

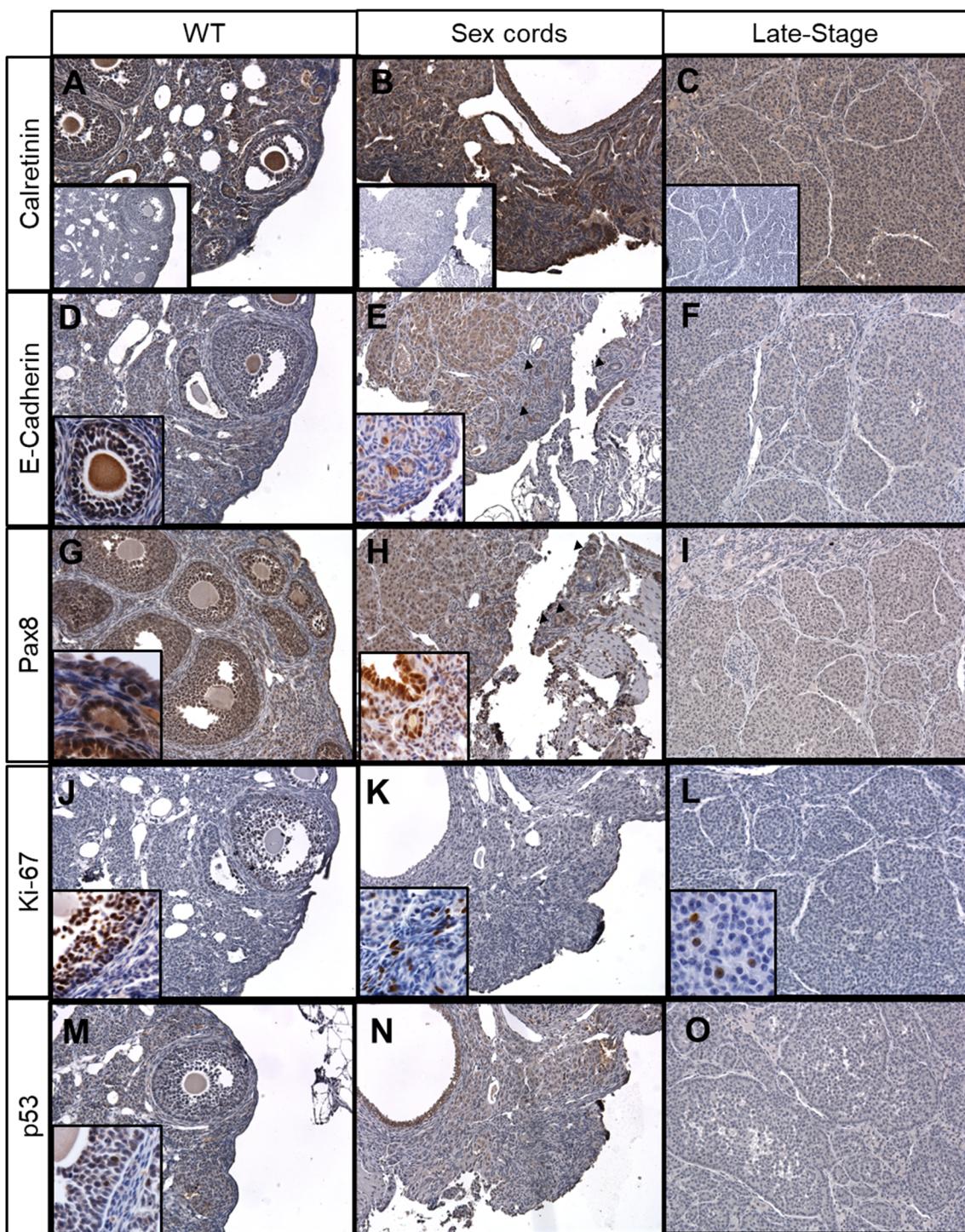


Article

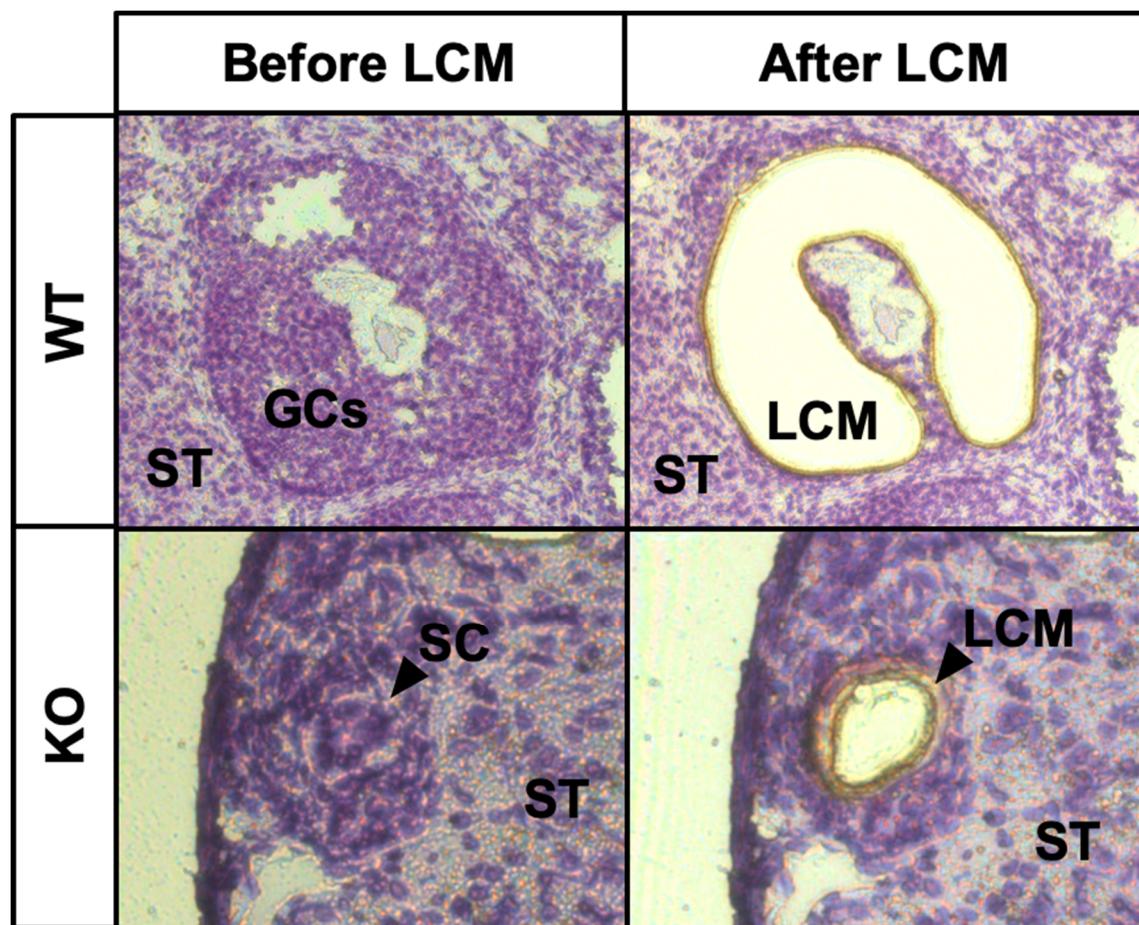
# The Validation of a Precursor Lesion of Epithelial Ovarian Cancer in *Fancd2*-KO Mice

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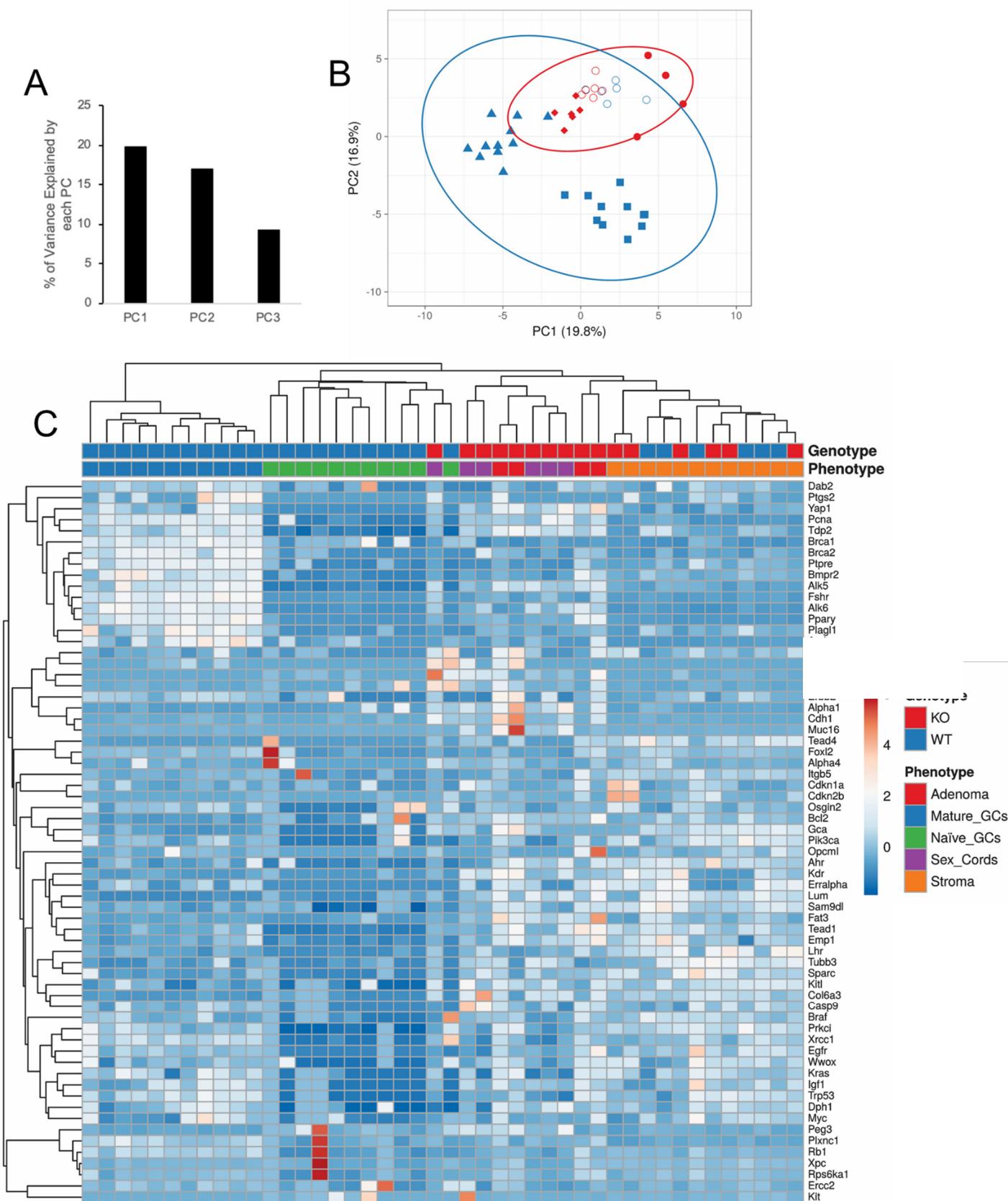
## Supplemental Figures



**Figure S1.** – Immunohistochemistry of common epithelial ovarian cancer markers to characterise a late-stage tumour phenotype in *Fancd2*<sup>-/-</sup> mice. (A,D,G,J,M) depicts ovarian tissue from a 3-month *Fancd2*<sup>+/+</sup> (wild-type; WT) mouse including primordial, primary and mature follicles, as well as stroma samples. (B,E,H,K,N) depicts ovarian tissue from a 3-month *Fancd2*<sup>-/-</sup> ovary that includes sex cords (E,H arrowheads) and stroma. (C,F,I,L,O) depicts ovarian tissue from a 1-year *Fancd2*<sup>-/-</sup> ovary with a later stage ovarian cancer neoplasm. (A,B,C) Calretinin is strongly expressed within all tissues and insets display secondary antibody only controls. (D) E-Cadherin and (G) Pax8 are both present in the ovarian surface epithelium and stroma, with Pax8 more strongly labelled in all granulosa cells, including primary follicles of WT mice. (E) E-Cadherin and (H) Pax8 are present in sex cords and stroma of 3-month *Fancd2*<sup>-/-</sup> animals, but (F,I) are only present as weak and diffuse staining in the 1-year *Fancd2*<sup>-/-</sup> phenotype. (J,K,L) Ki-67 staining is limited to a few cells within all samples and (M,N,O) p53 is present, and its weak and diffuse staining suggests normal function. Photomicrographs taken at 20x magnification or 40x for higher magnification in inset images in (D,E,G,H,J,K,L,M).

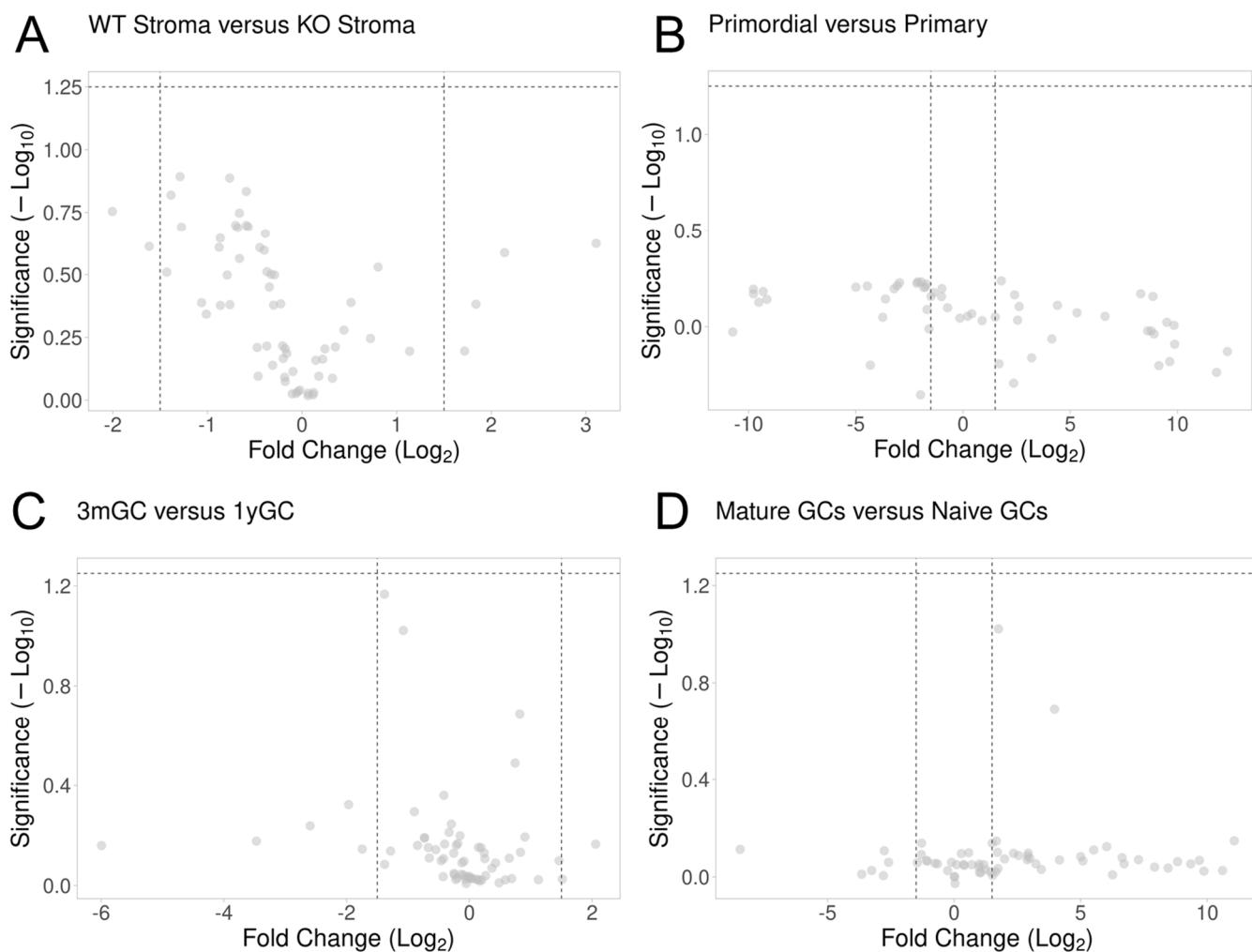


**Figure S2.** - LCM of mature GCs and sex cords. Representative images of tissue before and after laser capture micro-dissection (LCM) performed on *Fancd2*<sup>+/+</sup> (WT) and *Fancd2*<sup>-/-</sup> (KO) ovaries to collect mature GCs and sex cords (SC), respectively. ST = ovarian stroma.



**Figure S3.** Unbiased analyses of relative gene expression data of sex cords and ovarian stroma from 3-month-old *Fancd2*<sup>-/-</sup> (KO) mice, tubulostromal adenomas and ovarian stroma from 1-year old KO mice, naïve granulosa cells (GC), mature GC and stroma from 3-month-old *Fancd2*<sup>+/+</sup> (WT) mice and

mature GC and stroma from 1-year old WT mice. (A) Bar graph displays the percent variance represented by the first three principal components. (B) X and Y axis show principal components 1 and 2, which explain 19.8% and 16.9% of the total variance, respectively. Data segregation based on genotype, WT vs KO, however stromal controls are intermediately mixed. Prediction ellipses are such that with a probability of 0.95, a new observation from the same group will fall inside the ellipse. N = 44 data points (C) Heat map of unsupervised hierarchical clustering with minimal misclassification. Data mirrors results of PCA plot. Rows are centred; unit variance scaling is applied to rows. Imputation is used for missing value estimation. Both rows and columns are clustered using correlation distance and average linkage. 65 rows, 44 columns.



**Figure S4.** – Volcano plots representing non-significant gene expression changes following pairwise comparisons. (A) Ovarian stroma from 3-month-old *Fancd2<sup>+/+</sup>* (wild type; WT) and *Fancd2<sup>-/-</sup>* (knock out; KO) mice exhibited similar levels of expression of all genes tested and served as a genotype control. (B) Granulosa cells (GC) from primordial and primary follicles from WT mice exhibited similar levels of expression of all genes tested and confirmed that these sample types could be combined as naïve GCs. (C) GC from growing follicles of 3-month-old and 1-year old WT mice exhibited similar levels of expression of all genes tested and confirmed these sample types could be combined as mature GCs. (D) Naïve and mature GCs from WT mice exhibited similar levels of expression of all genes tested and supports the grouping of these phenotypes for further analyses. Thresholds represented as dashed lines, were set at a fold change of  $> 1.5$  and p-value of  $< 0.05$  for relative gene expression data from all GeXP gene sets. No genes for any of the control comparisons were significant based on designated threshold. .

## Supplemental Tables

**Table S1.** Genome GeXP Multiplex Assay Set 1.

Gene Symbol	Accession Number	Primer Sequences (5' – 3')	[Primer μM]		Amplicon Length (bp)
			F	R	
<i>Pax8</i>	NM_011040	F- AGGTGACACTATAGAATAAGATCCTGGCAGGTACT R- GTACGACTCACTATAGGGATGTAGTCTCCTATCTTCTCCA	2 0.5		145
<i>Rpl19</i>	NM_009078	F- AGGTGACACTATAGAATAACCCACAAGCTTTCC R- GTACGACTCACTATAGGAATCCAACCAGACCTTCTT	2 0.5		150
<i>Hprt</i>	NM_013556	F- AGGTGACACTATAGAATAACAGGTGTTCTAGTCCTGTG R- GTACGACTCACTATAGGGAGTGCAAATCAAAGTCTGG	2 0.0039		157
<i>Col6α3</i>	NM_001243008	F- AGGTGACACTATAGAATAGCCATTCTAACTCGTTCT R- GTACGACTCACTATAGGGATTAGCACCATATTGACAGG	2 0.0039		163
<i>Notch3</i>	NM_008716.3	F- AGGTGACACTATAGAATAACTCGTATGTACCAAGTAGCC R- GTACGACTCACTATAGGGACTTCCCAGTGAAGTCTACA	2 0.5		174
<i>Braf</i>	NM_139294	F- AGGTGACACTATAGAATAGTCTTCTGCCAACAA R- GTACGACTCACTATAGGGAGTTCTCTCTCCATCCTG	2 0.0078		182
<i>Dph1</i>	NM_144491	F- AGGTGACACTATAGAATAACACCCTAATATACTGCTT R- GTACGACTCACTATAGGGAAAGGTTCCCAGAATAAGG	2 0.5		187
<i>Erbb2</i>	NM_001003817	F- AGGTGACACTATAGAATAAACTGGTGTGTCAGATTG R- GTACGACTCACTATAGGAATTCACTCTCATCAATGTCC	2 0.0078		197
<i>Ccne1</i>	NM_007633	F- AGGTGACACTATAGAATAGAAATCTACCCTCCAAAG R- GTACGACTCACTATAGGGATGGACATAGACATTCAAGC	2 0.5		202
<i>Brca1</i>	NM_009764	F- AGGTGACACTATAGAATAGACCCCCAAAGAAGTAAT R- GTACGACTCACTATAGGGAACTATCCACTTCCCTCTG	2 0.5		208
<i>Cdkn1a</i>	NM_007669	F- AGGTGACACTATAGAATAAGACATTCAAGGCCACAG R- GTACGACTCACTATAGGGAGACGAAGTCAAAGTCCAC	2 1		219
<i>Casp3</i>	NM_001284409	F- AGGTGACACTATAGAATAGTGCAGATGAGGTGTT R- GTACGACTCACTATAGGGATCTTAGCGTACCGTTCC	2 0.5		224
<i>Myc</i>	NM_001177352	F- AGGTGACACTATAGAATAACTCAGTGGCTTCCCTAC R- GTACGACTCACTATAGGGATTCTGCTCTCTCAGAGT	2 0.5		233
<i>Igf1</i>	NM_010512	F- AGGTGACACTATAGAATATGTGACATTGCTCTAACATC R- GTACGACTCACTATAGGGACAGGTAGAAGAGGTGTGAAG	2 0.0039		240
<i>Ercc2</i>	NM_010512	F- AGGTGACACTATAGAATAGGAGGTGACCAAACCTATCTA R- GTACGACTCACTATAGGGAGACACTTCCATCAACATC	2 0.5		246
<i>Kit</i>	NM_010512	F- AGGTGACACTATAGAATAACAAAGAGCAAATCCAG R- GTACGACTCACTATAGGGAGGAAACTCCATTGTG	2 1		257
<i>Dlec1</i>	NM_177117	F- AGGTGACACTATAGAATAACTCTCGTCTGTATAACTCC R- GTACGACTCACTATAGGGAGGGACAGTGTGGCTAGA	2 1		263
<i>Aurka</i>	NM_011497	F- AGGTGACACTATAGAATAGAGGGTTGACTCCTATTTT R- GTACGACTCACTATAGGGAGGGAACAGTGGCTTAAAC	2 0.5		273
<i>Ppia</i>	NM_008907	F- AGGTGACACTATAGAATAGATGACGAGCCCTG R- GTACGACTCACTATAGGGAGGGACCTGTATGCTTAGGA	2 0.5		285
<i>Kitl</i>	NM_013598	F- AGGTGACACTATAGAATAGGTCCCTTAGTAAAAGTGAA R- GTACGACTCACTATAGGGATCCTTAGTAGCACCCATACAT	2 0.125		291
<i>Arl11</i>	NM_177337	F- AGGTGACACTATAGAATATGCCCTAACCAACTCG R- GTACGACTCACTATAGGGACCCTAGTAAATCCTCCTG	2 0.5		299
<i>Bax2</i>	NM_007527	F- AGGTGACACTATAGAATAACTACAGGGTTCATCCAG R- GTACGACTCACTATAGGGAAAAGTAGAAGAGGGCAACC	2 0.5		307
<i>Dab2</i>	NM_023118	F- AGGTGACACTATAGAATAAGTCCGTGTACTTGTGG R- GTACGACTCACTATAGGGACATCATCAATACCGATTAGC	2 0.0078		318
<i>Trp53</i>	NM_011640	F- AGGTGACACTATAGAATACCGAAGACTGGATGACT R- GTACGACTCACTATAGGGAGAAGGGACAAAAGATGAC	2 0.5		336
<i>Kras</i>	NM_021284	F- AGGTGACACTATAGAATAACCTGTCTTGGATATTCTC	2		345

Gene Symbol	Accession Number	Primer Sequences (5' – 3')	[Primer μM] F R	Amplicon Length (bp)
<i>Diras2</i>	NM_001024474	R- GTACGACTCACTATAGGAAACACCCGTCTTGTCTT	0.125	
		F- AGGTGACACTATAGAATACCTACCCCAAGTTGTAAC	2	
		R- GTACGACTCACTATAGGGAGCTGTAAGTGAGCTTATTG	0.5	363
<i>Casp9</i>	NM_015733	F- AGGTGACACTATAGAATATCCACTGAGTTAGATGG	2	
		R- GTACGACTCACTATAGGGACCTCTCTCCTGTGTGTC	0.5	377
<i>Brca2</i>	NM_001081001	F- AGGTGACACTATAGAATAGGGTGCTGTGATGTATCTAGT	2	
		R- GTACGACTCACTATAGGGAGATTCCCTGTGGTGTGTTT	0.5	383

Table S2. Genome GeXP Multiplex Assay Set 2.

Gene Symbol	Accession Number	Primer Sequences (5' – 3')	[Primer μM] F R	Amplicon Length (bp)
<i>Lum</i>	NM_008524	F- AGGTGACACTATAGAATATTGGCATTAGTCGGTAGT R- GTACGACTCACTATAGGGAGGGCAGTTACATTCTG	2 0.0078	142
<i>Rpl19</i>	NM_009078	F- AGGTGACACTATAGAATACCCACAAGCTTTCC R- GTACGACTCACTATAGGGAAATCCAACCAGACCTCTT	2 0.5	150
<i>Hprt</i>	NM_013556	F- AGGTGACACTATAGAATACAGGTGTTAGCCTGTG R- GTACGACTCACTATAGGGAGTGCAAATCAAAAGTCTGG	2 0.0156	157
<i>Prkci</i>	NM_008857	F- AGGTGACACTATAGAATAGTGAAGTTGTCAAGTTGT R- GTACGACTCACTATAAGGCACAGGCATCATCAGGTC	2 0.0156	169
<i>Peg3</i>	NM_008817	F- AGGTGACACTATAGAATAGTGCAGAGTGCTGTG R- GTACGACTCACTATAGGGACTGATCTCCCTTGCT	2 0.5	176
<i>Rb1</i>	NM_009029	F- AGGTGACACTATAGAATAGGCATCTGCATCTTATC R- GTACGACTCACTATAGGGACATAGCATTATCACCTTGG	2 0.5	184
<i>Samd9l</i>	NM_010156	F- AGGTGACACTATAGAATACGAAACTGGAAACTGAAG R- GTACGACTCACTATAGGGAGTCCGTGACAAACAACCTT	2 0.5	196
<i>Rab25</i>	NM_016899	F- AGGTGACACTATAGAATATCTCACCTCCCTACT R- GTACGACTCACTATAGGGACCAGGCACAATACAAAAC	2 0.5	206
<i>Tubb3</i>	NM_023279	F- AGGTGACACTATAGAATACAAGACAAGCAGCATCT R- GTACGACTCACTATAGGGAAATACAGAGGTGGCTAAAATG	2 0.5	212
<i>Bcl2</i>	NM_009741	F- AGGTGACACTATAGAATAGATGCCCTTGTGGAAC R- GTACGACTCACTATAGGGACTGCTTACTGAACCTTTG	2 0.5	221
<i>Kdr</i>	NM_010612	F- AGGTGACACTATAGAATATGAGCCAAGTGTAAAGT R- GTACGACTCACTATAGGGAGTAACCCACTGTGAAGA	2 0.5	230
<i>Osgin2</i>	NM_145950	F- AGGTGACACTATAGAATAGTTGGTATTGGCAGA R- GTACGACTCACTATAGGGAGCAGTAACACATTTCACAG	2 0.5	236
<i>Gca</i>	NM_145523	F- AGGTGACACTATAGAATAGATAACCTGGTTATTCTGAC R- GTACGACTCACTATAGGGATTCTGTGTAGTCTATCC	2 0.0156	245
<i>Xpc</i>	NM_009531	F- AGGTGACACTATAGAATAGGCTGTCCAGATGAC R- GTACGACTCACTATAGGGACCTTTCTCCTCTCCT	2 0.5	251
<i>Ptpre</i>	NM_001316678	F- AGGTGACACTATAGAATAATCCTTCATAGATGGCTAC R- GTACGACTCACTATAGGGAGGTGCGCTCTTATCTCT	2 0.5	263
<i>Itgb5</i>	NM_001145884	F- AGGTGACACTATAGAATATACAAAAGTCTAAGGACTG R- GTACGACTCACTATAGGGACGCTCACTTGGAACT	2 0.0156	270
<i>Sparc</i>	NM_009242	F- AGGTGACACTATAGAATACGAGACTTGAGAAGAACTAC R- GTACGACTCACTATAGGGAGATGTCCTGCTCCTGAT	2 0.0156	277
<i>Ppia</i>	NM_008907	F- AGGTGACACTATAGAATAGATGACGGAGCCCTTG R- GTACGACTCACTATAGGGAGGACCTGTATGCTTAGGA	2 0.5	285
<i>Plxnc1</i>	NM_018797	F- AGGTGACACTATAGAATAGTTGTGGAGGTGCTAGTG R- GTACGACTCACTATAGGGAAATTAGGAGCTAGAACCTG	2 0.5	290
<i>Xrcc1</i>	NM_009532	F- AGGTGACACTATAGAATATTGTGGAGGTGCTAGTG R- GTACGACTCACTATAGGGAAGGAGGGCTGTGAAACT	2 0.5	304

Gene Symbol	Accession Number	Primer Sequences (5' – 3')	[Primer µM]		Amplicon Length (bp)
			F	R	
<i>Egfr</i>	NM_207655	F- AGGTGACACTATAGAATATGTGCCCACTACATTGA R- GTACGACTCACTATAGGGAACGTCTCGCATGAATAG	2	1	313
<i>Cdkn2b</i>	NM_007670	F- AGGTGACACTATAGAATAAACCTGGAGAGTAGATGAGA R- GTACGACTCACTATAGGGAAAGAGCAAATGTAGGA	2	0.0156	331
<i>Pik3ca</i>	NM_008839	F- AGGTGACACTATAGAATAGAGGGAGAACCCTTATGTGA R- GTACGACTCACTATAGGGATGACCCAGTAACACCAATA	2	0.0156	342
<i>Opcml</i>	NM_177906	F- AGGTGACACTATAGAATAACCACATCTCATAGTCAAGTTC R- GTACGACTCACTATAGGGACCAGTGTCTTGGCTTT	2	0.5	349
<i>Plagl1</i>	NM_009538	F- AGGTGACACTATAGAATAACTGCTCTCACAGAAGATACC R- GTACGACTCACTATAGGGAGCTTAACTTGGAGACGAAG	2	0.5	358
<i>Rps5ka1</i>	NM_009097	F- AGGTGACACTATAGAATAACGACACCTTCTACTTGATA R- GTACGACTCACTATAGGGAGGATCTCTTGCTCTGT	2	0.5	367
<i>Wwox</i>	NM_019573	F- AGGTGACACTATAGAATAGCTGCTACCACTGTCTACTG R- GTACGACTCACTATAGGGATCCCTGCTCCACTA	2	0.5	372
<i>Rassf1</i>	NM_001243748	F- AGGTGACACTATAGAATAAGCCAGAACTCATTGAACTAC R- GTACGACTCACTATAGGGATCGTATCCGCTCTAGT	2	0.5	379
<i>Akt2</i>	NM_001110208	F- AGGTGACACTATAGAATATCCCTGGTCTACTACGTC R- GTACGACTCACTATAGGGAGCTGTACCTAGCTTGAG	2	0.5	387

Table S3. Genome GeXP Multiplex Assay Set 3.

Gene Symbol	Accession Number	Primer Sequences (5' – 3')	[Primer µM]		Amplicon Length (bp)
			F	R	
<i>Lhr</i>	NM_013582.2	F- AGGTGACACTATAGAATAGTGAAGCACAGTTAGAGAA R- GTACGACTCACTATAGGGATGGGTGAACAGAAAGTCATA	2	0.002	142
<i>Rpl19</i>	NM_009078	F- AGGTGACACTATAGAATAACCCACAAGCTCTTCC R- GTACGACTCACTATAGGAATCCAACCAGACCTCTT	2	0.5	150
<i>Hprt</i>	NM_013556	F- AGGTGACACTATAGAATACAGGTGTTCTAGTCCTGTG R- GTACGACTCACTATAGGGAGTGCAAATAAAAGTCTGG	2	0.0039	157
<i>Ahr</i>	NM_013464.4	F-AGGTGACACTATAGAATATCTGGTTCTGGCAAT R- GTACGACTCACTATAGGGAGGCTGAAGTGGAGTAGC	2	0.0015625	165
<i>Pcna</i>	NM_011045.2	F- AGGTGACACTATAGAATAGAACCAAAATCAAGAG R- GTACGACTCACTATAGGGAGCTAAGGTCTCGGCATA	2	0.0039	184
<i>Pgc1α1</i>	NM_008904.2	F- AGGTGACACTATAGAATAGGACATGTGCAGCCAAGACTCT R- GTACGACTCACTATAGGGACACTTCAATCCACCCAGAAAG CT	2	0.5	193
<i>Foxl2</i>	NM_012020.2	F- AGGTGACACTATAGAATAGCCCAGATGACACTCCTA R- GTACGACTCACTATAGGGACGAAAGACATTACTCAACCA	2	0.0039	201
<i>Pgc1α4</i>	NR_132764.1	F- AGGTGACACTATAGAATATCACACCAAAACCCACAGAAA R- GTACGACTCACTATAGGGACTGGAAGATATGGCACAT	2	0.5	209
<i>Alk6</i>	NM_007560.4	F- AGGTGACACTATAGAATATGCGTGTGCTAAATCTC R- GTACGACTCACTATAGGGATGTAAGAGGGCATACCAAG	2	0.5	216
<i>Tead1</i>	NM_001166584.1	F- AGGTGACACTATAGAATAGCTGAAGGTAACAAGCA R- GTACGACTCACTATAGGGATGGCTGTCTGTCTGTAT	2	0.5	224
<i>Cdh1</i>	NM_009864.3	F- AGGTGACACTATAGAATATCTCTGTCCCTTCCAC R- GTACGACTCACTATAGGGAAAGTGTCCCTCCAAATCC	2	0.5	232
<i>Tdp2</i>	NM_019551.2	F- AGGTGACACTATAGAATAGCACCTTATTCCCTCAAAGT R- GTACGACTCACTATAGGGATGTATTAGACCCAGATGTCC	2	0.125	237
<i>Fat3</i>	NM_001080814.1	F- AGGTGACACTATAGAATATGCTCTAACATAACCATC R- GTACGACTCACTATAGGAAATTACCTCACCATTAGAAC	2	0.5	247

Gene Symbol	Accession Number	Primer Sequences (5' – 3')	[Primer μM] F R	Amplicon Length (bp)
<i>Emp1</i>	NM_010128.4	F- AGGTGACACTATAGAATAATCTCCCTGTGGTCTTC R- GTACGACTCACTATAGGAAAGCTGAAACAGAACAGAG	2 0.5	252
<i>Muc16</i>	XM_011242634.1	F- AGGTGACACTATAGAATAATCACCATTGTCTACG R- GTACGACTCACTATAGGGAGTGAACATCCCATTCTCT	2 0.125	258
<i>Pparγ</i>	NM_001127330.2	F- AGGTGACACTATAGAATAATCACCATTGTCTACG R- GTACGACTCACTATAGGGAAACACCATACTGAGCAGAG	2 0.5	270
<i>Bmpr2</i>	NM_007561.4	F- AGGTGACACTATAGAATAAAATCAGACGAAGAGCAG R- GTACGACTCACTATAGGGATCTCACCTATCCCAAGAT	2 0.0039	279
<i>Ppia</i>	NM_008907	F- AGGTGACACTATAGAATAAGATGACGAGGCCCTTG R- GTACGACTCACTATAGGGAGGACCTGTATGCTTAGGA	2 0.125	285
<i>Ptgs2</i>	NM_011198.4	F- AGGTGACACTATAGAATAAGAACCAAGAACAAAG R- GTACGACTCACTATAGGAATGGGCTGGAAAGACATA	2 0.5	295
<i>Tead4</i>	NM_011567.2	F- AGGTGACACTATAGAATAAGATGTATGGTCGGAATG R- GTACGACTCACTATAGGGAAAACCTGAGATTGCTG	2 1	307
<i>Alk5</i>	NM_009370.3	F- AGGTGACACTATAGAATAAGCTTAGTGTCTGGAAAT R- GTACGACTCACTATAGGGACTGTTGGCTGAGTTGT	2 0.125	318
<i>Fshr</i>	NM_013523.3	F- AGGTGACACTATAGAATAACGTCAAGAGACACCAAGAT R- GTACGACTCACTATAGGGATTGTGGTAATAGATGAGGT	2 0.125	338
<i>Errα</i>	NM_007953.2	F- AGGTGACACTATAGAATAAAAGTGAATGCCAGGT R- GTACGACTCACTATAGGGACTGCTGGAGTTATTGCTT	2 1	343
<i>Mmp9</i>	NM_013599.4	F- AGGTGACACTATAGAATAACAGCACCTCCACTAT R- GTACGACTCACTATAGGGAGGGCTCCTCTATGATT	2 1	348
<i>Fcgbp</i>	NM_001122603.1	F- AGGTGACACTATAGAATAACCTGACAACAGCCACTAT R- GTACGACTCACTATAGGGACAAGGCACACTCCTCAC	2 0.5	355
<i>Yap1</i>	NM_001171147.1	F- AGGTGACACTATAGAATACTTGGTTATCTCCTCTCTACA R- GTACGACTCACTATAGGAAATCAGGCTAAGGGAAGTA	2 0.5	360
<i>Taz</i>	NM_001173547.2	F- AGGTGACACTATAGAATAACTGAAGTTGATGCGTTG R- GTACGACTCACTATAGGGAAAGTAGGGTGGGCTGTTAG	2 0.5	365
<i>Wfdc2</i>	NM_026323.2	F- AGGTGACACTATAGAATAAGCTCTAGGGACAGATACT R- GTACGACTCACTATAGGGAGGCAGAAACACCTCTCT	2 0.5	370
<i>Amh</i>	NM_007445.2	F- AGGTGACACTATAGAATAAGGGGAGACTGGAGAAC R- GTACGACTCACTATAGGGATCAGAGCCAATAGAAAGG	2 0.5	385

**Table S4.** Detailed information about antibodies used for immunohistochemistry.

Supplier	Catalogue Number	Target	Dilution	Antigen Retrieval Solution
<b>Abcam</b>	ab92341	Calretinin	1:100	Sodium Citrate
<b>Invitrogen</b>	PA585088	E-Cadherin	1:500	Sodium Citrate
<b>Abcam</b>	ab97477	Pax8	1:500	Tris-EDTA
<b>Abcam</b>	ab16667	Ki-67	1:200	Sodium Citrate
<b>Abcam</b>	ab131442	p53	1:200	Sodium Citrate
<b>Abcam</b>	ab6721	Goat α Rabbit IgG HRP	1:300	N/A

**Table S5.** – Summary of staining intensity of immunohistochemical markers in representative samples. Three-month wild-type (WT) samples represents staining of granulosa cells within primordial, primary and growing follicles as well as stroma. Sex cords represents both sex cord structures and stroma within 3-month old *Fancd2*<sup>-/-</sup> mice and late-stage represents adenomas from 1-year old *Fancd2*<sup>-/-</sup> mice.

Marker	3-month WT	Sex Cords	Late-Stage
Calretinin	+++	+++	+++

<b>E-Cadherin</b>	+	++	+
<b>Pax8</b>	++	++	++
<b>Ki-67</b>	++	+	+
<b>p53</b>	++	++	++

**Table S6.** – Summary of all pairwise comparisons made for Gene Set Enrichment Analysis and the resulting number of enriched gene ontology (GO) terms for each comparison.

		Compared to:			
		Naïve GCs	Mature GCs	Sex Cords	Adenoma
GO Terms Enriched in:	Naïve GCs	0	0	0	0
	Mature GCs	20	10	8	
	Sex Cords	23	0	1	p=0.053
		Adenoma	0	0	