

Supplementary Materials

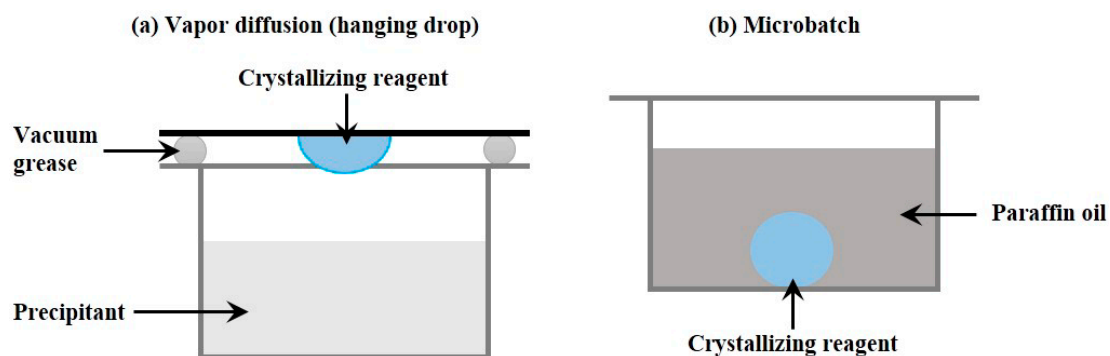


Figure S1. Schematic illustration of the traditional setups of (a) the vapor diffusion (hanging drop) method and (b) the microbatch method.

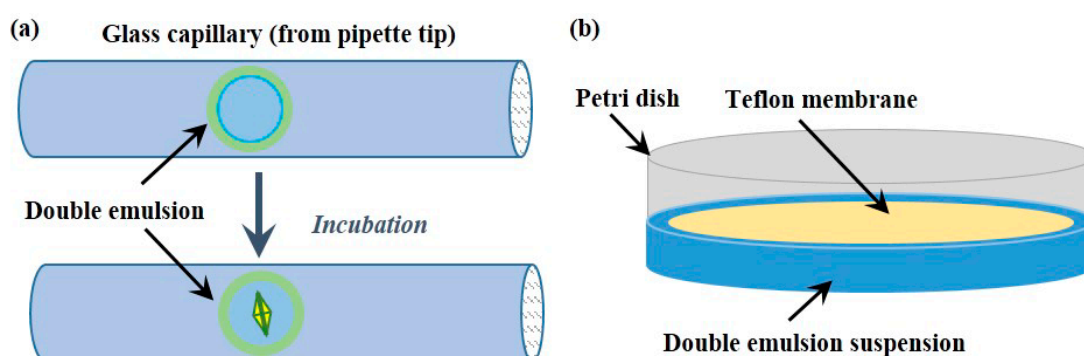


Figure S2. Schematic illustration of storing the double emulsions in (a) glass capillaries and under (b) a Teflon membrane.

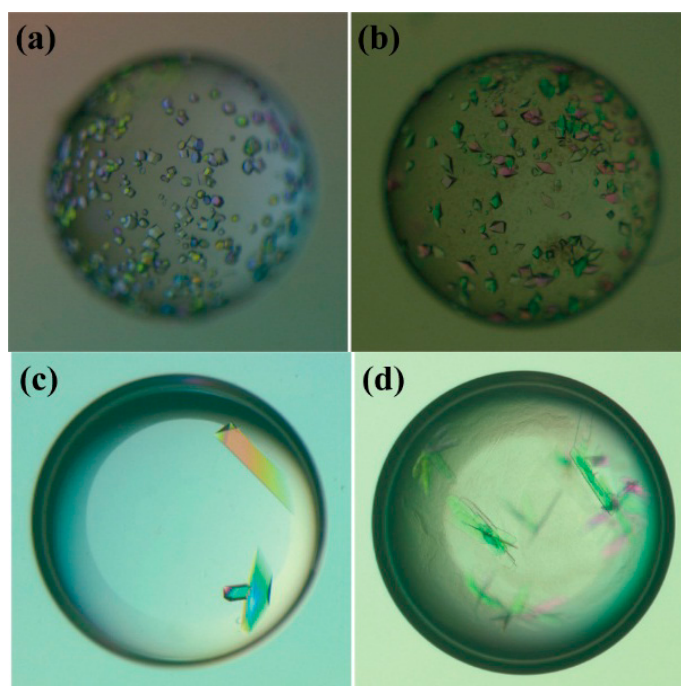


Figure S3. Crystallization results in droplets using the conventional microbatch method for (a) lysozyme, (b) thaumatin, (c) trypsin and (d) horseradish peroxidase. The total volume of one droplet is 1 μL of protein, and precipitant solutions are mixed at an equal volume ratio.

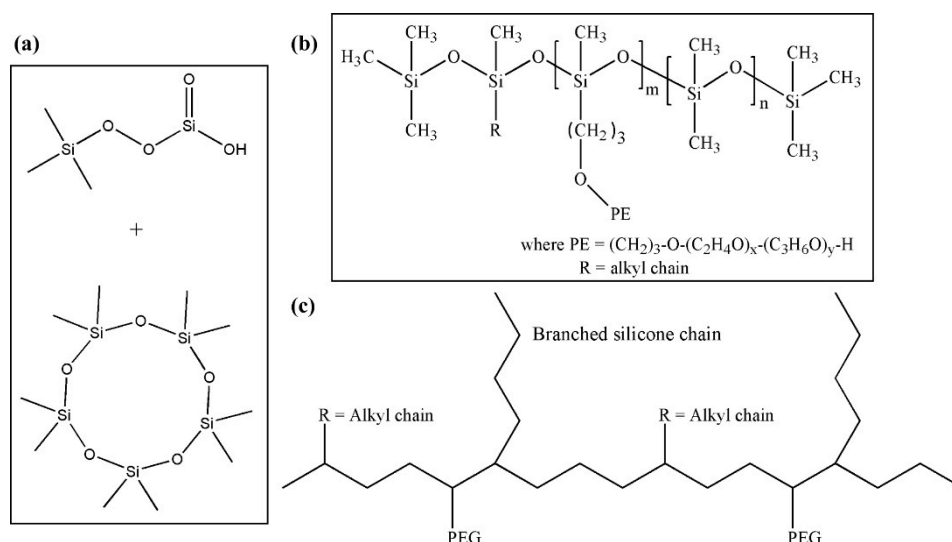


Figure S4. Molecular structures of: (a) 749 Fluid; (b) ABIL EM 90; and (c) KF-6038.

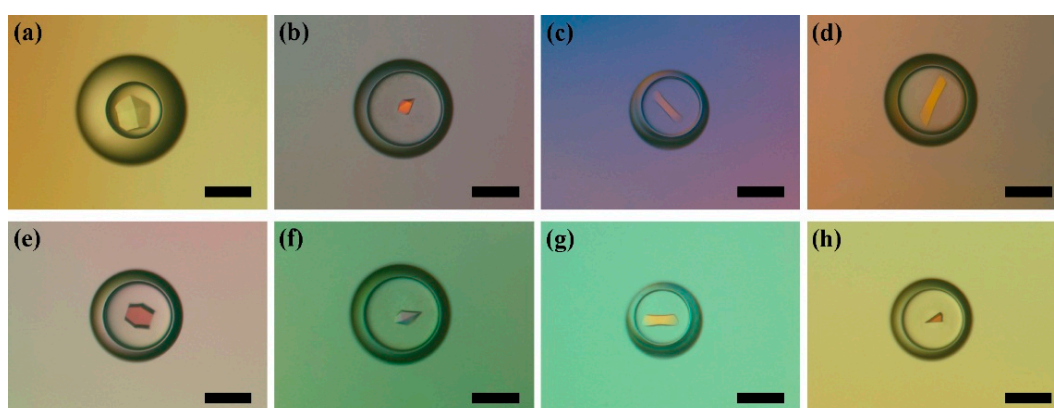


Figure S5. Crystallization results in double emulsion with KF-6038 as the surfactant using the vapor diffusion method (a–d) and the microbatch method (e–h) for (a,e) lysozyme, (b,f) thaumatin, (c,g) trypsin and (d,h) horseradish peroxidase. Scale bars: 300 μm .

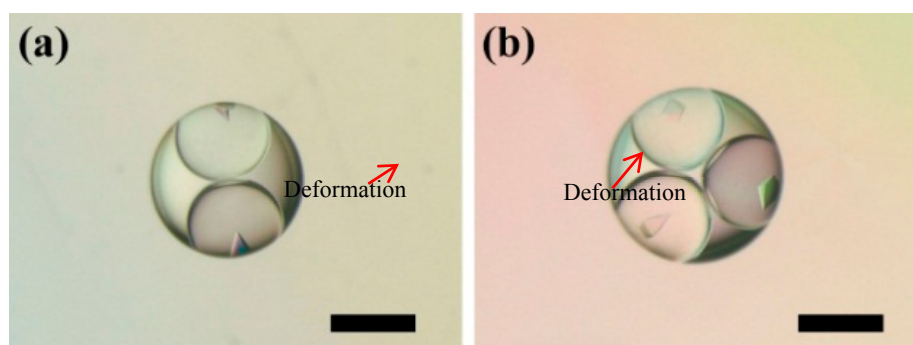


Figure S6. The effect of the non-homogeneous interface on protein crystallization in multiple emulsions containing (a) two and (b) three inner aqueous droplets, showing the crystals tend to occur in the deformed areas. The model protein: 20 mg/mL thaumatin. Scale bars: 500 μm .