



Article Involvement of HHV-4 (Epstein–Barr Virus) and HHV-5 (Cytomegalovirus) in Inflammatory Bowel Disease and Colorectal Cancer: A Meta-Analysis

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Simple Summary: The prevalence of colorectal cancer (CRC) and inflammatory bowel disease (IBD) is increasing worldwide. Understanding what factors foster these diseases' development can help reduce their burden. Viral infections have been suggested to be involved in the genesis of these diseases, but the data is contradictory. The present study analyzed 11,413 articles on the topic and identified 196 that could provide information on the relationship between a viral infection and these gastrointestinal diseases. The Epstein–Barr virus (EBV, HHV-4) was strongly associated with IBD, and cytomegalovirus (HHV-5) with both CRC and IBD.

Abstract: Gastrointestinal diseases (GDs) include colorectal cancer (CRC), gastric cancer (GC), and inflammatory bowel disease (IBD). CRC and GC are typically diagnosed at later stages of development, reducing patients' chances of survival. IBD is characterized by chronic intestinal inflammation and is a significant risk factor for the development of CRC. Chronic bacterial infections have been shown to promote some GDs, but the role of viruses in the etiology of these diseases is less clear. The present meta-analysis retrieved literature on the viral prevalence in GD patients, measuring the GD risk in odd ratios. By quantifying the study heterogeneity, the literature bias was fundamentally included in the analysis. The analysis also included 11 metagenomic studies. Our meta-analysis retrieved 11,413 studies, with 196 suitable for analysis. HHV-4 (Epstein–Barr virus) was identified as a significant risk factor for the development of IBD, and HHV-5 (cytomegalovirus) as a risk factor for both CRC and IBD. No relations withstanding the literature bias were identified for GC. The study discusses these findings, as well as the role of other viruses in the etiology of CRC and IBD.

Keywords: colorectal cancer; inflammatory bowel disease; Epstein–Barr virus; cytomegalovirus; meta-analysis; viral infection

1. Introduction

About 15% of all human cancers have been shown to have a microbial etiology [1]. Of the eleven micro-organisms recognized as class one carcinogens by the International Agency for Research on Cancer (IARC), seven are viruses: Human Herpesvirus 4 (HHV-4, also known as the Epstein–Barr virus), the Hepatitis B virus (HBV), the Hepatitis C virus (HCV), Human Herpesvirus 8 (HHV-8, also known as the Human Kaposi sarcoma virus), Human Immunodeficiency virus genotype 1 (HIV-1), the high risk genotypes of Human



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Papillomavirus (HPV), and the Human T-cell lymphotropic virus genotype 1 (HTLV-1) [2]. In some cases, the link between a certain virus and a given type of cancer is strong, in terms of both epidemiological data and experimental modeling. For instance, HPV have been recovered in virtually all cases of cervical cancer and the oncogenic process is well characterized at the molecular level [3]. In other cases, such relationships are less evident: HPV has been recovered only in 13–56 percent of oro-pharynx cancers and in 0–84 percent of tissues derived from the cancer of the colon and rectum, indicating that other causes are involved in the development of these typologies of cancer [4,5]. The reverse is also true: some types of cancer are not easily associated with microorganisms with transforming potential, even if the data are tantalizingly suggesting a relation. Among these, are the malignant diseases affecting the gastrointestinal tract.

Cancer of the stomach (better known as gastric cancer, GC) and colorectal cancer (CRC) share some similarities, including a high number of deaths caused worldwide, detection (at least in the western world) at rather late stages of the tumor development, and risk factors such as hereditary mutations, obesity, and sedentariness [6]. CRC is the second most common type of cancer worldwide, with a death toll of over half million people, yearly [6]. CRC is a group of heterogeneous malignancies, rather than a single type of tumor: about two thirds of cases are known as sporadic, further subdivided into hypermutable (16%) and non-hypermutable cancers (84%). The remaining one-third is classified as hereditary (or familial) [7,8]. The prevalence of GC has been decreasing in recent years, but it still remains the third cause of cancer-associated death after lung cancer and CRC, with over a million new cases yearly [9].

Every person has a four percent life-time risk of developing sporadic CRC [7], but some factors can increase this basal value: for instance, diet, sedentary lifestyle, and alcohol consumption [10], which are also risk factors for the development of GC [11,12]. Chronic inflammation of the gut is also a risk factor for the development of CRC that goes under the general term of inflammatory bowel diseases (IBD) and can be further differentiated into Crohn's Disease (CD) and ulcerative colitis (UC) [13–15]. Patients suffering from IBD, had a CRC risk increased by 40% in comparison to healthy people [16,17]. IBD incidences have escalated in the last few years and cause an economic burden of over six billion, in the US alone [18,19].

Recently, specific infections have been suggested to affect the risk of sporadic CRC [20]. For instance, the prevalence of a CMV infection has increased 350% in IBD cases, between 1998 and 2014, with indications of a higher mortality in the infected patients [21]. In the case of GC, an infection with *Helicobacter pylori* is known as a major risk factor, albeit the precise oncogenic mechanism is still poorly understood, and only a subset of *H. pylori* carriers develop GC [22]. It is believed that the abnormal activation of the NF- κ B signaling pathway induced by this bacterium might lead to chronic inflammation and, eventually, to cancer [23]. An infection with *Fusobacterium nucleatum* might put further strains on the aberrant activation of NF- κ B, further enhancing the gastrointestinal carcinoma risk [24] and a HHV-4 infection is observed in about 9% of cases [25]. Still, the situation is complex and hardly fits into a one cause-one effect model. In addition, most of the bacteria or viruses that have been associated with a higher risk of CRC, are commonly present in the healthy gut [26].

The idea of a relation between infection and CRC has been discussed in the last few decades [27]. Several experimental and epidemiological studies have provided support for the association between the presence of certain bacteria and CRC [28]. The current hypothesis is that several species of bacteria might be involved in CRC carcinogenesis and/or progression [29]. Moreover, the link between a viral infection and CRC remains less clear [26,30]. The odds ratios (OR) between infection with a given virus and development of CRC, ranged between 0.7 and 58.8 for HPV, 0.9 and 9.0 for the John Cunningham virus (JCV), 0.1 and 10.4 for Herpesviruses in different studies [31–34]. Furthermore, zur Hausen and collaborators have suggested the presence of a novel class of zoonotic vectors that might be involved in the development of CRC [35].

In relation with the establishment of the current paradigm linking oncogenic bacteria and CRC [29], bacteriophages (or simply phages) have been gaining interest in the most recent studies [36]. Using metagenomic approaches, research groups have shown differences in the prevalence of phages between the healthy and inflamed gut [37]. Specifically, an increased richness (number of species) of the phages belonging to the order *Caudovirales*, has been consistently reported [38].

Both CRC and GC develop rather slowly and symptoms are noticed only when the tumors have grown to a considerable size [10]. Thus, an early detection of the subjects at increased risk of developing either CRC and GC is paramount. To achieve such an early warning, a good comprehension of the risk factors that can facilitate CRC and GC carcinogenesis, progression or metastasis is fundamental. The present work had the objective to provide updated insights into the relationship between a viral infection and gastrointestinal diseases (CRC, GC, or IBD), in order to improve our understanding of their risk factors and to support the preventive measures.

Given the inconclusive data present in the public domain, we sought to provide an updated meta-analysis regarding the viral associations with CRC, GC, and IBD, with particular attention put on the phages. The research question posed herein was the following: *Is the presence of viruses associated with an increased risk of developing CRC/GC/IBD?* We answered this question by surveying the literature in the public domain and stratifying the eligible articles into groups that were used to determine the odds ratios (ORs) able to indicate the strength of such an association.

2. Materials and Methods

2.1. Protocol

The present study was conducted following the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) [39]. The complete search query is provided in Supplementary File 1. The search was aimed at identifying an increased risk of CRC, GC, or IBD. CRC included both carcinomas and adenocarcinomas. The viral families and prototypical viruses included in the analysis were:

- Adenoviridae: Adenovirus (ADV).
- Anelloviridae: Transfusion Transmitted Virus (or Torque Teno virus, TTV).
- *Astroviridae*: Astrovirus (AstV).
- *Bunyaviridae*: members of the Orthobunyavirus genus (BV).
- *Caliciviridae*: Norovirus (NV), Sapovirus (SaV).
- Herpesviridae: Human Herpesvirus 1 (HHV-1 or Herpes Simplex virus), Human Herpesvirus 3 (HHV-3, or Varicella Zoster virus), Human Herpesvirus 4 (HHV-4, or Epstein–Barr virus), Human Herpesvirus 5 (HHV-5, or Cytomegalovirus), Human Herpesvirus 6 (HHV-6), Human Herpesvirus 8 (HHV-8, or Kaposi's sarcoma-associated herpesvirus), Inoue–Melnick virus (IMV).
- *Hepadnaviridae*: Human Hepatitis B virus (HBV).
- *Flaviviridae*: Human Hepatitis C virus (HCV).
- *Matonaviridae*: Rubella virus (RuV).
- *Retroviridae*: Human Immunodeficiency virus (HIV), Human T-cell lymphotropic virus type 1 (HTLV-I).
- *Papillomaviridae*: Human Papilloma virus (HPV).
- *Parvoviridae*: Parvovirus B19 (B19), Bocaparvovirus (HuBV).
- Polyomaviridae: BK virus (BKV), John Cunningham virus (JCV), Merkel cell polyomavirus (MCPV), Polyovirus 5 (PyV6), Simian virus 40 (SV40).
- *Paramyxoviridae*: Measles virus (MeV), Mumps virus (MuV).
- *Reoviridae*: Reovirus (RV), Rotavirus (RoV).
- *Syphoviridae*: phages of the order *Caudovirales*.
- *Myoviridae*: phages of the order *Caudovirales*.
- *Inoviridae*: member of the Inovirus genus.

The viral families were clustered into the following groups:

- Herpesviruses (Herpesviridae).
- Respiratory viruses (Adenoviridae, Bocaviridae).
- Papillomaviruses (Papillomaviridae).
- Epithelial viruses (Anelloviridae, Bunyaviridae, Matonaviridae, Paramyxoviridae, Parvoviridae).
- Intestinal viruses (Astroviridae, Caliciviridae, Noroviridae, Reoviridae).
- Polyomaviruses (*Polyomaviridae*).
- Hepadnaviruses (Hepadnaviridae).
- Flaviruses (*Flaviviridae*).
- Retroviruses (*Retroviridae*).
- Phages (Syphoviridae, Myoviridae, Inoviridae).

2.2. Eligibility Criteria

The primary, original publications in international peer-reviewed journals, of human subjects, in the English language, and reporting the prevalence of viral infections in human patients were included.

2.3. Information Sources

PubMed, Cochrane Library, Web of Science Core Collection, Cinahl, and ClinicalTrial.gov.

2.4. Search

The search terms were subdivided into two groups: pathology and pathogen. These groups were combined with a logical AND operator. The terms were composed by the combination of the following terms.

- Pathology: "intestinal neoplasm", "colorectal cancer", "rectal neoplasia", "Crohn's disease", "inflammatory bowel disease", "ulcerative colitis", "regional enteritis", "granulomatous colitis", "terminal ileitis".
- Pathogen: "virus", "giant virus", "bacteriophage", "Siphoviridae", "Podoviridae", "T7-like virus", "φ-like virus", "Herellevirus", "Myoviridae", "Tristromavirus", "Bicaudavirus", "Pycodnaviridae", "Caudovirales", "Ackermannvirus", "Ampullavirus", "Clavavirus", "Corticoviridae", "Cystoviridae", "Fuselloviridae", "Globulovirus", "Guttaviridae", "Inoviridae", "Leviviridae", "Lipothrixviridae", "Microviridae", "Plasmavirus", "Pleolipovirus", "Rudiviridae", "Sphaerolipovirus", "Tectivirus", "Turrivirus", "Polyomaviridae", "Poxviridae", "Simian virus 40", "Papillomaviridae", "Herpesviridae".

2.5. Study Selection

In the first round of the assessment, the duplicated entries were removed; reviews, case reports, abstracts, clinical studies related to drugs or vaccines, and articles not written in the English language were also dismissed. In the second round, titles and abstracts were read; articles not related to a viral infection in the CRC or inflammatory diseases were withdrawn. In the third round, the whole articles were read and only those reporting the presence of viruses in intestinal lesions or metagenomic studies were included in the present review.

2.6. Data Collection Process

The publication repositories were queried with the strings reported in Supplementary File 1. The full texts of the selected articles were retrieved through the Central Library of the University of Heidelberg, Medical Faculty in Mannheim. Two investigators (L.M. and H.A.) selected the trials and independently reviewed the study titles and abstracts. The studies that met the inclusion criteria were retrieved for the full-text evaluation.

2.7. Data Items

The data related to articles, year of publication, number of patients having a pathology (cases) without infection, number of cases with infection, number of healthy patients

(controls) without infection, number of controls with infection, tissue type, analytical method and virus species, were tabulated for further analysis. The articles were also subdivided into virome and non-virome studies. The studies were further stratified, based on the source of the data: the tissues were obtained from the colon-rectum, stomach (gastrointestinal tract), stools, sera, and medical (archival) records.

2.8. Risk of Bias in the Individual Studies

The assessment of the literature bias was measured, taking the study heterogeneity (variation of the degree of association between the cause and effect) into account, and quantified with the Higgins I^2 index [40]. The values of the I^2 index were stratified, as suggested [41]: low (<30%), moderate (30–60), substantial (61–75), and considerable (>75). The heterogeneity of the studies was quantified using the Egger's regression test (ERT) of the funnel plot [42]. The ERT was preferred over the Cochran's Q test (CQT) because it was reported to be more reliable on small data sets [43]. However, the ERT requires at least six publications and could not be computed in many instances. Thus, the CQT was included to supply the additional information on the study heterogeneity. The significant ERT and CQT indicate a publication bias.

2.9. Summary Measures

The odds ratio (OR) statistics were calculated for the non-virome studies, using the random effects model implemented by Mantel and Haenszel [44]. The trim-and-fill method was employed to adjust for the funnel plot asymmetry [45]; this method removed the studies causing the funnel plot asymmetry and replaced the omitted studies with the data around the newly computed center of the distribution. The trim-and-fill method required at least three data points for its computation.

2.10. Synthesis of the Results

The data were summarized in the tables reporting the OR and l^2 with their 95% CI and associated *p*-value. The classes that did not contain data were not reported in the table to increase the readability. The most relevant data were also displayed in forest plots and funnel plots to increase the understanding of the underlying relations between the studies [40,46].

2.11. Risk of Bias across the Studies

The risk of bias for the observational studies was assessed with the Newcastle–Ottawa scale (NOS) [47] by preparing a table with the following fields:

- Representativeness: if the selected article included more than 50 participants, a cut-off accepted to provide the statistically solid results, a positive value was assigned.
- Control group: if the selected article included a control group, a positive value was assigned.
- Documentation: if the selected article was not a letter, a positive value was assigned.
- Outcome: if the selected article was not a virome study, a positive value was assigned.
- Co-infections: if the selected article did not report samples with co-infections, a positive
 value was assigned.

The sum of the assigned points was used to stratify the selected studies as follows:

- 5 points: High
- 3–4 points: Moderate
- 1–2 points: Low

2.12. Additional Analyses

For the virome studies, it was possible to do a qualitative description only, which was due to the absence of the clear number of cases and the controls infected with a given virus. Thus, the viral prevalence is provided for these cohorts as a whole.

2.13. Statistical Analysis

The meta-analysis was implement in *R* version 4.2.1 using the packages *metafor*, *meta*, and *metasens*, which provided both the statistical and graphical outputs.

3. Results

3.1. Study Selection

We inquired six scientific literature databases: PubMed, Cochrane Library, Web of Science, Cinahl, ClinicalTrial.gov, and the International Clinical Trials Registry Platform (ICTRP) on 3 March 2021 and identified 11,413 articles, with 1009 duplicated entries, matching our search algorithm (Figure 1a). Following the screening for titles and abstracts, we withdrew 9811 articles because they were not related to the presence of viruses in the tissues derived from either CRC or IBD patients. An additional 364 articles were removed after reading the full text because they were not related to the research question and 34 were withdrawn because they were related to anal cancer (false positives). We identified 196 articles that matched the research question (Supplementary Table S1). We subdivided the selected articles into two groups: one undergoing a quantitative analysis (n = 188, 95.9%) and another qualitative description (n = 8, 4.1%). This subdivision was due to the fact that the latter group comprised a MPS-derived virome analysis not reporting the number of patients infected with a given virus, but rather presenting an overview of the viral families observed in the whole sample set. It was therefore not possible to calculate the ORs for these studies but they were included because they represented the most recent advancements in CRC research and highlighted the presence of viruses that could not be determined by more conventional methods.

3.2. Study Characteristics

Since there were 38 articles of the 196 studies retrieved (19.4%) which reported results for more than one virus, there were collectively 255 data points, 211 of them (82.7%) performed after the year 2000 (Figure 1b). There were 117 data points out of 196 (45.9%) that did not provide a comparison group; thus, these articles were not suitable for the quantitative assessment.

The data points could be stratified, according to the following groups:

3.2.1. Disease

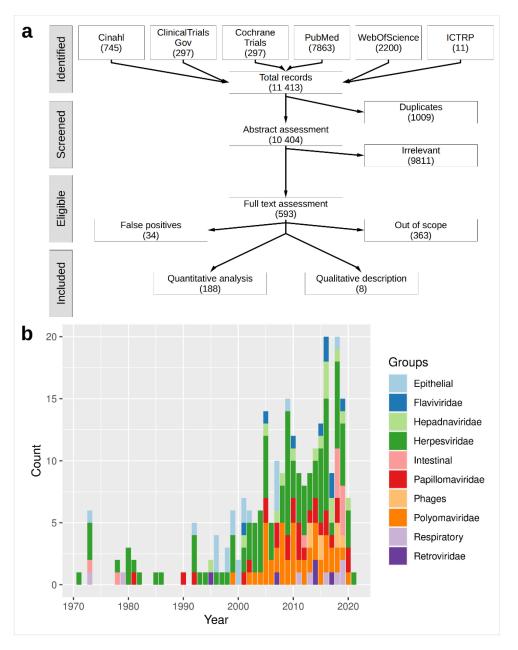
- CRC (n = 123, 48.2%).
- GC (n = 28, 11.0%).
- IBD (n = 104, 40.8%).

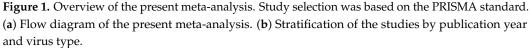
3.2.2. Detection Method

- Hybridization (n = 49, 18.8%). Fluorescent immuno-assay (IFA), in situ hybridization (ISH) or immuno-histochemistry (IHC).
- PCR (n = 118, 46.5%). Polymerase chain reaction (PCR), both end-point or quantitative.
- *Archival* (n = 25, 9.8%). Medical records obtained from the retrospective studies, and without direct detection of the virus within the specimens.
- Serology (n = 55, 21.6%). Enzyme-linked immunoassay (ELISA) and plaque assay.
- Virome (n = 9, 3.5%). Whole-genome sequencing (WGS) by massively parallel sequencing (MPS).

3.2.3. Type of Tissue

- Colon and rectum tissue (n = 114, 44.7%).
- Gastrointestinal (stomach) tissue (n = 43, 16.9%).
- Stools (n = 18, 7.1%).
- Sera (n = 56, 22.0%).
- Medical records (n = 24, 9.4%).





3.2.4. Viral Groups

- Respiratory viruses (n = 8, 3.1%).
- Intestinal viruses (n = 11, 4.3%).
- Epithelial viruses (n = 20, 7.8%).
- Herpesviruses (n = 107, 42.0%).
- Retroviruses (n = 5, 2.0%).
- Polyomaviruses (n = 44, 17.3%).
- Papillomaviruses (n = 28, 11.0%).
- Hepadnaviruses (n = 16, 6.3%).
- Flaviviruses (n = 9, 3.5%).
- Phage group (n = 7, 2.7%).

The majority of the 196 studies displayed a moderate NOS score (n = 127, 64.8%), followed by a high NOS score (n = 60, 30.6%), and only a fraction of the studies retrieved herein had a low score (n = 9, 4.6%). Therefore, it could be assumed that the quality of the dataset was good (Supplementary Table S2).

3.3. Quantitative Analysis: CRC

The risk of developing either CRC, GC or IBD due to a viral infection based on the random effects model, is reported on Table 1. Based on the colorectal tissues, there was a significant association between an infection and CRC for the viral groups herpes (OR = 2.62, 95% CI: 1.17–5.86, *p*-value = 0.0187), Polyomaviruses (OR = 2.17, 95% CI: 1.20–3.93, *p*-value = 0.0105), Hepadnaviruses (OR = 1.22, 95% CI: 1.16–1.28, *p*-value < 0.0001), and epithelial viruses (OR = 3.76, 95% CI: 1.21–11.68, p-value = 0.0221). An infection with Papillomaviruses was just above significance (OR = 4.56, 95% CI: 0.99–21.03, p-value = 0.0512). Based on the sera, none of the studies indicated a significant increase of risk of a gastrointestinal disease. Conversely, based on the archival records, the random effects model identified a significant association between infection with Flaviviruses (OR = 0.63, 95% CI: 0.04–0.10, p-value < 0.0001) and confirmed the association between an infection with Hepadnaviruses and CRC (OR = 0.12, 95% CI: 0.08–0.17, *p*-value < 0.0001). However, the results obtained from the archival records was based on individual studies and pointed to a decreased risk of cancer upon infection. None of the indices related to the quantification of study heterogeneity (I^2 index, ERT, and CQT) could be computed, confirming the low reliability of the results, based on archival data.

Similarly, the results regarding the increased risk of CRC upon infection with HBV, based on the colorectal tissues, was based on a single study and even in this case no measure of heterogeneity could be computed. The association between the increased rick of CRC and infection with epithelial viruses was slightly better because, based on two studies that consistently reported a higher cancer risk upon infection, only the CQT could be computed (*p*-value = 0.1803), suggesting a negligible publication bias.

The relation, based on the colorectal tissues and infection with either Herpesviruses and Polyomaviruses, was better because it was based on 12 and 19 studies, respectively. In both cases, the l^2 index was considerable (\geq 70.3%), indicating an issue of publication bias and corroborating the use of the random effects model for computing the meta-analysis because it is more reliable in the presence of such a bias. The ERT was slightly below significance, strengthening the issue of publication bias. The observation of the funnel plots for Herpesviruses, Papillomaviruses, and Polyomaviruses, showed that the reports related to an infection with HPV were seriously affected by an asymmetric distribution (Figure 2). The situation for Herpesviruses and Polyomaviruses was better than that of Papillomaviruses, but it was still possible to observe a higher representation of the studies reporting a positive association between an infection and CRC than the studies describing otherwise.

Given the influence of the publication bias in defining the association between viral infection and CRC, the trim and fill method was applied (Table 2). The ERT became significant for Herpesviruses, Papillomaviruses, and Polyomaviruses (*p*-value < 0.0001 in all cases), whereas the CQT became non-significant. Given the small sample set, the former metric should be considered more reliable. However, none of the viruses was associated with a significant risk of CRC; thus, the increased risk of CRC, based upon an infection with either Herpesviruses and Polyomaviruses, could not be confirmed by the trim-and-fill method.

Disease	Source	Group	Studies *	Cases ⁺	Controls ⁺	OR	<i>p</i> -Value	I ² (%)	ERT	CQT
CRC	Colon/Rectum	Herpes	12/28	375/1968	72/517	2.62 (1.17-5.86)	0.0187	70.3 (46.4–88.6)	0.0417	0.0001
		Papilloma	11/26	517/2402	177/888	4.56 (0.99-21.03)	0.0512	91.5 (86.8-94.5)	0.1270	< 0.0001
		Polyoma	19/36	1531/3980	768/2029	2.17 (1.20-3.93)	0.0105	79.0 (67.9–86.3)	0.0413	< 0.0001
		Hepadna	1/1	3536/69,478	2924/69,478	1.22 (1.16-1.28)	< 0.0001	—	—	
		Epithelial	2/3	116/132	86/119	3.76 (1.21-11.68)	0.0221	—	—	0.1803
		Papilloma	1/1	21/97	42/184	0.93 (0.52-1.69)	0.8221	_		_
		Hepadna	1/1	72/284	212/284	0.12 (0.08-0.17)	< 0.0001	—		_
		Flavivirus	1/1	51/255	204/255	0.63 (0.04-0.10)	< 0.0001	—		_
		Retrovirus	2/3	146/674	284/1542	0.82 (0.58-1.18)	0.2898	—	—	0.3099
	Serum	Herpes	5/5	64/89	37/79	3.06 (0.74-12.65)	0.1227	54.3 (0-83.2)	—	0.6740
		Polyoma	3/5	1029/1409	1008/1409	1.12 (0.89–1.42)	0.3224	30.9 (0-92.8)	—	0.2351
		Hepadna	1/3	674/6333	3168/96,000	1.31 (0.89–1.95)	0.1754	—	—	
		Flavivirus	1/3	10/820	806/96,000	1.58 (0.78–3.19)	0.2009	—		—
GC	Gastric	Herpes	1/22	502/5097	207/225	1.57 (0.60-4.06)	0.3567	_		_
		Papilloma	0/1	9/302	0/0	—	—	—	—	—
		Polyoma	2/2	18/80	4/82	5.22 (1.66-16.47)	0.0048	0	0.6804	_
		Epithelial	0/1	17/32	0/0	—	—	—	—	—
	Serum	Herpes	1/1	43/48	88/93	0.49 (0.13-1.78)	0.2774	—	—	—
		Hepadna	1/1	26/150	20/150	1.36 (0.72-2.57)	0.3375	—	—	—
		Retrovirus	0/1	0/150	0/150	—	—	_		—
IBD	Colon/Rectum	Herpes	9/16	173/882	10/211	2.71 (0.91-8.06)	0.0723	50.6 (0-76.9)		0.3980
	Stools	Herpes	0/1	9/400	0/0	—	—	—	—	—
		Intestinal	5/7	65/2609	3108/35,321	0.27 (0.09-0.08)	0.0236	92.0 (84.4–95.6)	—	< 0.0001
		Respiratory	1/2	3/711	106/8826	0.29 (0.07–1.16)	0.0801	—	—	
	Gastric	Herpes	7/10	235/591	56/259	2.90 (1.15-7.53)	0.0241	72.3 (40.2-87.2)	—	0.0014
		Epithelial	1/6	17/127	11/62	0.89 (0.27-2.97)	0.8530	—	—	—
	Record	Herpes	2/7	636/46,752	1083/33,355	1.22 (0.74-2.01)	0.4377	60.8 (0-90.9)	—	0.1100
		Intestinal	2/2	110/176,856	392/873,014	1.41 (0.92–2.17)	0.1174	75.7 (0–94.5)	—	0.0425
		Respiratory	1/1	6/88,428	11/436,507	2.69 (1.00-7.28)	0.0510	—		0.0000
		Hepadna	1/2	196/3776	4/67	0.37 (0.13-1.07)	0.0671	—	—	0.0000
		Flavivirus	1/2	34/3556	1/57	0.75 (0.10-5.64)	0.7799	—	—	0.0000
		Epithelial	2/2	38/12,738	36/15,264	1.27 (0.80-2.00)	0.3140	0	—	0.7617
		Retrovirus	1/1	2/1816	1/84	0.09 (0.01-1.02)	0.0519		_	0.0000

Table 1. Mixed-affect model for the risk of CRC, GC, and IBD predicted by the studies selected for the present meta-analysis. The studies are stratified by specimen type (Source) and virus type (Group). The ORs are reported with their 95% confidence interval and *n*-value, J^2 = Higgins index: ERT = Egger's regression test:

Disease	Source	Group	Studies *	Cases ⁺	Controls ⁺	OR	<i>p</i> -Value	I ² (%)	ERT	CQT
	Serum	Herpes	5/12	1926/2267	256/438	1.53 (0.52-4.46)	0.4379	82.7 (60.5–92.5)	_	0.0001
		Intestinal	2/3	62/200	23/89	1.08 (0.47-2.49)	0.8615	0	—	0.8542
		Respiratory	1/1	11/57	20/52	0.38 (0.16-0.91)	0.0291	—	—	0.0000
		Polyoma	1/2	158/244	6/53	9.47 (3.46-25.92)	< 0.0001	—	_	0.0000
		Hepadna	4/8	591/10,769	1249/12,140	0.77 (0.49-1.20)	0.2492	90.1 (77.6–95.6)	_	< 0.0001
		Flavivirus	2/3	104/8969	66/4264	0.99 (0.46-2.12)	0.9734	78.2 (4.9–95.0)	_	0.0323
		Epithelial	7/8	888/1372	729/1058	0.71 (0.37–1.36)	0.3027	70.4 (35.3–86.5)	—	0.0025

Table 1. Cont.

* The first number in front of the division sign '/' refers to the number of studies included in the statistical calculations; the second number refers to the total number of studies available for each specific combination of disease, source, and group. † The first number in front of the division sign '/' refers to the number of patients infected with a given virus; the second number refers to the total number of patients available for each specific combination of disease, source, and group.

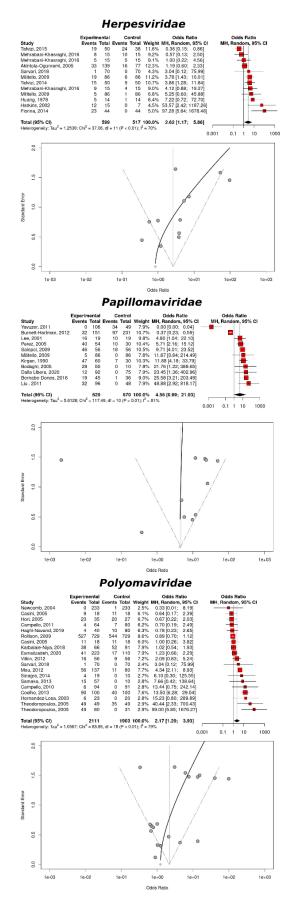


Figure 2. Association between the CRC risk and a viral infection. The upper, middle, and lower panels are related to the infection with members of the *Herpesviruses*, *Papillomaviridae*, and *Polyomaviridae*

families, respectively, for the colorectal tissues. Each panel includes a forest plot, based the random effects models and a funnel plot with a regression line.

Further stratification of the CRC risk, based on a Herpesviruses infection, was based on the viral genera of the *Herpesviridae* family (Table 3). Based on the colorectal tissues, HHV-4 showed an increased CRC risk (OR = 3.39, 95% CI: 0.64-18.10), but with a non-significant association (*p*-value = 0.1531). HHV-5 was instead just above significance (OR = 2.71, 95% CI: 0.99-7.45, *p*-value = 0.0519). HHV-1 (Herpes Simplex 1) and HHV-8 (Kaposi sarcoma herpes virus), with only one study each, did not provide valuable data to the analysis.

Table 2. Trim-and-fill model for the risk of CRC, GC, and IBD, predicted by the studies selected for the present meta-analysis. The model added a specified number of studies to adjust for the publication bias. The studies are stratified by specimen type (Source) and virus type (Group). The ORs are reported with their 95% confidence interval and *p*-value. I^2 = Higgins index; ERT = Egger's regression test; CQT = Cochran's Q test.

Disease	Source	Group	Added Studies	OR	<i>p</i> -Value	I ² (%)	ERT	CQT
CRC	Colorectal tissue	Herpes	4	1.45 (0.58–3.65)	0.4284	73.2 (56.0-83.7)	< 0.0001	0.7797
		Papilloma	4	1.59 (0.36-7.04)	0.5424	90.0 (85.2–93.2)	< 0.0001	0.5741
		Polyoma	6	1.13 (0.52-2.44)	0.7596	84.8 (78.6-89.1)	< 0.0001	0.7587
	Serum	Herpes	2	1.40 (0.26–7.69)	0.6958	66.8 (25.9-85.1)	0.0061	_
IBD	Colorectal tissue	Herpes	0	2.71 (0.91-8.06)	0.0723	50.6 (0-76.9)		0.398
		Intestinal	3	0.69 (0.19-2.45)	0.5630	94.2 (90.8-96.4)	_	< 0.0001
	Gastric tissue	Herpes	1	2.67 (1.10-6.52)	0.0307	69.1 (35.6-85.2)	_	0.0019
	Serum	Herpes	1	2.08 (0.60-7.20)	0.2472	81.8 (61.3-91.5)	_	< 0.0001
		Hepadna	2	0.55 (0.32-0.94)	0.0275	94.5 (90.6-96.8)	_	< 0.0001
		Epithelial	0	0.71 (0.37-1.36)	0.3027	70.4 (35.3–86.5)		< 0.0001

The heterogeneity associated with the studies related to HHV-4, was considerable ($I^2 = 83.3\%$) but moderate for HHV-5 ($I^2 = 60.1\%$), albeit the CQT was significant in both cases. The ERT could not be calculated for either sub-group. The application of the trim-and-fill method resulted in the addition of two studies with both HHV-4 and HHV-5. For the former, the association produced the values: OR = 1.24, 95% CI: 0.19–8.21, *p*-value = 0.8268, $I^2 = 84.3\%$ (95% CI: 69.3–91.9%), CQT < 0.0001; for the latter the values were: OR = 1.70, 95% CI: 0.55–5.25, *p*-value = 0.3573, $I^2 = 64.5\%$ (95% CI: 24.3–83.4%), CQT = 0.0062. The ERT could not be computed for either sub-group. Therefore, even in this occurrence, the fill-and-trim method did not substantiate the results of the random effects model. A visual inspection of the data (Figure 3) confirmed a publication bias due to the over-representation of the studies reporting a positive association between an infection and the CRC risk.

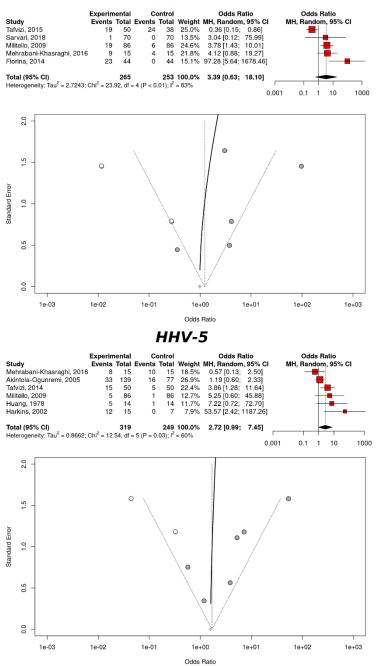
The vast majority of the studies related to the CRC risk and an infection with members of the *Polyomaviridae* family (n = 36) was due to JCV (n = 27, 75.0%). The other viruses involved in the infection model were BKV (n = 4, 11.1%), SV40 (n = 3, 8.3%), and MCPV (n = 2, 5.6%). Further analysis was focused on JCV because it was the most prevalent in the study set. The random effects model reported a significant association, based on 15 studies, between an infection with JCV and the CRC risk: OR = 2.64, 95% CI: 1.27–5.49, *p*-value = 0.0096, I^2 = 82.9% (95% CI: 73.0–89.1%), ERT = 0.0524, CQT < 0.0001. However, the trim-and-fill method, which added five studies to the set, did not confirm the association: OR = 1.19, 95% CI: 0.46–3.09, *p*-value = 0.7180, I^2 = 87.3% (95% CI: 81.8–91.2%), ERT = 0.7247, CQT < 0.0001.

The epithelial group associated with a high risk of CRC, was due to an infection with TTV (n = 2) and B19 (n = 1). Due to the paucity of data, it was not possible to further stratify the analysis. The other sources of information (gastric tissues, sera, and medical records) did not provide additional value to the analysis of the CRC risk.

Table 3. Mixed-affect model for the risk of CRC, and IBD, predicted by the studies related to an infection with Herpesviruses. The studies are stratified by specimen
type (Source) and Herpesvirus genotype (Group). The ORs are reported with their 95% confidence interval and <i>p</i> -value. <i>I</i> ² = Higgins index; ERT = Egger's regression
test; CQT = Cochran's Q test.

Disease	Source	Group	Studies *	Observations ⁺	Events ⁺	OR	<i>p</i> -Value	I ² (%)	ERT	CQT
CRC	Colorectal tissue	HHV-1	1/1	5/15	5/15	1 (0.22–4.56)	1			
		HHV-4	5/11	140/912	34/253	3.39 (0.64–18.10)	0.1531	83.3 (62.0–92.6)	—	< 0.0001
		HHV-5	6/15	228/855	33/249	2.71 (0.99–7.45)	0.0519	60.1 (2.2–83.8)	—	0.0281
		HHV-8	0/1	2/186	0/0	—	—	_		—
IBD	Gastric tissue	HHV-4	3/4	89/195	14/75	5.08 (2.57-10.02)	<0.0001	0 (0–89.6)		0.374
		HHV-5	3/5	117/346	24/163	3.82 (1.49-9.79)	0.0053	42.4 (0-82.6)	_	0.1761
		HHV-6	1/1	29/50	18/21	0.23 (0.06–0.88)	0.0323	—	—	—

* The first number in front of the division sign '/' refers to the number of studies included in the statistical calculations; the second number refers to the total number of studies available for each specific combination of disease, source, and group. + The first number in front of the division sign '/' refers to the number of patients infected with a given virus; the second number refers to the total number of patients available for each specific combination of disease, source, and group.



HHV-4

Figure 3. CRC risk and an infection with Herpesviruses, based on the colorectal tissues. The upper panel refers to HHV-4 (Epstein–Barr virus) and the lower panel refer to HHV-5 (Cytomegalovirus). Each panel includes a forest plot, based the random effects models and a funnel plot with a regression line. The studies added by the trim-and-fill method are represented by white dots.

3.4. Quantitative Analysis: GC

The number of studies regarding GC was very limited. Based on the gastric tissues, only Herpesviruses and Polyomaviruses had enough coverage to provide metrics for the meta-analysis (Table 2). However, only one study out of 22 could provide information regarding Herpesviruses (OR = 1.57, 95% CI: 0.60-4.06, *p*-value = 0.3567), resulting in a non-significant association and the absence of heterogeneity measures. Only two studies provided information regarding the GC risk upon infection with Polyomaviruses, but in this case with a significant association: OR = 5.22, 95% CI: 1.66-16.47, *p*-value = 0.0048, ERT = 0). It was not possible to perform the trim-and-fill method.

Serum samples provided information for Herpesviruses and HBV, but none of the associations was significant (Herpesviruses: OR = 0.49, 95% CI: 0.13–1.78, *p*-value = 0.2774; Hepadnaviruses: OR = 1.36, 95% CI: 0.72–2.57, *p*-value = 0.3375.

3.5. Quantitative Analysis: IBD

Based on the colorectal tissues, the only viruses providing information to compute in the meta-analysis, were the Herpesviruses, which, however, showed an association just above significance: OR = 2.71, 95% CI: 0.91–8.06, *p*-value = 0.0723, $I^2 = 50.6\%$ (95% CI: 0–76.9%), CQT = 0.3980. The trim-and-fill method did not add any study to the set, thus it confirmed the results of the random effects model. The moderate I^2 score and the non-significant CQT, highlighted that in this case, there was a negligible publication bias and, therefore, no requirement for an adjustment of the results.

Based on stool specimens, intestinal and respiratory viruses showed a significant association with the IBD risk. There were five studies substantiating the meta-analysis, however the trend reported was that of a reduction of IBD upon infection with intestinal viruses: OR = 0.27, 95% CI: 0.09–0.08, *p*-value = 0.0236. However, the heterogeneity was substantial ($I^2 = 92.0, 95\%$ CI: 84.4–95.6, CQT < 0.0001), suggesting that there was a strong publication bias. The trim-and-fill method added three articles to the set and did not confirm the association: OR = 0.69, CI: 0.19–2.45, *p*-value = 0.5630, $I^2 = 94.2\%$ (95% CI: 90.8–96.4%), CQT < 0.0001. Remarkably, the adjustment of the data showed that the IBD risk could be as high as 2.45 upon infection, suggesting that the possible beneficial role of an infection with intestinal viruses (Astrovirus, Norovirus, Rotavirus, and Sapovirus) was most probably a statistical artifact. With only one study supporting the association between an infection with respiratory viruses: OR = 0.29, CI: 0.07–1.16, *p*-value = 0.5630. Neither the measurement of the study heterogeneity nor the trim-and-fill method could be computed.

Based on gastric tissues, there was a significant involvement of Herpesviruses in increasing the IBD risk. The association was supported by seven studies and provided the following measures: OR = 2.90, 95% CI: 1.15–7.53, *p*-value = 0.0241, I^2 = 72.3% (95% CI: 40.2–87.2%), CQT = 0.0014. The trim-and-fill method added one study and confirmed the association between an infection with Herpesviruses and the high risk of IBD: OR = 2.67, 95% CI: 1.10–6.52, *p*-value = 0.0307, I^2 = 69.1% (95% CI: 35.6–85.2%), CQT = 0.0019. Figure 4 shows how the trim-and-fill method improve upon the publication bias associated with these measures.

Based on the sera, there was a significant association between a high IBD risk and an infection with either respiratory viruses and Polyomaviruses. The former association was, however, based on a single study: OR = 0.38, 95% CI: 0.16–0.91, *p*-value = 0.0291. Neither the measurement of the study heterogeneity nor the trim-and-fill method could be computed. As in the case of the association between an infection with respiratory viruses and the IBD risk based on stools, the reduced risk upon infection might simply be a statistical artifact. Similarly, the association between an infection with Polyomaviruses and the IBD risk, was based on a single study: OR = 9.47, 95% CI: 3.46–25.92, *p*-value < 0.0001. In this case, there was an elevated risk but the paucity of the study set made the association untrustful. Neither the measurement of the study heterogeneity nor the trim-and-fill method could be computed.

Interestingly, there was an association between an infection with Hepadnaviruses and the IBD risk. The random effects method did not find a significant association, based on four studies related to serum specimens: OR = 0.77, 95% CI: 0.49–1.20, *p*-value = 0.2492, $I^2 = 90.1\%$ (95% CI: 77.6–95.6%), CQT < 0.0001. However, the trim-and-fill method added two studies and reported a significant association: OR = 0.55, 95% CI: 0.32–0.94, *p*-value = 0.0275, $I^2 = 94.5\%$ (95% CI: 90.6–96.8%), CQT < 0.0001 (Figure 4). The trim-and-fill method improved upon the publication bias associated with these measures. As in the case

of intestinal and respiratory viruses, the reduced IBD risk upon infection might simply highlight a statistical artifact.

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Herpesviridae

Figure 4. IBD risk and a viral infection. The upper panel refers to Herpesviruses, based on the gastric tissues, and the lower panel refers to Hepadnaviruses on the sera. Each panel includes a forest plot, based the random effects models and a funnel plot with a regression line. The studies added by the trim-and-fill method are represented by white dots.

Further stratification of the IBD risk based on a Herpesviruses infection was based on the viral genera of the *Herpesviridae* family (Table 2). Based on three studies related to the gastric tissues, HHV-4 showed a significant increase in the IBD risk: OR = 5.08, 95% CI: 2.57–10.02,

p-value < 0.0001. The measure of heterogeneity indicated a negligible publication bias, further substantiating these results: $I^2 = 0\%$ (95% CI: 0–89.6%), CQT = 0.3740. The trim-and-fill method did not add any study, confirming the negligible publication bias associated with these measures (Figure 5). Based on three studies related to the gastric tissues, HHV-5 also showed a significant increase of the IBD risk: OR = 3.82, 95% CI: 1.49–9.79, *p*-value = 0.0053. As in the case of HHV-4, the measure of heterogeneity advocated for a low publication bias: $I^2 = 42.4\%$ (95% CI: 0–82.6%), CQT = 0.1761. The trim-and-fill method added two studies but the association was reduced to just above significance: OR = 2.26, 95% CI: 0.88–5.82, *p*-value = 0.0920, $I^2 = 56.4\%$ (95% CI: 0–83.8%), CQT = 0.0571 (Figure 5). There was only one study reporting about the IBD risk upon infection with HHV-6; the association was significant: OR = 0.23, 95% CI: 0.06–0.88, *p*-value = 0.0323. Neither the measurement of the study heterogeneity nor the trim-and-fill method could be computed.

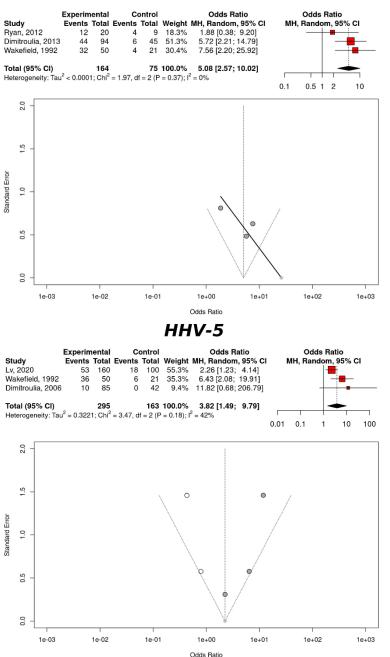
3.6. Qualitative Analysis of the Virome Studies

The majority of the virome studies were based upon stool samples: 8 out of 11 (72.7%). The microbiome analysis showed a higher viral richness in the controls (n = 28) than the CD patients (n = 31): there were 4399 viral species in the former group and 2161 viral species in the latter [48,49]. *Synechococcus* phage S CBS1 was characteristically observed only in inflammatory lesions [48]. Other studies reported a higher abundance (in terms of the total number of sequences) of phages, specifically those belonging to the order *Caudovirales*, in the inflammatory lesions (n = 12) than in the healthy controls (n = 12) (Dunn's test, *p*-value = 0.05); however, such an increase did not correspond to a rise in the phage richness [50]. An increased phage richness (specifically of the members of the *Caudovirales* family), together with a lower bacterial richness in the IBD patients (n = 52) over the healthy controls (n = 21), was instead reported by Norman and co-workers (Spearman correlation, *p*-value < 0.05) [38]. In particular, the authors identified the phages targeting the bacterial species *Lactococcus*, *Lactobacillus*, *Clostridium*, *Enterococcus* and *Streptococcus*, as specifically associated with this disease.

The comparison of the viral communities between the CRC cases (n = 74) and the healthy controls (n = 92) in the stools, showed an increased phage richness in the former (Wilcoxon rank test, *p*-value = 0.013) as well as an inverse association between the bacterial and viral richness [51]. Using the random forest analysis, the authors identified *Orthobunyavirus* as a taxon specifically associated with CRC specimens, followed by Inovirus and Tunalikevirus. Using a similar approach, Hannigan and collaborators [52] identified members of the viral species *Siphoviridae* and *Myoviridae* (both belonging to the order *Caudovirales*) as more associated with CRC (n = 60) than the healthy condition (n = 30) (Wilcoxon rank test, *p*-value < 0.01).

In one study based on colon biopsies (six CD patients and six healthy controls), the phages B40-8 and B124-14, both infecting *Bacteroidetes fragilis*, were identified as the most abundant; furthermore, the *Mycobacterium* phages TM4 and Wee were observed only in the inflammatory lesions [53]. In one study based upon the archival data, a high variability of virome communities between people was observed, although members of the *Partiviridae* and *Hepeviridae* families were the most prevalent [54]. Healthy specimens showed a higher predominance of the viral families *Polydnaviridae*, *Tymoviridae*, and *Virgaviridae* together with a significantly lower representation of *Hepeviridae*, compared to the inflammatory lesions (one-way ANOVA with Bonferroni correction, *p*-value < 0.05).

The work by Zapatka and collaborators was based on the re-analysis of the stored WGS-data, derived from 5354 pairs of tumors and normal tissues [55]. The tumors included 38 cancer types from 356 patients. The results showed that HHV-4 was the most common viral entity in the dataset, with a higher prevalence in the tumor tissues. The study also included 1057 RNA expression samples that indicated how GC sections were enriched in lytic HHV-4 transcripts, supporting a role of this virus in the gastric oncogenesis. The study did not report an infiltration of lymphocytes that could explain the presence of HHV-4 in the tumor tissues.



HHV-4

Figure 5. IBD risk and an infection with Herpesviruses based on the gastric tissues. The upper panel refers to HHV-4 (Epstein–Barr virus) and the lower panel refers to HHV-5 (Cytomegalovirus). Each panel includes a forest plot based on the random effects models and a funnel plot with a regression line. The studies added by the trim-and-fill method are represented by white dots.

4. Discussion

Gastrointestinal diseases represent a primary public health concern. In particular, CRC and GC are highly prevalent cancer types with a high mortality and delayed diagnosis. IBD is a significant cause of morbidity that has been pointed out as a risk factor for the development of CRC. The prevalence of CRC and GC is still increasing. A genetic predisposition, dietary habits, and sedentary lifestyle, amongst other causes, are associated with developing these gastrointestinal diseases. Bacterial infections have been recognized as additional risk factors as well. However, the role of viral infections in the development of CRC, GC, and IBD is less well understood. Our present meta-analysis suggests a role for

HHV-4 (EBV) as a risk factor for IBD and also, less significantly, for CRC, and of HHV-5 (CMV) as a less strong risk factor for CRC and IBD.

The present study sought to assess the role of viruses as risk factors for these main gastrointestinal diseases. This aim was fulfilled by applying a meta-analysis to the scientific publications related to the prevalence of viruses in people with CRC, GC, or IBD, against control subjects. We included all viruses associated with human diseases, including bacteriophages, to avoid the risk of missing possible links.

The most striking finding provided by the present study is the amount of literature bias on this subject. Among the causes of study heterogeneity, there might be, e.g., geographical variations or the use of disparate investigative methods [55]. In the vast majority of cases, the ERT and CQT were significant. In many cases, the groups contained too few studies to quantify heterogeneity, and heterogeneity of the studies was classified as significant in seven out of 17 studies (41.2%). Such a high study heterogeneity confirmed that the random effect model was correctly chosen for the data analysis. However, the presence of a publication bias also hampered the correct quantification of the association between a viral infection and the risk of gastrointestinal disease. Such a problem was reflected, for instance, in the possible role of HBV and HPV, in the development of CRC, based on the colorectal tissues: the former had a highly significant association but only one publication supported it, and the latter had an association just above significance, but was substantiated by 11 publications. The trim-and-fill method was used to account for the publication bias in this meta-analysis, but it could only be computed in 10 cases out of 137 (7.3%). Moreover, the employment of the trim-and-fill method did not confirm the results obtained with the random effect model, creating a contrast that made it more challenging to interpret the results. Moreover, the measures that were significant and using both methods, increase the reliability of the risks found between the particular infections and gastrointestinal diseases.

The data presented by our meta-analysis confirm the role of the particular viral infections as risk factors for CRC and IBD. The primary finding of the current meta-analysis was that, using both the random effect model and the trim-and-fill model, the OR of an infection over the absence of an infection was significantly greater than the unity, indicating that the Herpesvirus HHV-4 (EBV) was a significant risk factor in the development of IBD. The present study's second finding was less accurate than the primary result, since it was based on the random effect model only. Still, it suggests HHV-5 as a risk factor for both CRC and IBD. The meta-analysis supported the tertiary findings that were not only based on a single model, but also hampered by a small sample set and, consequently, a high publication bias. Such results suggested that: (a) Polyomaviruses could be associated with a higher risk of both CRC and IBD; (b) HBV could be associated with a higher risk of CRC; (c) Papillomaviruses could not be ruled out as CRC risk factors since the association was just slightly above significance; and (d) respiratory, intestinal, and epithelial viruses had a very limited prognostic value on the risk of these. In contrast, there was not sufficient evidence to provide an association between a viral infection and the GC risk, due to the absence of a high enough number of strong studies.

HHV-4 (EBV) is typically associated with lymphomas, rather than gastrointestinal diseases, whereas HHV-5 is believed to cause chronic inflammation and immune dysfunction in immune-compromised people [56,57]. However, a recent MPS-based survey of tumoral tissue, demonstrated the presence of *Herpesviridae* in colorectal (HHV-4), gastric (HHV-4 and HHV-5) and liver (HHV-4, HHV-5) tissues [58]. These findings were confirmed by our own recent metagenomic analysis of CRC and the CRC metastasis tissues, which identified the presence of both HHV-4 and HHV-5 [59]. Remarkably, further recent studies showed a concomitant presence of HPV and HHV-4 in CRC. In a group of 102 CRC patients, 17% showed a concomitant presence of HPV and HHV-4, with a higher risk of tumor progression compared to patients infected with only one virus type [60], suggesting that HPV and EBV might interact synergistically, fostering the development of several types of cancer, including CRC [61]. In this context, it is established that the immune suppression triggers the reactivation of the latent viruses, such as HHV-5 [62,63].

It is well known that HHV-4 can disrupt the cell cycle at several points, fostering the genesis of cancer and even metastasis [61,64]. For instance, the EBV-derived LMP1 disrupts several cellular signal pathways (such as NF- κ B, PI₃K, and MAPK) [65] and reduces the expression of E-cadherin [66], which is also targeted by HHV-4's micro interfering RNAs, such as miR-BART9 [67]. LMP2A is associated with increased metastasis [68]. Additionally, HHV-4 interacts with HER-2, fostering an increased metastatic capacity [69] and the EBV-associated EBNA-1 is usually over-expressed in metastases [70]. Based on these observations, it has been suggested that HPV and HHV-4 might interact synergically, increasing the risk for carcinogenesis and metastasis. Towards this end, HHV-4 would act in the first steps of the cellular transformation, and HPV would continue to induce metastatic abilities [71]. The oncogenic mechanism of action of HHV-5 is still not completely understood, but it has been shown that an infection with this virus increases the viability and migration of the CRC-derived cell lines [72].

HHV-4 (EBV) has been shown to be more prevalent in IBD patients [73,74]. For instance, in one study, the prevalence of HHV-4 in IBD patients was 33%, as compared to 7% in the healthy controls [75]. Moreover, the presence of HHV-4 in the intestinal tissues is known to exacerbate the symptoms associated with IBD [76,77]. It is assumed that HHV-4 infects the T cells rather than the epithelial cells, the T-cells then infiltrate the intestinal tissues; thus, the inflamed tissues have a higher probability of carrying HHV-4 genomes [78]. In analyzing the CRC tissues for the HHV-4 presence, using in situ hybridization, our group observed signals consistent with the infection of lymphocytes rather than colonocytes, indeed [79]. Still, the detection of HHV-4 in tissues is tricky and requires ISH approaches that are not routinely performed [78]. Similarly, the detection of HHV-5 has not been standardized [80], increasing the discrepancies in the viral prevalence between the studies. This might explain why Herpesviruses are still not consistently observed in gastric or colorectal tissues, and/or in different frequencies or quality.

IBD is a CRC precursor [81]. Since IBD is treated with immunosuppression (for instance, azathioprine and infliximab) and EBV fosters in the absence of a fully functional immune system, the risk of viral-driven oncogenesis might be increased [82,83]. The reactivation of latent HHV-4 infections has been shown to cause lymphomas in IBD patients treated with azathioprine [84]. Moreover, the immunosuppression that results from the therapy of IBD, stimulates HHV-4 replication, provoking more inflammation [74,85]. This not only explains why IBD treatment is sometimes ineffective, due to the recurrence of inflammation, but also clarifies why the risk of developing lymphomas in IBD patients under immunosuppressive therapy, is high [86].

Therapy with immune suppressants, such as infliximab, is also involved in the reactivation of Hepadnaviruses [87]. The main member of this family, HBV, is the causative agent of hepatocellular carcinoma, rather than GC or CRC [88]. HBV causes about 1.5 million deaths yearly and IBD patients are at an increased risk of infection [89], as the present study has also confirmed. The X protein encoded by this virus is understood to disrupt the expression of several cellular genes involved in apoptosis, DNA repair, oxidative stress control, and immune response [90]. Remarkably, a HBV infection is treated routinely with immune suppressants. The reactivation of HBV, due to immunosuppression might have deleterious consequences for the patients. For instance, it has been reported that the high HBV serum titers are associated with a higher risk of metastasis in CRC [91]. Therefore, it is plausible to hypothesize that the use of immune suppressants in IBD patients with a latent HBV infection, might reactivate the virus, causing, as in the case of EBV, the instauration of chronic inflammation that will neutralize the effectiveness of the therapy and, in turn, foster the development of CRC. Consequently, it is recommended to screen IBD patients for the presence of a HBV infection [92].

As in the case of Herpesviruses and Hepadnaviruses, Polyomaviruses can also be reactivated by immunosuppressive therapy [93–95]. It has been suggested that latent JCV might be reactivated in cases of IBD [96]. JCV can cause IBD by altering the immune system [97]. Similarly, Papillomaviruses are not typically associated with gastrointestinal

diseases. Papillomaviruses are observed in over 36% of cervical and oropharyngeal cancers, in about 12% of anal cancers, but in quite variable frequencies in CRCs, for example less than 2% in one study [98]. In contrast, Kirgan and collaborators reported a viral prevalence of 97% in the CRC tissues and 23% in matched normal colon mucosa for HPV, with a significant association between a HPV infection and the CRC risk [99]. Based on such heterogeneous data, other meta-analyses confirmed a Papillomavirus prevalence of 42–83% in the CRC tissues [100–103], and reported ORs of 5-10 for the CRC risk upon infection with HPV [104,105], which is in line with the value of the 4.56 (95% CI: 0.99–21.03) we observed in our study.

About a decade ago, HHV-4 was described as a causative agent of the Epstein–Barr virus-positive mucocutaneous ulcer (EBVMCU), a condition affecting the pharyngeal and gastrointestinal mucosa, with ulcers refractory to treatment and displaying histological features typical of HHV-4 associated lymphomas [106,107]. EBVMCU has also been reported in the colon and rectum [108] as has been a lymphoma of the colon, symptomatically resembling IBD, caused by HHV-4 [109]. A HHV-4 infection of the gastrointestinal tract might trigger symptoms resembling IBD, leading to mistreatment that actually fosters the establishment of IBD [110]. In general, it is becoming clear that a HHV-4 infection of the gastrointestinal tract might not only make it difficult to distinguish it from IBD, but also increases the risk of mistreatment, treatment resistance, and cancer [111,112].

Taken together, these findings suggest that intestinal damage can be caused by viruses that induce chronic inflammation without directly residing in the tissues of the gastrointestinal tract. HHV-4 might reside in the immune cells, HHV-5 in several tissues, and HBV in the liver, still able to affect the gastrointestinal tract by causing a generalized chronic inflammation and aberrant immune response. This framework might explain why these viruses are not consistently found in CRC and IBD cases. Another explanation for the lack of a clear relationship between a viral infection and CRC, might be that viruses can cause 'hit-and-run' mutagenesis [113,114]. Remarkably, 'hit-and-run' instances have been described for both Herpesviruses and Papillomaviruses [115]. Mouse models have demonstrated how HHV-4 can cause lymphomas, but then the virus is lost, making virus detection difficult. This emphasizes the need to shift the focus of virus research from the positivity of cancer tissues and cells [116]. It is possible to imagine viruses infecting colonocytes, pushing the infected cells to transformation, but then being cleared from the tissue. In addition, the virus-caused chronic inflammation per se might foster carcinomas. This might be achieved not only by known viruses, but of course by yet not well described species which we could not analyze in our present meta-analysis, due to the fact that they are not yet annotated sufficiently (e.g., [35]).

The present review included the analysis of the MPS-derived studies as well. These, being qualitative in essence, could not be used for the quantification of the risk of gastrointestinal diseases upon infection. Although MPS-based studies provide a broader overview of the presence of microbes in samples, they are, at the moment, not suitable for quantitative analysis because they are not designed to calculate ORs. The statistical methods applied in these studies are designed to indicate trends in the presence of microbes in the samples, but not to reveal a measured risk associated with a disease. Nevertheless, the MPS-based studies, including our own, consistently reported a higher viral richness (the number of species) in the inflamed or tumor tissues [117]. A weakened immune system might trigger the reactivation of latent viruses already present in a tissue, increasing their viral load over the detection limit. In addition, inflammation might cause tissue damage [118,119] that can facilitate the infection of tissues and the expansion of viruses.

Conversely, MPS-derived studies including our recent one, reported a decreased bacterial richness as well [38,51,117]. This feature raises an apparent contradiction: if a dysfunctional immune system can foster a viral infection, why would it not do the same for bacteria? It might be speculated that, on the one hand, the reduction in bacterial species might be due to the reduction of commensal species, which do not thrive in inflamed tissues. On the other hand, the result of a viral expansion includes bacteriophages; thus,

it is plausible that a higher proportion of bacteria might be lysed in an inflamed tissue. Although our understanding of the dynamics underlying intestinal microbiota is still poor, it is assumed that the gut microbiome is enriched in lysogenic viruses [120]. Thus, it is interesting to hypothesize that a significant rise of phage quantity might occur in the inflamed tissues, including the tumor regions, reducing the amount, and maybe diversity, of the bacteria.

The present study had some limitations. First, the present meta-analysis considered each viral group as a whole regardless of the detection method employed. However, such heterogeneity might have introduced a bias in the results. Ideally, the prevalence obtained with one technique should be confirmed by another method, but this is rarely the case. Herein, the subdivision of the results by detection method, would have reduced the number of studies in each class, hampering their statistical evaluation. The present study aimed to identify the possible trends in the association between a viral infection and intestinal disease that should be further investigated at the experimental level. Second, it was also impossible to include every type of virus. Despite using the general term "virus" as a keyword during the literature search, viruses known to cause gastroenteritis during the acute phase of infection, such as the Orthomyxoviridae family, did not appear in the results. While a comprehensive review of the relationship between all types of viruses and intestinal diseases may never be complete, the lack of reference between an infection with viruses, such as orthomyxoviruses and CRC, GC, or IBD, suggests that the link is tenuous. Third, the lack of longitudinal data made it impossible to determine whether the infection was a cause or result of intestinal diseases. The presented study only reported a statistical association between an infection and disease; more detailed studies will be required to determine the causality comprehensively.

5. Conclusions

The present study investigated the prevalence of viruses by a meta-analysis of the actual status of relevant scientific publications in CRC, GC, and IBD. The meta-analysis highlights a significant publication bias in the literature related to this topic. Taking this into account, the present study suggests that HHV-4 (EBV) is a significant risk factor in the development of IBD, and points to HHV-5 as a risk factor for both CRC and IBD. Furthermore, the meta-analysis suggests that, to a less significant extent, Polyomaviruses might play a role in the risk of both CRC and IBD, and that HBV might be a risk factor for CRC.

Supplementary Materials: The following supporting information can be downloaded at: https://www. mdpi.com/article/10.3390/cancers14205085/s1, Table S1: Studies included in the present meta-analysis, including quantitative and qualitative assessment. References [38,48,49,51–55,73,75,93,99,100,121–303]; Table S2: NOS scores of the studies included in the present meta-analysis, including quantitative and qualitative assessment.

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References

- 1. Plummer, M.; de Martel, C.; Vignat, J.; Ferlay, J.; Bray, F.; Franceschi, S. Global Burden of Cancers Attributable to Infections in 2012: A Synthetic Analysis. *Lancet Glob. Health* **2016**, *4*, e609–e616. [CrossRef]
- 2. De Martel, C.; Georges, D.; Bray, F.; Ferlay, J.; Clifford, G.M. Global Burden of Cancer Attributable to Infections in 2018: A Worldwide Incidence Analysis. *Lancet Glob. Health* **2020**, *8*, e180–e190. [CrossRef]
- Oyervides-Muoz, M.A.; Prez-Maya, A.A.; Rodrguez-Gutirrez, H.F.; Gmez-Macias, G.S.; Fajardo-Ramrez, O.R.; Trevio, V.; Barrera-Saldaa, H.A.; Garza-Rodrguez, M.L. Understanding the HPV Integration and Its Progression to Cervical Cancer. *Infect. Genet. Evol.* 2018, 61, 134–144. [CrossRef] [PubMed]
- 4. Bucchi, D.; Stracci, F.; Buonora, N.; Masanotti, G. Human Papillomavirus and Gastrointestinal Cancer: A Review. *World J. Gastroenterol.* **2016**, *22*, 7415–7430. [CrossRef] [PubMed]
- Forman, D.; de Martel, C.; Lacey, C.J.; Soerjomatarama, I.; Lortet-Tieulent, J.; Bruni, L.; Vignat, J.; Ferlay, J.; Bray, F.; Plummer, M.; et al. Global Burden of Human Papillomavirus and Related Diseases. *Vaccine* 2012, 30, F12–F23. [CrossRef] [PubMed]
- Ferlay, J.; Colombet, M.; Soerjomataram, I.; Dyba, T.; Randi, G.; Bettio, M.; Gavin, A.; Visser, O.; Bray, F. Cancer Incidence and Mortality Patterns in Europe: Estimates for 40 Countries and 25 Major Cancers in 2018. *Eur. J. Cancer* 2018, 103, 356–387. [CrossRef]
- Ma, H.; Brosens, L.A.A.; Offerhaus, G.J.A.; Giardiello, F.M.; de Leng, W.W.J.; Montgomery, E.A. Pathology and Genetics of Hereditary Colorectal Cancer. *Pathology* 2018, 50, 49–59. [CrossRef]
- Müller, M.F.; Ibrahim, A.E.; Arends, M.J. Molecular Pathological Classification of Colorectal Cancer. Virchows Arch. 2016, 469, 125–134. [CrossRef]
- Rawla, P.; Barsouk, A. Epidemiology of Gastric Cancer: Global Trends, Risk Factors and Prevention. *Przeglad Gastroenterol.* 2019, 14, 26–38. [CrossRef]
- 10. Simon, K. Colorectal Cancer Development and Advances in Screening. Clin. Interv. Aging 2016, 11, 967–976. [CrossRef]
- 11. Hruby, A.; Hu, F.B. The Epidemiology of Obesity: A Big Picture. *PharmacoEconomics* 2015, 33, 673–689. [CrossRef]
- 12. Machlowska, J.; Baj, J.; Sitarz, M.; Maciejewski, R.; Sitarz, R. Gastric Cancer: Epidemiology, Risk Factors, Classification, Genomic Characteristics and Treatment Strategies. *Int. J. Mol. Sci.* 2020, *21*, 4012. [CrossRef]
- 13. Axelrad, J.E.; Lichtiger, S.; Yajnik, V. Inflammatory Bowel Disease and Cancer: The Role of Inflammation, Immunosuppression, and Cancer Treatment. *World J. Gastroenterol.* **2016**, *22*, 4794. [CrossRef]
- 14. Murata, M. Inflammation and Cancer. Environ. Health Prev. Med. 2018, 23, 50. [CrossRef]
- 15. Weedon, D.D.; Shorter, R.G.; Ilstrup, D.M.; Huizenga, K.A.; Taylor, W.F. Crohn's Disease and Cancer. *N. Engl. J. Med.* **1973**, *289*, 1099–1103. [CrossRef]
- 16. Gyde, S.N.; Prior, P.; Macartney, J.C.; Thompson, H.; Waterhouse, J.A.; Allan, R.N. Malignancy in Crohn's Disease. *Gut* **1980**, *21*, 1024–1029. [CrossRef]
- 17. Kalla, R.; Ventham, N.T.; Satsangi, J.; Arnott, I.D.R. Crohn's Disease. BMJ Online 2014, 349, g6670. [CrossRef]
- 18. Bounthavong, M.; Li, M.; Watanabe, J.H. An Evaluation of Health Care Expenditures in Crohn's Disease Using the United States Medical Expenditure Panel Survey from 2003 to 2013. *Res. Soc. Adm. Pharm.* **2017**, *13*, 530–538. [CrossRef]
- 19. Kappelman, M.D.; Moore, K.R.; Allen, J.K.; Cook, S.F. Recent Trends in the Prevalence of Crohn's Disease and Ulcerative Colitis in a Commercially Insured US Population. *Dig. Dis. Sci.* **2013**, *58*, 519–525. [CrossRef]
- Lucas, C.; Barnich, N.; Nguyen, H.T.T. Microbiota, Inflammation and Colorectal Cancer. Int. J. Mol. Sci. 2017, 18, 1310–1337. [CrossRef]
- 21. Hendler, S.A.; Barber, G.E.; Okafor, P.N.; Chang, M.S.; Limsui, D.; Limketkai, B.N. Cytomegalovirus Infection Is Associated with Worse Outcomes in Inflammatory Bowel Disease Hospitalizations Nationwide. *Int. J. Colorectal Dis.* **2020**, *35*, 897–903. [CrossRef]
- 22. Waldum, H.; Fossmark, R. Gastritis, Gastric Polyps and Gastric Cancer. Int. J. Mol. Sci. 2021, 22, 6548. [CrossRef]
- 23. Peng, C.; Ouyang, Y.; Lu, N.; Li, N. The NF-KB Signaling Pathway, the Microbiota, and Gastrointestinal Tumorigenesis: Recent Advances. *Front. Immunol.* **2020**, *11*, 1387. [CrossRef]
- 24. Ng, C.; Li, H.; Wu, W.K.K.; Wong, S.H.; Yu, J. Genomics and Metagenomics of Colorectal Cancer. J. Gastrointest. Oncol. 2019, 10, 1164–1170. [CrossRef] [PubMed]
- 25. Murphy, G.; Pfeiffer, R.; Camargo, M.C.; Rabkin, C.S. Meta-Analysis Shows That Prevalence of Epstein-Barr Virus-Positive Gastric Cancer Differs Based on Sex and Anatomic Location. *Gastroenterology* **2009**, *137*, 824–833. [CrossRef] [PubMed]
- 26. Collins, D.; Hogan, A.M.; Winter, D.C. Microbial and Viral Pathogens in Colorectal Cancer. *Lancet Oncol.* 2011, 12, 504–512. [CrossRef]
- 27. Tamboli, C.P.; Neut, C.; Desreumaux, P.; Colombel, J.F. Dysbiosis in Inflammatory Bowel Disease. *Gut* **2004**, *53*, 1. [CrossRef] [PubMed]

- Cougnoux, A.; Dalmasso, G.; Martinez, R.; Buc, E.; Delmas, J.; Gibold, L.; Sauvanet, P.; Darcha, C.; Déchelotte, P.; Bonnet, M.; et al. Bacterial Genotoxin Colibactin Promotes Colon Tumour Growth by Inducing a Senescence-Associated Secretory Phenotype. *Gut* 2014, 63, 1932–1942. [CrossRef] [PubMed]
- Tjalsma, H.; Boleij, A.; Marchesi, J.R.; Dutilh, B.E. A Bacterial Driver-Passenger Model for Colorectal Cancer: Beyond the Usual Suspects. *Nat. Rev. Microbiol.* 2012, 10, 575–582. [CrossRef]
- De Paoli, P.; Carbone, A. Carcinogenic Viruses and Solid Cancers without Sufficient Evidence of Causal Association. *Int. J. Cancer* 2013, 133, 1517–1529. [CrossRef]
- 31. Baandrup, L.; Thomsen, L.T.; Olesen, T.B.; Andersen, K.K.; Norrild, B.; Kjaer, S.K. The Prevalence of Human Papillomavirus in Colorectal Adenomas and Adenocarcinomas: A Systematic Review and Meta-Analysis. *Eur. J. Cancer* **2014**, *50*, 1446–1461. [CrossRef]
- Burnett-Hartman, A.N.; Newcomb, P.A.; Potter, J.D. Infectious Agents and Colorectal Cancer: A Review of *Helicobacter pylori*, Streptococcus Bovis, JC Virus, and Human Papillomavirus. *Cancer Epidemiol. Biomarkers Prev.* 2008, 17, 2970–2979. [CrossRef]
- Chen, H.; Chen, X.-Z.; Waterboer, T.; Castro, F.A.; Brenner, H. Viral Infections and Colorectal Cancer: A Systematic Review of Epidemiological Studies. Int. J. Cancer 2015, 137, 12–24. [CrossRef]
- Coelho, T.R.; Almeida, L.; Lazo, P.A. JC Virus in the Pathogenesis of Colorectal Cancer, an Etiological Agent or Another Component in a Multistep Process? *Virol. J.* 2010, 7, 1–8. [CrossRef]
- De Villiers, E.-M.; Gunst, K.; Chakraborty, D.; Ernst, C.; Bund, T.; zur Hausen, H. A Specific Class of Infectious Agents Isolated from Bovine Serum and Dairy Products and Peritumoral Colon Cancer Tissue. *Emerg. Microbes Infect.* 2019, *8*, 1205–1218. [CrossRef]
- Lepage, P.; Colombet, J.; Marteau, P.; Sime-Ngando, T. Dor Dysbiosis in Inflammatory Bowel Disease: A Role for Bacteriophages? Gut 2008, 57, 424–425. [CrossRef]
- 37. Zuo, T.; Lu, X.J.; Zhang, Y.; Cheung, C.P.; Lam, S.; Zhang, F.; Tang, W.; Ching, J.Y.L.; Zhao, R.; Chan, P.K.S.; et al. Gut Mucosal Virome Alterations in Ulcerative Colitis. *Gut* **2019**, *68*, 1169–1179. [CrossRef]
- Norman, J.M.; Handley, S.A.; Baldridge, M.T.; Droit, L.; Liu, C.Y.; Keller, B.C.; Kambal, A.; Monaco, C.L.; Zhao, G.; Fleshner, P.; et al. Disease-Specific Alterations in the Enteric Virome in Inflammatory Bowel Disease. *Cell* 2015, 160, 447–460. [CrossRef]
- 39. Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D.G. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. *J. Clin. Epidemiol.* **2009**, *62*, 1006–1012. [CrossRef]
- Higgins, J.P.T.; Thompson, S.G.; Deeks, J.J.; Altman, D.G. Measuring Inconsistency in Meta-Analyses. *BMJ* 2003, 327, 557–560. [CrossRef]
- Nguyen, N.H.; Singh, S. A Primer on Systematic Reviews and Meta-Analyses. Semin. Liver Dis. 2018, 38, 103–111. [CrossRef] [PubMed]
- 42. Egger, M.; Davey Smith, G.; Schneider, M.; Minder, C. Bias in Meta-Analysis Detected by a Simple, Graphical Test. *BMJ* **1997**, *315*, 629–634. [CrossRef] [PubMed]
- 43. Sedgwick, P. Meta-Analyses: What Is Heterogeneity? BMJ 2015, 350, h1435. [CrossRef] [PubMed]
- 44. Mantel, N.; Haenszel, W. Statistical Aspects of the Analysis of Data from Retrospective Studies of Disease. *J. Natl. Cancer Inst.* **1959**, *22*, 719–748.
- 45. Lin, L.; Chu, H. Quantifying Publication Bias in Meta-Analysis. *Biometrics* **2018**, *74*, 785–794. [CrossRef]
- Van der Willik, E.M.; van Zwet, E.W.; Hoekstra, T.; van Ittersum, F.J.; Hemmelder, M.H.; Zoccali, C.; Jager, K.J.; Dekker, F.W.; Meuleman, Y. Funnel Plots of Patient-Reported Outcomes to Evaluate Health-Care Quality: Basic Principles, Pitfalls and Considerations. *Nephrol. Carlton Vic.* 2021, 26, 95–104. [CrossRef]
- Lo, C.K.L.; Mertz, D.; Loeb, M. Newcastle-Ottawa Scale: Comparing Reviewers' to Authors' Assessments. BMC Med. Res. Methodol. 2014, 14, 45. [CrossRef]
- Perez-Brocal, V.; Garcia-Lopez, R.; Vazquez-Castellanos, J.F.; Nos, P.; Beltran, B.; Latorre, A.; Moya, A. Study of the Viral and Microbial Communities Associated with Crohn's Disease: A Metagenomic Approach. *Clin. Transl. Gastroenterol.* 2013, 4, e36. [CrossRef]
- Perez-Brocal, V.; Garcia-Lopez, R.; Nos, P.; Beltran, B.; Moret, I.; Moya, A. Metagenomic Analysis of Crohn's Disease Patients Identifies Changes in the Virome and Microbiome Related to Disease Status and Therapy, and Detects Potential Interactions and Biomarkers. *Inflamm. Bowel Dis.* 2015, *21*, 2515–2532. [CrossRef]
- Fernandes, M.A.; Verstraete, S.G.; Phan, T.G.; Deng, X.; Stekol, E.; LaMere, B.; Lynch, S.V.; Heyman, M.B.; Delwart, E. Enteric Virome and Bacterial Microbiota in Children with Ulcerative Colitis and Crohn Disease. *J. Pediatr. Gastroenterol. Nutr.* 2019, 68, 30–36. [CrossRef]
- Nakatsu, G.; Zhou, H.; Wu, W.K.K.; Wong, S.H.; Coker, O.O.; Dai, Z.; Li, X.; Szeto, C.H.; Sugimura, N.; Lam, T.Y.T.; et al. Alterations in Enteric Virome Are Associated with Colorectal Cancer and Survival Outcomes. *Gastroenterology* 2018, 155, 529–541.e5. [CrossRef]
- 52. Hannigan, G.D.; Duhaime, M.B.; Ruffin, M.T.; Koumpouras, C.C.; Schloss, P.D. Diagnostic Potential and Interactive Dynamics of the Colorectal Cancer Virome. *mBio* **2018**, *9*, 1–13. [CrossRef]
- 53. Wagner, J.; Maksimovic, J.; Farries, G.; Sim, W.H.; Bishop, R.F.; Cameron, D.J.; Catto-Smith, A.G.; Kirkwood, C.D. Bacteriophages in Gut Samples from Pediatric Crohn's Disease Patients. *Inflamm. Bowel Dis.* **2013**, *19*, 1598–1608. [CrossRef]

- Ungaro, F.; Massimino, L.; Furfaro, F.; Rimoldi, V.; Peyrin-Biroulet, L.; D'Alessio, S.; Danese, S. Metagenomic Analysis of Intestinal Mucosa Revealed a Specific Eukaryotic Gut Virome Signature in Early-Diagnosed Inflammatory Bowel Disease. *Gut Microbes* 2019, 10, 149–158. [CrossRef]
- 55. Zapatka, M.; Borozan, I.; Brewer, D.S.; Iskar, M.; Grundhoff, A.; Alawi, M.; Desai, N.; Sültmann, H.; Moch, H.; Cooper, C.S.; et al. The Landscape of Viral Associations in Human Cancers. *Nat. Genet.* **2020**, *52*, 320–330. [CrossRef]
- 56. Dickersin, K.; Min, Y.-I. Publication Bias: The Problem That Won't Go Away. Ann. N. Y. Acad. Sci. 1993, 703, 135–148. [CrossRef]
- 57. DeVito, N.J.; Goldacre, B. Catalogue of Bias: Publication Bias. BMJ Evid.-Based Med. 2019, 24, 53–54. [CrossRef]
- 58. Spineli, L.M.; Pandis, N. Publication Bias: Graphical and Statistical Methods. Am. J. Orthod. Dentofac. Orthop. Off. Publ. Am. Assoc. Orthod. Its Const. Soc. Am. Board Orthod. 2021, 159, 248–251. [CrossRef]
- 59. Fletcher, J. What Is Heterogeneity and Is It Important? BMJ 2007, 334, 94–96. [CrossRef]
- 60. Ibragimova, M.K.; Tsyganov, M.M.; Litviakov, N.V. Human Papillomavirus and Colorectal Cancer. *Med. Oncol. Northwood Lond. Engl.* **2018**, *35*, 140. [CrossRef]
- 61. Akram, N.; Imran, M.; Noreen, M.; Ahmed, F.; Atif, M.; Fatima, Z.; Bilal Waqar, A. Oncogenic Role of Tumor Viruses in Humans. *Viral Immunol.* 2017, *30*, 20–27. [CrossRef] [PubMed]
- 62. Sezgin, E.; An, P.; Winkler, C.A. Host Genetics of Cytomegalovirus Pathogenesis. Front. Genet. 2019, 10, 616. [CrossRef] [PubMed]
- Malki, M.I.; Gupta, I.; Fernandes, Q.; Aboulkassim, T.; Yasmeen, A.; Vranic, S.; Al Moustafa, A.-E.; Al-Thawadi, H.A. Co-Presence of Epstein-Barr Virus and High-Risk Human Papillomaviruses in Syrian Colorectal Cancer Samples. *Hum. Vaccines Immunother*. 2020, 16, 2403–2407. [CrossRef] [PubMed]
- 64. Marongiu, L.; Allgayer, H. Viruses in Colorectal Cancer. Mol. Oncol. 2022, 16, 1423–1450. [CrossRef]
- 65. Tsai, C.-L.; Li, H.-P.; Lu, Y.-J.; Hsueh, C.; Liang, Y.; Chen, C.-L.; Tsao, S.W.; Tse, K.-P.; Yu, J.-S.; Chang, Y.-S. Activation of DNA Methyltransferase 1 by EBV LMP1 Involves C-Jun NH(2)-Terminal Kinase Signaling. *Cancer Res.* 2006, 66, 11668–11676. [CrossRef]
- 66. Jeon, Y.K.; Lee, B.Y.; Kim, J.E.; Lee, S.S.; Kim, C.W. Molecular Characterization of Epstein-Barr Virus and Oncoprotein Expression in Nasopharyngeal Carcinoma in Korea. *Head Neck* 2004, 26, 573–583. [CrossRef]
- 67. Hsu, C.-Y.; Yi, Y.-H.; Chang, K.-P.; Chang, Y.-S.; Chen, S.-J.; Chen, H.-C. The Epstein-Barr Virus-Encoded MicroRNA MiR-BART9 Promotes Tumor Metastasis by Targeting E-Cadherin in Nasopharyngeal Carcinoma. *PLoS Pathog.* **2014**, *10*, e1003974. [CrossRef]
- 68. Lin, Z.; Wan, X.; Jiang, R.; Deng, L.; Gao, Y.; Tang, J.; Yang, Y.; Zhao, W.; Yan, X.; Yao, K.; et al. Epstein-Barr Virus-Encoded Latent Membrane Protein 2A Promotes the Epithelial-Mesenchymal Transition in Nasopharyngeal Carcinoma via Metastatic Tumor Antigen 1 and Mechanistic Target of Rapamycin Signaling Induction. J. Virol. 2014, 88, 11872–11885. [CrossRef]
- 69. Cyprian, F.S.; Al-Antary, N.; Al Moustafa, A.-E. HER-2/Epstein-Barr Virus Crosstalk in Human Gastric Carcinogenesis: A Novel Concept of Oncogene/Oncovirus Interaction. *Cell Adhes. Migr.* **2018**, *12*, 1–4. [CrossRef]
- Gaur, N.; Gandhi, J.; Robertson, E.S.; Verma, S.C.; Kaul, R. Epstein-Barr Virus Latent Antigens EBNA3C and EBNA1 Modulate Epithelial to Mesenchymal Transition of Cancer Cells Associated with Tumor Metastasis. *Tumour Biol. J. Int. Soc. Oncodevelopmen. Biol. Med.* 2015, *36*, 3051–3060. [CrossRef]
- Cyprian, F.S.; Al-Farsi, H.F.; Vranic, S.; Akhtar, S.; Al Moustafa, A.-E. Epstein-Barr Virus and Human Papillomaviruses Interactions and Their Roles in the Initiation of Epithelial-Mesenchymal Transition and Cancer Progression. *Front. Oncol.* 2018, *8*, 111. [CrossRef]
- 72. Teo, W.H.; Chen, H.-P.; Huang, J.C.; Chan, Y.-J. Human Cytomegalovirus Infection Enhances Cell Proliferation, Migration and Upregulation of EMT Markers in Colorectal Cancer-Derived Stem Cell-like Cells. *Int. J. Oncol.* **2017**, *51*, 1415–1426. [CrossRef]
- 73. Ryan, J.L.; Shen, Y.J.; Morgan, D.R.; Thorne, L.B.; Kenney, S.C.; Dominguez, R.L.; Gulley, M.L. Epstein-Barr Virus Infection Is Common in Inflamed Gastrointestinal Mucosa. *Dig. Sci.* 2012, *57*, 1887–1898. [CrossRef]
- Magro, F.; Santos-Antunes, J.; Albuquerque, A.; Vilas-Boas, F.; Macedo, G.N.; Nazareth, N.; Lopes, S.; Sobrinho-Simões, J.; Teixeira, S.; Dias, C.C.; et al. Epstein-Barr Virus in Inflammatory Bowel Disease-Correlation with Different Therapeutic Regimens. *Inflamm. Bowel Dis.* 2013, 19, 1710–1716. [CrossRef]
- 75. Li, X.; Chen, N.; You, P.; Peng, T.; Chen, G.; Wang, J.; Li, J.; Liu, Y. The Status of Epstein-Barr Virus Infection in Intestinal Mucosa of Chinese Patients with Inflammatory Bowel Disease. *Digestion* **2019**, *99*, 126–132. [CrossRef]
- 76. Núñez Ortiz, A.; Rojas Feria, M.; de la Cruz Ramírez, M.D.; Gómez Izquierdo, L.; Trigo Salado, C.; Herrera Justiniano, J.M.; Leo Carnerero, E. Impact of Epstein-Barr Virus Infection on Inflammatory Bowel Disease (IBD) Clinical Outcomes. *Rev. Espanola Enfermedades Dig. Organo Of. Soc. Espanola Patol. Dig.* 2022, 114, 259–265. [CrossRef]
- 77. Ciccocioppo, R.; Racca, F.; Paolucci, S.; Campanini, G.; Pozzi, L.; Betti, E.; Riboni, R.; Vanoli, A.; Baldanti, F.; Corazza, G.R. Human Cytomegalovirus and Epstein-Barr Virus Infection in Inflammatory Bowel Disease: Need for Mucosal Viral Load Measurement. World J. Gastroenterol. 2015, 21, 1915–1926. [CrossRef]
- 78. Lapsia, S.; Koganti, S.; Spadaro, S.; Rajapakse, R.; Chawla, A.; Bhaduri-McIntosh, S. Anti-TNFα Therapy for Inflammatory Bowel Diseases Is Associated with Epstein-Barr Virus Lytic Activation. *J. Med. Virol.* **2016**, *88*, 312–318. [CrossRef]
- Nissen, L.H.C.; Nagtegaal, I.D.; de Jong, D.J.; Kievit, W.; Derikx, L.A.A.P.; Groenen, P.J.T.A.; van Krieken, J.H.J.M.; Hoentjen, F. Epstein-Barr Virus in Inflammatory Bowel Disease: The Spectrum of Intestinal Lymphoproliferative Disorders. *J. Crohns Colitis* 2015, *9*, 398–403. [CrossRef]
- Ambelil, M.; Saulino, D.M.; Ertan, A.; DuPont, A.W.; Younes, M. The Significance of So-Called Equivocal Immunohistochemical Staining for Cytomegalovirus in Colorectal Biopsies. *Arch. Pathol. Lab. Med.* 2019, 143, 985–989. [CrossRef]

- Shah, S.C.; Itzkowitz, S.H. Colorectal Cancer in Inflammatory Bowel Disease: Mechanisms and Management. *Gastroenterology* 2022, 162, 715–730.e3. [CrossRef]
- Wu, S.; He, C.; Tang, T.-Y.; Li, Y.-Q. A Review on Co-Existent Epstein-Barr Virus-Induced Complications in Inflammatory Bowel Disease. *Eur. J. Gastroenterol. Hepatol.* 2019, *31*, 1085–1091. [CrossRef]
- Afzal, M.; Nigam, G.B. EBV Colitis with Ulcerative Colitis: A Double Whammy. BMJ Case Rep. 2018, 2018, bcr-2018-224963. [CrossRef]
- 84. Losco, A.; Gianelli, U.; Cassani, B.; Baldini, L.; Conte, D.; Basilisco, G. Epstein-Barr Virus-Associated Lymphoma in Crohn's Disease. *Inflamm. Bowel Dis.* 2004, *10*, 425–429. [CrossRef]
- Sankaran-Walters, S.; Ransibrahmanakul, K.; Grishina, I.; Hung, J.; Martinez, E.; Prindiville, T.; Dandekar, S. Epstein-Barr Virus Replication Linked to B Cell Proliferation in Inflamed Areas of Colonic Mucosa of Patients with Inflammatory Bowel Disease. J. Clin. Virol. Off. Publ. Pan Am. Soc. Clin. Virol. 2011, 50, 31–36. [CrossRef]
- 86. Subramaniam, K.; D'Rozario, J.; Pavli, P. Lymphoma and Other Lymphoproliferative Disorders in Inflammatory Bowel Disease: A Review. J. Gastroenterol. Hepatol. 2013, 28, 24–30. [CrossRef]
- Morisco, F.; Castiglione, F.; Rispo, A.; Stroffolini, T.; Sansone, S.; Vitale, R.; Guarino, M.; Biancone, L.; Caruso, A.; D'Inca, R.; et al. Effect of Immunosuppressive Therapy on Patients with Inflammatory Bowel Diseases and Hepatitis B or C Virus Infection. *J. Viral Hepat.* 2013, 20, 200–208. [CrossRef]
- 88. Seeger, C.; Mason, W.S. Molecular Biology of Hepatitis B Virus Infection. Virology 2015, 479–480, 672–686. [CrossRef]
- Jiang, H.-Y.; Wang, S.-Y.; Deng, M.; Li, Y.-C.; Ling, Z.-X.; Shao, L.; Ruan, B. Immune Response to Hepatitis B Vaccination among People with Inflammatory Bowel Diseases: A Systematic Review and Meta-Analysis. *Vaccine* 2017, 35, 2633–2641. [CrossRef]
- 90. Liu, S.; Koh, S.S.Y.; Lee, C.G.L. Hepatitis B Virus X Protein and Hepatocarcinogenesis. Int. J. Mol. Sci. 2016, 17, 940. [CrossRef]
- 91. Li, Z.; Li, S.; Tao, H.; Zhan, Y.; Ni, K.; Gong, J.; Li, G. Higher Titer Hepatitis B Core Antibody Predicts a Higher Risk of Liver Metastases and Worse Survival in Patients with Colorectal Cancer. *World J. Surg. Oncol.* **2021**, *19*, 251. [CrossRef] [PubMed]
- López-Serrano, P.; Pérez-Calle, J.L.; Sánchez-Tembleque, M.D. Hepatitis B and Inflammatory Bowel Disease: Role of Antiviral Prophylaxis. World J. Gastroenterol. 2013, 19, 1342–1348. [CrossRef]
- 93. Boltin, D.; Vilkin, A.; Levi, Z.; Elkayam, O.; Niv, Y. JC Virus T-Antigen DNA in Gastrointestinal Mucosa of Immunosuppressed Patients: A Prospective, Controlled Study. *Dig. Sci.* **2010**, *55*, 1975–1981. [CrossRef]
- Bellaguarda, E.; Keyashian, K.; Pekow, J.; Rubin, D.T.; Cohen, R.D.; Sakuraba, A. Prevalence of Antibodies Against JC Virus in Patients with Refractory Crohn's Disease and Effects of Natalizumab Therapy. *Clin. Gastroenterol. Hepatol. Off. Clin. Pract. J. Am. Gastroenterol. Assoc.* 2015, 13, 1919–1925. [CrossRef] [PubMed]
- 95. Verbeeck, J.; van Assche, G.; Ryding, J.; Wollants, E.; Rans, K.; Vermeire, S.; Pourkarim, M.R.; Noman, M.; Dillner, J.; van Ranst, M.; et al. JC Viral Loads in Patients with Crohn's Disease Treated with Immunosuppression: Can We Screen for Elevated Risk of Progressive Multifocal Leukoencephalopathy? *Gut* 2008, *57*, 1393–1397. [CrossRef] [PubMed]
- Bellizzi, A.; Barucca, V.; Di Nardo, G.; Fioriti, F.; Iebba, V.; Schippa, S.; Conte, M.P.; Proietti Checchi, M.; Colosimo, M.T.; Cucchiara, S.; et al. JC Viral Reactivation in a Pediatric Patient with Crohn's Disease. *Int. J. Immunopathol. Pharmacol.* 2010, 23, 955–959. [CrossRef] [PubMed]
- Altschuler, E.L. Is JC Polyoma Virus the Cause of Ulcerative Colitis and Multiple Sclerosis? *Med. Hypotheses* 2000, 55, 335–336. [CrossRef] [PubMed]
- 98. Razzaghi, H.; Saraiya, M.; Thompson, T.D.; Henley, S.J.; Viens, L.; Wilson, R. Five-Year Relative Survival for Human Papillomavirus-Associated Cancer Sites. *Cancer* **2018**, *124*, 203–211. [CrossRef]
- Kirgan, D.; Manalo, P.; McGregor, B. Immunohistochemical Demonstration of Human Papilloma Virus Antigen in Human Colon Neoplasms. J. Surg. Res. 1990, 48, 397–402. [CrossRef]
- Damin, D.C.; Caetano, M.B.; Rosito, M.A.; Schwartsmann, G.; Damin, A.S.; Frazzon, A.P.; Ruppenthal, R.D.; Alexandre, C.O.P. Evidence for an Association of Human Papillomavirus Infection and Colorectal Cancer. *Eur. J. Surg. Oncol.* 2007, 33, 569–574. [CrossRef]
- 101. Pelizzer, T.; Dias, C.P.; Poeta, J.; Torriani, T.; Roncada, C. Colorectal Cancer Prevalence Linked to Human Papillomavirus: A Systematic Review with Meta-Analysis. *Rev. Bras. Epidemiol. Braz. J. Epidemiol.* **2016**, *19*, 791–802. [CrossRef]
- 102. Lorenzon, L.; Ferri, M.; Pilozzi, E.; Torrisi, M.R.; Ziparo, V.; French, D. Human Papillomavirus and Colorectal Cancer: Evidences and Pitfalls of Published Literature. *Int. J. Colorectal Dis.* **2011**, *26*, 135–142. [CrossRef]
- 103. Roesch-Dietlen, F.; Cano-Contreras, A.D.; Sánchez-Maza, Y.J.; Espinosa-González, J.M.; Vázquez-Prieto, M.Á.; Valdés-de la, O.E.J.; Díaz-Roesch, F.; Carrasco-Arroniz, M.Á.; Cruz-Palacios, A.; Grube-Pagola, P.; et al. Frequency of human papillomavirus infection in patients with gastrointestinal cancer. *Rev. Gastroenterol. Mex. Engl.* 2018, *83*, 253–258. [CrossRef]
- Zhang, X.-H.; Wang, W.; Wang, Y.-Q.; Jia, D.-F.; Zhu, L. Human Papillomavirus Infection and Colorectal Cancer in the Chinese Population: A Meta-Analysis. *Colorectal Dis. Off. J. Assoc. Coloproctol. G. B. Irel.* 2018, 20, 961–969. [CrossRef]
- Chao, G.; Hong, X.; Chen, X.; Zhang, S. The Prevalence of Human Papillomavirus in Colorectal Cancer and Adenoma: A Meta-Analysis. J. Cancer Res. Ther. 2020, 16, 1656–1663. [CrossRef]
- 106. Dojcinov, S.D.; Venkataraman, G.; Raffeld, M.; Pittaluga, S.; Jaffe, E.S. EBV Positive Mucocutaneous Ulcer–a Study of 26 Cases Associated with Various Sources of Immunosuppression. *Am. J. Surg. Pathol.* **2010**, *34*, 405–417. [CrossRef]
- 107. Moran, N.R.; Webster, B.; Lee, K.M.; Trotman, J.; Kwan, Y.-L.; Napoli, J.; Leong, R.W. Epstein Barr Virus-Positive Mucocutaneous Ulcer of the Colon Associated Hodgkin Lymphoma in Crohn's Disease. World J. Gastroenterol. 2015, 21, 6072–6076. [CrossRef]

- 108. Juan, A.; Lobatón, T.; Tapia, G.; Mañosa, M.; Cabré, E.; Domènech, E. Epstein-Barr Virus-Positive Mucocutaneous Ulcer in Crohn's Disease. A Condition to Consider in Immunosuppressed IBD Patients. Dig. Liver Dis. Off. J. Ital. Soc. Gastroenterol. Ital. Assoc. Study Liver 2017, 49, 934–937. [CrossRef]
- Wang, Y.; Li, Y.; Meng, X.; Duan, X.; Wang, M.; Chen, W.; Tang, T.; Li, Y. Epstein-Barr Virus-Associated T-Cell Lymphoproliferative Disorder Presenting as Chronic Diarrhea and Intestinal Bleeding: A Case Report. *Front. Immunol.* 2018, *9*, 2583. [CrossRef]
- 110. Liu, Y.; Li, Y.; Li, Y.; Wu, S.; Tian, X.; Tang, T.; Sun, H.; He, C. Clinical Features of Intestinal Ulcers Complicated by Epstein-Barr Virus Infection: Importance of Active Infection. *Dis. Markers* **2021**, 2021, 6627620. [CrossRef]
- 111. Wang, Z.; Zhang, W.; Luo, C.; Zhu, M.; Zhen, Y.; Mu, J.; Zhang, Y.; Hu, R.; Wang, Y.; Wen, Z.; et al. Primary Intestinal Epstein-Barr Virus-Associated Natural Killer/T-Cell Lymphoproliferative Disorder: A Disease Mimicking Inflammatory Bowel Disease. J. Crohns Colitis 2018, 12, 896–904. [CrossRef] [PubMed]
- 112. Xu, W.; Jiang, X.; Chen, J.; Mao, Q.; Zhao, X.; Sun, X.; Zhong, L.; Rong, L. Chronic Active Epstein-Barr Virus Infection Involving Gastrointestinal Tract Mimicking Inflammatory Bowel Disease. *BMC Gastroenterol.* **2020**, *20*, 257. [CrossRef] [PubMed]
- 113. Ambinder, R.F. Gammaherpesviruses and "Hit-and-Run" Oncogenesis. Am. J. Pathol. 2000, 156, 1–3. [CrossRef]
- Skinner, G.R. Transformation of Primary Hamster Embryo Fibroblasts by Type 2 Simplex Virus: Evidence for a "Hit and Run" Mechanism. Br. J. Exp. Pathol. 1976, 57, 361–376. [PubMed]
- Gao, Y.; Lu, Y.-J.; Xue, S.-A.; Chen, H.; Wedderburn, N.; Griffin, B.E. Hypothesis: A Novel Route for Immortalization of Epithelial Cells by Epstein-Barr Virus. Oncogene 2002, 21, 825–835. [CrossRef] [PubMed]
- 116. Stevenson, P.G.; Simas, J.P.; Efstathiou, S. Immune Control of Mammalian Gamma-Herpesviruses: Lessons from Murid Herpesvirus-4. J. Gen. Virol. 2009, 90, 2317–2330. [CrossRef]
- Marongiu, L.; Landry, J.J.M.; Rausch, T.; Abba, M.L.; Delecluse, S.; Delecluse, H.-J.; Allgayer, H. Metagenomic Analysis of Primary Colorectal Carcinomas and Their Metastases Identifies Potential Microbial Risk Factors. *Mol. Oncol.* 2021, 15, 3363–3384. [CrossRef]
- Tatiya-Aphiradee, N.; Chatuphonprasert, W.; Jarukamjorn, K. Immune Response and Inflammatory Pathway of Ulcerative Colitis. J. Basic Clin. Physiol. Pharmacol. 2018, 30, 1–10. [CrossRef]
- 119. Glauser, M.P.; Meylan, P.; Bille, J. The Inflammatory Response and Tissue Damage. The Example of Renal Scars Following Acute Renal Infection. *Pediatr. Nephrol. Berl. Ger.* **1987**, *1*, 615–622. [CrossRef]
- 120. Mills, S.; Shanahan, F.; Stanton, C.; Hill, C.; Coffey, A.; Ross, R.P. Movers and Shakers: Influence of Bacteriophages in Shaping the Mammalian Gut Microbiota. *Gut Microbes* **2013**, *4*, 4–16. [CrossRef]
- 121. Aarnio, M.T.; Bohm, J.P.; Nuorva, K.P.; Pitkanen, R.I.; Kuopio, T.H.; Voutilainen, M.E. Absence of Cytomegalovirus from the Gastrointestinal Tract of Patients with Active Crohn's Disease. *In Vivo* **2012**, *26*, 151–155.
- 122. Abdel-Moneim, A.S.; El-Fol, H.A.; Kamel, M.M.; Soliman, A.S.; Mahdi, E.A.; El-Gammal, A.S.; Mahran, T.Z. Screening of Human Bocavirus in Surgically Excised Cancer Specimens. Arch. Virol 2016, 161, 2095–2102. [CrossRef]
- 123. Abdirad, A.; Ghaderi-Sohi, S.; Shuyama, K.; Koriyama, C.; Nadimi-Barforoosh, H.; Emami, S.; Mosavi-Jarrahi, A.; Nahvijou, A.; Akiba, S. Epstein-Barr Virus Associated Gastric Carcinoma: A Report from Iran in the Last Four Decades. *Diagn. Pathol.* 2007, 2, 25. [CrossRef]
- 124. Adams, D.J.; Nylund, C.M. Hospitalization for Varicella and Zoster in Children with Inflammatory Bowel Disease. J. Pediatr 2016, 171, 140–145. [CrossRef]
- 125. Afzal, M.A.; Armitage, E.; Ghosh, S.; Williams, L.C.; Minor, P.D. Further Evidence of the Absence of Measles Virus Genome Sequence in Full Thickness Intestinal Specimens from Patients with Crohn's Disease. J. Med. Virol. 2000, 62, 377–382. [CrossRef]
- 126. Afzal, M.A.; Armitage, E.; Begley, J.; Bentley, M.L.; Minor, P.D.; Ghosh, S.; Ferguson, A. Absence of Detectable Measles Virus Genome Sequence in Inflammatory Bowel Disease Tissues and Peripheral Blood Lymphocytes. J. Med. Virol. 1998, 55, 243–249. [CrossRef]
- 127. Aghakhani, A.; Hamkar, R.; Ramezani, A.; Bidari-Zerehpoosh, F.; Sabeti, S.; Ghavami, N.; Banifazl, M.; Rashidi, N.; Eslamifar, A. Lack of Human Papillomavirus DNA in Colon Adenocarcinama and Adenoma. *J. Cancer Res. Ther.* **2014**, *10*, 531–534. [CrossRef]
- 128. Ahmad, W.; Nguyen, N.H.; Boland, B.S.; Dulai, P.S.; Pride, D.T.; Bouland, D.; Sandborn, W.J.; Singh, S. Comparison of Multiplex Gastrointestinal Pathogen Panel and Conventional Stool Testing for Evaluation of Diarrhea in Patients with Inflammatory Bowel Diseases. *Dig. Sci.* **2019**, *64*, 382–390. [CrossRef]
- 129. Akintola-Ogunremi, O.; Luo, Q.; He, T.C.; Wang, H.L. Is Cytomegalovirus Associated with Human Colorectal Tumorigenesis? *Am. J. Clin. Pathol* 2005, 123, 244–249. [CrossRef]
- Alacam, S.; Karabulut, N.; Bakir, A.; Onel, M.; Buyuk, M.; Gulluoglu, M.; Agacfidan, A. Diagnostic Significance of Cytomegalovirus DNA Quantitation in Gastrointestinal Biopsies: Comparison with Histopathological Data and Blood Cytomegalovirus DNA. *Eur. J. Gastroenterol. Hepatol.* 2021, 33, 40–45. [CrossRef]
- 131. Alain, S.; Ducancelle, A.; Le Pors, M.J.S.; Mazeron, M.C.; de Saussure, P.; Bouhnik, Y.; Lavergne, A. Cytomegalovirus Infection in Patients with Active Inflammatory Bowel Disease. *J. Clin. Virol.* **2005**, *33*, 180–182. [CrossRef]
- 132. Alipov, G.; Nakayama, T.; Nakashima, M.; Wen, C.Y.; Niino, D.; Kondo, H.; Pruglo, Y.; Sekine, I. Epstein-Barr Virus-Associated Gastric Carcinoma in Kazakhstan. *World J. Gastroenterol.* **2005**, *11*, 27–30. [CrossRef]
- 133. Audeau, A.; Han, H.W.; Johnston, M.J.; Whitehead, M.W.; Frizelle, F.A. Does Human Papilloma Virus Have a Role in Squamous Cell Carcinoma of the Colon and Upper Rectum? *Eur. J. Surg. Oncol.* **2002**, *28*, 657–660. [CrossRef]

- 134. Axelrad, J.E.; Joelson, A.; Green, P.H.R.; Lawlor, G.; Lichtiger, S.; Cadwell, K.; Lebwohl, B. Enteric Infections Are Common in Patients with Flares of Inflammatory Bowel Disease. *Am. J. Gastroenterol.* **2018**, *113*, 1530–1539. [CrossRef]
- Axelrad, J.E.; Olen, O.; Askling, J.; Lebwohl, B.; Khalili, H.; Sachs, M.C.; Ludvigsson, J.F. Gastrointestinal Infection Increases Odds of Inflammatory Bowel Disease in a Nationwide Case-Control Study. *Clin. Gastroenterol. Hepatol.* 2019, 17, 1311. [CrossRef]
- 136. Balzola, F.A.; Khan, K.; Pera, A.; Bonino, F.; Pounder, R.E.; Wakefield, A.J. Measles IgM Immunoreactivity in Patients with Inflammatory Bowel Disease. *Ital. J. Gastroenterol. Hepatol.* **1998**, *30*, 378–382.
- 137. Baran, M.; Aksoy, B.; Vardı, K.; Appak, Y.Ç.; Öncel, E.K.; Çiftdoğan, D.Y. The Frequency and Importance of Cytomegalovirus and Epstein-Barr Virus Infections in Children with Inflammatory Bowel Disease: Single Center Experience. J. Pediatr. Infect. Cocuk Enfeksiyon Derg. 2018, 12, e140–e146. [CrossRef]
- Bellaguarda, E.; Pekow, J.; Cohen, R.D.; Rubin, D.T.; Sakuraba, A. Prevalence of Serum JC Virus Antibody in Refractory Crohn's Disease Patients. *Gastroenterology* 2014, 146, S580. [CrossRef]
- 139. Bender, C.; Zipeto, D.; Bidoia, C.; Costantini, S.; Zamo, A.; Menestrina, F.; Bertazzoni, U. Analysis of Colorectal Cancers for Human Cytomegalovirus Presence. *Infect. Agent Cancer* **2009**, *4*, 6. [CrossRef]
- Bernabe-Dones, R.D.; Gonzalez-Pons, M.; Villar-Prados, A.; Lacourt-Ventura, M.; Rodriguez-Arroyo, H.; Fonseca-Williams, S.; Velazquez, F.E.; Diaz-Algorri, Y.; Lopez-Diaz, S.M.; Rodriguez, N.; et al. High Prevalence of Human Papillomavirus in Colorectal Cancer in Hispanics: A Case-Control Study. *Gastroenterol Res. Pract.* 2016, 2016, 7896716. [CrossRef]
- 141. Bernstein, C.N.; Rawsthorne, P.; Blanchard, J.F. Population-Based Case-Control Study of Measles, Mumps, and Rubella and Inflammatory Bowel Disease. *Inflamm. Bowel Dis.* 2007, *13*, 759–762. [CrossRef] [PubMed]
- 142. Bertalot, G.; Villanacci, V.; Gramegna, M.; Orvieto, E.; Negrini, R.; Saleri, A.; Terraroli, C.; Ravelli, P.; Cestari, R.; Viale, G. Evidence of Epstein-Barr Virus Infection in Ulcerative Colitis. *Dig. Liver Dis.* 2001, *33*, 551–558. [CrossRef]
- Biancone, L.; DelleMonache, M.; Ricci, G.L.; Pallone, F. Hepatitis B and C Virus Infection in Crohn's Disease. *Gastroenterology* 2001, 7, 287–294. [CrossRef]
- 144. Bodaghi, S.; Yamanegi, K.; Xiao, S.Y.; Da Costa, M.; Palefsky, J.M.; Zheng, Z.M. Colorectal Papillomavirus Infection in Patients with Colorectal Cancer. *Clin. Cancer Res.* 2005, *11*, 2862–2867. [CrossRef] [PubMed]
- 145. Brichacek, B.; Hirsch, I.; Zavadova, H.; Prochazka, M.; Faltyn, J.; Vonka, V. Absence of Cytomegalovirus DNA from Adenocarcinoma of the Colon. *Intervirology* **1980**, *14*, 223–227. [CrossRef]
- 146. Burnett-Hartman, A.N.; Newcomb, P.A.; Mandelson, M.T.; Galloway, D.A.; Madeleine, M.M.; Wurscher, M.A.; Carter, J.J.; Makar, K.W.; Potter, J.D.; Schwartz, S.M. No Evidence for Human Papillomavirus in the Etiology of Colorectal Polyps. *Cancer Epidemiol Biomark. Prev.* 2011, 20, 2288–2297. [CrossRef] [PubMed]
- Burnett-Hartman, A.N.; Newcomb, P.A.; Schwartz, S.M.; Bostick, R.M.; Pawlita, M.; Waterboer, T.; Potter, J.D. No Association between Antibodies to Sexually Transmitted Infections and Colorectal Hyperplastic Polyps in Men: Minnesota Cancer Prevention Research Unit Polyp Study. *Cancer Epidemiol. Biomark. Prev.* 2012, 21, 1599–1601. [CrossRef]
- Burnett-Hartman, A.N.; Feng, Q.; Popov, V.; Kalidindi, A.; Newcomb, P.A. Human Papillomavirus DNA Is Rarely Detected in Colorectal Carcinomas and Not Associated with Microsatellite Instability: The Seattle Colon Cancer Family Registry. *Cancer Epidemiol. Biomark. Prev.* 2013, 22, 317–319. [CrossRef]
- Butt, J.; Romero-Hernandez, B.; Perez-Gomez, B.; Willhauck-Fleckenstein, M.; Holzinger, D.; Martin, V.; Moreno, V.; Linares, C.; Dierssen-Sotos, T.; Barricarte, A.; et al. Association of Streptococcus Gallolyticus Subspecies Gallolyticus with Colorectal Cancer: Serological Evidence. *Int. J. Cancer* 2016, 138, 1670–1679. [CrossRef]
- 150. Campello, C.; Comar, M.; Zanotta, N.; Minicozzi, A.; Rodella, L.; Poli, A. Detection of SV40 in Colon Cancer: A Molecular Case-Control Study from Northeast Italy. *J. Med. Virol.* **2010**, *82*, 1197–1200. [CrossRef]
- 151. Campello, C.; Comar, M.; D'Agaro, P.; Minicozzi, A.; Rodella, L.; Poli, A. A Molecular Case-Control Study of the Merkel Cell Polyomavirus in Colon Cancer. J. Med. Virol. 2011, 83, 721–724. [CrossRef]
- 152. Cardenas-Mondragon, M.G.; Torres, J.; Flores-Luna, L.; Camorlinga-Ponce, M.; Carreon-Talavera, R.; Gomez-Delgado, A.; Kasamatsu, E.; Fuentes-Panana, E.M. Case-Control Study of Epstein-Barr Virus and *Helicobacter pylori* Serology in Latin American Patients with Gastric Disease. *Br. J. Cancer* 2015, *112*, 1866–1873. [CrossRef]
- 153. Carrascal, E.; Koriyama, C.; Akiba, S.; Tamayo, O.; Itoh, T.; Eizuru, Y.; Garcia, F.; Sera, M.; Carrasquilla, G.; Piazuelo, M.B.; et al. Epstein-Barr Virus-Associated Gastric Carcinoma in Cali, Colombia. *Oncol Rep.* **2003**, *10*, 1059–1062. [CrossRef]
- 154. Casini, B.; Borgese, L.; Del Nonno, F.; Galati, G.; Izzo, L.; Caputo, M.; Donnorso, R.P.; Castelli, M.; Risuleo, G.; Visca, P. Presence and Incidence of DNA Sequences of Human Polyomaviruses BKV and JCV in Colorectal Tumor Tissues. *Anticancer Res.* 2005, 25, 1079–1085.
- 155. Chan, H.C.; Wong, V.W.; Wong, G.L.; Tang, W.; Wu, J.C.; Ng, S.C. Prevalence of Hepatitis B and Clinical Outcomes in Inflammatory Bowel Disease Patients in a Viral-Endemic Region. *BMC Gastroenterol.* **2016**, *16*, 100. [CrossRef]
- 156. Chen, H.P.; Jiang, J.K.; Chen, C.Y.; Yang, C.Y.; Chen, Y.C.; Lin, C.H.; Chou, T.Y.; Cho, W.L.; Chan, Y.J. Identification of Human Cytomegalovirus in Tumour Tissues of Colorectal Cancer and Its Association with the Outcome of Non-Elderly Patients. *J. Gen. Virol.* **2016**, *97*, 2411–2420. [CrossRef]
- Chen, H.P.; Jiang, J.K.; Lai, P.Y.; Teo, W.H.; Yang, C.Y.; Chou, T.Y.; Lin, C.H.; Chan, Y.J. Serological and Viraemic Status of Human Cytomegalovirus Infection in Patients with Colorectal Cancer Is Not Correlated with Viral Replication and Transcription in Tumours. J. Gen. Virol. 2016, 97, 152–159. [CrossRef]

- 158. Chen, D.; Luo, S.; Ben, Q.; Lu, L.; Wan, X.; Wu, J. Prevalence of Hepatitis B and C and Factors for Infection and Nonimmune in Inflammatory Bowel Disease Patients in China. *Eur. J. Gastroenterol. Hepatol.* **2017**, *29*, 509–515. [CrossRef]
- 159. Chevaux, J.B.; Nani, A.; Oussalah, A.; Venard, V.; Bensenane, M.; Belle, A.; Gueant, J.L.; Bigard, M.A.; Bronowicki, J.P.; Peyrin-Biroulet, L. Prevalence of Hepatitis B and C and Risk Factors for Nonvaccination in Inflammatory Bowel Disease Patients in Northeast France. *Inflamm. Bowel Dis.* 2010, *16*, 916–924. [CrossRef]
- 160. Cho, Y.J.; Chang, M.S.; Park, S.H.; Kim, H.S.; Kim, W.H. In Situ Hybridization of Epstein-Barr Virus in Tumor Cells and Tumor-Infiltrating Lymphocytes of the Gastrointestinal Tract. *Hum. Pathol.* **2001**, *32*, 297–301. [CrossRef]
- Coelho, T.R.; Gaspar, R.; Figueiredo, P.; Mendonca, C.; Lazo, P.A.; Almeida, L. Human JC Polyomavirus in Normal Colorectal Mucosa, Hyperplastic Polyps, Sporadic Adenomas, and Adenocarcinomas in Portugal. *J. Med. Virol.* 2013, 85, 2119–2127. [CrossRef]
- 162. Cohen, S.; Martinez-Vinson, C.; Aloi, M.; Turner, D.; Assa, A.; de Ridder, L.; Wolters, V.M.; de Meij, T.; Alvisi, P.; Bronsky, J.; et al. Cytomegalovirus Infection in Pediatric Severe Ulcerative Colitis-A Multicenter Study from the Pediatric Inflammatory Bowel Disease Porto Group of the European Society of Pediatric Gastroenterology, Hepatology and Nutrition. *Pediatr. Infect. J.* 2018, 37, 197–201. [CrossRef] [PubMed]
- 163. Dalla Libera, L.S.; de Siqueira, T.; Santos, I.L.; Porto Ramos, J.E.; Milhomen, A.X.; Alencar, R. de C.G. de; Rabelo Santos, S.H.; Dos Santos Carneiro, M.A.; Figueiredo Alves, R.R.; Saddi, V.A. Detection of Human Papillomavirus and the Role of P16INK4a in Colorectal Carcinomas. *PLoS ONE* 2020, *15*, e0235065. [CrossRef] [PubMed]
- 164. De Francisco, R.; Castaño-García, A.; Martínez-González, S.; Pérez-Martínez, I.; González-Huerta, A.J.; Morais, L.R.; Fernández-García, M.S.; Jiménez, S.; Díaz-Coto, S.; Flórez-Díez, P.; et al. Impact of Epstein-Barr Virus Serological Status on Clinical Outcomes in Adult Patients with Inflammatory Bowel Disease. *Aliment. Pharmacol. Ther.* 2018, 48, 723–730. [CrossRef] [PubMed]
- De Lima, M.A.; Ferreira, M.V.; Barros, M.A.; Pardini, M.I.; Ferrasi, A.C.; Rabenhorst, S.H. Epstein-Barr Virus-Associated Gastric Carcinoma in Brazil: Comparison between in Situ Hybridization and Polymerase Chain Reaction Detection. *Braz. J. Microbiol.* 2012, 43, 393–404. [CrossRef]
- 166. De Saussure, P.; Lavergne-Slove, A.; Mazeron, M.C.; Alain, S.; Matuchansky, C.; Bouhnik, Y. A Prospective Assessment of Cytomegalovirus Infection in Active Inflammatory Bowel Disease. *Aliment Pharmacol. Ther.* 2004, 20, 1323–1327. [CrossRef]
- De Souza, C.R.T.; Almeida, M.C.A.; Khayat, A.S.; da Silva, E.L.; Soares, P.C.; Chaves, L.C.; Burbano, R.M.R. Association between *Helicobacter pylori*, Epstein-Barr Virus, Human Papillomavirus and Gastric Adenocarcinomas. *World J. Gastroenterol.* 2018, 24, 4928–4938. [CrossRef]
- 168. De Villiers, E.M.; Schmidt, R.; Delius, H.; zur Hausen, H. Heterogeneity of TT Virus Related Sequences Isolated from Human Tumour Biopsy Specimens. *J. Mol. Med. Berl.* **2002**, *80*, 44–50. [CrossRef]
- 169. De Villiers, E.M. TTV Infection in Colorectal Cancer Tissues and Normal Mucosa 1. Int. J. Cancer 2007, 121, 2109–2112. [CrossRef]
- 170. Del Moral-Hernández, O.; Castañón-Sánchez, C.A.; Reyes-Navarrete, S.; Martínez-Carrillo, D.N.; Betancourt-Linares, R.; Jiménez-Wences, H.; de la Peña, S.; Román-Román, A.; Hernández-Sotelo, D.; Fernández-Tilapa, G. Multiple Infections by EBV, HCMV and *Helicobacter pylori* Are Highly Frequent in Patients with Chronic Gastritis and Gastric Cancer from Southwest Mexico: An Observational Study. *Med. Baltim.* 2019, 98, e14124. [CrossRef]
- 171. Deschoolmeester, V.; Van Marck, V.; Baay, M.; Weyn, C.; Vermeulen, P.; Van Marck, E.; Lardon, F.; Fontaine, V.; Vermorken, J.B. Detection of HPV and the Role of P16INK4A Overexpression as a Surrogate Marker for the Presence of Functional HPV Oncoprotein E7 in Colorectal Cancer. *BMC Cancer* **2010**, *10*, 117. [CrossRef]
- 172. Dimitroulia, E.; Spanakis, N.; Konstantinidou, A.E.; Legakis, N.J.; Tsakris, A. Frequent Detection of Cytomegalovirus in the Intestine of Patients with Inflammatory Bowel Disease. *Inflamm. Bowel Dis.* **2006**, *12*, 879–884. [CrossRef]
- 173. Dimitroulia, E.; Pitiriga, V.C.; Piperaki, E.T.; Spanakis, N.E.; Tsakris, A. Inflammatory Bowel Disease Exacerbation Associated with Epstein-Barr Virus Infection. *Colon Rectum.* **2013**, *56*, 322–327. [CrossRef]
- 174. Dimberg, J.; Hong, T.T.; Skarstedt, M.; Lofgren, S.; Zar, N.; Matussek, A. Detection of Cytomegalovirus DNA in Colorectal Tissue from Swedish and Vietnamese Patients with Colorectal Cancer. *Anticancer Res.* **2013**, *33*, 4947–4950.
- 175. Do Carmo, A.M.; Santos, F.M.; Ortiz-Agostinho, C.L.; Nishitokukado, I.; Frota, C.S.; Gomes, F.U.; Leite, A.Z.; Pannuti, C.S.; Boas, L.S.; Teixeira, M.G.; et al. Cytomegalovirus Infection in Inflammatory Bowel Disease Is Not Associated with Worsening of Intestinal Inflammatory Activity. *PLoS ONE* 2014, 9, e111574. [CrossRef]
- 176. El-Matary, W.; Stefanovici, C.; Van Caeseele, P.; Deora, V.; McCurdy, J. Detection of Cytomegalovirus in Colonic Mucosa of Children with Inflammatory Bowel Disease: Inflammatory Bowel Disease. J. Pediatr. Gastroenterol. Nutr. 2018, 67, 221–224. [CrossRef]
- 177. Enam, S.; Del Valle, L.; Lara, C.; Gan, D.D.; Ortiz-Hidalgo, C.; Palazzo, J.P.; Khalili, K. Association of Human Polyomavirus JCV with Colon Cancer: Evidence for Interaction of Viral T-Antigen and Beta-Catenin. *Cancer Res.* 2002, *62*, 7093–7101.
- 178. Esmailzadeh, N.; Ranaee, M.; Alizadeh, A.; Khademian, A.; Saber Amoli, S.; Sadeghi, F. Presence of JC Polyomavirus in Nonneoplastic Inflamed Colon Mucosa and Primary and Metastatic Colorectal Cancer. *Gastrointest. Tumors* 2020, 7, 30–40. [CrossRef]
- 179. Eyre-Brook, I.A.; Dundas, S. Incidence and Clinical Significance of Colonic Cytomegalovirus Infection in Idiopathic Inflammatory Bowel Disease Requiring Colectomy. *Gut* **1986**, 27, 1419–1425. [CrossRef]
- 180. Fahal, A.H.; el Razig, S.A.; Suliman, S.H.; Ibrahim, S.Z.; Tigani, A.E. Gastrointestinal Tract Cancer in Association with Hepatitis and HIV Infection. *East. Afr. Med. J.* **1995**, 72, 424–426.

- Farmer, G.W.; Vincent, M.M.; Fuccillo, D.A.; Horta-Barbosa, L.; Ritman, S.; Sever, J.L.; Gitnick, G.L. Viral Investigations in Ulcerative Colitis and Regional Enteritis. *Gastroenterology* 1973, 65, 8–18. [CrossRef]
- 182. Fiorina, L.; Ricotti, M.; Vanoli, A.; Luinetti, O.; Dallera, E.; Riboni, R.; Paolucci, S.; Brugnatelli, S.; Paulli, M.; Pedrazzoli, P.; et al. Systematic Analysis of Human Oncogenic Viruses in Colon Cancer Revealed EBV Latency in Lymphoid Infiltrates. *Infect. Agent Cancer* 2014, *9*, 18. [CrossRef]
- Flores, V.; Rodriguez-sanchez, B.; Marin-Jimenez, I.; Bouza, E.; Menchen, L.; Munoz, P. Prospective Study of Bk Virus Infection in Inflammatory Bowel Disease Patients. *Gastroenterology* 2014, 136, A190. [CrossRef]
- 184. Gauss, A.; Rosenstiel, S.; Schnitzler, P.; Hinz, U.; Rehlen, T.; Kadmon, M.; Ehehalt, R.; Stremmel, W.; Zawierucha, A. Intestinal Cytomegalovirus Infection in Patients Hospitalized for Exacerbation of Inflammatory Bowel Disease: A 10-Year Tertiary Referral Center Experience. Eur. J. Gastroenterol. Hepatol. 2015, 27, 712–720. [CrossRef]
- Gazzaz, F.; Mosli, M.H.; Jawa, H.; Sibiany, A. Detection of Human Papillomavirus Infection by Molecular Tests and Its Relation to Colonic Polyps and Colorectal Cancer. *Saudi Med. J.* 2016, *37*, 256–261. [CrossRef]
- Genitsch, V.; Novotny, A.; Seiler, C.A.; Kroll, D.; Walch, A.; Langer, R. Epstein-Barr Virus in Gastro-Esophageal Adenocarcinomas— Single Center Experiences in the Context of Current Literature. *Front. Oncol.* 2015, *5*, 73. [CrossRef]
- Giuliani, L.; Ronci, C.; Bonifacio, D.; Di Bonito, L.; Favalli, C.; Perno, C.F.; Syrjänen, K.; Ciotti, M. Detection of Oncogenic DNA Viruses in Colorectal Cancer. *Anticancer Res.* 2008, 28, 1405–1410.
- 188. Goel, A.; Li, M.S.; Nagasaka, T.; Shin, S.K.; Fuerst, F.; Ricciardiello, L.; Wasserman, L.; Boland, C.R. Association of JC Virus T-Antigen Expression with the Methylator Phenotype in Sporadic Colorectal Cancers. *Gastroenterology* 2006, 130, 1950–1961. [CrossRef]
- Gong, S.S.; Fan, Y.H.; Han, Q.Q.; Lv, B.; Xu, Y. Nested Case-Control Study on Risk Factors for Opportunistic Infections in Patients with Inflammatory Bowel Disease. World J. Gastroenterol. 2019, 25, 2240–2250. [CrossRef]
- Gonzalez, H.C.; Lamerato, L.; Rogers, C.G.; Gordon, S.C. Chronic Hepatitis C Infection as a Risk Factor for Renal Cell Carcinoma. Dig. Sci. 2015, 60, 1820–1824. [CrossRef]
- 191. Gordon, J.; Ramaswami, A.; Beuttler, M.; Jossen, J.; Pittman, N.; Lai, J.; Dunkin, D.; Benkov, K.; Dubinsky, M. EBV Status and Thiopurine Use in Pediatric IBD. *J. Pediatr. Gastroenterol. Nutr.* **2016**, *62*, 711–714. [CrossRef] [PubMed]
- 192. Gornick, M.C.; Castellsague, X.; Sanchez, G.; Giordano, T.J.; Vinco, M.; Greenson, J.K.; Capella, G.; Raskin, L.; Rennert, G.; Gruber, S.B.; et al. Human Papillomavirus Is Not Associated with Colorectal Cancer in a Large International Study. *Cancer Causes Control* **2010**, *21*, 737–743. [CrossRef] [PubMed]
- 193. Green, M.; Orth, G.; Wold, W.S.; Sanders, P.R.; Mackey, J.K.; Favre, M.; Croissant, O. Analysis of Human Cancers, Normal Tissues, and Verruce Plantares for DNA Sequences of Human Papillomavirus Types 1 and 2. *Virology* **1981**, *110*, 176–184. [CrossRef]
- Greenberg, H.B.; Gebhard, R.L.; McClain, C.J.; Soltis, R.D.; Kapikian, A.Z. Antibodies to Viral Gastroenteritis Viruses in Crohn's Disease. *Gastroenterology* 1979, 76, 349–350. [CrossRef]
- 195. Grinstein, S.; Preciado, M.V.; Gattuso, P.; Chabay, P.A.; Warren, W.H.; De Matteo, E.; Gould, V.E. Demonstration of Epstein-Barr Virus in Carcinomas of Various Sites. *Cancer Res.* 2002, 62, 4876–4878.
- 196. Gupta, I.; Al Farsi, H.; Jabeen, A.; Skenderi, F.; Al-Thawadi, H.; AlAhmad, Y.M.; Al Moustafa, A.-E.; Vranic, S. High-Risk Human Papillomaviruses and Epstein-Barr Virus in Colorectal Cancer and Their Association with Clinicopathological Status. *Pathogens* 2020, 9, 452. [CrossRef]
- 197. Haga, Y.; Funakoshi, O.; Kuroe, K.; Kanazawa, K.; Nakajima, H.; Saito, H.; Murata, Y.; Munakata, A.; Yoshida, Y. Absence of Measles Viral Genomic Sequence in Intestinal Tissues from Crohn's Disease by Nested Polymerase Chain Reaction. *Gut* 1996, 38, 211–215. [CrossRef]
- 198. Haghi-Navand, A.; Teimoori, A.; Makvandi, M.; Nisi, N.; Pourjabari, K. Study on JV Virus in Patients with Colon Cancer Type Adenocarcinoma. *Asian Pac. J. Cancer Prev.* 2019, 20, 1147–1151. [CrossRef]
- 199. Hamada, Y.; Nagata, N.; Nishijima, T.; Shimbo, T.; Asayama, N.; Kishida, Y.; Sekine, K.; Tanaka, S.; Aoki, T.; Watanabe, K.; et al. Impact of HIV Infection on Colorectal Tumors: A Prospective Colonoscopic Study of Asian Patients. J. Acquir. Immune Defic. Syndr. 2014, 65, 312–317. [CrossRef]
- Hampras, S.S.; Viscidi, R.P.; Helzlsouer, K.J.; Lee, J.H.; Fulp, W.J.; Giuliano, A.R.; Platz, E.A.; Rollison, D.E. Prospective Study of Seroreactivity to JC Virus T-Antigen and Risk of Colorectal Cancers and Adenomas. *Cancer Epidemiol. Biomark. Prev.* 2014, 23, 2591–2596. [CrossRef]
- 201. Harkins, L.; Volk, A.L.; Samanta, M.; Mikolaenko, I.; Britt, W.J.; Bland, K.I.; Cobbs, C.S. Specific Localisation of Human Cytomegalovirus Nucleic Acids and Proteins in Human Colorectal Cancer. *Lancet* 2002, *360*, 1557–1563. [CrossRef]
- Harsh, P.; Gupta, V.; Kedia, S.; Bopanna, S.; Pilli, S.; Surendernath; Makharia, G.K.; Ahuja, V. Prevalence of Hepatitis B, Hepatitis C and Human Immunodeficiency Viral Infections in Patients with Inflammatory Bowel Disease in North India. *Intest. Res.* 2017, 15, 97–102. [CrossRef]
- 203. Hart, H.; Neill, W.A.; Norval, M. Lack of Association of Cytomegalovirus with Adenocarcinoma of the Colon. *Gut* 1982, 23, 21–30. [CrossRef]
- 204. Hayashi, K.; Teramoto, N.; Akagi, T.; Sasaki, Y.; Suzuki, T. In Situ Detection of Epstein-Barr Virus in the Gastric Glands with Intestinal Metaplasia. *Am. J. Gastroenterol.* **1996**, *91*, 1481.
- 205. He, Y.; Xu, P.; Chen, Y.; Yang, R.; Chen, B.; Zeng, Z.; Chen, M. Prevalence and Influences of Hepatitis B Virus Infection on Inflammatory Bowel Disease: A Retrospective Study in Southern China. *Int. J. Clin. Exp. Med.* **2015**, *8*, 8078–8085.

- 206. Hernandez-Losa, J.; Fernandez-Soria, V.; Parada, C.; Sanchez-Prieto, R.; Ramon y Cajal, S. JC Virus and Human Colon Carcinoma: An Intriguing and Inconclusive Association. *Gastroenterology* **2003**, *124*, 268–270. [CrossRef]
- 207. Herrera-Goepfert, R.; Akiba, S.; Koriyama, C.; Ding, S.; Reyes, E.; Itoh, T.; Minakami, Y.; Eizuru, Y. Epstein-Barr Virus-Associated Gastric Carcinoma: Evidence of Age-Dependence among a Mexican Population. *World J. Gastroenterol.* 2005, 11, 6096–6103. [CrossRef]
- Hirata, T.; Nakamoto, M.; Nakamura, M.; Kinjo, N.; Hokama, A.; Kinjo, F.; Fujita, J. Low Prevalence of Human T Cell Lymphotropic Virus Type 1 Infection in Patients with Gastric Cancer. J. Gastroenterol. Hepatol. 2007, 22, 2238–2241. [CrossRef]
- 209. Hori, R.; Murai, Y.; Tsuneyama, K.; Abdel-Aziz, H.O.; Nomoto, K.; Takahashi, H.; Cheng, C.M.; Kuchina, T.; Harman, B.V.; Takano, Y. Detection of JC Virus DNA Sequences in Colorectal Cancers in Japan. *Virchows Arch.* **2005**, 447, 723–730. [CrossRef]
- Hradsky, O.; Copova, I.; Zarubova, K.; Durilova, M.; Nevoral, J.; Maminak, M.; Hubacek, P.; Bronsky, J. Seroprevalence of Epstein-Barr Virus, Cytomegalovirus, and Polyomaviruses in Children with Inflammatory Bowel Disease. *Dig. Sci.* 2015, 60, 3399–3407. [CrossRef]
- Hsieh, L.L.; Lin, P.J.; Chen, T.C.; Ou, J.T. Frequency of Epstein-Barr Virus-Associated Gastric Adenocarcinoma in Taiwan. *Cancer Lett.* 1998, 129, 125–129. [CrossRef]
- Huang, E.S.; Roche, J.K. Cytomegalovirus D.N.A. and Adenocarcinoma of the Colon: Evidence for Latent Viral Infection. *Lancet* 1978, 1, 957–960. [CrossRef]
- Iizuka, M.; Saito, H.; Yukawa, M.; Itou, H.; Shirasaka, T.; Chiba, M.; Fukushima, T.; Watanabe, S. No Evidence of Persistent Mumps Virus Infection in Inflammatory Bowel Disease. *Gut* 2001, 48, 637–641. [CrossRef]
- Ito, M.; Rodriguez-Bigas, M.A.; Creaven, P.J.; Petrelli, N.J. High Prevalence of Inoue-Melnick Virus Antibodies in Patients with Colorectal Carcinoma. *Cancer Lett.* 1992, 65, 233–237. [CrossRef]
- Jarzynski, A.; Zajac, P.; Zebrowski, R.; Boguszewska, A.; Polz-Dacewicz, M. Occurrence of BK Virus and Human Papilloma Virus in Colorectal Cancer. Ann. Agric. Env. Med. 2017, 24, 440–445. [CrossRef]
- Jung, W.T.; Li, M.S.; Goel, A.; Boland, C.R. JC Virus T-Antigen Expression in Sporadic Adenomatous Polyps of the Colon. *Cancer* 2008, 112, 1028–1036. [CrossRef]
- 217. Jung, Y.S.; Kim, N.H.; Park, J.H.; Park, D.I.; Sohn, C.I. Correlation between Hepatitis B Virus Infection and Colorectal Neoplasia. *J. Clin. Med.* 2019, *8*, 2085. [CrossRef]
- Kambham, N.; Vij, R.; Cartwright, C.A.; Longacre, T. Cytomegalovirus Infection in Steroid-Refractory Ulcerative Colitis: A Case-Control Study. Am. J. Surg. Pathol. 2004, 28, 365–373. [CrossRef]
- Kamiza, A.B.; Su, F.H.; Wang, W.C.; Sung, F.C.; Chang, S.N.; Yeh, C.C. Chronic Hepatitis Infection Is Associated with Extrahepatic Cancer Development: A Nationwide Population-Based Study in Taiwan. *BMC Cancer* 2016, 16, 861. [CrossRef]
- 220. Kane, S.P.; Nye, F.J.E.B. Virus Antibody in Crohn's Disease. Lancet 1971, 1, 233. [CrossRef]
- Karbalaie Niya, M.; Tameshkel, F.; Alemrajabi, M.; Taherizadeh, M.; Keshavarz, M.; Rezaee, M.; Kevyani, H. Molecular Survey on Merkel Cell Polyomavlrus in Patients with Colorectal Cancer. *Med. J. Indones.* 2018, 27, 229–236. [CrossRef]
- Karim, N.; Pallesen, G. Epstein-Barr Virus (EBV) and Gastric Carcinoma in Malaysian Patients. *Malays J. Pathol.* 2003, 25, 45–47. [PubMed]
- Karpinski, P.; Myszka, A.; Ramsey, D.; Kielan, W.; Sasiadek, M.M. Detection of Viral DNA Sequences in Sporadic Colorectal Cancers in Relation to CpG Island Methylation and Methylator Phenotype. *Tumour Biol.* 2011, 32, 653–659. [CrossRef] [PubMed]
- 224. Kattoor, J.; Koriyama, C.; Akiba, S.; Itoh, T.; Ding, S.; Eizuru, Y.; Abraham, E.K.; Chandralekha, B.; Amma, N.S.; Nair, M.K. Epstein-Barr Virus-Associated Gastric Carcinoma in Southern India: A Comparison with a Large-Scale Japanese Series. J. Med. Virol. 2002, 68, 384–389. [CrossRef] [PubMed]
- 225. Kawashima, H.; Mori, T.; Kashiwagi, Y.; Takekuma, K.; Hoshika, A.; Wakefield, A. Detection and Sequencing of Measles Virus from Peripheral Mononuclear Cells from Patients with Inflammatory Bowel Disease and Autism. *Dig. Sci.* 2000, 45, 723–729. [CrossRef] [PubMed]
- 226. Kayamba, V.; Monze, M.; Asombang, A.W.; Zyambo, K.; Kelly, P. Serological Response to Epstein-Barr Virus Early Antigen Is Associated with Gastric Cancer and Human Immunodeficiency Virus Infection in Zambian Adults: A Case-Control Study. *Pan. Afr. Med. J.* 2016, 23, 45. [CrossRef]
- 227. Keller, S.C.; Momplaisir, F.; Lo Re, V.; Newcomb, C.; Liu, Q.; Ratcliffe, S.J.; Long, J.A. Colorectal Cancer Incidence and Screening in US Medicaid Patients with and without HIV Infection. *Aids Care-Psychol. Socio-Med. Asp. AidsHiv* 2014, 26, 716–722. [CrossRef]
- 228. Khabaz, M.N.; Nedjadi, T.; Gari, M.A.; Atta, H.M.; Basuni, A.A.; Elderzi, D.A.; Bakarman, M. Simian Virus 40 Is Not Involved in the Development of Colorectal Adenocarcinoma. *Future Virol.* **2016**, *11*, 175–180. [CrossRef]
- 229. Kiewe, P.; Wojtke, S.; Thiel, E.; Nagorsen, D. Antiviral Cellular Immunity in Colorectal Cancer Patients. *Hum. Immunol.* 2009, 70, 85–88. [CrossRef]
- Kim, J.J.; Simpson, N.; Klipfel, N.; Debose, R.; Barr, N.; Laine, L. Cytomegalovirus Infection in Patients with Active Inflammatory Bowel Disease. *Dig. Sci.* 2010, 55, 1059–1065. [CrossRef]
- 231. Kim, E.S.; Cho, K.B.; Park, K.S.; Jang, B.I.; Kim, K.O.; Jeon, S.W.; Kim, E.Y.; Yang, C.H.; Kim, W.J. Prevalence of Hepatitis-B Viral Markers in Patients with Inflammatory Bowel Disease in a Hepatitis-B-Endemic Area: Inadequate Protective Antibody Levels in Young Patients. J. Clin. Gastroenterol. 2014, 48, 553–558. [CrossRef]

- Kishore, J.; Ghoshal, U.; Ghoshal, U.C.; Krishnani, N.; Kumar, S.; Singh, M.; Ayyagari, A. Infection with Cytomegalovirus in Patients with Inflammatory Bowel Disease: Prevalence, Clinical Significance and Outcome. *J. Med. Microbiol.* 2004, 53, 1155–1160. [CrossRef]
- Knoell, K.A.; Hendrix, J.D.; Stoler, M.H.; Patterson, J.W.; Montes, C.M. Absence of Human Herpesvirus 8 in Sarcoidosis and Crohn Disease Granulomas. Arch. Dermatol. 2005, 141, 909–910. [CrossRef]
- Knosel, T.; Schewe, C.; Dietel, M.; Petersen, I. Cytomegalovirus Is Not Associated with Progression and Metastasis of Colorectal Cancer. *Cancer Lett.* 2004, 211, 243–247. [CrossRef]
- Knosel, T.; Schewe, C.; Petersen, N.; Dietel, M.; Petersen, I. Prevalence of Infectious Pathogens in Crohn's Disease. *Pathol. Res. Pr.* 2009, 205, 223–230. [CrossRef]
- Kocoglu, H.; Karaca, M.; Tural, D.; Hocaoglu, E.; Okuturlar, Y.; Fetullahoglu, Z.; Gunaldi, M.; Ciftci, R.; Tuna, S.; Yucil, O.K.; et al. Hepatitis B and C Rates Are Significantly Increased in Certain Solid Tumors: A Large Retrospective Study. *J. Cancer Res. Ther.* 2018, 14, S774–S778. [CrossRef]
- 237. Kojima, T.; Watanabe, T.; Hata, K.; Shinozaki, M.; Yokoyama, T.; Nagawa, H. Cytomegalovirus Infection in Ulcerative Colitis. Scand J. Gastroenterol. 2006, 41, 706–711. [CrossRef]
- Kolho, K.L.; Klemola, P.; Simonen-Tikka, M.L.; Ollonen, M.L.; Roivainen, M. Enteric Viral Pathogens in Children with Inflammatory Bowel Disease. J. Med. Virol. 2012, 84, 345–347. [CrossRef]
- Kong, C.S.; Welton, M.L.; Longacre, T.A. Role of Human Papillomavirus in Squamous Cell Metaplasia-Dysplasia-Carcinoma of the Rectum. Am. J. Surg. Pathol. 2007, 31, 919–925. [CrossRef]
- 240. Ksiaa, F.; Ziadi, S.; Mokni, M.; Korbi, S.; Trimeche, M. The Presence of JC Virus in Gastric Carcinomas Correlates with Patient's Age, Intestinal Histological Type and Aberrant Methylation of Tumor Suppressor Genes. *Mod. Pathol.* 2010, 23, 522–530. [CrossRef]
- Ksiaa, F.; Allous, A.; Ziadi, S.; Mokni, M.; Trimeche, M. Assessment and Biological Significance of JC Polyomavirus in Colorectal Cancer in Tunisia. J. Buon 2015, 20, 762–769.
- 242. Laghi, L.; Randolph, A.E.; Chauhan, D.P.; Marra, G.; Major, E.O.; Neel, J.V.; Boland, C.R. JC Virus DNA Is Present in the Mucosa of the Human Colon and in Colorectal Cancers. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 7484–7489. [CrossRef]
- 243. Lavy, A.; Broide, E.; Reif, S.; Keter, D.; Niv, Y.; Odes, S.; Eliakim, R.; Halak, A.; Ron, Y.; Patz, J.; et al. Measles Is More Prevalent in Crohn's Disease Patients. A Multicentre Israeli Study. *Dig. Liver Dis.* **2001**, *33*, 472–476. [CrossRef]
- 244. Lee, Y.M.; Leu, S.Y.; Chiang, H.; Fung, C.P.; Liu, W.T. Human Papillomavirus Type 18 in Colorectal Cancer. J. Microbiol. Immunol. Infect. 2001, 34, 87–91.
- Leveque, N.; Brixi-Benmansour, H.; Reig, T.; Renois, F.; Talmud, D.; Brodard, V.; Coste, J.F.; De Champs, C.; Andréoletti, L.; Diebold, M.D. Low Frequency of Cytomegalovirus Infection during Exacerbations of Inflammatory Bowel Diseases. *J. Med. Virol.* 2010, 82, 1694–1700. [CrossRef]
- 246. Li, Y.; Wang, J.; Zhu, G.; Zhang, X.; Zhai, H.; Zhang, W.; Wang, W.; Huang, G. Detection of Parvovirus B19 Nucleic Acids and Expression of Viral VP1/VP2 Antigen in Human Colon Carcinoma. *Am. J. Gastroenterol.* **2007**, *102*, 1489–1498. [CrossRef]
- 247. Lin, P.Y.; Fung, C.Y.; Chang, F.P.; Huang, W.S.; Chen, W.C.; Wang, J.Y.; Chang, D. Prevalence and Genotype Identification of Human JC Virus in Colon Cancer in Taiwan. *J. Med. Virol.* **2008**, *80*, 1828–1834. [CrossRef]
- 248. Liu, F.; Mou, X.; Zhao, N.; Lin, J.; Teng, L.; Xiang, C. Prevalence of Human Papillomavirus in Chinese Patients with Colorectal Cancer. *Colorectal. Dis.* 2011, 13, 865–871. [CrossRef]
- 249. Lu, T.; Yang, Q.; Li, M.; Zhang, J.; Zou, J.; Huang, L.; Lin, J.; Jin, H.; He, J. HBV Infection and Extra-Hepatic Cancers in Adolescents and 20s: A Retrospective Study in China. *Cancer Epidemiol.* **2018**, *55*, 149–155. [CrossRef]
- Lundstig, A.; Stattin, P.; Persson, K.; Sasnauskas, K.; Viscidi, R.P.; Gislefoss, R.E.; Dillner, J. No Excess Risk for Colorectal Cancer among Subjects Seropositive for the JC Polyomavirus. *Int. J. Cancer* 2007, 121, 1098–1102. [CrossRef]
- 251. Lv, Y.-L.; Han, F.-F.; An, Z.-L.; Jia, Y.; Xuan, L.-L.; Gong, L.-L.; Zhang, W.; Ren, L.-L.; Yang, S.; Liu, H.; et al. Cytomegalovirus Infection Is a Risk Factor in Gastrointestinal Cancer: A Cross-Sectional and Meta-Analysis Study. *Intervirology* 2020, 63, 10–16. [CrossRef] [PubMed]
- Mackey, J.K.; Green, M.; Wold, W.S.; Rigden, P. Analysis of Human Cancer DNA for DNA Sequences of Human Adenovirus Type
 I. Natl. Cancer Inst. 1979, 62, 23–26. [PubMed]
- 253. Maconi, G.; Colombo, E.; Zerbi, P.; Sampietro, G.M.; Fociani, P.; Bosani, M.; Cassinotti, A.; Casini, V.; Russo, A.; Ardizzone, S.; et al. Prevalence, Detection Rate and Outcome of Cytomegalovirus Infection in Ulcerative Colitis Patients Requiring Colonic Resection. *Dig. Liver Dis.* 2005, *37*, 418–423. [CrossRef] [PubMed]
- 254. Malekpour Afshar, R.; Deldar, Z.; Mollaei, H.; Iranpour, M. Evaluation of HPV DNA Positivity in Colorectal Cancer Patients in Kerman, Southeast Iran. *Asian Pac. J. Cancer Prev.* 2018, 19, 193–198. [CrossRef]
- 255. Mariguela, V.C.; Chacha, S.G.; Cunha, A.; Troncon, L.E.; Zucoloto, S.; Figueiredo, L.T. Cytomegalovirus in Colorectal Cancer and Idiopathic Ulcerative Colitis. *Rev. Inst. Med. Trop Sao Paulo* 2008, *50*, 83–87. [CrossRef]
- Mehrabani-Khasraghi, S.; Ameli, M.; Khalily, F. Demonstration of Herpes Simplex Virus, Cytomegalovirus, and Epstein-Barr Virus in Colorectal Cancer. *Iran Biomed. J.* 2016, 20, 302–306. [CrossRef]
- 257. Militello, V.; Trevisan, M.; Squarzon, L.; Biasolo, M.A.; Rugge, M.; Militello, C.; Palu, G.; Barzon, L. Investigation on the Presence of Polyomavirus, Herpesvirus, and Papillomavirus Sequences in Colorectal Neoplasms and Their Association with Cancer. *Int. J. Cancer* 2009, 124, 2501–2503. [CrossRef]

- 258. Montgomery, S.M.; Morris, D.L.; Pounder, R.E.; Wakefield, A.J. Paramyxovirus Infections in Childhood and Subsequent Inflammatory Bowel Disease. *Gastroenterology* **1999**, *116*, 796–803. [CrossRef]
- Morewaya, J.; Koriyama, C.; Akiba, S.; Shan, D.; Itoh, T.; Eizuru, Y. Epstein-Barr Virus-Associated Gastric Carcinoma in Papua New Guinea. Oncol. Rep. 2004, 12, 1093–1098. [CrossRef]
- Mou, X.; Chen, L.; Liu, F.; Lin, J.; Diao, P.; Wang, H.; Li, Y.; Lin, J.; Teng, L.; Xiang, C. Prevalence of JC Virus in Chinese Patients with Colorectal Cancer. *PLoS ONE* 2012, 7, e35900. [CrossRef]
- 261. Newcomb, P.A.; Bush, A.C.; Stoner, G.L.; Lampe, J.W.; Potter, J.D.; Bigler, J. No Evidence of an Association of JC Virus and Colon Neoplasia. *Cancer Epidemiol. Biomark. Prev.* 2004, 13, 662–666. [CrossRef]
- Niv, Y.; Vilkin, A.; Brenner, B.; Kendel, Y.; Morgenstern, S.; Levi, Z. HMLH1 Promoter Methylation and JC Virus T Antigen Presence in the Tumor Tissue of Colorectal Cancer Israeli Patients of Different Ethnic Groups. *Eur. J. Gastroenterol. Hepatol.* 2010, 22, 938–941. [CrossRef]
- 263. Nosho, K.; Yamamoto, H.; Takahashi, T.; Mikami, M.; Hizaki, K.; Maehata, T.; Taniguchi, H.; Yamaoka, S.; Adachi, Y.; Itoh, F.; et al. Correlation of Laterally Spreading Type and JC Virus with Methylator Phenotype Status in Colorectal Adenoma. *Hum. Pathol.* 2008, 39, 767–775. [CrossRef]
- 264. Nosrati, A.; Naghshvar, F.; Torabizadeh, Z.; Haghshenas, M.; Sangsefidi, H. Relationship between Human Papilloma Virus and Colorectal Cancer in Northern Iran. *Middle East J. Cancer* **2015**, *6*, 237–241.
- Oda, K.; Koda, K.; Takiguchi, N.; Nunomura, M.; Seike, K.; Miyazaki, M. Detection of Epstein-Barr Virus in Gastric Carcinoma Cells and Surrounding Lymphocytes. *Gastric. Cancer* 2003, *6*, 173–178. [CrossRef]
- Perez, L.O.; Abba, M.C.; Laguens, R.M.; Golijow, C.D. Analysis of Adenocarcinoma of the Colon and Rectum: Detection of Human Papillomavirus (HPV) DNA by Polymerase Chain Reaction. *Colorectal. Dis.* 2005, 7, 492–495. [CrossRef]
- 267. Pironi, L.; Bonvicini, F.; Gionchetti, P.; D'Errico, A.; Rizzello, F.; Corsini, C.; Foroni, L.; Gallinella, G. Parvovirus B19 Infection Localized in the Intestinal Mucosa and Associated with Severe Inflammatory Bowel Disease. J. Clin. Microbiol. 2009, 47, 1591–1595. [CrossRef]
- Roblin, X.; Pillet, S.; Oussalah, A.; Berthelot, P.; Del Tedesco, E.; Phelip, J.M.; Chambonnière, M.L.; Garraud, O.; Peyrin-Biroulet, L.; Pozzetto, B. Cytomegalovirus Load in Inflamed Intestinal Tissue Is Predictive of Resistance to Immunosuppressive Therapy in Ulcerative Colitis. *Am. J. Gastroenterol.* 2011, 106, 2001–2008. [CrossRef]
- Roblin, X.; Pillet, S.; Berthelot, P.; Del Tedesco, E.; Phelip, J.M.; Chambonnière, M.L.; Peyrin-Biroulet, L.; Pozzetto, B. Prevalence of Cytomegalovirus Infection in Steroid-Refractory Crohn's Disease. *Inflamm. Bowel Dis.* 2012, *18*, E1396–E1397. [CrossRef]
- 270. Roche, J.K.; Cheung, K.S.; Boldogh, I.; Huang, E.S.; Lang, D.J. Cytomegalovirus: Detection in Human Colonic and Circulating Mononuclear Cells in Association with Gastrointestinal Disease. *Int. J. Cancer* **1981**, 27, 659–667. [CrossRef]
- 271. Rollison, D.E.; Helzlsouer, K.J.; Lee, J.H.; Fulp, W.; Clipp, S.; Hoffman-Bolton, J.A.; Giuliano, A.R.; Platz, E.A.; Viscidi, R.P. Prospective Study of JC Virus Seroreactivity and the Development of Colorectal Cancers and Adenomas. *Cancer Epidemiol. Biomark. Prev.* 2009, *18*, 1515–1523. [CrossRef]
- 272. Ruger, R.; Fleckenstein, B. Cytomegalovirus DNA in Colorectal Carcinoma Tissues. Klin. Wochenschr. 1985, 63, 405–408. [CrossRef]
- 273. Salepci, T.; Yazici, H.; Dane, F.; Topuz, E.; Dalay, N.; Onat, H.; Aykan, F.; Seker, M.; Aydiner, A. Detection of Human Papillomavirus DNA by Polymerase Chain Reaction and Southern Blot Hybridization in Colorectal Cancer Patients. J. Buon 2009, 14, 495–499.
- 274. Samaka, R.M.; Abd El-Wahed, M.M.; Aiad, H.A.; Kandil, M.A.; Al-Sharaky, D.R. Does JC Virus Have a Role in the Etiology and Prognosis of Egyptian Colorectal Carcinoma? *Apmis* 2013, 121, 316–328. [CrossRef]
- 275. Sarvari, J.; Mahmoudvand, S.; Pirbonyeh, N.; Safaei, A.; Hosseini, S.Y. The Very Low Frequency of Epstein-Barr JC and BK Viruses DNA in Colorectal Cancer Tissues in Shiraz, Southwest Iran. *Pol. J. Microbiol.* 2018, 67, 73–79. [CrossRef]
- Schildgen, V.; Malecki, M.; Tillmann, R.L.; Brockmann, M.; Schildgen, O. The Human Bocavirus Is Associated with Some Lung and Colorectal Cancers and Persists in Solid Tumors. *PLoS ONE* 2013, 8, e68020. [CrossRef]
- Shah, K.V.; Daniel, R.W.; Simons, J.W.; Vogelstein, B. Investigation of Colon Cancers for Human Papillomavirus Genomic Sequences by Polymerase Chain Reaction. J. Surg. Oncol. 1992, 51, 5–7. [CrossRef]
- 278. Shibata, D.; Hawes, D.; Stemmermann, G.N.; Weiss, L.M. Epstein-Barr Virus-Associated Gastric Adenocarcinoma among Japanese Americans in Hawaii. *Cancer Epidemiol. Biomark. Prev.* **1993**, *2*, 213–217.
- 279. Sinagra, E.; Raimondo, D.; Gallo, E.; Stella, M.; Cottone, M.; Orlando, A.; Rossi, F.; Orlando, E.; Messina, M.; Tomasello, G.; et al. Could JC Virus Provoke Metastasis in Colon Cancer? *World J. Gastroenterol.* 2014, 20, 15745–15749. [CrossRef]
- 280. Su, F.-H.; Le, T.N.; Muo, C.-H.; Te, S.A.; Sung, F.-C.; Yeh, C.-C. Chronic Hepatitis B Virus Infection Associated with Increased Colorectal Cancer Risk in Taiwanese Population. *Viruses* **2020**, *12*, 97. [CrossRef]
- Sura, R.; Gavrilov, B.; Flamand, L.; Ablashi, D.; Cartun, R.; Colombel, J.F.; Van Kruiningen, H.J. Human Herpesvirus-6 in Patients with Crohn's Disease. *Applies* 2010, 118, 394–400. [CrossRef] [PubMed]
- Tafvizi, F.; Fard, Z.T. Detection of Human Cytomegalovirus in Patients with Colorectal Cancer by Nested-PCR. Asian Pac. J. Cancer Prev 2014, 15, 1453–1457. [CrossRef] [PubMed]
- Tafvizi, F.; Fard, Z.T.; Assareh, R. Epstein-Barr Virus DNA in Colorectal Carcinoma in Iranian Patients. *Pol. J. Pathol* 2015, 66, 154–160. [CrossRef] [PubMed]
- Taherian, H.; Tafvizi, F.; Fard, Z.T.; Abdirad, A. Lack of Association between Human Papillomavirus Infection and Colorectal Cancer. Prz. Gastroenterol. 2014, 9, 280–284. [CrossRef] [PubMed]

- Theodoropoulos, G.; Panoussopoulos, D.; Papaconstantinou, I.; Gazouli, M.; Perdiki, M.; Bramis, J.; Lazaris, A. Assessment of JC Polyoma Virus in Colon Neoplasms. *Colon. Rectum* 2005, 48, 86–91. [CrossRef] [PubMed]
- Tolentino, Y.F.; Fogaca, H.S.; Zaltman, C.; Ximenes, L.L.; Coelho, H.S. Hepatitis B Virus Prevalence and Transmission Risk Factors in Inflammatory Bowel Disease Patients at Clementino Fraga Filho University Hospital. World J. Gastroenterol. 2008, 14, 3201–3206. [CrossRef]
- 287. Toumi, W.; Ripalti, A.; Ricciardiello, L.; Cherif, A.; Gargouri, D.; Bouhafa, A.; Kharrat, J.; Jarboui, S.; Benrhouma, H.; Zili, M.; et al. Detection of a New JCV Strain of Genotype A in a Subpopulation of Colorectal Adenocarcinomas in Tunisia. *New Microbiol.* 2017, 40, 99–106.
- 288. Trimeche, M.; Ksiaa, F.; Ziadi, S.; Mestiri, S.; Hachana, M.; Gacem, R.B.; Sriha, B.; Korbi, S. Prevalence and Characteristics of Epstein-Barr Virus-Associated Gastric Carcinomas in Tunisia. *Eur. J. Gastroenterol. Hepatol.* 2009, 21, 1001–1007. [CrossRef]
- Truong, C.D.; Feng, W.; Li, W.; Khoury, T.; Li, Q.; Alrawi, S.; Yu, Y.; Xie, K.; Yao, J.; Tan, D. Characteristics of Epstein-Barr Virus-Associated Gastric Cancer: A Study of 235 Cases at a Comprehensive Cancer Center in U.S.A. *J. Exp. Clin. Cancer Res.* 2009, 28, 14. [CrossRef]
- 290. Tsai, S.Y.; Yang, T.Y.; Lin, C.L.; Tsai, Y.H.; Kuo, C.F.; Kao, C.H. Increased Risk of Varicella Zoster Virus Infection in Inflammatory Bowel Disease in an Asian Population: A Nationwide Population-Based Cohort Study. Int. J. Clin. Pract. 2015, 69, 228–234. [CrossRef]
- 291. Vanoli, A.; Di Sabatino, A.; Martino, M.; Dallera, E.; Furlan, D.; Mescoli, C.; Macciomei, M.C.; Biancone, L.; Neri, B.; Grillo, F.; et al. Epstein-Barr Virus-Positive Ileal Carcinomas Associated with Crohn's Disease. *Virchows Arch.* 2017, 471, 549–552. [CrossRef]
- 292. Vega, R.; Bertran, X.; Menacho, M.; Domenech, E.; de Vega, V.M.; Hombrados, M.; Ojanguren, I.; Gassull, M.A. Cytomegalovirus Infection in Patients with Inflammatory Bowel Disease. *Am. J. Gastroenterol.* **1999**, *94*, 1053–1056. [CrossRef]
- 293. Vilkin, A.; Ronen, Z.; Levi, Z.; Morgenstern, S.; Halpern, M.; Niv, Y. Presence of JC Virus DNA in the Tumor Tissue and Normal Mucosa of Patients with Sporadic Colorectal Cancer (CRC) or with Positive Family History and Bethesda Criteria. *Dig. Sci.* 2012, 57, 79–84. [CrossRef]
- 294. Wakefield, A.J.; Fox, J.D.; Sawyerr, A.M.; Taylor, J.E.; Sweenie, C.H.; Smith, M.; Emery, V.C.; Hudson, M.; Tedder, R.S.; Pounder, R.E. Detection of Herpesvirus DNA in the Large Intestine of Patients with Ulcerative Colitis and Crohn's Disease Using the Nested Polymerase Chain Reaction. J. Med. Virol. 1992, 38, 183–190. [CrossRef]
- 295. Weinreb, D.B. Re: Detection of JC Virus Sequences in Colorectal Cancers in Japan. Virchows Arch. 2006, 448, 239. [CrossRef]
- 296. Wong, N.A.; Herbst, H.; Herrmann, K.; Kirchner, T.; Krajewski, A.S.; Moorghen, M.; Niedobitek, F.; Rooney, N.; Shepherd, N.A.; Niedobitek, G. Epstein-Barr Virus Infection in Colorectal Neoplasms Associated with Inflammatory Bowel Disease: Detection of the Virus in Lymphomas but Not in Adenocarcinomas. J. Pathol. 2003, 201, 312–318. [CrossRef]
- Yanai, H.; Takada, K.; Shimizu, N.; Mizugaki, Y.; Tada, M.; Okita, K. Epstein-Barr Virus Infection in Non-Carcinomatous Gastric Epithelium. J. Pathol. 1997, 183, 293–298. [CrossRef]
- Yanai, H.; Shimizu, N.; Nagasaki, S.; Mitani, N.; Okita, K. Epstein-Barr Virus Infection of the Colon with Inflammatory Bowel Disease. Am. J. Gastroenterol. 1999, 94, 1582–1586. [CrossRef]
- Yavuzer, D.; Karadayi, N.; Salepci, T.; Baloglu, H.; Dabak, R.; Bayramicli, O.U. Investigation of Human Papillomavirus DNA in Colorectal Carcinomas and Adenomas. *Med. Oncol.* 2011, 28, 127–132. [CrossRef]
- 300. Yi, F.; Zhao, J.; Luckheeram, R.V.; Lei, Y.; Wang, C.; Huang, S.; Song, L.; Wang, W.; Xia, B. The Prevalence and Risk Factors of Cytomegalovirus Infection in Inflammatory Bowel Disease in Wuhan, Central China. *Virol. J.* 2013, 10, 43. [CrossRef]
- Yuen, S.T.; Chung, L.P.; Leung, S.Y.; Luk, I.S.; Chan, S.Y.; Ho, J. In Situ Detection of Epstein-Barr Virus in Gastric and Colorectal Adenocarcinomas. *Am. J. Surg. Pathol.* 1994, 18, 1158–1163. [CrossRef]
- Yunos, A.M.; Jaafar, H.; Idris, F.M.; Kaur, G.; Mabruk, M.J. Detection of Epstein-Barr Virus in Lower Gastrointestinal Tract Lymphomas: A Study in Malaysian Patients. *Mol. Diagn. Ther.* 2006, 10, 251–256. [CrossRef]
- 303. Zagorowicz, E.; Bugajski, M.; Wieszczy, P.; Pietrzak, A.; Magdziak, A.; Mroz, A. Cytomegalovirus Infection in Ulcerative Colitis Is Related to Severe Inflammation and a High Count of Cytomegalovirus-Positive Cells in Biopsy Is a Risk Factor for Colectomy. J. Crohns Colitis 2016, 10, 1205–1211. [CrossRef]