

# Optimised Degradation of Lignocelluloses by Edible Filamentous Fungi for the Efficient Biorefinery of Sugar Beet Pulp

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## Supplementary Materials

### S1. Chemical analysis

The raw material was analysed for moisture, fibre, protein, sucrose, and ash contents according to the AOAC Official Methods [51]. For the moisture content determination an oven-drying method was used (drying at 135 °C for 2 h) (method 930.15). The crude protein content was determined by the Kjeldahl nitrogen (method 920.152, nitrogen to protein conversion factor was 6.25). Sucrose was determined according to the method 942.20.

### S2. Determination of soluble and insoluble dietary fibre

The quantification of soluble (SDF) and insoluble (IDF) dietary fibres was measured by using an integrated total dietary fibre assay procedure (K-TDFR, Megazyme, Co. Wicklow, Ireland), including the key attributes of the validated AOAC Official Methods 2002.02, 985.29, and 991.43 [51]. Shortly, powdered SBP sample ( $1.00 \pm 0.01$  g) was mixed with MES-TRIS (2(N-morpholino)ethanesulfonic acid tris(hydroxymethyl)aminomethane) buffer (pH 8.2) and subjected to sequential enzymatic hydrolysis by heat-stable  $\alpha$ -amylase (100 °C; 30 min). After amylase hydrolysis, sample was cooled to 60 °C and hydrolysed with protease (60 °C; 30 min). Further, the pH was adjusted to 4.8 and sample was hydrolysed with amyloglucosidase for 30 min., filtered using a vacuum through the crucible and washed twice with 60°C deionised water. The filtrate was collected and used for the SDF determination. SDF was precipitated from filtrate with 95% ethanol heated to 60 °C and recovered on the crucible by washing with distilled water, 78% ethanol, 95% ethanol, and acetone. For the determination of IDF, the sediment after enzymatic hydrolysis was then further washed with 95% ethanol and acetone. Crucibles containing residues were dried overnight in 105 °C oven and weighed. Obtained SDF and IDF contents were corrected for protein and ash. The content of each fibre was expressed as g/100 g d.w. of SBP. The analysis were performed in triplicate.

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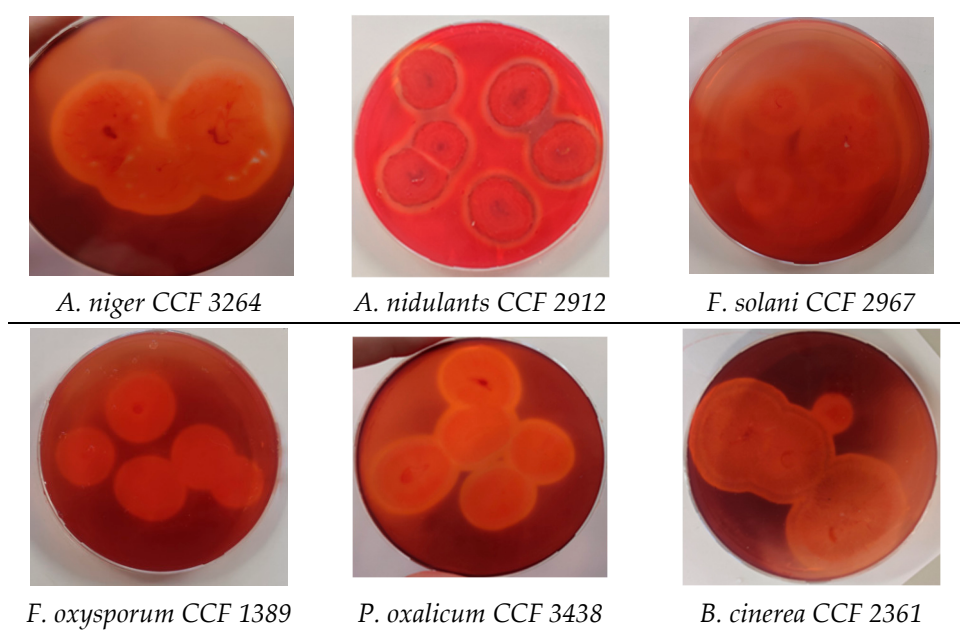
### S3. Lignin determination

The determination of acid-insoluble lignin was based on the NREL methodology [52]. Some modifications have been made to the sample size and heating procedure [53]. The sample was prepared by mixing  $1.0 \pm 0.01$  g ( $m_0$ ) of the SPB sample with  $15.00 \pm 0.01$  mL of 72% sulfuric acid and kept in a  $30^\circ\text{C}$  water bath for 1 h. Then, 575 mL of deionised water were added, and the sample was boiled under reflux for 4 h. After heating, the solution was filtered through a pre-weighted fritted glass filter ( $m_1$ ) and washed with boiling water until neutral pH (7.0). The neutralised filter was then oven-dried overnight at  $105^\circ\text{C}$  and weighed ( $m_2$ ). Insoluble lignin was calculated according to the eq.:

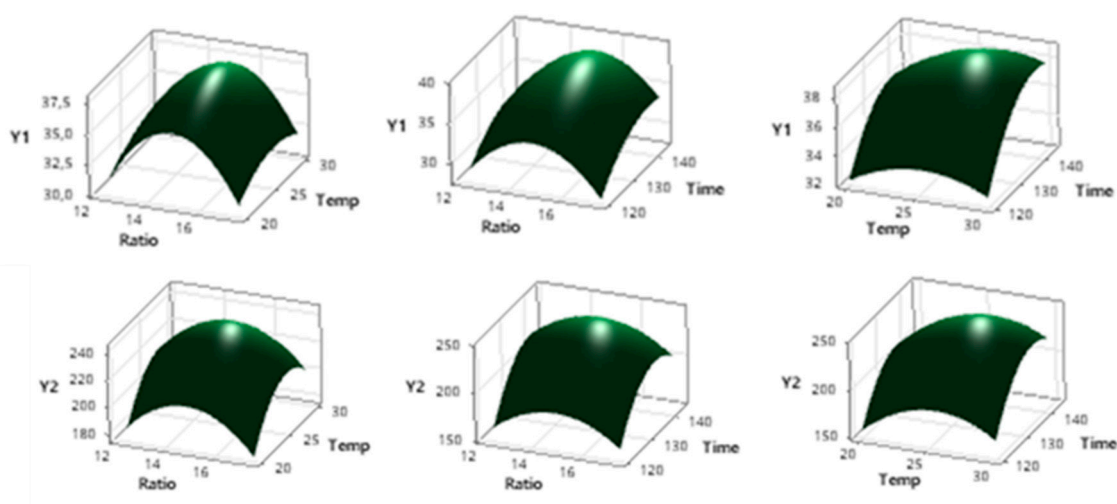
$$\text{Insoluble lignin, \%} = \frac{m_2 - m_1}{m_0} \times 100 \quad (1)$$

where:  $m_0$  – initial sample weight, g;  $m_1$  – initial filter weight, g;  $m_2$  – final dry weight, g.

### Figures and Tables



**Figure S1.** Hydrolytic activity of tested fungal strains on CMC-based agar.



**Figure S2.** Response surface plots of the interactions between process parameters for RS yield (Y1) and cellulase activity (Y2).

**Table S1.** Coded coefficients of the mathematical models for Y1 and Y2 responses.

Model Y1					Model Y2			
Term	Coef	SE Coef	T-Value	P-Value	Coef	SE Coef	T-Value	P-Value
Constant	37.909	0.235	161.25	0.000	6.5285	0.0181	361.65	0.000
Ratio	-0.167	0.216	-0.77	0.458	-0.0601	0.0200	-3.00	0.013
Time	0.527	0.216	2.44	0.035	0.5148	0.0193	26.64	0.000
Temp	2.150	0.216	9.94	0.000	0.2300	0.0193	11.90	0.000
Ratio*Ratio	-5.152	0.412	-12.49	0.000	-0.6164	0.0318	-19.36	0.000
Time*Time	-1.462	0.412	-3.55	0.005	-0.8661	0.0318	-27.21	0.000
Temp*Temp	-1.747	0.412	-4.24	0.002	-0.6651	0.0318	-20.89	0.000
Ratio*Time	-0.010	0.242	-0.04	0.968	-0.1997	0.0232	-8.59	0.000
Ratio*Temp	0.535	0.242	2.21	0.051	0.1961	0.0232	8.44	0.000
Time*Temp	0.440	0.242	1.82	0.099	-0.0823	0.0223	-3.69	0.004

**Table S2.** ANOVA of mathematical models.

Mathematical model for Y1					Mathematical model for Y2			
Term	Estimated Value	SE	T-Value	P-Value	Estimated Value	SE	T-Value	P-Value
Const.	37.909	0.235	161.25	0.000	6.5285	0.0181	361.65	0.000
X1	-0.167	0.216	-0.77	0.045	-0.0601	0.0200	-3.00	0.013
X2	2.150	0.216	9.94	0.000	0.5148	0.0193	26.64	0.000
X3	0.527	0.216	2.44	0.035	0.2300	0.0193	11.90	0.000
X1 <sup>2</sup>	-5.152	0.412	-12.49	0.000	-0.6164	0.0318	-19.36	0.000
X2 <sup>2</sup>	-1.747	0.412	-4.24	0.002	-0.8661	0.0318	-27.21	0.000
X3 <sup>2</sup>	-1.462	0.412	-3.55	0.005	-0.6651	0.0318	-20.89	0.000
X1X3	-0.010	0.242	-0.04	0.968	-0.1997	0.0232	-8.59	0.000
X1X2	0.535	0.242	2.21	0.051	0.1961	0.0232	8.44	0.000
X2X3	0.440	0.242	1.82	0.049	-0.0823	0.0223	-3.69	0.004
<b>R<sup>2</sup></b>	<b>R<sup>2</sup><sub>adj</sub></b>		<b>R<sup>2</sup><sub>pred</sub></b>		<b>R<sup>2</sup></b>	<b>R<sup>2</sup><sub>adj</sub></b>		<b>R<sup>2</sup><sub>pred</sub></b>
98.59%	97.32%		92.41%		98.89%	97.89%		96.99%
<i>Lack of fit</i>	<b>F-Value</b>		<b>P-Value</b>		<i>Lack of fit</i>	<b>F-Value</b>		<b>P-Value</b>
	6.81		0.089			53.61		0.985