef\]](#)
23. Cosgrove, D.J. Wall structure and wall loosening: A look backwards and forwards. *Plant Physiol.* **2001**, *125*, 131–134. [\[CrossRef\]](#)
24. Mandalari, G.; Faulks, R.M.; Rich, G.T.; Lo Turco, V.; Picout, D.R.; Lo Curto, R.B.; Bisignano, G.; Dugo, P.; Dugo, G.; Waldron, K.W.; et al. Release of protein, lipid, and vitamin E from almond seeds during digestion. *J. Agric. Food Chem.* **2008**, *56*, 3409–3416. [\[CrossRef\]](#)
25. Tydeman, E.A.; Parker, M.L.; Wickham, M.S.; Rich, G.T.; Faulks, R.M.; Gidley, M.J.; Fillery-Travis, A.; Waldron, K.W. Effect of carrot (*Daucus carota*) microstructure on carotene bioaccessibility in the upper gastrointestinal tract. 1. In vitro simulations of carrot digestion. *J. Agric. Food Chem.* **2010**, *58*, 9847–9854. [\[CrossRef\]](#)
26. Gill, S.K.; Rossi, M.; Bajka, B.; Whelan, K. Dietary fibre in gastrointestinal health and disease. *Nat. Rev. Gastroenterol. Hepatol.* **2021**, *18*, 101–116. [\[CrossRef\]](#) [\[PubMed\]](#)
27. Harland, B.F. Dietary fibre and mineral bioavailability. *Nutr. Res. Rev.* **1989**, *2*, 133–147. [\[CrossRef\]](#)

28. Abrams, S.A.; Griffin, I.J.; Hawthorne, K.M. Young adolescents who respond to an inulin-type fructan substantially increase total absorbed calcium and daily calcium accretion to the skeleton. *J. Nutr.* **2007**, *137*, 2524–2526. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Kasper, H.; Rabast, U.; Fassl, H.; Fehle, F. The effect of dietary fiber on the postprandial serum vitamin A concentration in man. *Am. J. Clin. Nutr.* **1979**, *32*, 1847–1849. [\[CrossRef\]](#)
30. Basu, T.K.; Donaldson, D. Intestinal absorption in health and disease: Micronutrients. *Best Pract. Res. Clin. Gastroenterol.* **2003**, *17*, 957–979. [\[CrossRef\]](#)
31. Adams, S.; Sello, C.T.; Qin, G.X.; Che, D.; Han, R. Does dietary fiber affect the levels of nutritional components after feed formulation? *Fibers* **2018**, *6*, 29. [\[CrossRef\]](#)
32. Chan, Y.M.; Aufreiter, S.; O’Keefe, S.J.; O’Connor, D.L. Switching to a fibre-rich and low-fat diet increases colonic folate contents among African Americans. *Appl. Physiol. Nutr. Metab.* **2019**, *44*, 127–132. [\[CrossRef\]](#)
33. Riedl, J.; Linseisen, J.; Hoffmann, J.; Wolfram, G. Some dietary fibers reduce the absorption of carotenoids in women. *J. Nutr.* **1999**, *129*, 2170–2176. [\[CrossRef\]](#)
34. Ribichini, E.; Stigliano, S.; Rossi, S.; Zaccari, P.; Sacchi, M.C.; Bruno, G.; Badiali, D.; Severi, C. Role of fibre in nutritional management of pancreatic diseases. *Nutrients* **2019**, *11*, 2219. [\[CrossRef\]](#)
35. Cummings, J.H.; Stephen, A.M. Carbohydrate terminology and classification. *Eur. J. Clin. Nutr.* **2007**, *61*, S5–S18. [\[CrossRef\]](#)
36. Eastwood, M.A.; Morris, E.R. Physical properties of dietary fiber that influence physiological function: A model for polymers along the gastrointestinal tract. *Am. J. Clin. Nutr.* **1992**, *55*, 436–442. [\[CrossRef\]](#)
37. Wursch, P.; Pi-Sunyer, X. The role of viscous soluble fiber in the metabolic control of diabetes. *Diabetes Care* **1997**, *20*, 1774–1780. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Ko, J.; Kimita, W.; Skudder-Hill, L.; Li, X.; Priya, S.; Bharmal, S.H.; Cho, J.; Petrov, M.S. Dietary carbohydrate intake and insulin traits in individuals after acute pancreatitis: Effect modification by intra-pancreatic fat deposition. *Pancreatology* **2021**, *21*, 353–362. [\[CrossRef\]](#) [\[PubMed\]](#)
39. Salamone, D.; Rivellese, A.A.; Vetrani, C. The relationship between gut microbiota, short-chain fatty acids and type 2 diabetes mellitus: The possible role of dietary fibre. *Acta Diabetol.* **2021**, *58*, 1131–1138. [\[CrossRef\]](#) [\[PubMed\]](#)
40. Weickert, M.O.; Pfeiffer, A.F. Impact of dietary fiber consumption on insulin resistance and the prevention of type 2 diabetes. *J. Nutr.* **2018**, *148*, 7–12. [\[CrossRef\]](#) [\[PubMed\]](#)
41. Food, J.M.; Dahl, W.J.; Lockert, E.A.; Cammer, A.L.; Whiting, S.J. Effects of flax fiber on laxation and glycemic response in healthy volunteers. *J. Med. Food* **2005**, *8*, 508–511.
42. Samra, R.A.; Anderson, G.H. Insoluble cereal fiber reduces appetite and short-term food intake and glycemic response to food consumed 75 min later by healthy men. *Am. J. Clin. Nutr.* **2007**, *86*, 972–979. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Weickert, M.O.; Möhlig, M.; Schöfl, C.; Arafat, A.M.; Otto, B.; Viehoff, H.; Koebnick, C.; Kohl, A.; Spranger, J.; Pfeiffer, A.F. Cereal fiber improves whole-body insulin sensitivity in overweight and obese women. *Diabetes Care* **2006**, *29*, 775–780. [\[CrossRef\]](#)
44. Schulze, M.B.; Schulz, M.; Heidemann, C.; Schienkiewitz, A.; Hoffmann, K.; Boeing, H. Fiber and magnesium intake and incidence of type 2 diabetes: A prospective study and meta-analysis. *Arch. Intern. Med.* **2007**, *167*, 956–965. [\[CrossRef\]](#)
45. InterAct Consortium. Dietary fibre and incidence of type 2 diabetes in eight European countries: The EPIC-InterAct Study and a meta-analysis of prospective studies. *Diabetologia* **2015**, *58*, 1394–1408. [\[CrossRef\]](#)
46. Reynolds, A.; Mann, J.; Cummings, J.; Winter, N.; Mete, E.; Te Morenga, L. Carbohydrate quality and human health: A series of systematic reviews and meta-analyses. *Lancet* **2019**, *393*, 434–445. [\[CrossRef\]](#)
47. Kabisch, S.; Honsek, C.; Kemper, M.; Gerbracht, C.; Arafat, A.M.; Birkenfeld, A.L.; Dambeck, U.; Osterhoff, M.A.; Weickert, M.O.; Pfeiffer, A.F. Dose-dependent effects of insoluble fibre on glucose metabolism: A stratified post hoc analysis of the Optimal Fibre Trial (OptiFiT). *Acta Diabetol.* **2021**, *58*, 1649–1658. [\[CrossRef\]](#)
48. Ginos, B.N.; Navarro, S.L.; Schwarz, Y.; Gu, H.; Wang, D.; Randolph, T.W.; Shojaie, A.; Hullar, M.A.; Lampe, P.D.; Kratz, M.; et al. Circulating bile acids in healthy adults respond differently to a dietary pattern characterized by whole grains, legumes and fruits and vegetables compared to a diet high in refined grains and added sugars: A randomized, controlled, crossover feeding study. *Metabolism* **2018**, *83*, 197–204. [\[CrossRef\]](#)
49. Skudder-Hill, L.; Sequeira-Bisson, I.R.; Ko, J.; Cho, J.; Poppitt, S.D.; Petrov, M.S. Remnant cholesterol, but not low-density lipoprotein cholesterol, is associated with intra-pancreatic fat deposition. *Diabetes Obes. Metab.* **2023**, *25*, 3337–3346. [\[CrossRef\]](#)
50. Skudder-Hill, L.; Coffey, S.; Sequeira-Bisson, I.R.; Ko, J.; Poppitt, S.D.; Petrov, M.S. Comprehensive analysis of dyslipidemia states associated with fat in the pancreas. *Diabetes Metab. Syndr.* **2023**, *17*, 102881. [\[CrossRef\]](#)
51. Ghavami, A.; Ziaei, R.; Talebi, S.; Barghchi, H.; Nattagh-Eshstivani, E.; Moradi, S.; Rahbarinejad, P.; Mohammadi, H.; Ghasemi-Tehrani, H.; Marx, W.; et al. Soluble fiber supplementation and serum lipid profile: A systematic review and dose-response meta-analysis of randomized controlled trials. *Adv. Nutr.* **2023**, *14*, 465–474. [\[CrossRef\]](#)
52. Petrov, M.S. Fatty change of the pancreas: The Pandora’s box of pancreatology. *Lancet Gastroenterol. Hepatol.* **2023**, *8*, 671–682. [\[CrossRef\]](#)
53. de Vries, J.; Miller, P.E.; Verbeke, K. Effects of cereal fiber on bowel function: A systematic review of intervention trials. *World J. Gastroenterol.* **2015**, *21*, 8952–8963. [\[CrossRef\]](#)
54. Brodribb, A.J.M.; Groves, C. Effect of bran particle size on stool weight. *Gut* **1978**, *19*, 60–63. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Suares, N.C.; Ford, A.C. Systematic review: The effects of fibre in the management of chronic idiopathic constipation. *Aliment. Pharmacol. Ther.* **2011**, *33*, 895–901. [\[CrossRef\]](#) [\[PubMed\]](#)

56. Christodoulides, S.; Dimidi, E.; Fragkos, K.C.; Farmer, A.D.; Whelan, K.; Scott, S.M. Systematic review with meta-analysis: Effect of fibre supplementation on chronic idiopathic constipation in adults. *Aliment. Pharmacol. Ther.* **2016**, *44*, 103–116. [\[CrossRef\]](#)
57. Fleury, N.; Lahaye, M. Chemical and physico-chemical characterisation of fibres from *Laminaria digitata* (kombu breton): A physiological approach. *J. Sci. Food Agric.* **1991**, *55*, 389–400. [\[CrossRef\]](#)
58. Tomlin, J.; Read, N.W. Laxative properties of indigestible plastic particles. *Br. Med. J.* **1988**, *6657*, 1175–1176. [\[CrossRef\]](#)
59. Lewis, S.J.; Heaton, K.W. Roughage revisited: The effect on intestinal function of inert plastic particles of different sizes and shape. *Dig. Dis. Sci.* **1999**, *44*, 744–748. [\[CrossRef\]](#)
60. Petrov, M.S. Metabolic trifecta after pancreatitis: Exocrine pancreatic dysfunction, altered gut microbiota, and new-onset diabetes. *Clin. Transl. Gastroenterol.* **2019**, *10*, e00086. [\[CrossRef\]](#)
61. Flint, H.J.; Scott, K.P.; Duncan, S.H.; Louis, P.; Forano, E. Microbial degradation of complex carbohydrates in the gut. *Gut Microbes* **2012**, *3*, 289–306. [\[CrossRef\]](#)
62. Jarchum, I.; Pamer, E.G. Regulation of innate and adaptive immunity by the commensal microbiota. *Curr. Opin. Immunol.* **2011**, *23*, 353–360. [\[CrossRef\]](#)
63. Salonen, A.; de Vos, W.M. Impact of diet on human intestinal microbiota and health. *Annu. Rev. Food Sci. Technol.* **2014**, *5*, 239–262. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Mortensen, P.B.; Clausen, M.R. Short-chain fatty acids in the human colon: Relation to gastrointestinal health and disease. *Scand. J. Gastroenterol.* **1996**, *31*, 132–148. [\[CrossRef\]](#)
65. Broekaert, W.F.; Courtin, C.M.; Verbeke, K.; van de Wiele, T.; Verstraete, W.; Delcour, J.A. Prebiotic and other health-related effects of cereal-derived arabinoxylans, arabinoxylan-oligosaccharides, and xylooligosaccharides. *Crit. Rev. Food Sci. Nutr.* **2011**, *51*, 178–194. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Gong, J.; Yang, C. Advances in the methods for studying gut microbiota and their relevance to the research of dietary fiber functions. *Food Res. Int.* **2012**, *48*, 916–929. [\[CrossRef\]](#)
67. Brownawell, A.M.; Caers, W.; Gibson, G.R.; Kendall, C.W.; Lewis, K.D.; Ringel, Y.; Slavin, J.L. Prebiotics and the health benefits of fiber: Current regulatory status, future research, and goals. *J. Nutr.* **2012**, *142*, 962–974. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Gibson, G.R.; Roberfroid, M.B. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J. Nutr.* **1995**, *125*, 1401–1412. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Swanson, K.S.; Gibson, G.R.; Hutkins, R.; Reimer, R.A.; Reid, G.; Verbeke, K.; Scott, K.P.; Holscher, H.D.; Azad, M.B.; Delzenne, N.M.; et al. The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of synbiotics. *Nat. Rev. Gastroenterol. Hepatol.* **2020**, *17*, 687–701. [\[CrossRef\]](#)
70. Roberfroid, M.B.; Gibson, G.R.; Hoyle, L.; McCartney, A.L.; Rastall, R.; Rowland, I.; Wolvers, D.; Watzl, B.; Szajewska, H.; Stahl, B.; et al. Prebiotic effects: Metabolic and health benefits. *Br. J. Nutr.* **2010**, *104*, 1–63. [\[CrossRef\]](#)
71. Beserra, B.T.; Fernandes, R.; do Rosario, V.A.; Mocellin, M.C.; Kuntz, M.G.; Trindade, E.B. A systematic review and meta-analysis of the prebiotics and synbiotics effects on glycaemia, insulin concentrations and lipid parameters in adult patients with overweight or obesity. *Clin. Nutr.* **2015**, *34*, 845–858. [\[CrossRef\]](#)
72. Bock, P.M.; Telo, G.H.; Ramalho, R.; Sbaraini, M.; Leivas, G.; Martins, A.F.; Schaan, B.D. The effect of probiotics, prebiotics or synbiotics on metabolic outcomes in individuals with diabetes: A systematic review and meta-analysis. *Diabetologia* **2020**, *64*, 26–41. [\[CrossRef\]](#)
73. Cuervo, A.; Salazar, N.; Ruas-Madiedo, P.; Gueimonde, M.; González, S. Fiber from a regular diet is directly associated with fecal short-chain fatty acid concentrations in the elderly. *Nutr. Res.* **2013**, *33*, 811–816. [\[CrossRef\]](#)
74. Titgemeyer, E.C.; Bourquin, L.D.; Fahey, G.C.; Garleb, K.A. Fermentability of various fiber sources by human fecal bacteria in vitro. *Am. J. Clin. Nutr.* **1991**, *53*, 1418–1424. [\[CrossRef\]](#)
75. Mortensen, P.B.; Nordgaard-Andersen, I. The dependence of the in vitro fermentation of dietary fibre to short-chain fatty acids on the contents of soluble non-starch polysaccharides. *Scand. J. Gastroenterol.* **1993**, *28*, 418–422. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Bourquin, L.D.; Titgemeyer, E.C.; Fahey, G.C.; Garleb, K.A. Fermentation of dietary fibre by human colonic bacteria: Disappearance of, short-chain fatty acid production from, and potential water-holding capacity of, various substrates. *Scand. J. Gastroenterol.* **1993**, *28*, 249–255. [\[CrossRef\]](#)
77. Pylkas, A.M.; Juneja, L.R.; Slavin, J.L. Comparison of different fibers for in vitro production of short chain fatty acids by intestinal microflora. *J. Med. Food* **2005**, *8*, 113–116. [\[CrossRef\]](#)
78. Walker, A.W.; Duncan, S.H.; Carol McWilliam Leitch, E.; Child, M.W.; Flint, H.J. pH and peptide supply can radically alter bacterial populations and short-chain fatty acid ratios within microbial communities from the human colon. *Appl. Environ. Microbiol.* **2005**, *71*, 3692–3700. [\[CrossRef\]](#)
79. Duncan, S.H.; Louis, P.; Thomson, J.M.; Flint, H.J. The role of pH in determining the species composition of the human colonic microbiota. *Environ. Microbiol.* **2009**, *11*, 2112–2122. [\[CrossRef\]](#) [\[PubMed\]](#)
80. Soret, R.; Chevalier, J.; De Coppet, P.; Poupeau, G.; Derkinderen, P.; Segain, J.P.; Neunlist, M. Short-chain fatty acids regulate the enteric neurons and control gastrointestinal motility in rats. *Gastroenterology* **2010**, *138*, 1772–1782. [\[CrossRef\]](#)
81. Smith, P.M.; Howitt, M.R.; Panikov, N.; Michaud, M.; Gallini, C.A.; Bohlooly-Y, M.; Glickman, J.N.; Garrett, W.S. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* **2013**, *6145*, 569–573. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Hiippala, K.; Jouhten, H.; Ronkainen, A.; Hartikainen, A.; Kainulainen, V.; Jalanka, J.; Satokari, R. The potential of gut commensals in reinforcing intestinal barrier function and alleviating inflammation. *Nutrients* **2018**, *10*, 988. [\[CrossRef\]](#)

83. Arora, T.; Sharma, R.; Frost, G. Propionate. Anti-obesity and satiety enhancing factor? *Appetite* **2011**, *56*, 511–515. [[CrossRef](#)]
84. Mithieux, G. Metabolic effects of portal vein sensing. *Diabetes Obes. Metab.* **2014**, *16*, 56–60. [[CrossRef](#)]
85. Abreu Y Abreu, A.T.; Milke-García, M.P.; Argüello-Arévalo, G.A.; Calderón-de la Barca, A.M.; Carmona-Sánchez, R.I.; Consuelo-Sánchez, A.; Coss-Adame, E.; García-Cedillo, M.F.; Hernández-Rosiles, V.; Icaza-Chávez, M.E.; et al. Dietary fiber and the microbiota: A narrative review by a group of experts from the Asociación Mexicana de Gastroenterología. *Rev. Gastroenterol. Mex. Engl. Ed.* **2021**, *86*, 287–304. [[CrossRef](#)]
86. Barber, T.M.; Valsamakis, G.; Mastorakos, G.; Hanson, P.; Kyrou, I.; Randeva, H.S.; Weickert, M.O. Dietary influences on the microbiota-gut-brain axis. *Int. J. Mol. Sci.* **2021**, *22*, 3502. [[CrossRef](#)]
87. Holscher, H.D. Dietary fiber and prebiotics and the gastrointestinal microbiota. *Gut Microbes* **2017**, *8*, 172–184. [[CrossRef](#)] [[PubMed](#)]
88. Calatayud, M.; Van den Abbeele, P.; Ghyselinck, J.; Marzorati, M.; Rohs, E.; Birkett, A. Comparative effect of 22 dietary sources of fiber on gut microbiota of healthy humans in vitro. *Front. Nutr.* **2021**, *8*, 700571. [[CrossRef](#)] [[PubMed](#)]
89. Pendharkar, S.A.; Singh, R.G.; Cervantes, A.; DeSouza, S.V.; Bharmal, S.H.; Petrov, M.S. Gut hormone responses to mixed meal test in new-onset prediabetes/diabetes after acute pancreatitis. *Horm. Metab. Res.* **2019**, *51*, 191–199. [[CrossRef](#)]
90. Bharmal, S.H.; Cho, J.; Stuart, C.E.; Alarcon Ramos, G.C.; Ko, J.; Petrov, M.S. Oxyntomodulin may distinguish new-onset diabetes after acute pancreatitis from type 2 diabetes. *Clin. Transl. Gastroenterol.* **2020**, *11*, e00132. [[CrossRef](#)]
91. Singaram, K.; Gold-Smith, F.D.; Petrov, M.S. Motilin: A panoply of communications between the gut, brain, and pancreas. *Expert Rev. Gastroenterol. Hepatol.* **2020**, *14*, 103–111. [[CrossRef](#)] [[PubMed](#)]
92. Charles, S.; Liu, Y.; Kimita, W.; Ko, J.; Bharmal, S.H.; Petrov, M.S. Effect of D-β-hydroxybutyrate-(R)-1,3 butanediol on plasma levels of asprosin and leptin: Results from a randomised controlled trial. *Food Funct.* **2023**, *14*, 759–768.
93. Nauck, M.A.; Meier, J.J. Incretin hormones: Their role in health and disease. *Diabetes Obes. Metab.* **2018**, *20*, 5–21. [[CrossRef](#)]
94. Dupre, J.; Ross, S.A.; Watson, D.; Brown, J.C. Stimulation of insulin secretion by gastric inhibitory polypeptide in man. *J. Clin. Endocrinol. Metab.* **1973**, *37*, 826–828. [[CrossRef](#)] [[PubMed](#)]
95. Eissele, R.; Goke, R.; Willemer, S.; Harthus, H.P.; Vermeer, H.; Arnold, R.; Göke, B. Glucagon-like peptide-1 cells in the gastrointestinal tract and pancreas of rat, pig and man. *Eur. J. Clin. Investig.* **1992**, *22*, 283–291. [[CrossRef](#)]
96. Vilsbøll, T.; Holst, J.J. Incretins, insulin secretion and type 2 diabetes mellitus. *Diabetologia* **2004**, *47*, 357–366. [[CrossRef](#)] [[PubMed](#)]
97. Drucker, D.J.; Nauck, M.A. The incretin system: Glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors in type 2 diabetes. *Lancet* **2006**, *368*, 1696–1705. [[CrossRef](#)]
98. Sánchez-Garrido, M.A.; Brandt, S.J.; Clemmensen, C.; Müller, T.D.; DiMarchi, R.D.; Tschöp, M.H. GLP-1/glucagon receptor co-agonism for treatment of obesity. *Diabetologia* **2017**, *60*, 1851–1861. [[CrossRef](#)]
99. Eckel, R.H.; Fujimoto, W.Y.; Brunzell, J.D. Gastric inhibitory polypeptide enhanced lipoprotein lipase activity in cultured preadipocytes. *Diabetes* **1979**, *28*, 1141–1142. [[CrossRef](#)]
100. Wasada, T.; McCorkle, K.; Harris, V.; Kawai, K.; Howard, B.; Unger, R.H. Effect of gastric inhibitory polypeptide on plasma levels of chylomicron triglycerides in dogs. *J. Clin. Investig.* **1981**, *68*, 1106–1107. [[CrossRef](#)]
101. Nauck, M.A.; Niedereichholz, U.; Ettler, R.; Holst, J.J.; Ørskov, C.; Ritzel, R.; Schmiegel, W.H. Glucagon-like peptide 1 inhibition of gastric emptying outweighs its insulinotropic effects in healthy humans. *Am. J. Physiol. Endocrinol. Metab.* **1997**, *273*, 981–988. [[CrossRef](#)]
102. Deane, A.M.; Nguyen, N.Q.; Stevens, J.E.; Fraser, R.J.L.; Holloway, R.H.; Besanko, L.K.; Burgstad, C.; Jones, K.L.; Chapman, M.J.; Rayner, C.K.; et al. Endogenous glucagon-like peptide-1 slows gastric emptying in healthy subjects, attenuating postprandial glycemia. *J. Clin. Endocrinol. Metab.* **2010**, *95*, 215–221. [[CrossRef](#)]
103. Meier, J.J.; Gethmann, A.; Götze, O.; Gallwitz, B.; Holst, J.J.; Schmidt, W.E.; Nauck, M.A. Glucagon-like peptide 1 abolishes the postprandial rise in triglyceride concentrations and lowers levels of non-esterified fatty acids in humans. *Diabetologia* **2006**, *49*, 452–458. [[CrossRef](#)]
104. Tolessa, T.; Gutniak, M.; Holst, J.J.; Efendic, S.; Hellström, P.M. Glucagon-like peptide-1 retards gastric emptying and small bowel transit in the rat: Effect mediated through central or enteric nervous mechanisms. *Dig. Dis. Sci.* **1998**, *43*, 2284–2290. [[CrossRef](#)]
105. Tsukiyama, K.; Yamada, Y.; Yamada, C.; Harada, N.; Kawasaki, Y.; Ogura, M.; Bessho, K.; Li, M.; Amizuka, N.; Sato, M.; et al. Gastric inhibitory polypeptide as an endogenous factor promoting new bone formation after food ingestion. *Mol. Endocrinol.* **2006**, *20*, 1644–1651. [[CrossRef](#)]
106. Weickert, M.O.; Mohlig, M.; Koebsnick, C.; Holst, J.J.; Namsolleck, P.; Ristow, M.; Osterhoff, M.; Rochlitz, H.; Rudovich, N.; Spranger, J.; et al. Impact of cereal fibre on glucose-regulating factors. *Diabetologia* **2005**, *48*, 2343–2353. [[CrossRef](#)]
107. Ames, N.; Blewett, H.; Storsley, J.; Thandapilly, S.J.; Zahradka, P.; Taylor, C. A double-blind randomised controlled trial testing the effect of a barley product containing varying amounts and types of fibre on the postprandial glucose response of healthy volunteers. *Br. J. Nutr.* **2015**, *113*, 1373–1383. [[CrossRef](#)] [[PubMed](#)]
108. Bataille, D.; Coudray, A.M.; Carlqvist, M.; Rosselin, G.; Mutt, V. Isolation of glucagon-37 (bioactive enteroglucagon/oxyntomodulin) from porcine jejunum-ileum. Isolation of the peptide. *FEBS. Lett.* **1982**, *146*, 73–78. [[CrossRef](#)] [[PubMed](#)]
109. Petrov, M.S. Post-pancreatitis diabetes mellitus: Investigational drugs in preclinical and clinical development and therapeutic implications. *Expert Opin. Investig. Drugs* **2021**, *30*, 737–747. [[CrossRef](#)] [[PubMed](#)]
110. Goodarzi, M.O.; Petrov, M.S. Diabetes of the exocrine pancreas: Implications for pharmacological management. *Drugs* **2023**, *83*, 1077–1090. [[CrossRef](#)]

111. Pocai, A. Unraveling oxyntomodulin, GLP1's enigmatic brother. *J. Endocrinol.* **2012**, *215*, 335–346. [[CrossRef](#)]
112. Di Mauro, A.; Tuccinardi, D.; Watanabe, M.; Del Toro, R.; Monte, L.; Giorgino, R.; Rampa, L.; Rossini, G.; Kyanvash, S.; Soare, A.; et al. The Mediterranean diet increases glucagon-like peptide 1 and oxyntomodulin compared with a vegetarian diet in patients with type 2 diabetes: A randomized controlled cross-over trial. *Diabetes Metab. Res. Rev.* **2021**, *37*, e3406. [[CrossRef](#)] [[PubMed](#)]
113. Kabisch, S.; Weickert, M.O.; Pfeiffer, A.F. The role of cereal soluble fiber in the beneficial modulation of glycometabolic gastrointestinal hormones. *Crit. Rev. Food Sci. Nutr.* **2022**, *epub ahead of print*. [[CrossRef](#)]
114. Petrov, M.S.; Olesen, S.S. Metabolic sequelae—The pancreatitis zeitgeist of the 21st century. *Gastroenterology* **2023**, *165*, 1122–1135. [[CrossRef](#)]
115. Koulouris, A.I.; Luben, R.; Banim, P.; Hart, A.R. Dietary fiber and the risk of pancreatic cancer. *Pancreas* **2019**, *48*, 121–125. [[CrossRef](#)]
116. Kirkegård, J.; Mortensen, F.V.; Cronin-Fenton, D. Chronic pancreatitis and pancreatic cancer risk: A systematic review and meta-analysis. *Am. J. Gastroenterol.* **2017**, *112*, 1366–1372. [[CrossRef](#)]
117. Kirkegård, J.; Cronin-Fenton, D.; Heide-Jørgensen, U.; Mortensen, F.V. Acute pancreatitis and pancreatic cancer risk: A nationwide matched-cohort study in Denmark. *Gastroenterology* **2018**, *154*, 1729–1736. [[CrossRef](#)]
118. Sadr-Azodi, O.; Oskarsson, V.; Discacciati, A.; Videhult, P.; Askling, J.; Ekblom, A. Pancreatic cancer following acute pancreatitis: A population-based matched cohort study. *Am. J. Gastroenterol.* **2018**, *113*, 1711–1719. [[CrossRef](#)]
119. Petrov, M.S. Post-pancreatitis diabetes mellitus: Prime time for secondary disease. *Eur. J. Endocrinol.* **2021**, *184*, R137–R149. [[CrossRef](#)] [[PubMed](#)]
120. Petrov, M.S.; Yadav, D. Global epidemiology and holistic prevention of pancreatitis. *Nat. Rev. Gastroenterol. Hepatol.* **2019**, *16*, 175–184. [[CrossRef](#)] [[PubMed](#)]
121. Petrov, M.S.; Basina, M. Diagnosing and classifying diabetes in diseases of the exocrine pancreas. *Eur. J. Endocrinol.* **2021**, *184*, R151–R163. [[CrossRef](#)] [[PubMed](#)]
122. Bharmal, S.H.; Kimita, W.; Ko, J.; Petrov, M.S. Pancreatic and gut hormones as predictors of new-onset prediabetes after non-necrotising acute pancreatitis: A prospective longitudinal cohort study. *Endocr. Connect.* **2021**, *10*, 715–724. [[CrossRef](#)] [[PubMed](#)]
123. Petrov, M.S. Panorama of mediators in postpancreatitis diabetes mellitus. *Curr. Opin. Gastroenterol.* **2020**, *36*, 443–451. [[CrossRef](#)]
124. Petrov, M.S.; Taylor, R. Intra-pancreatic fat deposition: Bringing hidden fat to the fore. *Nat. Rev. Gastroenterol. Hepatol.* **2022**, *19*, 153–168. [[CrossRef](#)]
125. Bharmal, S.H.; Cho, J.; Alarcon Ramos, G.C.; Ko, J.; Stuart, C.E.; Modesto, A.E.; Singh, R.G.; Petrov, M.S. Trajectories of glycaemia following acute pancreatitis: A prospective longitudinal cohort study with 24 months follow-up. *J. Gastroenterol.* **2020**, *55*, 775–788. [[CrossRef](#)] [[PubMed](#)]
126. Das, S.L.; Singh, P.P.; Phillips, A.R.; Murphy, R.; Windsor, J.A.; Petrov, M.S. Newly diagnosed diabetes mellitus after acute pancreatitis: A systematic review and meta-analysis. *Gut* **2014**, *63*, 818–831. [[CrossRef](#)]
127. Lee, Y.K.; Huang, M.Y.; Hsu, C.Y.; Su, Y.C. Bidirectional relationship between diabetes and acute pancreatitis: A population-based cohort study in Taiwan. *Medicine* **2016**, *95*, e2448. [[CrossRef](#)] [[PubMed](#)]
128. Shen, H.N.; Yang, C.C.; Chang, Y.H.; Lu, C.L.; Li, C.Y. Risk of diabetes mellitus after first-attack acute pancreatitis: A national population-based study. *Am. J. Gastroenterol.* **2015**, *110*, 1698–1706. [[CrossRef](#)]
129. Woodmansey, C.; McGovern, A.P.; McCullough, K.A.; Whyte, M.B.; Munro, N.M.; Correa, A.C.; Gatenby, P.A.; Jones, S.A.; de Lusignan, S. Incidence, demographics, and clinical characteristics of diabetes of the exocrine pancreas (type 3c): A retrospective cohort study. *Diabetes Care* **2017**, *40*, 1486–1493. [[CrossRef](#)] [[PubMed](#)]
130. Cho, J.; Scragg, R.; Pandol, S.J.; Goodarzi, M.O.; Petrov, M.S. Antidiabetic medications and mortality risk in individuals with pancreatic cancer-related diabetes and postpancreatitis diabetes: A nationwide cohort study. *Diabetes Care* **2019**, *42*, 1675–1683. [[CrossRef](#)]
131. Cho, J.; Scragg, R.; Petrov, M.S. Use of insulin and the risk of progression of pancreatitis: A population-based cohort study. *Clin. Pharmacol. Ther.* **2020**, *107*, 580–587. [[CrossRef](#)]
132. Cho, J.; Scragg, R.; Petrov, M.S. Risk of mortality and hospitalization after post-pancreatitis diabetes mellitus vs. type 2 diabetes mellitus: A population-based matched cohort study. *Am. J. Gastroenterol.* **2019**, *114*, 804–812. [[CrossRef](#)]
133. Cho, J.; Scragg, R.; Petrov, M.S. Postpancreatitis diabetes confers higher risk for pancreatic cancer than type 2 diabetes: Results from a nationwide cancer registry. *Diabetes Care* **2020**, *43*, 2106–2112. [[CrossRef](#)]
134. Oláh, A.; Belágyi, T.; Pótó, L.; Romics, L.; Bengmark, S. Synbiotic control of inflammation and infection in severe acute pancreatitis: A prospective, randomized, double blind study. *Hepatogastroenterology* **2007**, *54*, 590–594.
135. Karakan, T.; Ergun, M.; Dogan, I.; Cindoruk, M.; Unal, S. Comparison of early enteral nutrition in severe acute pancreatitis with prebiotic fiber supplementation versus standard enteral solution: A prospective randomized double-blind study. *World J. Gastroenterol.* **2007**, *13*, 2733–2737. [[CrossRef](#)] [[PubMed](#)]
136. Chen, T.; Ma, Y.; Xu, L.; Sun, C.; Xu, H.; Zhu, J. Soluble dietary fiber reduces feeding intolerance in severe acute pancreatitis: A randomized study. *J. Parenter. Enter. Nutr.* **2021**, *45*, 125–135. [[CrossRef](#)] [[PubMed](#)]
137. Verspreet, J.; Damen, B.; Broekaert, W.F.; Verbeke, K.; Delcour, J.A.; Courtin, C.M. A critical look at prebiotics within the dietary fiber concept. *Annu. Rev. Food Sci. Technol.* **2016**, *7*, 167–190. [[CrossRef](#)]

138. Oláh, A.; Belágyi, T.; Issekutz, Á.; Gamal, M.E.; Bengmark, S. Randomized clinical trial of specific lactobacillus and fibre supplement to early enteral nutrition in patients with acute pancreatitis. *Br. J. Surg.* **2002**, *89*, 1103–1107. [[CrossRef](#)]
139. Petrov, M.S.; Atduev, V.A.; Zagainov, V.E. Advanced enteral therapy in acute pancreatitis: Is there a room for immunonutrition? A meta-analysis. *Int. J. Surg.* **2008**, *6*, 119–124. [[CrossRef](#)] [[PubMed](#)]
140. Rasmussen, H.H.; Irtun, Ø.; Olesen, S.S.; Drewes, A.M.; Holst, M. Nutrition in chronic pancreatitis. *World J. Gastroenterol.* **2013**, *19*, 7267–7275. [[CrossRef](#)]
141. O'Brien, S.J.; Omer, E. Chronic pancreatitis and nutrition therapy. *Nutr. Clin. Pract.* **2019**, *34*, S13–S26. [[CrossRef](#)]
142. Schneeman, B.O.; Richter, B.D.; Jacobs, L.R. Response to dietary wheat bran in the exocrine pancreas and intestine of rats. *J. Nutr.* **1982**, *112*, 283–286. [[CrossRef](#)]
143. Singh, A.; Aggarwal, M.; Garg, R.; Stevens, T.; Chahal, P. Post-pancreatitis diabetes mellitus: Insight on optimal management with nutrition and lifestyle approaches. *Ann. Med.* **2022**, *54*, 1776–1786. [[CrossRef](#)]
144. Li, X.; Kimita, W.; Cho, J.; Ko, J.; Bharmal, S.H.; Petrov, M.S. Dietary fibre intake in type 2 and new-onset prediabetes/diabetes after acute pancreatitis: A nested cross-sectional study. *Nutrients* **2021**, *13*, 1112. [[CrossRef](#)]
145. Yu, K.; Ke, M.Y.; Li, W.H.; Zhang, S.Q.; Fang, X.C. The impact of soluble dietary fibre on gastric emptying, postprandial blood glucose and insulin in patients with type 2 diabetes. *Asia Pac. J. Clin. Nutr.* **2014**, *23*, 210–218.
146. Xavier-Santos, D.; Bedani, R.; Dorea Lima, E.; Isay Saad, S.M. Impact of probiotics and prebiotics targeting metabolic syndrome. *J. Funct. Foods* **2020**, *64*, 103666. [[CrossRef](#)]
147. Eslamparast, T.; Zamani, F.; Hekmatdoost, A.; Sharafkhah, M.; Eghtesad, S.; Malekzadeh, R.; Poustchi, H. Effects of synbiotic supplementation on insulin resistance in subjects with the metabolic syndrome: A randomised, double-blind, placebo-controlled pilot study. *Br. J. Nutr.* **2014**, *112*, 438–445. [[CrossRef](#)]
148. Asemi, Z.; Khorrami-Rad, A.; Alizadeh, S.A.; Shakeri, H.; Esmailzadeh, A. Effects of synbiotic food consumption on metabolic status of diabetic patients: A double-blind randomized cross-over controlled clinical trial. *Clin. Nutr.* **2014**, *33*, 198–203. [[CrossRef](#)]
149. Shakeri, H.; Hadaegh, H.; Abedi, F.; Tajabadi-Ebrahimi, M.; Mazroei, N.; Ghandi, Y.; Asemi, Z. Consumption of synbiotic bread decreases triacylglycerol and VLDL levels while increasing HDL levels in serum from patients with type-2 diabetes. *Lipids* **2014**, *49*, 695–701. [[CrossRef](#)]
150. Levy, J.; Atkinson, A.B.; Bell, P.M.; McCance, D.R.; Hadden, D.R. Beta-cell deterioration determines the onset and rate of progression of secondary dietary failure in type 2 diabetes mellitus: The 10-year follow-up of the belfast diet study. *Diabet. Med.* **1998**, *15*, 290–296. [[CrossRef](#)]
151. Kahn, S.E. The importance of β -cell failure in the development and progression of type 2 diabetes. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 4047–4058.
152. DeFronzo, R.A.; Bonadonna, R.C.; Ferrannini, E. Pathogenesis of NIDDM: A balanced overview. *Diabetes Care* **1992**, *15*, 318–368. [[CrossRef](#)]
153. Robertson, R.P.; Olson, L.K.; Zhang, H.J. Differentiating glucose toxicity from glucose desensitization: A new message from the insulin gene. *Diabetes* **1994**, *43*, 1085–1089. [[CrossRef](#)]
154. Yki-Jarvinen, H. Glucose toxicity. *Endocr. Rev.* **1992**, *13*, 415–431.
155. UK Prospective Diabetes Study Group. U.K. Prospective diabetes study 16, Overview of 6 years' therapy of type II diabetes: A progressive disease. *Diabetes* **1995**, *44*, 1249–1258. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.