

Supplementary Text S1

Features of in-house written macro for processing images from microtubule regrowth experiment

Images of γ -tubulin (centrosomal marker, 1st channel) and α -tubulin (microtubule asters, 2nd channel) were stored separately as z-stacks in tif format. The size of the image matrix was 256×256 pixels, using 16-bit depth. The size of voxel was $0.106 \mu\text{m} \times 0.106 \mu\text{m} \times 200 \mu\text{m}$. Macro written in ImageJ macro language using Fiji works under a directory with files in tif format to process automatically all contained images.

Tasks for individual channels

1st channel: Creating Maximum Intensity projection from a z-stack data. Smoothing the resulting picture by a 5x5 mean filter to remove high-intensity noise particles. Finding the brightest point in the smoothed picture corresponding to a center of a centrosome. Creating a circle $1 \mu\text{m}$ in a diameter as a ROI around this center. Measuring a mean intensity in this circle in original non-smoothed data.

2nd channel: Creating Maximum Intensity projection from a z-stack data. Removing non-homogenous background from this picture. Creating a circle $1 \mu\text{m}$ in a diameter as a ROI around the center point position found in the 1st channel. Measuring a mean intensity in this circle.

For checking the quality of analysis after the computation, an image pair of both channels is stored with overlays of found ROIs for each original picture. Computed mean image intensities are stored in an Excel file.