

1 *Supplement*

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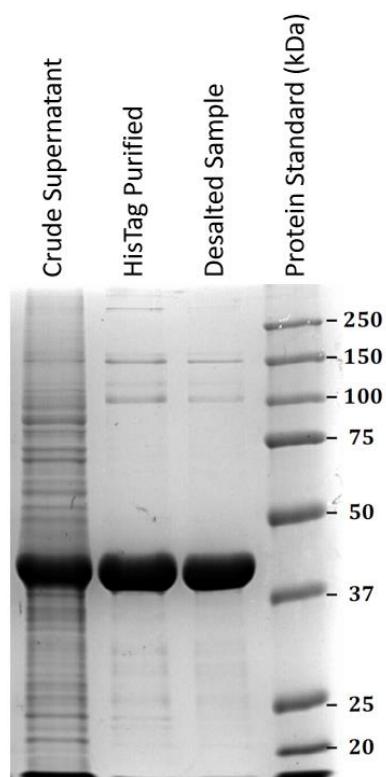
3 **Ppo-Er1 a novel ene reductase from *Paenibacillus*
4 *polymyxa***5 **Supporting Information**

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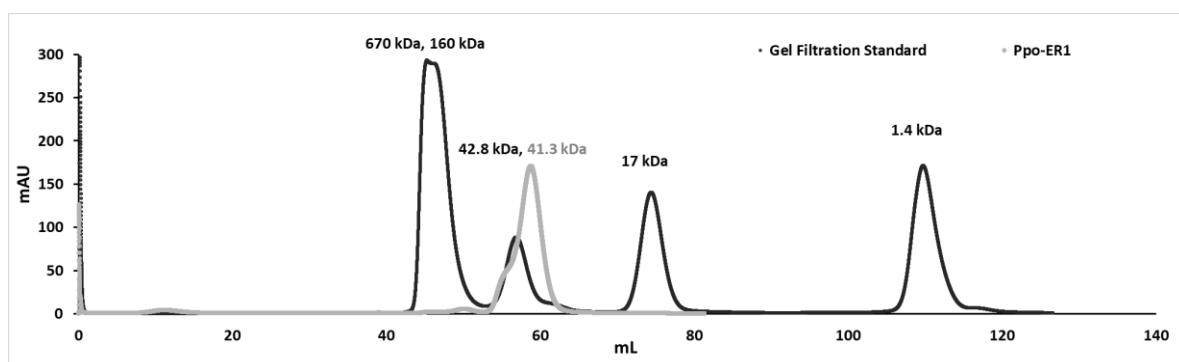
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29 **Figure S1:** SDS-PAGE showing the purity of ene reductase Ppo-ER1 after each purification
30 step.

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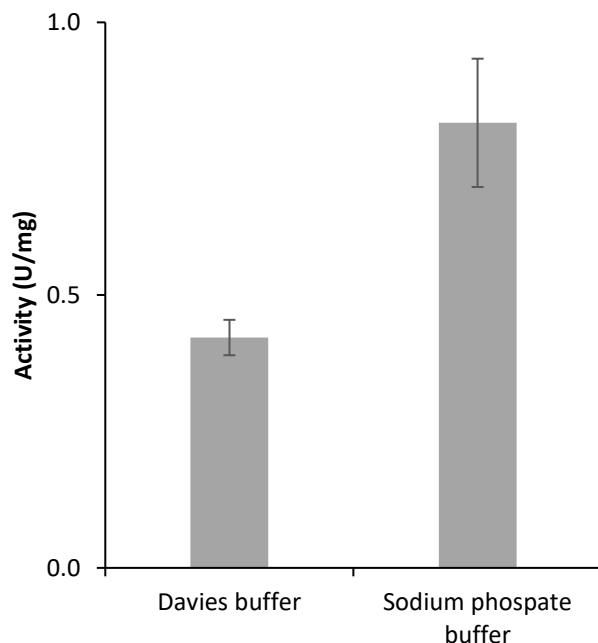


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35 **Figure S2:** Gel filtration of Ppo-ER1 in comparison with the Gel Filtration Standard (1.4-670
36 kDa) from Bio Rad.

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45 **Figure S3:** Comparison of the activity of Ppo-ER1 in Davies buffer and sodium phosphate
46 buffer.
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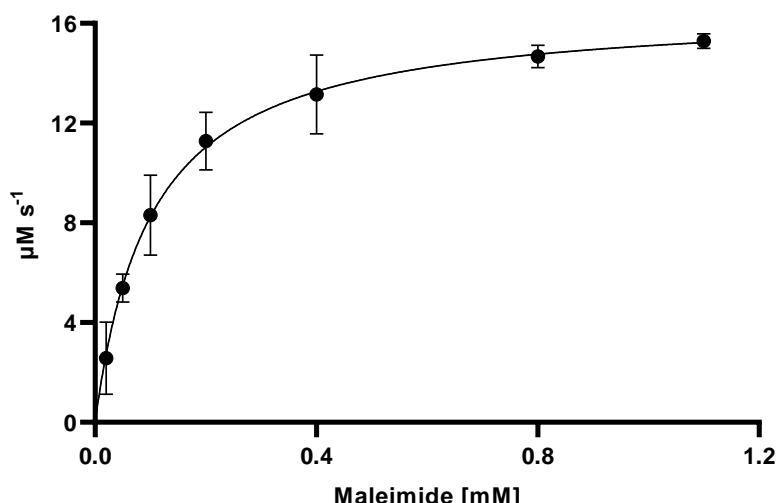
48 **Table S1:** Overview of the used GC-methods. For all measurements a 7890B GC System
 49 Agilent with FID detector was used.

Method	Column	Temperature profile	Constant Flow/Pressure	Split	Substrate Retention Time (min)	Product Retention Time (min)
A	DB5	40 °C 5 min -> 3 °C/min -> 60 °C -> 40 °C/min -> 300 °C	25 psi	10:01	Cyclohexenone (8.8) 2-Methyl-2-cyclohexenone (12.0) 3-Methyl-2-cyclohexenone (13.5) Cyclopentenone (4.7) 3-Methyl-2-cyclopentenone (10.6) Hexenal (5.5) 2-Methyl-2-pentenal (4.7)	Cyclohexanone (7.0) 2-Methyl-2-cyclohexanone (9.6) 3-Methyl-2-cyclohexanone (9.7) Cyclopentanone (3.5) 3-Methyl-2-cyclopentanone (5.2) Hexanal (3.8) 2-Methyl-2-pentanal (2.9)
B	DB5	110 °C 10 min -> 60 °C/min -> 300 °C	25 psi	10:01	4-Phenyl-3-buten-2-one (9.8)	4-Phenyl-3-butanone (5.6)
C	DB5	40 °C 10 min -> 3 °C/min -> 60 °C -> 40 °C/min -> 300 °C	15.8 psi	10:01	Butyl acrylate (11.9) Ethyl crotonate (8.1)	Butyl propionate (12.9) Ethyl butyrate (5.9)
D	DB5	50 °C -> 10 °C/min -> 90 °C 10 -> 15 °C/min -> 120 °C -> 60 °C/min -> 300 °C	25 psi	10:01	Carvone (13.2)	Dihydrocarvone (10.5/10.9)
E	DB5	100 °C 8 min -> 10 °C/min -> 190 °C -> 60 °C/min -> 300 °C	25 psi	10:01	<i>trans</i> -β-Methyl-β-nitrosyrene (13.2)	2-Methyl-2-nitroethylbenzen (10.6)
F	DB5	110 °C 10 min -> 10 °C/min -> 180 °C -> 60 °C/min -> 300 °C	25 psi	10:01	Cinnamic acid (13.1)	3-Phenylpropionic acid (9.6)
G	DB5	40 °C 10 min -> 11 °C/min -> 110 °C -> 60 °C/min -> 300 °C	25 psi	10:01	Citraconic acid (11.1) Crotonic acid (6.6)	
H	DB5	100 °C 10 min -> 5 °C/min -> 140 °C -> 20 °C/min -> 220 °C -> 40 °C/min -> 300 °C	26.3 psi	10:01	Citral (7.2/8.5)	Citronellal (4.6)
I	DB5	40 °C 8 min -> 15 °C/min -> 140 °C -> 60 °C/min -> 300 °C	26.3 psi	10:01	Maleimide (11.6)	Succinimide (13.7)

Method	Column	Temperature profile	Constant Flow/Pressure	Split	Substrate Retention Time (min)	Product Retention Time (min)
J	DB5	100 °C 7 min -> 10 °C/min -> 200 °C -> 40 °C/min -> 300 °C	26.3 psi	10:01	Diethylbenzylidienmalonate (16.5)	Diethylbenzylmalonate (12.3)
K	DB5	95 °C 10 min -> 15 °C/min -> 180 °C -> 60 °C/min -> 300 °C	25 psi	10:01	Cinnamic aldehyde (10.6)	3-Phenylpropanal (5.9)
L	BGB-175	120 °C 10 min -> 5 °C/min -> 160 °C -> 20 °C/min -> 220 °C	27.9 psi	10:01	Carvone (8.2)	Dihydrocarvone ((R) 6.2/ (S) 6.9)
M	BGB-175	100 °C 10 min -> 5 °C/min -> 140 °C -> 20 °C/min -> 220 °C	26.3 psi	10:01	Citral (15.8/16.1)	Citronellal ((S) 8.3/ (R) 9.0))
N	BGB-175	90 °C 10 min -> 5 °C/min -> 140 °C -> 20 °C/min -> 220 °C	25.4 psi	10:01	2-Methyl-2-cyclohexenone (10.6)	2-Methyl-2-cyclohexenone ((S) 8.2/ (R) 9.7)
O	BGB-175	45 °C 10 min -> 5 °C/min -> 140 °C -> 20 °C/min -> 220 °C	16.9 psi	10:01	2-Methyl-2-pentenal 15.3)	2-Methyl-2-pentanal ((R) 13.1/ (S) 13.2)

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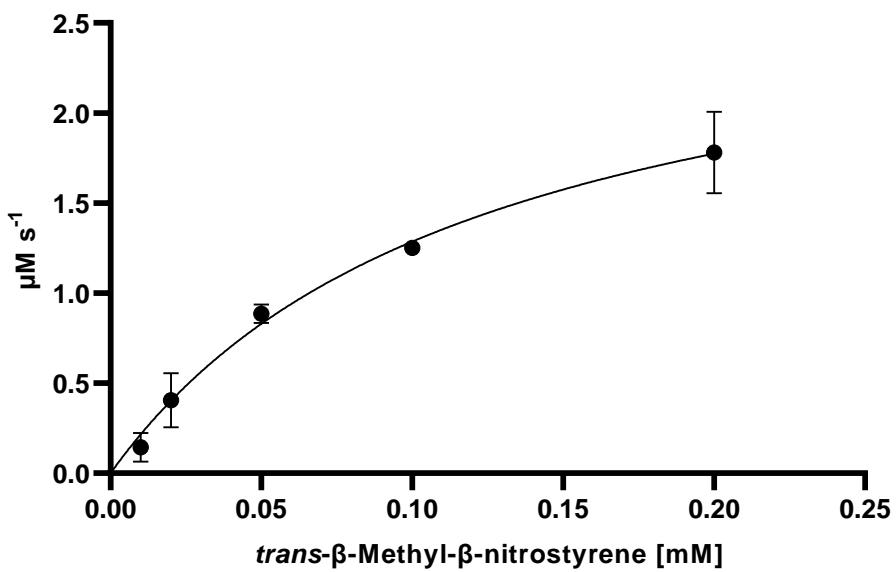
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54 Figure S4: Michaelis-Menten kinetic for maleimide

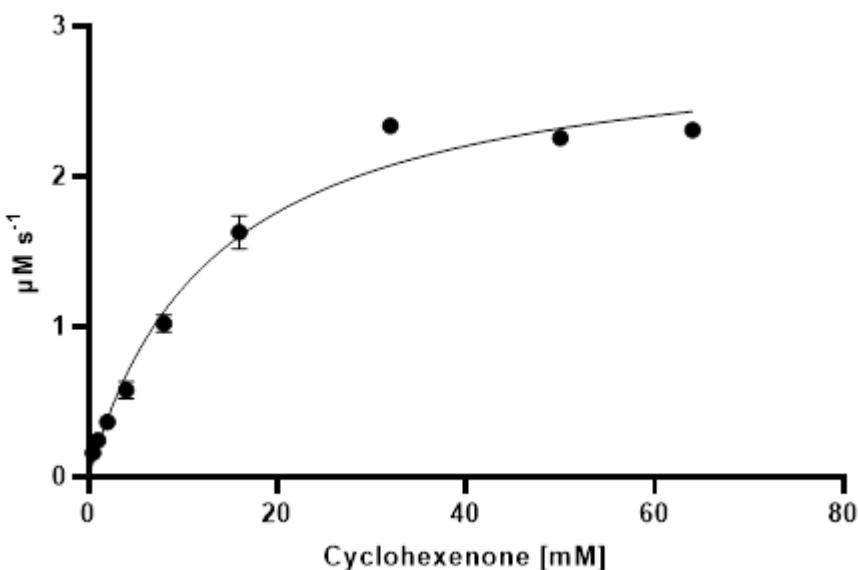
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57 **Figure S5:** Michaelis-Menten kinetic for *trans*- β -Methyl- β -nitrostyrene

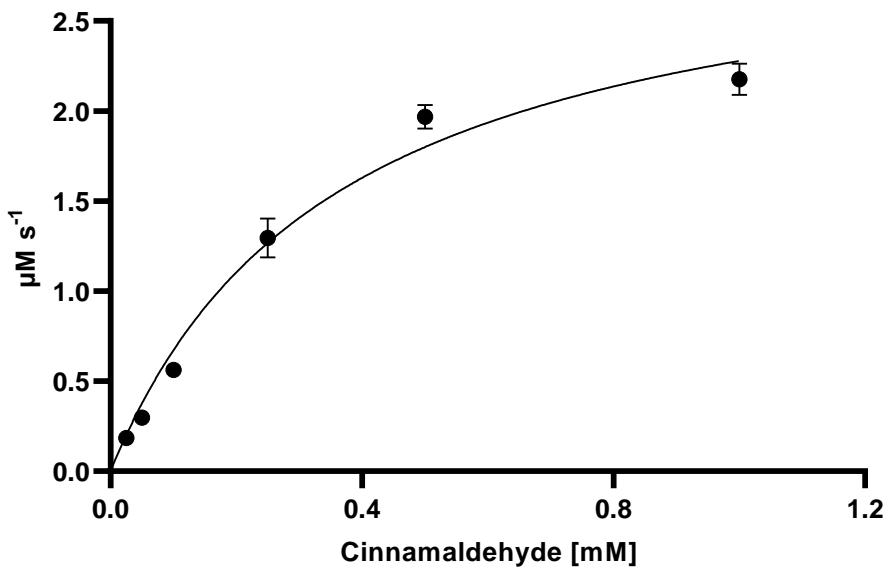
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60 **Figure S6:** Michaelis-Menten kinetic for cyclohexenone

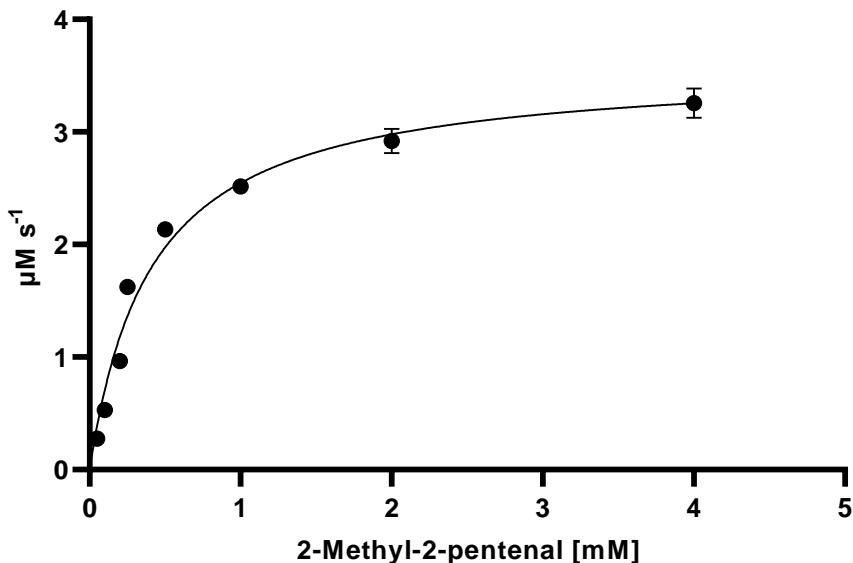
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63 **Figure S7:** Michaelis-Menten kinetic for cinnamaldehyde

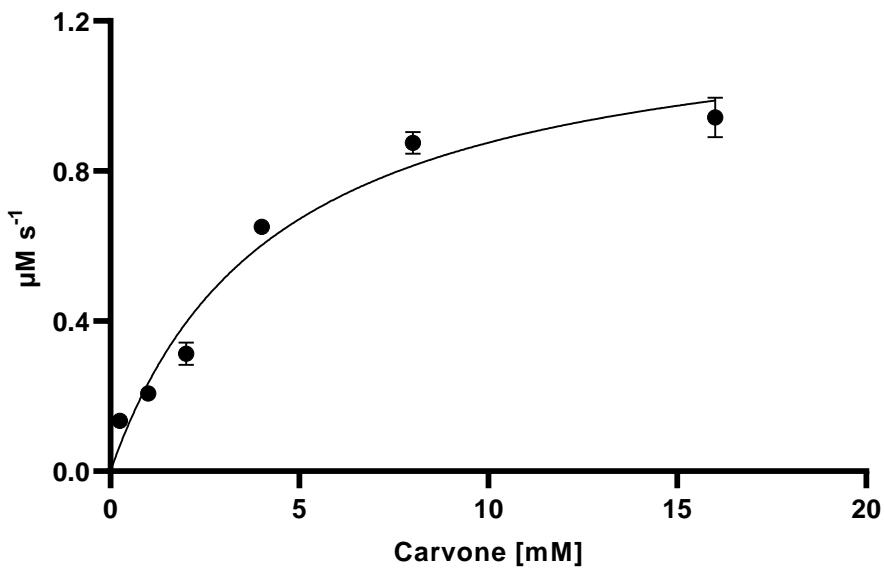
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66 **Figure S8:** Michaelis-Menten kinetic for 2-methyl-2-pentenal.

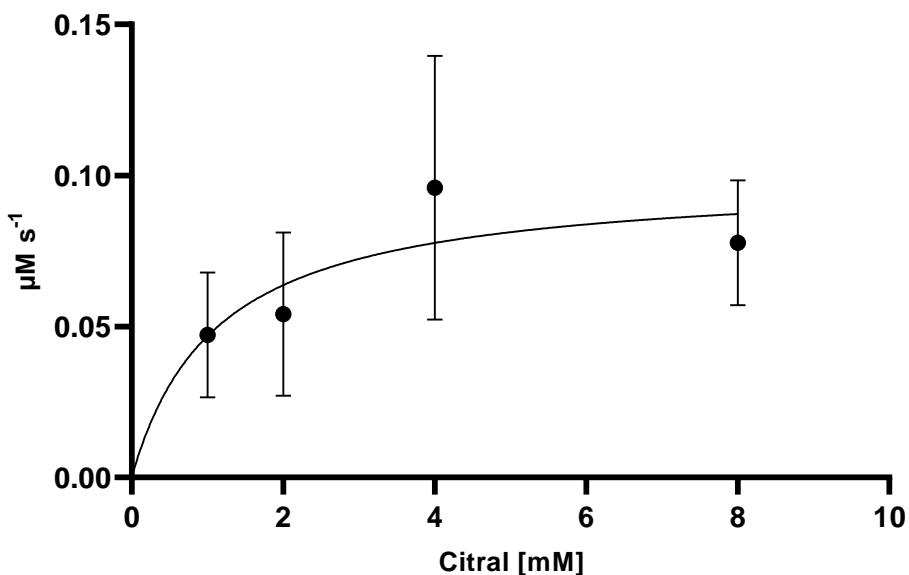
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69 **Figure S9:** Michaelis-Menten kinetic for carvone.

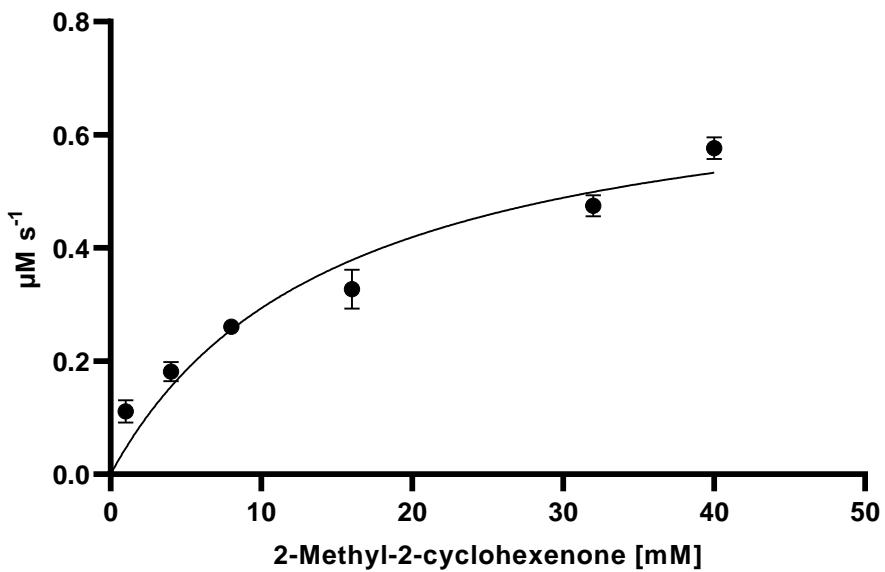
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72 **Figure S10:** Michaelis-Menten kinetic for citral.

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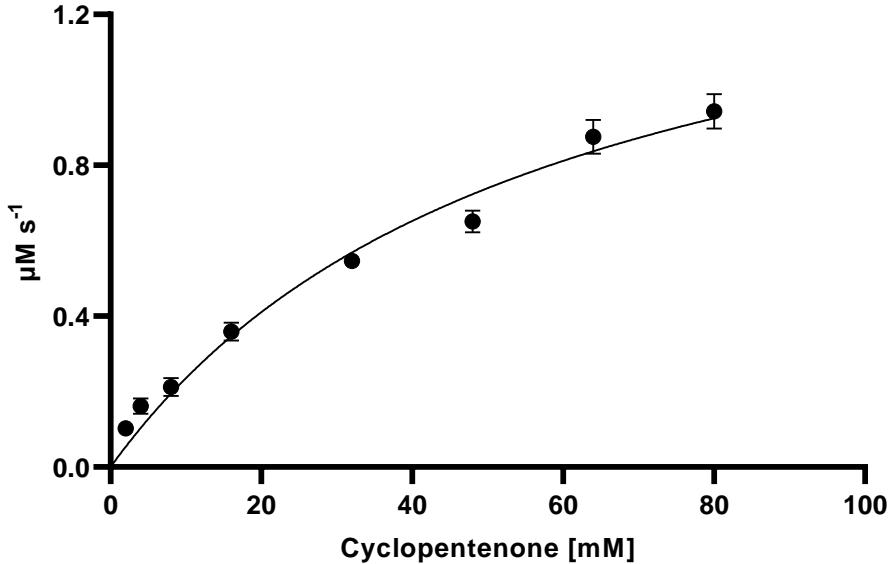


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75 **Figure S11:** Michaelis-Menten kinetic for 2-methyl-2-cyclohexenone.

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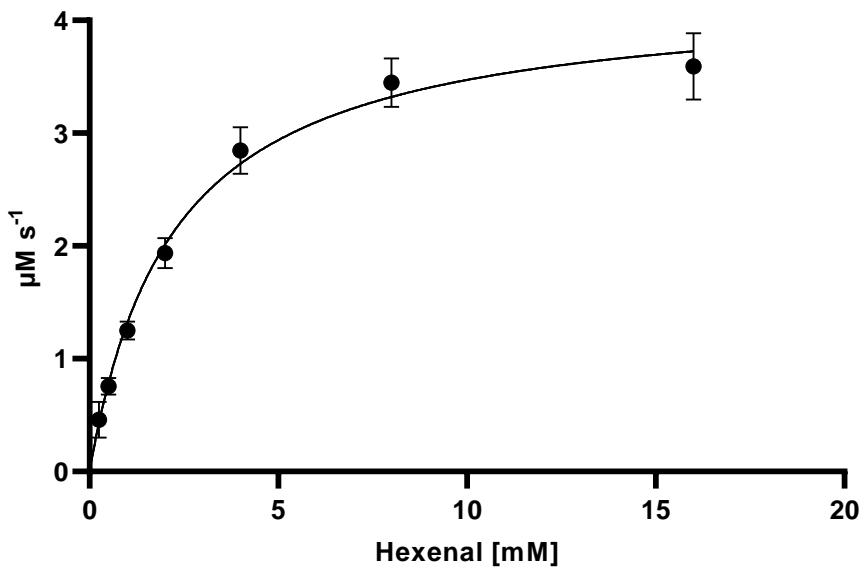
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79 **Figure S12:** Michaelis-Menten kinetic for cyclopentenone.

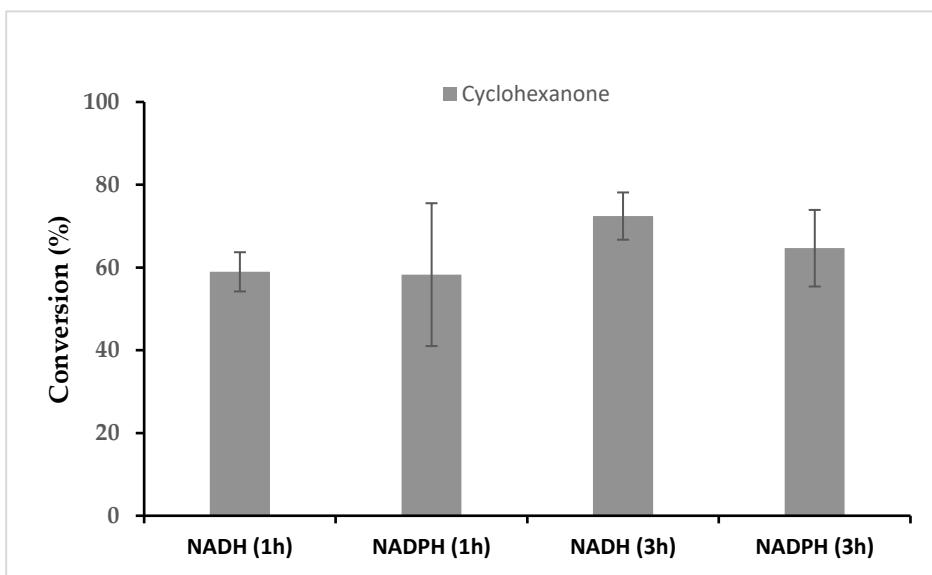
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82 **Figure S13:** Michaelis-Menten kinetic for hexenal.

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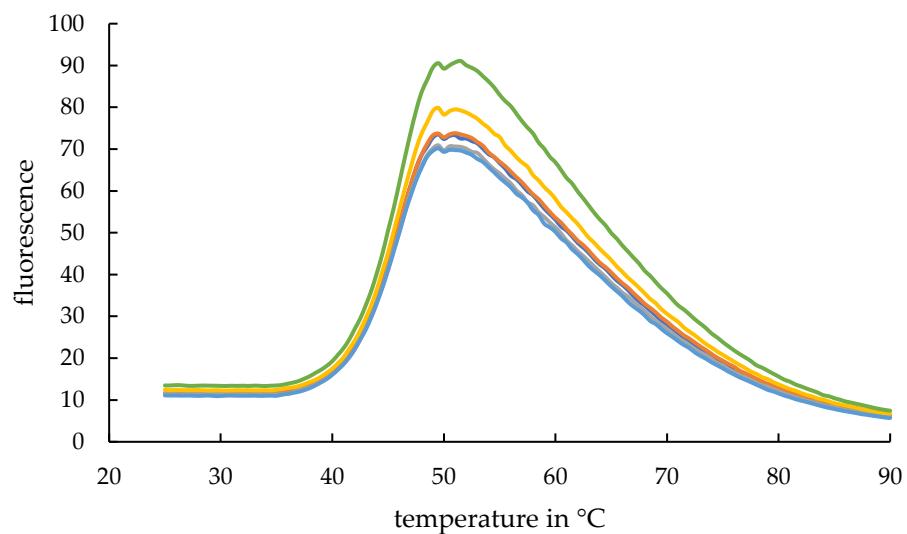


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85 **Figure S14:** Comparison of cyclohexanone conversion using either NADH or NADPH as
86 cofactor. Biocatalysis reactions were performed in triplicates with 5 mM cyclohexanone and
87 equimolar NADH/NADPH.

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91 **Figure S15:** Melting curve measured with the *ThermoFAD* assay.

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