Supplementary files



Supplementary Figure S1: Microtubule targeting drugs (MTDs) influence AP-1 activity and AP-1 DNA-binding activity. (a) AP-1 luciferase reporter gene assays with a stable Mel Ju AP-1 LUC cell clone and the control clone (stable Mel Ju pGL2) after incubation with paclitaxel (PX; 10 μ M) or nocodazole (NX; 30 μ M). Bars show the means ± s.d. of three independent experiments (*:P<0.05 compared to ctrl). (b) EMSA with nuclear extracts of melanoma cells (Mel Juso) treated with DMSO, PX (5 μ M) or NX (30 μ M) using the classical AP-1 consensus sequence (AP-1 Oligo). Supershift experiments with an anti-c-Jun antibody show the direct involvement of c-JUN in the AP-1-DNA-binding complex. The experiment was repeated three times.



Supplementary Figure S2: Co-sedimentation by ultra-centrifugal spin-down assays show no binding between c-Jun and polymerized microtubules. Western blot analysis of the supernatant (S) and cell pellet (P) of Mel Im protein lysates after microtubule spin-down assay. In the microtubule-positive samples (+MT), c-Jun was not detectable in the pellet, and direct interactions between c-Jun and polymerized microtubules can therefore be excluded. The microtubule-negative samples (-MT) served as a negative control. CYLD served as a positive control for microtubule interaction[26]. The experiment was repeated three times.



Supplementary Figure S3: Determination of the input protein amounts used for immunoprecipitation (experiments shown in Fig. 2a, 2b, 3d-h). Western blot analyses show the Input protein amounts for immunoprecipitations (IP) of (a) the melanoma cell lines Mel Juso and Mel Ju and (b) of Mel Juso Input protein amounts after sictrl, siIPO13 or siTub1A transfection. (c) Western blot analyses of TUB1A in nuclear extracts and cytoplasmic fractions of Mel Juso and Mel Ju cells, respectively. IkappaB was used as a loading control.



Supplementary Figure S4: Determination of TUB1A-suppression efficiency by qRT-PCR and Western blot analysis. (a) Quantification of the mRNA expression of TUB1A in Mel Juso cells after 96 h of TUB1A si- RNA (siTub1A) and control si-RNA (sictrl) transfection by qRT-PCR. Decreased levels of TUB1A mRNA after siTub1A transfection could be detected compared to the control transfected cells. Bars show the means \pm s.d. of three independent experiments; (*: P<0.05 compared to sictrl). (b) Western blot analysis showing protein expression of TUB1A in Mel Juso cell lysates after siTub1A transfection. Reduced amounts of TUB1A protein were observed after siTub1A transfection compared to control-transfected cells. β -Actin was used as a loading control. The measurements were performed in replicates and each experiment was repeated at least three times.



Supplementary Figure S5: α -Tubulin knockdown resulted in a reduced AP-1 activity in a stable AP-1 LUC clone. AP-1 luciferase reporter gene assays after transfection with α -Tubulin siRNA (siTub1A) compared to control transfected cells (sictrl) of stable transfected Mel Ju cells (Mel Ju (pGL2/AP-1) LUC cell clones) with AP-1 LUC constructs. Bar graph shows the mean± s.d. of three independent experiments; (*: P<0.05 compared to sictrl).



Oligo c-Jun AK TUB1A AK

Supplementary Figure S6: c-Jun, but not TUB1A, is involved in the AP-1-DNA binding complex. EMSA with nuclear extracts of Mel Juso cells using the classical AP-1 consensus sequence (Oligo). Supershift experiments with an anti-c-Jun antibody and an anti-TUB1A antibody show the direct involvement of c-Jun in the AP-1-DNA-binding complex, but no involvement of TUB1A. The experiment was repeated three times.



Supplementary Figure S7: Determination of Importin si-RNA transfection efficiency by qRT-PCR and Western blot analysis. (**a**) Quantification of the expression of Importin mRNA (IPO7, IPO8, IPO9, IPO13, IPOβ) in Mel Juso cells after 96 h of Importin si-RNA and control si-RNA (sictrl) transfection by qRT-PCR. The levels of all Importins decreased in the siRNA-transfected cells compared to controltransfected cells. Bars show the means ± s.d. of three independent experiments; (*: P<0.05 compared to sictrl). (**b**) Western blot analysis showing the expression of IPO13 in Mel Juso cell lysates after IPO13

si-RNA transfection. Reduced amounts of IPO13 protein were observed after IPO13 si-RNA transfection compared to control transfected cells. β -Actin was used as a loading control. The measurements were performed in replicates and each experiment was repeated at least three times.



Supplementary Figure S8: Control immunoprecipitations (IP). (a) Western Blot analysis after coimmunoprecipitation with an anti-CYLD or anti-TUB1A antibody, respectively, showed an interaction between CYLD protein and TUB1A[26]. (b) Co-immunoprecipitation with an anti-IgG antibody served as a negative control for all performed co-immunoprecipitations. Western Blot analysis after co-immunoprecipitation with an anti-IgG antibody showed no detectable protein amount of TUB1A, Actin, c-Jun, CYLD, IPO13 or HA-tagged c-Jun, respectively. Each experiment was repeated at least three times.

Full unedited gel for Figure 1b:



AP-1 Oligo DMSO paclitaxel nocodazole DMSO /c-JUN AK

Mel Ju

Full unedited gel for Figure S1b:



Mel Juso



Full unedited gel for Figure 1c:



Full unedited gel for Figure S3:

e . COIP - Input IP Input Mel Juso Mel Ju Mel Juso Mel Ju Fig. S3a α-Tubulin Imp-E tubulin E JUN actin-Fig. S3a c-Jun Fig. S3a β-Actin Ju uso Juso Ju



Full unedited gel for Figure 2d:



Full unedited gel for Figure 2e:



Full unedited gel for Figure 2f:





Full unedited gel for Figure 3b; 3c:

+sictrl.



Full unedited gel for Figure 3 d, e, f:



Full unedited gel for Figure 3 d, f:





Full unedited gel for Figure 3h:









Full unedited gel for Supplement Figure S2:





S P (+)microtubule Full unedited gel for Figure Supplement Figure S4 a,b:



Mel Juso

Full unedited gel for Figure Supplement Figure S7:



Full unedited gel for Figure Supplement Figure S8a,b:









IP: Actin

