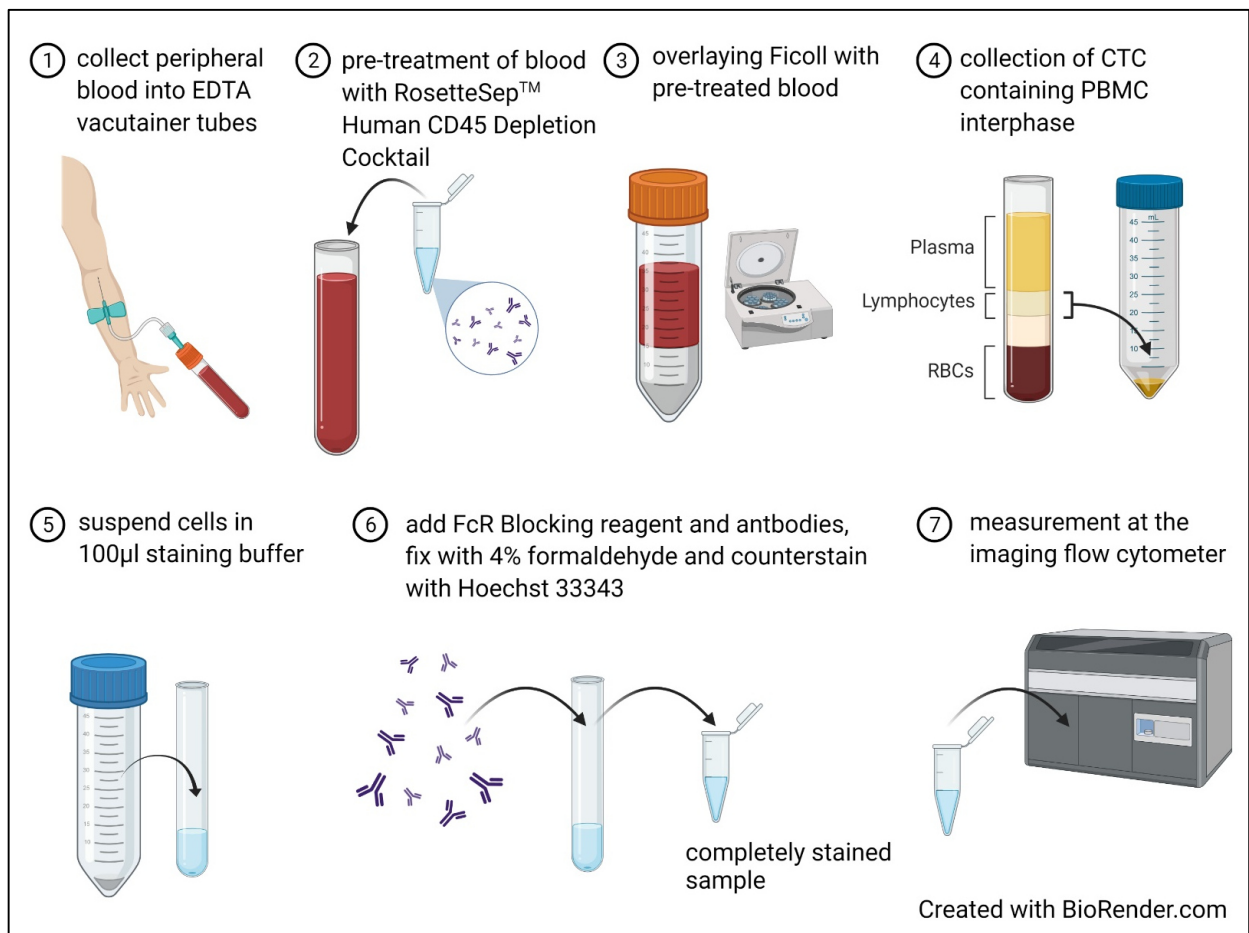


Cell line	Culture media
FaDu UD-SCC-4 UM-SCC-22B (HNSCC)	MEM Medium <i>(Life Technologies, cat. no.: 31095052)</i> 12% FCS <i>(Life Technologies, cat. no.: 10270-106)</i> 1x non-essential amino acids <i>(Life Technologies, order: 11140035)</i>
SCC-25 (HSNCC)	DMEM/F12 +GlutaMax <i>(ThermoFisher, cat. no.: 31331093)</i> 10% FCS <i>(Life Technologies, order: 10270-106)</i> 2mM L-Glutamine <i>(ThermoFisher, cat. no.: 25030149)</i> 1% Penicillin/ Streptomycin <i>(Fisher BioReagent, cat. no.: BP2959-50)</i> 15mM HEPES <i>(sigma, order: H4034-100g)</i>
MDA-MB-231 (TN breast cancer)	Advanced DMEM/F12 <i>(ThermoFisher, cat. no.: 12634010)</i> 10% FCS <i>(Life Technologies, cat. no.: 10270-106)</i> 2mM L-Glutamine <i>(ThermoFisher, cat. no.: 25030149)</i>
SW620 (colon cancer)	RPMI Medium 1640 <i>(Life Technologies cat. no.: 21875-034)</i> 20% FCS <i>(Life Technologies, cat. no.: 10270-106)</i> 2mM L-Glutamine <i>(ThermoFisher, cat. no.: 25030149)</i> 1% Penicillin/ Streptomycin <i>(Fisher BioReagent, cat. no.: BP2959-50)</i>

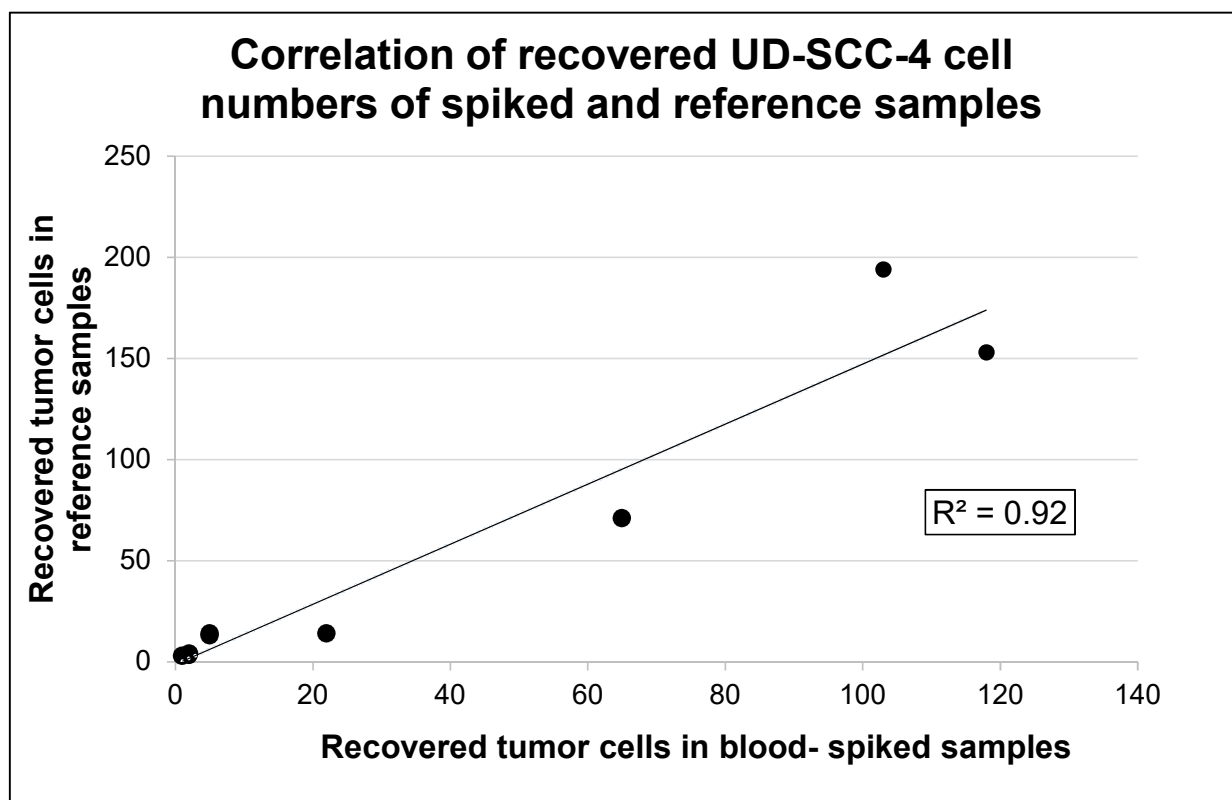
Supplementary Table S1: Media composition.



Supplementary Figure S1: Workflow of the CTC analysis. (1) Blood samples were processed within four hours after collection. (2) Enrichment of CTCs by adding RosetteSep™ Human CD45 Depletion Cocktail was done, (3) followed by Ficoll density centrifugation and (4) harvest of the PBMC interphase containing CTCs. (5-6) Next, cells were stained for extracellular markers, fixed with 4% formaldehyde and counterstained with Hoechst 33342 and (7) acquired at the Amnis® brand ImageStream®X MkII. ***Created with BioRender.com***

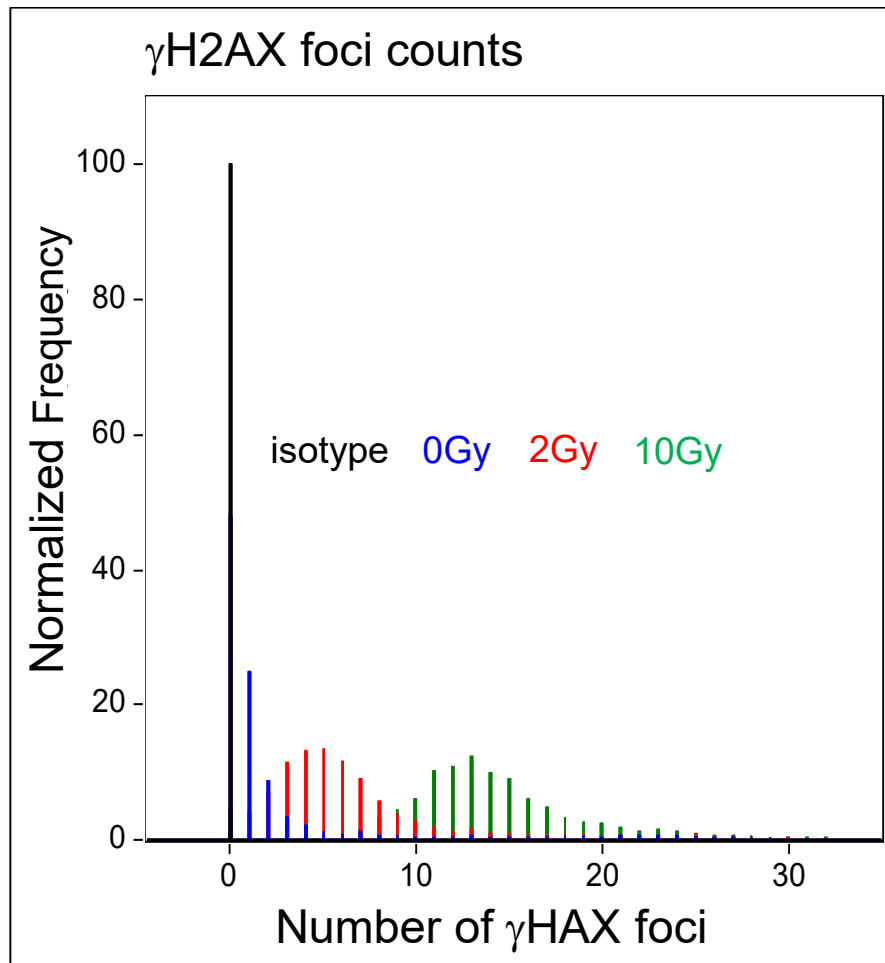
Laser (nm)	Target	Fluorophore	Channel	Laser power (mW)
405	Hoechst 33342	UV	7	30
488	EpCAM-EGFR	AF488	2	100
561	PD-L1	PE	3	200
561	phospho-EGFR	TexasRed	4	200
561	PD-L2	PE-Vio770	6	200
642	CD45 / γ H2AX	AF647	11	150/130
642	CD45	APC/Fire	12	130
782	SCC		12	0.9
bright field			1 and 9	

Supplementary Table S2: Laser settings. For data acquisition the 40x magnification was used and the fluidics were set to low speed and high sensitivity, EDF-module was switched off and focus/ centering were set to automatic. CD45 signal was acquired in channel 12 (instead of 11) for samples stained for γ H2AX.



Supplementary Figure S2: Results from spiking experiments using the UD-SCC-4 cell line.

Analysis of samples (n=3 replicates per dilution) revealed a correlation of $R^2 = 0.92$ of recovered cells in reference and spiked samples. The median recovery rate was 50%.



Supplementary Figure S3: Quantitative analysis of γ H2AX foci. The distribution of the number of γ H2AX foci detected in FaDu cells irradiated with the indicated dose is presented in the histogram.