

Review

Human Genetic Adaptation to High Altitude: Evidence from the Andes

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Received: 2 November 2018; Accepted: 11 February 2019; Published: 15 February 2019



Abstract: Whether Andean populations are genetically adapted to high altitudes has long been of interest. Initial studies focused on physiological changes in the O₂ transport system that occur with acclimatization in newcomers and their comparison with those of long-resident Andeans. These as well as more recent studies indicate that Andeans have somewhat larger lung volumes, narrower alveolar to arterial O₂ gradients, slightly less hypoxic pulmonary vasoconstrictor response, greater uterine artery blood flow during pregnancy, and increased cardiac O₂ utilization, which overall suggests greater efficiency of O₂ transfer and utilization. More recent single nucleotide polymorphism and whole-genome sequencing studies indicate that multiple gene regions have undergone recent positive selection in Andeans. These include genes involved in the regulation of vascular control, metabolic hemostasis, and erythropoiesis. However, fundamental questions remain regarding the functional links between these adaptive genomic signals and the unique physiological attributes of highland Andeans. Well-designed physiological and genome association studies are needed to address such questions. It will be especially important to incorporate the role of epigenetic processes (i.e., non-sequence-based features of the genome) that are vital for transcriptional responses to hypoxia and are potentially heritable across generations. In short, further exploration of the interaction among genetic, epigenetic, and environmental factors in shaping patterns of adaptation to high altitude promises to improve the understanding of the mechanisms underlying human adaptive potential and clarify its implications for human health.

Keywords: adaptation; hypoxia; altitude; genomics; epigenomics

1. Introduction

Since the early 1900s, anthropologists and physiologists alike have sought to determine if there has been genetic adaptation to high altitude, conventionally defined as above 2500 m or 8250 ft since that is where O₂ saturation levels in the arterial blood begin to fall in most persons. Some of the first studies took place in the Andes, where approximately 6 million indigenous Aymara and Quechua (whom we shall refer to here as Andeans) populations reside, chiefly in Peru and Bolivia, however also in neighboring countries. Andeans are derived from the early settlers of the Americas who reached South America 15 to 16 thousand years ago (kya) and then split into two branches, one that settled in the Pacific coastal and Andean regions and the other that moved along the Atlantic coast and then eastward [1]. Of interest, there has been little admixture between Andeans with the descendants of the more easterly groups as attested to by mitochondrial and autosomal genetic markers, demonstrating the Andeans' genetic continuity and substantial isolation from other South American groups [1,2].

Initial debate as to whether genetic adaptation to high altitude has taken place was driven by differences in theoretical orientation and the sources of evidence being considered. Theoretical

orientation influenced the way in which the term “adaptation” was being employed. Physiologists used the term to refer to any response regardless of whether it was likely to be beneficial or otherwise affect the chance(s) of being able to live or reproduce, whereas evolutionary biologists or geneticists restricted its usage to those responses likely to influence reproductive success [3]. Evidence for adaptation was predominantly sought from studies separating short-term physiological responses or those occurring over hours to days or even weeks, termed acclimatization, from those occurring across lifetimes, termed developmental responses, and from those that persisted independent of duration of high-altitude exposure and were inferred to be genetic. The migration model, introduced in the 1960s, was productively used to distinguish between acclimatization, developmental, and presumed genetic responses [4–6]. The advent of single nucleotide polymorphism technologies and statistical methods for detecting evidence of natural selection constituted a paradigm shift and resulted in an exponential rise in the number of publications reporting genetic adaptation [7]. While multiple studies have shown that Andean and other high-altitude populations have undergone natural selection in several gene regions influencing O₂-sensitive pathways, numerous questions remain regarding the biological processes driving human adaptation to the chronic hypoxia of high altitude and their importance for human health.

This review discusses the kinds of evidence by which adaptation to high altitude has been assessed and which have led to widespread acceptance of the idea that genetic adaptation to high altitude has occurred. Studies in Andean residents of high altitude are summarized with respect to the physiological characteristics distinguishing them from acclimatized newcomers and the genomic or genetic factors that are potentially involved. While further study is needed, such studies offer the opportunity to identify the importance of interactions between genomic and epigenomic processes for human adaptation to limited oxygen availability.

2. Genetic Adaptation of Andean High-Altitude Populations

Two kinds of information support the existence of Andean genetic adaptation to high altitude. First, indirect evidence provided by physiologic studies demonstrates that native highland populations exhibit unique O₂ transport traits when compared with acclimatized newcomers that cannot be attributed to developmental processes. Second, direct evidence comes from genomic studies that show signals of recent positive selection in specific gene regions. However, despite the remarkable progress in recent years for identifying targets of natural selection and the recognition that many are involved in O₂-sensitive signaling pathways, few investigations have been able to show how these gene regions affect specific physiological characteristics and how, in turn, such effects influence reproductive success. From an evolutionary point of view, these relationships are essential since, by definition, only genes with effects on reproductive success are acted upon by natural selection.

2.1. Physiologic Evidence of Genetic Adaptation to High Altitude

Since the hypoxia of high altitude challenges O₂ homeostasis, there has been a long history of studies of the O₂ transport system and its components (arterial O₂ content, distribution, and utilization) in acclimatized newcomers, Andeans, and other long-term residents of high altitude for establishing the physiologic mechanisms underlying human adaptation to high altitude.

2.1.1. O₂ Content

The partial pressure of O₂ in the arterial blood (P_aO₂) is determined by alveolar ventilation and the alveolar-arterial (A-a) O₂ gradient (Figure 1). Since the A-a O₂ gradient is minimal in healthy persons, alveolar or end-tidal PCO₂ (P_ACO₂) can serve as a proxy for arterial PCO₂ (P_aCO₂). Additionally, since according to the alveolar air equation P_aCO₂ is inversely related to alveolar ventilation, P_ACO₂ can serve as an index of alveolar ventilation per unit of CO₂ production (or metabolic rate given that, normally, CO₂ production and O₂ consumption are closely coupled). At low altitude, P_ACO₂ averages ~40 mmHg, however it falls to ~10 mmHg at elevations above 3000 m and even further at

extreme altitudes. $P_A\text{CO}_2$ or alveolar ventilation is lower in Andeans and acclimatized newcomers than sea-level residents, however Andean values are somewhat higher than those of acclimatized newcomers, indicating lower levels of alveolar ventilation (Table 1) [8–10]. Consistent with this, the hypoxic ventilatory response of Andeans is blunted compared to acclimatized newcomers. Indicating a genetic component, greater indigenous (Quechua) ancestry is directly related to the blunting that is observed [11].

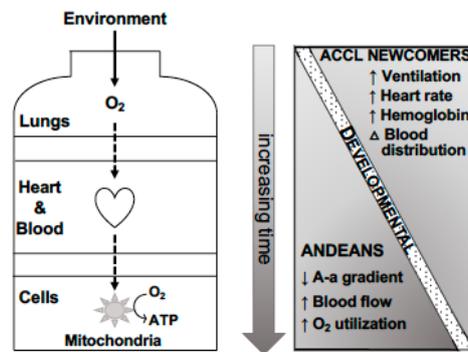


Figure 1. The O_2 transport system and its temporal changes. The O_2 transport system consists of two pumps (the lungs and the heart), two diffusion steps (alveoli to arterial blood, capillary blood to the mitochondria), and the mitochondria where O_2 is consumed to generate chemical energy, adenosine triphosphate (ATP). Increases in the levels of ventilation, heart rate, and hemoglobin as well as changes in blood flow distribution to favor high demand organs occur with acclimatization. Developmental changes increase lung volume. In Andeans, multigenerational high-altitude residence produces further changes in the alveoli to arterial O_2 gradient, regional blood flow, and O_2 utilization. See text for details and references. Adapted from [12].

Acclimatization does not appreciably change the A-a O_2 gradient, however the gradient is lower in Andeans (and other lifelong of high- compared to low-altitude residents), enabling Andeans to achieve an increased efficiency of O_2 transfer [13–15]. This enhanced efficiency is likely the result of greater total lung capacity and, especially, residual volume in Andean populations compared with those residing at low altitudes [16] and gives rise to the “barrel-chest” Andean morphology. Developmental processes play a key role in such lung-volume expansion with increased lung growth being apparent from infancy through adolescence not only in Andeans, however also other high-altitude residents, including Coloradans and even experimental animals born and raised under hypoxic conditions [17,18]. Genetic factors are also implicated insofar as Andean total lung volumes are greater than values seen in lowlanders born and raised at high altitude [19], and the proportion of indigenous American ancestry is directly related to residual volume yet, interestingly, not chest dimensions [20]. However, there appears to be an interaction between Andean ancestry and high-altitude residence that acts to increase the altitude-associated expansion of chest dimensions and lung volumes [19] and reduction in stature and limb measurements [21]. While more studies are needed with controls for confounding factors, the improved efficiency of O_2 transfer is likely important for maintaining arterial O_2 saturation and thus, blood O_2 content during exercise [16,22].

The position of the hemoglobin- O_2 dissociation curve determines the level of arterial O_2 saturation (SaO_2) for a given PaO_2 . Little change occurs in the *in vivo* curve position with acclimatization since the left curve-shifting effect of respiratory alkalosis is offset by an increase in red blood cell 2,3-bisphosphoglyceric acid levels. Current evidence does not support differences in Andean versus acclimatized newcomer curve position (J. Prchal, personal communication) [23–25] (Table 1). Of interest, the I-allele of the angiotensin-converting enzyme has been associated with higher SaO_2 in Quechua regardless of whether they were born at high altitude or at low altitude yet were exposed transiently to high altitude [26], however this allele is not unique to highlanders as it is found in human populations worldwide.

Hemoglobin levels rise with ascent to high altitude in both men and women, due initially to a contraction in plasma volume and subsequently to a rise in red blood cell production. Generally, hemoglobin levels in acclimatized newcomers are similar to those present in healthy Andeans (Table 1). Urban values are higher than those of rural residents [27,28], perhaps due to dust exposure [29] or the greater levels of admixture present in Andean mining communities [30]. While modest increases in hemoglobin and red cell production are considered beneficial at high altitude, excessive erythrocytosis is maladaptive; a detailed discussion of this topic is provided in Section 2.2.2.

Arterial O₂ content (CaO₂) is either measured directly or calculated using hemoglobin values (in gm/dL multiplied by 1.36, the mls of O₂ bound per gm) multiplied by SaO₂, using a correction factor for dissolved O₂. Acclimatized newcomers re-establish the sea-level value of ~21 vol % due to their hyperventilation, which helps to overcome the initial fall in PaO₂, and increased hemoglobin. Andean and acclimatized newcomers achieve similar CaO₂ levels due to the Andeans' narrower alveolar-arterial O₂ gradient offsetting their somewhat lower levels of ventilation.

2.1.2. O₂ Distribution

Acclimatized newcomers and Andeans have similar levels of cardiac output at a given workload, however values are lower at maximal exercise in both groups compared to sea level [31]. Despite considerable research, the cause of the reduction in maximal cardiac output remains unclear, leading some to suggest that cardiac output is actively suppressed by central nervous system factors [31,32]. Decreased filling is unlikely since low- and high-altitude residents do not differ in terms of blood volume [33]. Increased afterload due to higher pulmonary arterial pressures could be a factor, however values are lower in healthy Andeans compared with acclimatized newcomers and systemic (left heart) pressures are modestly lower in Andeans yet cardiac output is similar [34,35]. While continuing to be debated, a substantial number of studies indicate that Andeans have higher maximal O₂ consumption than acclimatized newcomers and less altitude-associated decrement [36].

Table 1. Determinants of O₂ transport in long-term highland groups and acclimatized newcomers at ~3600–4300 m.

Variable	Acclimatized Newcomer	Andean	Andean versus Accl newcomer
P _A CO ₂ , mmHg	30 [8]	Higher [8,32]	↑ω
A-a O ₂ , mmHg	7–11 [14,15]	Lower [4,15]	↓
SaO ₂ , %	92 [16,19,37]	Same [9,37–40]	≈
Hemoglobin, g/dL	17.6 [16,19]	Same [9,17,21]	≈
CaO ₂ , vol% ¹¹	21 [16,19]	Same [19]	≈
Ppa hypoxic response	Present	Intermediate [41]	↓
Brain blood flow velocity, cm/s	27 [42]	18% [39]	↓
Uterine artery blood flow, mL/min	269 [43]	Higher [43]	↑

Abbreviations: A-a DO₂ = alveolar to arterial O₂ diffusion gradient, Accl = acclimatized, CaO₂ = arterial O₂ content, P_ACO₂ or P_{ET}CO₂ = alveolar or end-tidal PCO₂, Ppa = pulmonary arterial pressure, SaO₂ = arterial O₂ saturation. Numbers in the table are mean values or, in cases where few data are available, ranges. References are given in parentheses.

O₂ distribution is determined by regional blood flow. There is increased sympathetic nervous system stimulation in acclimatized lowlanders at high altitude which likely reduced blood flow to the periphery [44,45]. Blood flow to the leg and fractional O₂ extraction during exercise are reduced in Andeans compared with acclimatized lowlanders as a result of blood being diverted to other tissues [46]. Blood flow velocity through the internal carotid, middle cerebral, and vertebral arteries has been used as indices of brain blood flow; however it should be recognized that blood flow is also a function of the vessel diameter or cross-sectional area. Highland Andeans appear to have lower resting middle cerebral flow velocities than at low altitude, however unchanged O₂ delivery due

to higher hemoglobin levels [39,47] (Table 1); the adaptive significance of reduced middle cerebral blood flow, however, is not clear. Cerebral blood vessels are highly responsive to blood gas changes or bioactive molecules such as NO. Andeans had less middle cerebral artery vasodilator response to hypoxia or NO at high compared to low altitude, and less vasoconstrictor response to hypocapnia than Sherpa [48]; however whether such responses differed from those of acclimatized newcomers was not studied. Compared to acclimatized lowlanders, Andeans distribute a larger proportion of pelvic blood flow to the uteroplacental circulation during pregnancy, which in turn raises uterine artery blood flow and uteroplacental O₂ delivery [49–51] (Table 1). Andean protection was accompanied by greater antioxidant levels and more angiogenic relative to anti-angiogenic substances [52,53]. Developmental factors were not responsible since the Andeans' uterine artery blood flow was greater than that seen in Europeans who were born and raised at high altitude [54]. Cortisol levels are also lower in pregnant Andeans than acclimatized newcomers [55], perhaps reflecting less sympathetic stimulation. Greater vascularity, as observed in the placental [56] and the skin microcirculation in neonates [57], both could also increase blood flow.

2.1.3. O₂ Utilization

O₂ delivery to the mitochondria generates chemical energy or ATP (adenosine triphosphate) (Figure 1). Of interest is that the efficiency with which ATP is produced varies by fuel source. Specifically, the metabolism of carbohydrates (glucose, glycogen) generates 25–50% more ATP per mole of O₂ consumed than is the case with the use of free fatty acids or lipids [58]. Carbohydrates become the preferred fuel in males after three weeks of altitude acclimatization [59]; however, interestingly, not for females [60]. Only a few studies have been performed in long-term high-altitude residents. Specifically, using positron emission tomography to measure heart metabolism, Hochachka and co-workers found greater reliance on carbohydrate metabolism in Quechua males studied at sea level and 50–60% more ATP produced per mole of O₂ consumption compared with lowlanders [58]. The authors concluded that Quechua hearts displayed increased O₂ efficiency, representing a biochemical adaptation for defending against hypoxia [32]. Residence at high altitude also alters glucose metabolism. Glucose uptake is increased, glucose tolerance is improved, and consequently, venous glucose levels are lower at high altitude [61,62] as well as in pregnant Andeans [63]; this was interpreted as reflecting greater placenta glucose uptake in order to spare O₂ for fetal consumption [64].

In summary, the several unique O₂ transport characteristics of Andeans compared to acclimatized newcomers that are not due to developmental factors—namely, lower alveolar ventilation, lower pulmonary vasoconstrictor response, larger lung volumes, higher uterine artery and possibly lower middle cerebral blood flow, less altitude decrement in maximal exercise O₂ consumption, and more efficient cardiac O₂ utilization—suggest a greater efficiency of O₂ transfer and utilization and are consistent with the likelihood of Andean genetic adaptation to high altitude.

2.2. Genomic Evidence of Andean High Altitude Adaptation

Direct evidence for Andean genetic adaptation to high altitude comes from single nucleotide polymorphism (SNP) genome scans and sequencing studies that have identified genomic regions with evidence of recent positive selection (Table 2). Genes that regulate or are regulated by the hypoxia-inducible factor (HIF) pathway have been of particular interest. HIF consists of two α -subunits (HIF1 α and HIF2 α) and a constitutively expressed β -subunit [65,66]. In normoxia, O₂-dependent negative regulators of HIF called prolyl hydroxylases (PHDs) enable the hydroxylation of proline residues of HIF1/2 α subunits [67]; this promotes the binding of von Hippel–Lindau tumor suppressor (vHL) protein and, subsequently, degrades the HIF1/2 α [68,69]. In a hypoxic environment, HIF1/2 α are not hydroxylated by PH and therefore escape recognition by vHL, allowing these subunits to bind with hypoxia responsive elements (HRE) within gene promoters and associated cofactors to initiate HIF-regulated gene transcription [70]. While there are more than 100 genes containing response elements to which HIF can bind, existing SNP data indicate that the HIF-pathway has not been

disproportionately acted upon by natural selection [71]. Further, not all O₂-sensitive genes contain HREs. Therefore HIF, while certainly central for governing transcriptional responses to hypoxia, is not the *only* regulator of molecular responses to changes in O₂ tension.

Table 2. Autosomal gene regions acted upon by natural selection in Andean populations.

<i>AS3MT</i> [72]	<i>EDNRA</i> [71,74]	<i>NOS2</i> [71,73,74]	<i>TBX5</i> [73]
<i>BRINP3</i> [73]	<i>EGLN1</i> [71,74]	<i>PRKAA1</i> [71,74]	<i>TMEM38B</i> [72]
<i>CLC</i> [72]	<i>ELTD1</i> [75]	<i>SFTPD</i> [77]	TP53 pathway [78]
<i>DUOX2</i> [72]	<i>ET-1</i> [76]	<i>SP100</i> [72]	<i>VEGFB</i> [75]
	<i>FAM213A</i> [77]		

Abbreviations: *AS3MT* = arsenite 3 methyltransferase; *BRINP3* = BMP/retinoic acid inducible neural specific 3; *CLC* = galectin-10; *DUOX2* = dual oxidase 2; *EDNRA* = endothelin receptor type A; *EGLN1* = egl-9 family hypoxia inducible factor 1; *ELTD1* = adhesion G protein-coupled receptor L4; *FAM213A* = family with sequence similarity 213 member A; *NOS2* = nitric oxide synthase 2; *PRKAA1* = protein kinase AMP-activated, α 1 catalytic subunit; *SFTPD* = surfactant protein D; *SP100* = SP100 nuclear antigen; *TBX5* = T-box 5; *TMEM38B* = transmembrane protein 38B; *TP53* = tumor protein p53; *VEGFB* = vascular endothelial growth factor B.

The first genome scan to study high-altitude adaptation was performed in Andeans residing in Bolivia [76]; however to date, there have been fewer studies in Andeans than Tibetans. The peopling of the Andes appears to have begun 12,000 or more years ago [1,2,79], a timeframe that would be expected to permit the natural selection of genes that have at least a modest effect on reproductive success. Just one gene, *EGLN1*, has thus far been identified as being acted upon by natural selection in both Andeans and Tibetans [71]. Among Andeans, several other genes showing evidence of natural selection have been identified, including some involved in vasoregulation (*PRKAA1*, *NOS2*), vascular growth (*VEGFB*, *ELTD1*), cerebral blood flow (*CBS*), and oxidative defense (*FAM213A*) [71,75,77] (Table 2). There has just been one whole-genome sequencing study in Andeans to date; it identified three gene regions—*BRINP3*, *NOS2*, and *TBX5*—with just one (*NOS2*) having been identified previously in a SNP scan [71]. These genes have previously been associated with cardiovascular function, however not hypoxia-sensing [73]. Of note, while we commented above on the role of genetic and developmental factors for the larger lung volumes seen in Andeans, no study to the best of our knowledge has sought to determine the relationship of such morphological variation with any of the gene regions identified as having been acted upon by natural selection.

To determine whether genomic regions acted upon by natural selection provide an adaptive advantage in the high-altitude environment, it is essential to understand the functional consequences of the variants identified. Residence at high altitudes poses several challenges for reproductive success; such challenges occur during the perinatal (i.e., from conception through infancy), adolescent, and adult periods, with the heaviest concentration occurring during perinatal life (Figure 2).

ADAPTIVE CHALLENGES OF HIGH ALTITUDE

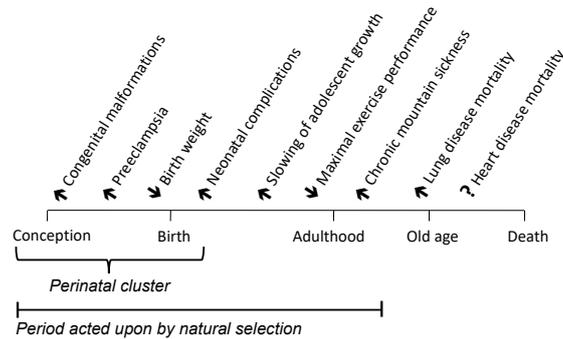


Figure 2. Adaptive challenges or those affecting reproductive success occur at high altitudes at multiple times across the lifespan. About half the cluster during the perinatal period or that from gestation through to the first week of postnatal life, with the remainder occurring during adolescence or adulthood (see text for references). Adapted from [12].

2.2.1. Perinatal Complications

Residence at high altitude reduces birth weight as the result of slowed fetal growth rather than shortened gestation [80,81]. Andean infants have half as much birth-weight reduction at high altitude as acclimatized newcomers (Figure 3A), with the magnitude of protection being greater in the Andean populations in the southern (southern Peru, Bolivian) compared to the more northerly region, likely reflecting the duration of high-altitude residence and the extent of forced migration by Incan rulers as well as foreign admixture [82]. Andean protection from altitude-associated reductions in birth weight is directly related to the amount of indigenous American, specifically Andean, ancestry [83–85], suggesting genetic involvement. Enhanced uteroplacental blood flow and O₂ delivery, resulting in part from a larger pregnancy-associated rise in uterine artery diameter [50,51,86,87], is an important factor contributing to Andean protection from fetal growth reductions at high altitude. Numerous studies have shown associations between low uterine artery blood flows, decreased birth weights, and fetal demise [51,88–90], supporting the likelihood that maintenance of high uterine artery blood flow is important for normal fetal growth at high altitude. Greater blood flow, not CaO₂, is responsible for raising uteroplacental O₂ delivery since CaO₂ is similar in Andeans and newcomers [51,86]. Placental factors may also be involved since placenta weight, both absolute and relative to fetal weight, is greater in Andeans than acclimatized newcomers [40], and Andean placentas have enhanced villous capillarization and vascular remodeling [91].

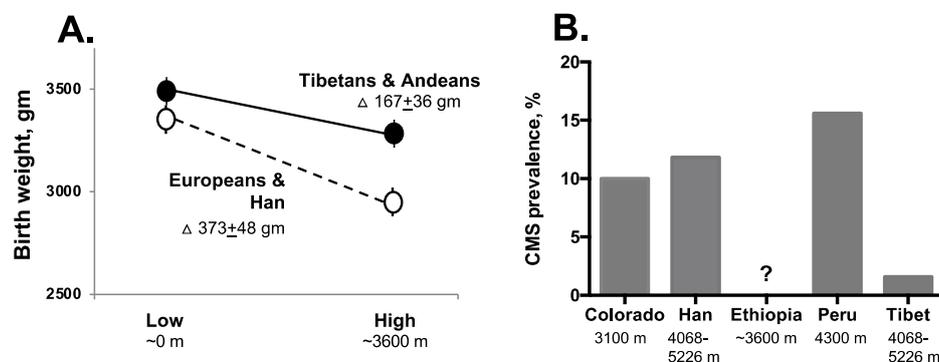


Figure 3. A. Tibetans and Andeans have approximately half the altitude-associated reduction in infant birth weight compared with Europeans or Han Chinese (see text for details and [78] for original references). **B:** prevalence of chronic mountain sickness (CMS) is markedly less in Tibetans than similarly aged men from various ancestry groups residing at the altitudes shown. [Adapted from Niermeyer et al. [78] with permission from SAGE Publications.].

High altitude also increases the incidence of preeclampsia, an effect that contributes to the altitude-associated decline in birth weight. Preeclampsia is a multisystem vascular disease of placental origin that complicates roughly 8.5 million pregnancies worldwide each year. It not only poses a threat to maternal and perinatal survival but also increases the risk of cardiovascular disease in affected mothers and offspring later in life [92–97]. Existing evidence suggests that high-altitude residence increases the incidence of preeclampsia in Andeans as well as acclimatized newcomers [98–100]. However, the lack of systematic assessment of diagnostic criteria and vital-statistic databases in Andean countries has thus far prevented determination as to whether indigenous women are relatively protected compared with acclimatized newcomers. Supporting such a possibility, sFlt-1 levels and sFlt-1/PLGF ratios are lower in pregnant Andeans compared with Europeans living at the same altitudes [53], both of which are protective against preeclampsia [101]. Andeans may also benefit from higher antioxidants levels [52] as well as higher levels of progesterone, estrone, 17- β estradiol, and estriol [55]. Andean pregnant women also have lower cortisol levels than Europeans, with their lower cortisol and higher estriol levels being associated with greater uterine artery diameter and blood flows [55].

Given that compromised fetal growth raises the risk of perinatal mortality, an outcome of direct relevance for reproductive success, we tested the relationship between birth weight and 63 single nucleotide polymorphisms in 16 genes with evidence of natural selection at high altitude while making corrections for multiple comparisons [102]. Several SNPs near *PRKAA1* (coding for the α -1 catalytic subunit of adenosine monophosphate kinase, AMPK) and *EDNRA* (coding the vascular smooth muscle cell endothelin receptor A) were associated with the preservation of birth weight at high altitude; however only *PRKAA1* was also associated with larger uterine artery diameters. In addition, the expression of mTOR-pathway genes in circulating peripheral blood mononuclear cells—a pathway known to play a crucial role in mediating the effects of hypoxia, nutrient restriction, and other factors on fetal growth—differed in women with versus without the selected-for maternal *PRKAA1* genotype, suggesting that AMPK may play an important role in vascular adaptation to pregnancy [103–105].

Limited data suggest that native compared with acclimatized newcomer groups have better neonatal outcomes. Perinatal mortality is generally higher at high than low altitudes in South America, with the altitude-associated increase being least in the regions of Peru where populations have lived the longest [82]. Infants of mixed Native American and European ancestry residing at high altitudes in Bolivia spent ~80% of the night with SaO₂ values below 90%, with lower proportions of the night being seen in children and adolescents [106], but sample sizes and composition were not sufficient to address the possible differences between ancestry groups.

2.2.2. Chronic Mountain Sickness

Slight elevations of red cell mass increase arterial O₂ content under conditions of ambient hypoxia. In contrast, however, excessive red blood cell production, as observed in chronic mountain sickness (CMS), increases in blood viscosity and impairs blood flow and O₂ delivery to tissues [73]; for this reason, excessive erythrocytosis is considered to be maladaptive. CMS has long been known to occur at high altitudes [107] and can lead to pulmonary hypertension and right or left heart failure. While such deaths typically occur after the end of the reproductive period, the disease begins in early adulthood and may impact fitness given that affected individuals are no longer able to engage in normal daily activities [108,109].

CMS prevalence varies between highland resident populations (Figure 3B). For instance, CMS has been reported to occur in ~10% of Andean, Coloradan, or Han males over the age of 30 or post-menopausal females, while a smaller proportion of Tibetans are affected [28,110] and, to date, CMS has not yet been reported in Ethiopians [38,111]. CMS has a gradual onset, being seen in 15–25 year old males as preclinical CMS, defined as >2 standard deviations above the mean hemoglobin level or 18.3 g/dL, together with accompanying signs or symptoms [108] and worsening with advancing age [109,112,113]. Persons with CMS have lower levels of ventilation than acclimatized newcomers;

however this is also true for healthy Andeans, suggesting that hypoventilation may be necessary yet not sufficient for the development of CMS. Breathing during sleep is likely a key component, with sleep-disordered breathing (apneas, hypopneas) more common in both clinical and preclinical patients [108,109,114–116]. Cerebral blood flow is also affected by sleep-disordered breathing [117]. The middle cerebral artery vasodilator response to NO is blunted, carotid artery intimal thickness greater, and flow-mediated brachial artery vasodilation is impaired in Andean men with versus without CMS [118]. Early-life hypoxic exposures may also play a role. Adults with exaggerated hypoxia as neonates showed higher pulmonary arterial pressures during acute-altitude exposure [119]. Compared with healthy controls, high-altitude residents with CMS were more often small-for-gestational age [112], born to a preeclamptic mother, or to have experienced exaggerated neonatal hypoxia [109]. Thus, perhaps hypoxic or oxidative injury during perinatal life predisposes individuals to develop CMS later in life due to impaired pulmonary or cerebral vascular development.

With respect to genetic factors in CMS, *EGLN1*, and *EPAS1*, variants related to hemoglobin levels in Tibetans are not so related in Andeans [120], suggesting that these specific variants may not be involved in increasing Andean susceptibility. A limited whole genome sequencing comparison of 10 men with versus 10 men without CMS identified 11 regions that differed by CMS status [121]. Using a fibroblast cell-culture model, acute hypoxia upregulated two of these genes' transcriptional responses (*SENP1* and *ANP32D*, known to play roles in regulating erythropoiesis and cellular metabolism, respectively) in CMS patients, but not in controls. The association between *SENP1* and CMS (however not *ANP32D*) was replicated in a larger sample of CMS and control residents of 4338 m [122]. Other genes, such as *SENP1* which codes for a protease that rescues HIF1alpha from degradation, have also been suggested to play a role in increasing susceptibility to CMS [123].

3. Speculation on the Role of Epigenetics for Andean High-Altitude Adaptation

Genomic studies are well positioned to reveal functional links between genetic regions that appear to have been subject to recent positive selection and adaptive phenotypes of highland populations. It is critical, however, to recognize that phenotypes are the objects on which selective pressures act and are seldom the product of genetic factors alone. Complex phenotypes most often arise through gene-gene and gene-environment interactions, as well as the functional interaction of the genome and epigenome. Epigenetic marks are non-sequence-based features of the genome that are vital for coordinating transcriptional responses to environmental stimuli. In this way, the epigenome acts as an interface through which genetic sequence is “translated” to generate physiological responses to shifting biological or environmental conditions. This section presents evidence supporting the possibility that epigenetic processes contribute to human high-altitude adaptation, emphasizing the role of epigenetics for transcriptional and developmental responses to limited oxygen availability, epigenetic inheritance, and genome-epigenome interactions. Existing literature largely focuses on transient epigenetic effects. However, several recent investigations have explored mechanisms for epigenetic inheritance and the importance of genome-epigenome interactions for driving physiologic responses and phenotype. Taken together, this work indicates that epigenetic modifications could provide a mechanism for the rapid acquisition of potentially heritable features. This flexibility, itself, could be viewed as a selective advantage during periods of rapid environmental change or periods of the lifespan, such as the perinatal life or pregnancy which require widespread physiological changes over a short time period [124].

Numerous epigenetic mechanisms exist, such as DNA methylation, histone modification, RNA-based mechanisms, and histone variants. DNA methylation, the most well-studied epigenetic modification in humans, is defined by the addition of a methyl group to cytosine residues within CpG dinucleotides. Research has predominantly focused on DNA methylation because of its central involvement in the regulation of gene transcription, genomic imprinting, and the silencing of repetitive DNA elements [125,126]. While the majority of CpG sites across the human genome are methylated [127,128], genomic “islands” of high CpG density (“CpG islands”) are scattered throughout

the genome. These regions are generally devoid of methylation, thereby allowing for transcription factor binding and active gene transcription. Hypermethylation of CpG sites within CpG islands typically impedes transcription factor binding, thereby establishing a dormant chromatin state [129]. However, the methylation state of CpG sites within enhancers, gene bodies [130,131], and low-density CpG regions [132] also influences gene expression and alternative splicing.

3.1. Epigenetics and Transcriptional Responses to Hypoxia

Epigenetic processes are essential to the regulation of the HIF transcriptional program by, for instance, silencing HIF-stabilization genes, including von Hippel-Lindau (*VHL*) and *EPAS1* [133,134]. De novo methylation of *EPAS1* promoter CpG sites by DNA methyltransferase 3a also prohibits HIF2 α -mediated gene expression under hypoxic conditions [134]. DNA methylation events also govern the hypoxic-induction of erythropoietin, a pleiotropic cytokine that is recognized as the central driver of red blood cell production [135]. Moreover, enzymes that alter the epigenetic status of histones and cytosine residues (histone acetyltransferases and demethylases, respectively) are regulated, in part, by hypoxia and are involved in determining chromatin conformation within and around HIF-binding sites [136–138]. In this way, epigenomic marks would be expected to influence the “translation” of genomic sequence into physiological responses to acute hypoxic exposure and, potentially, durable phenotypic traits at high altitude [129,139]. Following the same logic, interruption of epigenetic processes that are essential to regulate the HIF-transcriptional program could compromise or augment transcriptional responses that are important to sustain oxygenation under conditions of limited O₂ supply, such as at high altitude.

3.2. Epigenetics and the Developmental Programming of Physiological Responses to Hypoxia

Epigenetic processes are considered central for the effects of intrauterine or early-life exposures on organ system development given the well-established role of epigenetics for determining cellular identity, their responsiveness to environmental and biological cues, and the particular vulnerability of the epigenome to environmental insults in early life. For instance, during embryonic development, the differentiation of genetically-identical pluripotent cells into hundreds of distinct cell types is driven largely via epigenetic mechanisms [140,141]. Existing evidence suggests that the epigenome is involved in the effect of environmental exposures occurring during developmental periods to influence physiological responses to hypoxia in later life. In mice, intrauterine hypoxia induces hypermethylation of CpG motifs located within the protein kinase C epsilon promoter, a gene that encodes a protein known to enhance cardiovascular hemodynamics in ischemia-reperfusion injury, thereby reducing cardiac protein kinase C epsilon expression and, ultimately, increasing the risk of ischemia-reperfusion injury in later life [142–144]. Other studies also support the involvement of epigenetic factors for the fetal programming of pulmonary vascular dysfunction. Maternal undernutrition in pregnancy, for example, exaggerates the affected offspring’s pulmonary vascular response to hypoxia and modifies global DNA methylation of the lung [145]. Moreover, treatment of offspring with histone deacetylase inhibitors normalized pulmonary vascular function and DNA methylation status [145].

Existing evidence indicates that perinatal hypoxia may also influence pulmonary vascular function at high altitude in humans. Specifically, lowlanders who experienced transient perinatal hypoxic pulmonary hypertension had an exaggerated pulmonary artery pressure response with high-altitude exposure (4559 m) as adults compared to lowlanders who did not experience hypoxic pulmonary hypertension during perinatal life [119]. Among Andeans residing in La Paz or El Alto, Bolivia (3600–4100 m), adverse oxygenation during perinatal life increases the risk of a preclinical form of CMS and attendant pulmonary vascular dysfunction during young adulthood [109]. Infants born to preeclamptic women at high altitudes also have higher basal pulmonary artery pressure [146]. Further, infants born to preeclamptic women who went on to develop abnormal pulmonary vascular function at high altitude during later life show unique methylation-expression relationships within numerous genes that are important for vascular function, suggesting that epigenetic effects may influence the

relationship between pulmonary hypertension and preeclampsia [109]. While much work remains to be done, existing evidence supports the hypothesis that impaired perinatal oxygenation induces epigenetic modifications influencing physiological responses to hypoxia during adulthood.

3.3. Inheritance of Epigenetic Marks

From an evolutionary point of view, the relevance of epigenetic marks or the capacity for epigenetic modification for human adaptation depends upon the heritability of epigenetic features themselves and/or the capacity for epigenetic modification in particular regions of the genome. Epigenetic heritability, that is the inheritance of epigenetic marks themselves, remains contentious primarily because non-imprinted genes undergo widespread, yet incomplete, epigenetic reprogramming prior to implantation [147]. However, existing literature supports the persistence of environmentally-induced DNA methylation changes across generations [145,148,149] and the transmission of DNA methylation marks through the germline and somatic pathways [150]. Potential mechanisms for pure transgenerational epigenetic inheritance include constitutional epialleles (epigenetic marks that originate from the early embryo or parental germ line) that are, in some instances, retained across meiotic division [150]. One report further reveals that somatic epigenetic modifications may not need to be carried through the gamete intact, but may rather be transmitted via epigenetic-modifying RNA species [151]. Through this mechanism, heritable DNA methylation marks could avoid the widespread epigenetic reprogramming that occurs during early development. DNA methylation status is also heavily influenced by genetic variation, particularly within CpG motifs [152–156]. One report indicated that up to 80% of genetic variants that disrupt CpG sites alter the methylation status of local CpG sites as well as those located up to 10 kb distant [157].

In short, evidence supporting epigenetic heritability raises novel questions about how genetic sequence orchestrates physiological responses and durable adaptations to environmental exposures such as high altitude. Much work remains to be done in this area, particularly with respect to direct epigenetic inheritance. In the context of human adaptation, understanding the impact of putatively adaptive genetic variants that modify CpG motifs on the epigenetic regulation of gene expression should be of particular interest.

3.4. Querying Genomic-Epigenomic Interactions in High-Altitude Populations

As discussed above, SNPs that disrupt (or create) CpG sites are important determinants of epigenetic capacity or, in other words, the potential for epigenetic regulation of gene expression. Prior work has speculated that epigenetics may be involved in high-altitude acclimatization and adaptation [158] and the development of hypoxia-associated pulmonary vascular dysfunction in high-altitude Andeans [159]. However, only three publications report site-specific DNA methylation differences in high-altitude populations [160–162], including one paper that presented the hypothesis that genetic variants showing evidence of recent positive selection in high-altitude populations could affect the capacity for the epigenetic modification of gene transcription under hypoxic conditions [162]. Specifically, Julian notes that nearly 40% of the putatively adaptive *EPAS1* SNPs in high-altitude populations modified CpG content [162]. This observation is important in the context of high altitude for several reasons. First, putatively adaptive *EPAS1* SNPs are apparent in high-altitude populations and have been associated with reduced hemoglobin concentrations in some native highland populations (i.e., Tibetans) [163]. Second, the *EPAS1* promoter lies entirely within a CpG island and is epigenetically regulated under hypoxic conditions [134]. Third, *EPAS1* encodes HIF-2 α and therefore may be of functional importance for adaptation to hypoxia. Finally, given that CpG-modifying SNPs can influence methylation [157], heritable differences in CpG density may promote or inhibit the epigenetic modifications that influence transcriptional responses to environmental hypoxia. For instance, if a SNP were to decrease CG content in regulatory regions of the genome, there would be less (or no) opportunity for epigenetic regulation of gene expression via DNA methylation (i.e., less plasticity). Alternatively, a SNP that increased CG content in regulatory

regions may be more permissive of epigenetic regulation via DNA methylation (i.e., more plasticity). While this concept is provocative, it is also somewhat premature and requires further investigation.

Epigenomic processes may also contribute to maladaptive phenotypes of high-altitude populations, including CMS in highland Andeans. Julian contrasted peripheral blood mononuclear cell DNA methylation patterns between Andean men living in La Paz-El Alto, Bolivia who presented with a preclinical form of CMS and healthy controls [162]. Of the numerous differentially methylated regions identified, the most notable differentially methylated region (DMR) associated with preclinical CMS was the hypermethylation of *EGLN1* [162], a gene that encodes PHD2. Given that PHD2 negatively regulates the HIF-transcriptional program via promoting the proteasomal degradation of HIF1/2 α [67], the hypermethylation of *EGLN1* would be anticipated to diminish PHD2 expression and thereby enable the transcription of HIF-regulated genes such as erythropoietin. In support of the hypothesis that hypermethylation of *EGLN1* contributes to the excessive production of red blood cells in CMS, *EGLN1* inactivation in mice results in an overproduction of erythropoietin and polycythemia, and familial polycythemia in humans has been linked to *EGLN1* mutations [164–166]. Further investigations are needed to not only test this hypothesis, however also to evaluate the functional importance of the DMRs identified.

4. Summary, Conclusions, and Directions for Future Work

In summary, there are several unique O₂-transport characteristics of Andeans compared to acclimatized or lifelong newcomer residents of high altitude. As reviewed above, these are lower alveolar ventilation, lower hypoxic pulmonary vasoconstrictor response, slightly larger lung volumes, higher uterine artery and possibly lower middle cerebral blood flows, less altitude decrement in maximal exercise O₂ consumption, and more efficient cardiac O₂ utilization. Collectively, these are suggestive of greater efficiency in O₂ transfer and utilization; in turn, such differences between acclimatized or lifelong high-altitude residents support the existence of Andean genetic adaptation to high altitude.

Direct support for Andean genetic adaptation to high altitude comes from SNP genome scans and whole-genome sequencing studies. Genome scans can be performed at relatively low cost and in large numbers of persons, but only sample a small portion of the genome [1]. They have shown that natural selection has acted on a gene region that is involved in regulating the HIF-pathway, *EGLN1*, and on others that are not in the HIF-pathway yet are O₂ sensitive, underscoring the importance of looking broadly at the range of genetic factors potentially involved. Whereas whole-genome scans are necessarily more complete, they are considerably more expensive and hence difficult to conduct in large numbers of persons. Thus more, especially high-coverage whole-genome sequencing studies, are needed. The one whole-genome scan to date indicates, intriguingly, that selection has acted not only on genes that are involved in O₂ sensing, but also on those regulating cardiovascular responses to hypoxia [73].

Future studies are also required to provide deeper exploration of the associations between selected-for genotypes and phenotypic traits that are likely to influence reproductive success. The inclusion of epigenomic factors in such studies is also vital as the few studies conducted to date indicate potentially key roles for epigenetic regulation of gene transcription in ways that could affect reproductive fitness. While functional studies on the impact of locus-specific methylation status remains challenging, the advent, for example, of genome-editing technologies such as the CRISPR/Cas-9 system permit the induction of targeted CpG methylation and demethylation events *in vitro* as well as *in vivo* experimental animal models [167]. Transcription Activator-Like Effector Nucleases (TALENs), another genome-editing technique, can also be used to target locus-specific CpG methylation sites [168,169]. Using these strategies, future experimental models could be developed to determine whether hypomethylation or hypermethylation of specific CpG sites affect molecular and, ultimately, physiological function.

In short, the singular nature of the hypoxic stress posed by residence at high altitudes together with the central role played by oxygenation for health and disease states during intrauterine and postnatal life continues to provide a unique study environment for advancing our understanding of the mechanisms underlying human adaptive potential and of human evolutionary processes. Ultimately, such studies can also benefit biomedical research with the identification of new therapeutic targets for treating or preventing O₂ related diseases.

Funding: The authors are supported by the National Institutes of Health (R01 HL138181, R01 HD088590, and FIC R21 TW010797) and the University of Colorado Denver Anschutz Medical Campus.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Gomez-Carballa, A.; Pardo-Seco, J.; Brandini, S.; Achilli, A.; Perego, U.A.; Coble, M.D.; Diegoli, T.M.; Alvarez-Iglesias, V.; Martinon-Torres, F.; Olivieri, A.; et al. The peopling of South America and the trans-Andean gene flow of the first settlers. *Genome Res.* **2018**, *28*, 767–779. [[CrossRef](#)] [[PubMed](#)]
- Harris, D.N.; Song, W.; Shetty, A.C.; Levano, K.S.; Caceres, O.; Padilla, C.; Borda, V.; Tarazona, D.; Trujillo, O.; Sanchez, C.; et al. Evolutionary genomic dynamics of Peruvians before, during, and after the Inca Empire. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E6526–E6535. [[CrossRef](#)] [[PubMed](#)]
- Dobzhansky, T. Adaptedness and fitness. In *Population Biology and Evolution*; Lewontin, R., Ed.; Syracuse University Press: Syracuse, NY, USA, 1968; p. 111.
- Frisancho, A.R.; Frisancho, H.G.; Milotich, M.; Brutsaert, T.; Albalak, R.; Spielvogel, H.; Villain, M.; Vargas, E.; Soria, R. Developmental, genetic, and environmental components of aerobic capacity at high altitude. *Am. J. Phys. Anthropol.* **1995**, *96*, 431–442. [[CrossRef](#)]
- Frisancho, A.R. Developmental responses to high altitude hypoxia. *Am. J. Phys. Anthropol.* **1970**, *32*, 401–407. [[CrossRef](#)] [[PubMed](#)]
- Frisancho, A.R. Developmental adaptation: Where we go from here. *Am. J. Hum. Biol.* **2009**, *21*, 694–703. [[CrossRef](#)] [[PubMed](#)]
- Moore, L.G. Human genetic adaptation to high altitude: Current status and future prospects. *Quat. Int.* **2017**, *461*, 4–13. [[CrossRef](#)] [[PubMed](#)]
- Zhuang, J.; Droma, T.; Sun, S.; Janes, C.; McCullough, R.E.; McCullough, R.G.; Cymerman, A.; Huang, S.Y.; Reeves, J.T.; Moore, L.G. Hypoxic ventilatory responsiveness in Tibetan compared with Han residents of 3,658 m. *J. Appl. Physiol.* **1993**, *74*, 303–311. [[CrossRef](#)] [[PubMed](#)]
- Beall, C.M. Two routes to functional adaptation: Tibetan and Andean high-altitude natives. *Proc. Natl. Acad. Sci. USA* **2007**, *104* (Suppl. 1), 8655–8660. [[CrossRef](#)]
- Brutsaert, T.D. Population genetic aspects and phenotypic plasticity of ventilatory responses in high altitude natives. *Respir. Physiol. Neurobiol.* **2007**, *158*, 151–160. [[CrossRef](#)] [[PubMed](#)]
- Brutsaert, T.D.; Parra, E.J.; Shriver, M.D.; Gamboa, A.; Rivera-Ch, M.; Leon-Velarde, F. Ancestry explains the blunted ventilatory response to sustained hypoxia and lower exercise ventilation of Quechua altitude natives. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2005**, *289*, R225–R234. [[CrossRef](#)] [[PubMed](#)]
- Moore, L.G. Measuring high-altitude adaptation. *J. Appl. Physiol.* **2017**, *123*, 1371–1385. [[CrossRef](#)] [[PubMed](#)]
- Schoene, R.B.; Roach, R.C.; Lahiri, S.; Peters, R.M., Jr.; Hackett, P.H.; Santolaya, R. Increased diffusion capacity maintains arterial saturation during exercise in the Quechua Indians of Chilean Altiplano. *Am. J. Hum. Biol. Off. J. Hum. Biol. Counc.* **1990**, *2*, 663–668. [[CrossRef](#)] [[PubMed](#)]
- Zhuang, J.; Droma, T.; Sutton, J.R.; Groves, B.M.; McCullough, R.E.; McCullough, R.G.; Sun, S.; Moore, L.G. Smaller alveolar-arterial O₂ gradients in Tibetan than Han residents of Lhasa (3658 m). *Respir. Physiol.* **1996**, *103*, 75–82. [[CrossRef](#)]
- Wagner, P.D.; Araoz, M.; Boushel, R.; Calbet, J.A.; Jessen, B.; Radegran, G.; Spielvogel, H.; Sondegaard, H.; Wagner, H.; Saltin, B. Pulmonary gas exchange and acid-base state at 5,260 m in high-altitude Bolivians and acclimatized lowlanders. *J. Appl. Physiol.* **2002**, *92*, 1393–1400. [[CrossRef](#)] [[PubMed](#)]
- Brutsaert, T.D.; Araoz, M.; Soria, R.; Spielvogel, H.; Haas, J.D. Higher arterial oxygen saturation during submaximal exercise in Bolivian Aymara compared to European sojourners and Europeans born and raised at high altitude. *Am. J. Phys. Anthr.* **2000**, *113*, 169–181. [[CrossRef](#)]

17. DeGraff, A.C., Jr.; Grover, R.F.; Johnson, R.L., Jr.; Hammond, J.W., Jr.; Miller, J.M. Diffusing capacity of the lung in Caucasians native to 3,100 m. *J. Appl. Physiol.* **1970**, *29*, 71–76. [[CrossRef](#)] [[PubMed](#)]
18. Johnson, R.L., Jr.; Cassidy, S.S.; Grover, R.F.; Schutte, J.E.; Epstein, R.H. Functional capacities of lungs and thorax in beagles after prolonged residence at 3,100 m. *J. Appl. Physiol.* **1985**, *59*, 1773–1782. [[CrossRef](#)]
19. Brutsaert, T.D.; Soria, R.; Caceres, E.; Spielvogel, H.; Haas, J.D. Effect of developmental and ancestral high altitude exposure on chest morphology and pulmonary function in Andean and European/North American natives. *Am. J. Hum. Biol. Off. J. Hum. Biol. Council.* **1999**, *11*, 383–395. [[CrossRef](#)]
20. Kiyamu, M.; Bigham, A.; Parra, E.; Leon-Velarde, F.; Rivera-Chira, M.; Brutsaert, T.D. Developmental and genetic components explain enhanced pulmonary volumes of female Peruvian Quechua. *Am. J. Phys. Anthr.* **2012**, *148*, 534–542. [[CrossRef](#)]
21. Kiyamu, M.; León-Velarde, F.; Rivera-Chira, M.; Brutaser, T.D. Developmental effects determine submaximal arterial oxygen saturation in Peruvian Quechua. *High Alt. Med. Biol.* **2015**, *16*, 138–146. [[CrossRef](#)]
22. Pomeroy, E.; Wells, J.C.; Stanojevic, S.; Miranda, J.J.; Moore, L.G.; Cole, T.J.; Stock, J.T. Surname-inferred Andean ancestry is associated with child stature and limb lengths at high altitude in Peru, but not at sea level. *Am. J. Hum. Biol.* **2015**, *27*, 798–806. [[CrossRef](#)] [[PubMed](#)]
23. Moore, L.G. Comparative human ventilatory adaptation to high altitude. *Respir. Physiol.* **2000**, *121*, 257–276. [[CrossRef](#)]
24. Storz, J.F. Hemoglobin-oxygen affinity in high-altitude vertebrates: Is there evidence for an adaptive trend? *J. Exp. Biol.* **2016**, *219*, 3190–3203. [[CrossRef](#)] [[PubMed](#)]
25. Winslow, R.M.; Chapman, K.W.; Gibson, C.C.; Samaja, M.; Monge, C.C.; Goldwasser, E.; Sherpa, M.; Blume, F.D.; Santolaya, R. Different hematologic responses to hypoxia in Sherpas and Quechua Indians. *J. Appl. Physiol.* **1989**, *66*, 1561–1569. [[CrossRef](#)] [[PubMed](#)]
26. Bigham, A.W.; Kiyamu, M.; Leon-Velarde, F.; Parra, E.J.; Rivera-Ch, M.; Shriver, M.D.; Brutsaert, T.D. Angiotensin-converting enzyme genotype and arterial oxygen saturation at high altitude in Peruvian Quechua. *High Alt. Med. Biol.* **2008**, *9*, 167–178. [[CrossRef](#)] [[PubMed](#)]
27. Monge, C.; Bonavia, D.; León-Velarde, F.; Arregui, A. High altitude populations in Nepal and the Andes. In *Hypoxia: The Adaptations*; Sutton, J., Coates, G., Remmers, J., Eds.; B.C. Decker: Toronto, ON, Canada, 1990; pp. 53–58.
28. Leon-Velarde, F.; Ramos, M.A.; Hernandez, J.A.; De Idiaquez, D.; Munoz, L.S.; Gaffo, A.; Cordova, S.; Durand, D.; Monge, C. The role of menopause in the development of chronic mountain sickness. *Am. J. Physiol.* **1997**, *272*, R90–R94. [[CrossRef](#)] [[PubMed](#)]
29. Ballew, C.; Garruto, R.M.; Haas, J. High altitude hematology: Paradigm or enigma. In *Human Population Biology*; Little, M.A., Haas, J.D., Eds.; Oxford University Press: Oxford, UK, 1989; pp. 239–262.
30. Rupert, J.L.; Hochachka, P.W. The evidence for hereditary factors contributing to high altitude adaptation in Andean natives: A review. *High Alt. Med. Biol.* **2001**, *2*, 235–256. [[CrossRef](#)]
31. Vogel, J.A.; Hartley, L.H.; Cruz, J.C. Cardiac output during exercise in altitude natives at sea level and high altitude. *J. Appl. Physiol.* **1974**, *36*, 173–176. [[CrossRef](#)]
32. Hochachka, P.W.; Clark, C.M.; Holden, J.E.; Stanley, C.; Ugurbil, K.; Menon, R.S. 31P magnetic resonance spectroscopy of the Sherpa heart: A phosphocreatine/adenosine triphosphate signature of metabolic defense against hypobaric hypoxia. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 1215–1220. [[CrossRef](#)]
33. Sawka, M.N.; Young, A.J.; Rock, P.B.; Lyons, T.P.; Boushel, R.; Freund, B.J.; Muza, S.R.; Cymerman, A.; Dennis, R.C.; Pandolf, K.B.; et al. Altitude acclimatization and blood volume: Effects of exogenous erythrocyte volume expansion. *J. Appl. Physiol.* **1996**, *81*, 636–642. [[CrossRef](#)]
34. Banchemo, N.; Sime, F.; Peñaloza, D.; Cruz, J.; Gamboa, R.; Marticorena, E. Pulmonary pressure, cardiac output, and arterial oxygen saturation during exercise at high altitude and at sea level. *Circulation* **1966**, *33*, 249–262. [[CrossRef](#)] [[PubMed](#)]
35. Cruz-Jibaja, J.; Banchemo, N.; Sime, F.; Peñaloza, D.; Gamboa, R.; Marticorena, E. Correlation between pulmonary artery pressure and level of altitude. *Dis. Chest* **1964**, *46*, 446–451. [[CrossRef](#)]
36. Brutsaert, T.D. Do high-altitude natives have enhanced exercise performance at altitude? *Appl. Physiol. Nutr. Metab.* **2008**, *33*, 582–592. [[CrossRef](#)] [[PubMed](#)]
37. Soria, R.; Egger, M.; Scherrer, U.; Bender, N.; Rimoldi, S.F. Pulmonary artery pressure and arterial oxygen saturation in people living at high or low altitude: Systematic review and meta-analysis. *J. Appl. Physiol.* **2016**, *121*, 1151–1159. [[CrossRef](#)]

38. Beall, C.M.; Decker, M.J.; Brittenham, G.M.; Kushner, I.; Gebremedhin, A.; Strohl, K.P. An Ethiopian pattern of human adaptation to high-altitude hypoxia. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 17215–17218. [[CrossRef](#)]
39. Jansen, G.F.; Basnyat, B. Brain blood flow in Andean and Himalayan high-altitude populations: Evidence of different traits for the same environmental constraint. *J. Cereb. Blood Flow Metab.* **2011**, *31*, 706–714. [[CrossRef](#)] [[PubMed](#)]
40. Jackson, M.R.; Mayhew, T.M.; Haas, J.D. The volumetric composition of human term placentae: Altitudinal, ethnic and sex differences in Bolivia. *J. Anat.* **1987**, *152*, 173–187. [[PubMed](#)]
41. Groves, B.M.; Droma, T.; Sutton, J.R.; McCullough, R.G.; McCullough, R.E.; Zhuang, J.; Rapmund, G.; Sun, S.; Janes, C.; Moore, L.G. Minimal hypoxic pulmonary hypertension in normal Tibetans at 3,658 m. *J. Appl. Physiol.* **1993**, *74*, 312–318. [[CrossRef](#)] [[PubMed](#)]
42. Huang, S.Y.; Sun, S.; Droma, T.; Zhuang, J.; Tao, J.X.; McCullough, R.G.; McCullough, R.E.; Micco, A.J.; Reeves, J.T.; Moore, L.G. Internal carotid arterial flow velocity during exercise in Tibetan and Han residents of Lhasa (3,658 m). *J. Appl. Physiol.* **1992**, *73*, 2638–2642. [[CrossRef](#)]
43. Moore, L.G. Uterine blood flow as a determinant of feto-placental development. In *The Placenta and Human Developmental Programming*; Burton, G.J., Barker, D.J.P., Moffett, A., Thornburg, K.L., Eds.; Cambridge University Press: Cambridge, UK, 2011; pp. 126–146.
44. Tymko, M.M.; Tremblay, J.C.; Hansen, A.B.; Howe, C.A.; Willie, C.K.; Stembridge, M.; Green, D.J.; Hoiland, R.L.; Subedi, P.; Anholm, J.D.; et al. The effect of alpha1-adrenergic blockade on post-exercise brachial artery flow-mediated dilatation at sea level and high altitude. *J. Physiol.* **2017**, *595*, 1671–1686. [[CrossRef](#)]
45. Dhar, P.; Sharma, V.K.; Hota, K.B.; Das, S.K.; Hota, S.K.; Srivastava, R.B.; Singh, S.B. Autonomic cardiovascular responses in acclimatized lowlanders on prolonged stay at high altitude: A longitudinal follow up study. *PLoS ONE* **2014**, *9*, e84274. [[CrossRef](#)] [[PubMed](#)]
46. Lundby, C.; Sander, M.; van Hall, G.; Saltin, B.; Calbet, J.A. Maximal exercise and muscle oxygen extraction in acclimatizing lowlanders and high altitude natives. *J. Physiol.* **2006**, *573*, 535–547. [[CrossRef](#)] [[PubMed](#)]
47. Hoiland, R.L.; Bain, A.R.; Rieger, M.G.; Bailey, D.M.; Ainslie, P.N. Hypoxemia, oxygen content, and the regulation of cerebral blood flow. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2016**, *310*, R398–R413. [[CrossRef](#)] [[PubMed](#)]
48. Norcliffe, L.J.; Rivera-Ch, M.; Claydon, V.E.; Moore, J.P.; Leon-Velarde, F.; Appenzeller, O.; Hainsworth, R. Cerebrovascular responses to hypoxia and hypocapnia in high-altitude dwellers. *J. Physiol.* **2005**, *566*, 287–294. [[CrossRef](#)] [[PubMed](#)]
49. Moore, L.G.; Zamudio, S.; Zhuang, J.; Sun, S.; Droma, T. Oxygen transport in Tibetan women during pregnancy at 3,658 m. *Am. J. Phys. Anthr.* **2001**, *114*, 42–53. [[CrossRef](#)]
50. Wilson, M.J.; Lopez, M.; Vargas, M.; Julian, C.; Tellez, W.; Rodriguez, A.; Bigham, A.; Armaza, J.F.; Niermeyer, S.; Shriver, M.; et al. Greater uterine artery blood flow during pregnancy in multigenerational (Andean) than shorter-term (European) high-altitude residents. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2007**, *293*, R1313–R1324. [[CrossRef](#)] [[PubMed](#)]
51. Julian, C.G.; Wilson, M.J.; Lopez, M.; Yamashiro, H.; Tellez, W.; Rodriguez, A.; Bigham, A.W.; Shriver, M.D.; Rodriguez, C.; Vargas, E.; et al. Augmented uterine artery blood flow and oxygen delivery protect Andeans from altitude-associated reductions in fetal growth. *Am. J. Physiol.* **2009**, *296*, R1564–R1575. [[CrossRef](#)]
52. Julian, C.G.; Vargas, E.; Browne, V.A.; Wilson, M.J.; Bigham, A.W.; Rodriguez, C.; McCord, J.M.; Moore, L.G. Potential role for elevated maternal enzymatic antioxidant status in Andean protection against altitude-associated SGA. *J. Matern. Fetal Neonatal Med.* **2012**, *25*, 1233–1240. [[CrossRef](#)]
53. Davila, R.D.; Julian, C.G.; Wilson, M.J.; Browne, V.A.; Rodriguez, C.; Bigham, A.W.; Shriver, M.D.; Vargas, E.; Moore, L.G. Do anti-angiogenic or angiogenic factors contribute to the protection of birth weight at high altitude afforded by Andean ancestry? *Reprod. Sci.* **2010**, *17*, 861–870. [[CrossRef](#)]
54. Julian, C.G.; Hageman, J.L.; Wilson, M.J.; Vargas, E.; Moore, L.G. Lowland origin women raised at high altitude are not protected against lower uteroplacental O₂ delivery during pregnancy or reduced birth weight. *Am. J. Hum. Biol.* **2011**, *23*, 509–516. [[CrossRef](#)]
55. Charles, S.M.; Julian, C.G.; Vargas, E.; Moore, L.G. Higher estrogen levels during pregnancy in Andean than European residents of high altitude suggest differences in aromatase activity. *J. Clin. Endocrinol. Metab.* **2014**, *99*, 2908–2916. [[CrossRef](#)] [[PubMed](#)]

56. Tissot van Patot, M.C.; Bendrick-Peart, J.; Beckey, V.E.; Serkova, N.; Zwerdinger, L. Greater vascularity, lowered HIF-1/DNA binding, and elevated GSH as markers of adaptation to in vivo chronic hypoxia. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2004**, *287*, L525–L532. [[CrossRef](#)] [[PubMed](#)]
57. Gassmann, N.N.; van Elteren, H.A.; Goos, T.G.; Morales, C.R.; Rivera-Ch, M.; Martin, D.S.; Cabala Peralta, P.; Passano Del Carpio, A.; Aranibar Machaca, S.; Huicho, L.; et al. Pregnancy at high altitude in the Andes leads to increased total vessel density in healthy newborns. *J. Appl. Physiol.* **2016**, *121*, 709–715. [[CrossRef](#)] [[PubMed](#)]
58. Holden, J.E.; Stone, C.K.; Clark, C.M.; Brown, W.D.; Nickles, R.J.; Stanley, C.; Hochachka, P.W. Enhanced cardiac metabolism of plasma glucose in high-altitude natives: Adaptation against chronic hypoxia. *J. Appl. Physiol.* **1995**, *79*, 222–228. [[CrossRef](#)] [[PubMed](#)]
59. Brooks, G.A.; Butterfield, G.E.; Wolfe, R.R.; Groves, B.M.; Mazzeo, R.S.; Sutton, J.R.; Wolfel, E.E.; Reeves, J.T. Increased dependence on blood glucose after acclimatization to 4,300 m. *J. Appl. Physiol.* **1991**, *70*, 919–927. [[CrossRef](#)]
60. Braun, B.; Mawson, J.T.; Muza, S.R.; Dominick, S.B.; Brooks, G.A.; Horning, M.A.; Rock, P.B.; Moore, L.G.; Mazzeo, R.S.; Ezeji-Okoye, S.C.; et al. Women at altitude: Carbohydrate utilization during exercise at 4,300 m. *J Appl Physiol* **2000**, *88*, 246–256. [[CrossRef](#)]
61. Woolcott, O.O.; Ader, M.; Bergman, R.N. Glucose homeostasis during short-term and prolonged exposure to high altitudes. *Endocr. Rev.* **2015**, *36*, 149–173. [[CrossRef](#)]
62. Horscroft, J.A.; Kotwica, A.O.; Laner, V.; West, J.A.; Hennis, P.J.; Levett, D.Z.H.; Howard, D.J.; Fernandez, B.O.; Burgess, S.L.; Ament, S.; et al. Metabolic basis to Sherpa altitude adaptation. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 6382–6387. [[CrossRef](#)]
63. Krampfl, E.; Kametas, N.A.; Nowotny, P.; Roden, M.; Nicolaidis, K.H. Glucose metabolism in pregnancy at high altitude. *Diabetes Care* **2001**, *24*, 817–822. [[CrossRef](#)]
64. Zamudio, S.; Torricos, T.; Fik, E.; Oyala, M.; Echalar, L.; Pullockaran, J.; Tutino, E.; Martin, B.; Belliappa, S.; Balanza, E.; et al. Hypoglycemia and the origin of hypoxia-induced reduction in human fetal growth. *PLoS ONE* **2010**, *5*, e8551. [[CrossRef](#)]
65. Semenza, G.L. Oxygen sensing, homeostasis, and disease. *N. Engl. J. Med.* **2011**, *365*, 537–547. [[CrossRef](#)] [[PubMed](#)]
66. Wang, G.L.; Jiang, B.H.; Rue, E.A.; Semenza, G.L. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O₂ tension. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 5510–5514. [[CrossRef](#)] [[PubMed](#)]
67. Bruick, R.K.; McKnight, S.L. A conserved family of prolyl-4-hydroxylases that modify HIF. *Science* **2001**, *294*, 1337–1340. [[CrossRef](#)] [[PubMed](#)]
68. Ivan, M.; Kondo, K.; Yang, H.; Kim, W.; Valiando, J.; Ohh, M.; Salic, A.; Asara, J.M.; Lane, W.S.; Kaelin, W.G., Jr. HIF α targeted for VHL-mediated destruction by proline hydroxylation: Implications for O₂ sensing. *Science* **2001**, *292*, 464–468. [[CrossRef](#)] [[PubMed](#)]
69. Jaakkola, P.; Mole, D.R.; Tian, Y.M.; Wilson, M.I.; Gielbert, J.; Gaskell, S.J.; von Kriegsheim, A.; Hebestreit, H.F.; Mukherji, M.; Schofield, C.J.; et al. Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* **2001**, *292*, 468–472. [[CrossRef](#)] [[PubMed](#)]
70. Majmundar, A.J.; Wong, W.J.; Simon, M.C. Hypoxia-inducible factors and the response to hypoxic stress. *Mol. Cell* **2010**, *40*, 294–309. [[CrossRef](#)]
71. Bigham, A.W.; Mao, X.; Mei, R.; Brutsaert, T.; Wilson, M.J.; Julian, C.G.; Parra, E.J.; Akey, J.M.; Moore, L.G.; Shriver, M.D. Identifying positive selection candidate loci for high-altitude adaptation in Andean populations. *Hum. Genom.* **2009**, *4*, 79–90.
72. Jacovas, V.C.; Couto-Silva, C.M.; Nunes, K.; Lemes, R.B.; de Oliveira, M.Z.; Salzano, F.M.; Bortolini, M.C.; Hunemeier, T. Selection scan reveals three new loci related to high altitude adaptation in Native Andeans. *Sci. Rep.* **2018**, *8*, 12733. [[CrossRef](#)]
73. Crawford, J.E.; Amaru, R.; Song, J.; Julian, C.G.; Racimo, F.; Cheng, J.Y.; Guo, X.; Yao, J.; Ambale-Venkatesh, B.; Lima, J.A.; et al. Natural selection on genes related to cardiovascular health in high-altitude adapted Andeans. *Am. J. Hum. Genet.* **2017**, *101*, 752–767. [[CrossRef](#)] [[PubMed](#)]
74. Bigham, A.; Bauchet, M.; Pinto, D.; Mao, X.; Akey, J.M.; Mei, R.; Scherer, S.W.; Julian, C.G.; Wilson, M.J.; Lopez Herraes, D.; et al. Identifying signatures of natural selection in Tibetan and Andean populations using dense genome scan data. *PLoS Genet.* **2010**, *6*, e1001116. [[CrossRef](#)]

75. Eichstaedt, C.A.; Antao, T.; Pagani, L.; Cardona, A.; Kivisild, T.; Mormina, M. The Andean adaptive toolkit to counteract high altitude maladaptation: Genome-wide and phenotypic analysis of the Collas. *PLoS ONE* **2014**, *9*, e93314. [[CrossRef](#)] [[PubMed](#)]
76. Moore, L.G.; Shriver, M.; Bemis, L.; Hickler, B.; Wilson, M.; Brutsaert, T.; Parra, E.; Vargas, E. Maternal adaptation to high-altitude pregnancy: An experiment of nature. *Placenta* **2004**, *25*, S60–S71. [[CrossRef](#)]
77. Valverde, G.; Zhou, H.; Lippold, S.; de Filippo, C.; Tang, K.; Lopez Herraiez, D.; Li, J.; Stoneking, M. A novel candidate region for genetic adaptation to high altitude in Andean populations. *PLoS ONE* **2015**, *10*, e0125444. [[CrossRef](#)] [[PubMed](#)]
78. Jacovas, V.C.; Rovaris, D.L.; Perez, O.; de Azevedo, S.; Macedo, G.S.; Sandoval, J.R.; Salazar-Granara, A.; Villena, M.; Dugoujon, J.M.; Bisso-Machado, R.; et al. Genetic variations in the TP53 pathway in Native Americans strongly suggest adaptation to the high altitudes of the Andes. *PLoS ONE* **2015**, *10*, e0137823. [[CrossRef](#)] [[PubMed](#)]
79. Rademaker, K.; Hodgins, G.; Moore, K.; Zarrillo, S.; Miller, C.; Bromley, G.R.; Leach, P.; Reid, D.A.; Alvarez, W.Y.; Sandweiss, D.H. Paleoindian settlement of the high-altitude Peruvian Andes. *Science* **2014**, *346*, 466–469. [[CrossRef](#)] [[PubMed](#)]
80. Moore, L.G. Human genetic adaptation to high altitude. *High Alt. Med. Biol.* **2001**, *2*, 257–279. [[CrossRef](#)] [[PubMed](#)]
81. Niermeyer, S.; Andrade, M.M.; Vargas, E.; Moore, L.G. Neonatal oxygenation, pulmonary hypertension, and evolutionary adaptation to high altitude (2013 Grover Conference series). *Pulm Circ.* **2015**, *5*, 48–62. [[CrossRef](#)]
82. Gonzales, G.F. Peruvian contributions to the study on human reproduction at high altitude: From the chronicles of the Spanish conquest to the present. *Respir. Physiol. Neurobiol.* **2007**, *158*, 172–179. [[CrossRef](#)]
83. Julian, C.G.; Vargas, E.; Armaza, J.F.; Wilson, M.J.; Niermeyer, S.; Moore, L.G. High-altitude ancestry protects against hypoxia-associated reductions in fetal growth. *Arch. Dis. Child Fetal Neonatal Ed.* **2007**, *92*, F372–F377. [[CrossRef](#)]
84. Bennett, A.; Sain, S.R.; Vargas, E.; Moore, L.G. Evidence that parent-of-origin affects birth-weight reductions at high altitude. *Am. J. Hum. Biol.* **2008**, *20*, 592–597. [[CrossRef](#)]
85. Soria, R.; Julian, C.; Vargas, E.; Moore, L.; Giussani, D. Graduated effects of high-altitude hypoxia and highland ancestry on birth size. *Pediatr. Res.* **2013**, *74*, 633–638. [[CrossRef](#)] [[PubMed](#)]
86. Moore, L.G.; Young, D.; McCullough, R.E.; Droma, T.; Zamudio, S. Tibetan protection from intrauterine growth restriction (IUGR) and reproductive loss at high altitude. *Am. J. Hum. Biol.* **2001**, *13*, 635–644. [[CrossRef](#)] [[PubMed](#)]
87. Zamudio, S.; Postigo, L.; Illsley, N.P.; Rodriguez, C.; Heredia, G.; Brimacombe, M.; Echalar, L.; Torricos, T.; Tellez, W.; Maldonado, I.; et al. Maternal oxygen delivery is not related to altitude- and ancestry-associated differences in human fetal growth. *J. Physiol.* **2007**, *582*, 883–895. [[CrossRef](#)] [[PubMed](#)]
88. Zamudio, S.; Palmer, S.K.; Droma, T.; Stamm, E.; Coffin, C.; Moore, L.G. Effect of altitude on uterine artery blood flow during normal pregnancy. *J. Appl. Physiol.* **1995**, *79*, 7–14. [[CrossRef](#)] [[PubMed](#)]
89. Julian, C.G.; Galan, H.L.; Wilson, M.J.; Desilva, W.; Cioffi-Ragan, D.; Schwartz, J.; Moore, L.G. Lower uterine artery blood flow and higher endothelin relative to nitric oxide metabolite levels are associated with reductions in birth weight at high altitude. *Am. J. Physiol.* **2008**, *295*, R906–R915. [[CrossRef](#)] [[PubMed](#)]
90. Browne, V.A.; Toledo-Jaldin, L.; Davila, R.D.; Lopez, L.P.; Yamashiro, H.; Cioffi-Ragan, D.; Julian, C.G.; Wilson, M.J.; Bigham, A.W.; Shriver, M.D.; et al. High-end arteriolar resistance limits uterine artery blood flow and restricts fetal growth in preeclampsia and gestational hypertension at high altitude. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2011**, *300*, R1221–R1229. [[CrossRef](#)] [[PubMed](#)]
91. Mayhew, T.M.; Ohadike, C.; Baker, P.N.; Crocker, I.P.; Mitchell, C.; Ong, S.S. Stereological investigation of placental morphology in pregnancies complicated by pre-eclampsia with and without intrauterine growth restriction. *Placenta* **2003**, *24*, 219–226. [[CrossRef](#)]
92. Bellamy, L.; Casas, J.P.; Hingorani, A.D.; Williams, D.J. Pre-eclampsia and risk of cardiovascular disease and cancer in later life: Systematic review and meta-analysis. *BMJ* **2007**, *335*, 974. [[CrossRef](#)]
93. Irgens, H.U.; Reisaeter, L.; Irgens, L.M.; Lie, R.T. Long term mortality of mothers and fathers after pre-eclampsia: Population based cohort study. *BMJ* **2001**, *323*, 1213–1217. [[CrossRef](#)]

94. Lykke, J.A.; Langhoff-Roos, J.; Sibai, B.M.; Funai, E.F.; Triche, E.W.; Paidas, M.J. Hypertensive pregnancy disorders and subsequent cardiovascular morbidity and type 2 diabetes mellitus in the mother. *Hypertension* **2009**, *53*, 944–951. [[CrossRef](#)]
95. Roberts, J.M.; Hubel, C.A. Pregnancy: A screening test for later life cardiovascular disease. *Womens Health Issues* **2010**, *20*, 304–307. [[CrossRef](#)] [[PubMed](#)]
96. Smith, G.C.; Pell, J.P.; Walsh, D. Pregnancy complications and maternal risk of ischaemic heart disease: A retrospective cohort study of 129,290 births. *Lancet* **2001**, *357*, 2002–2006. [[CrossRef](#)]
97. Wikstrom, A.K.; Haglund, B.; Olovsson, M.; Lindeberg, S.N. The risk of maternal ischaemic heart disease after gestational hypertensive disease. *BJOG Int. J. Obstet. Gynaecol.* **2005**, *112*, 1486–1491. [[CrossRef](#)]
98. Palmer, S.K.; Moore, L.G.; Young, D.; Cregger, B.; Berman, J.C.; Zamudio, S. Altered blood pressure course during normal pregnancy and increased preeclampsia at high altitude (3100 meters) in Colorado. *Am. J. Obs. Gynecol.* **1999**, *180*, 1161–1168. [[CrossRef](#)]
99. Keyes, L.E.; Armaza, J.F.; Niermeyer, S.; Vargas, E.; Young, D.A.; Moore, L.G. Intrauterine growth restriction, preeclampsia, and intrauterine mortality at high altitude in Bolivia. *Pediatr. Res.* **2003**, *54*, 20–25. [[CrossRef](#)] [[PubMed](#)]
100. Miller, S.; Tudor, C.; Thorsten, V.R.; Craig, S.; Le, P.; Wright, L.L.; Varner, M.W. Maternal and neonatal outcomes of hospital vaginal deliveries in Tibet. *Int. J. Gynaecol. Obstet.* **2007**, *98*, 217–221. [[CrossRef](#)]
101. Chaiworapongsa, T.; Chaemsaihong, P.; Korzeniewski, S.J.; Yeo, L.; Romero, R. Pre-eclampsia part 2: Prediction, prevention and management. *Nat. Rev. Nephrol.* **2014**, *10*, 531–540. [[CrossRef](#)] [[PubMed](#)]
102. Bigham, A.W.; Julian, C.G.; Wilson, M.J.; Vargas, E.; Browne, V.A.; Shriver, M.D.; Moore, L.G. Maternal *PRKAA1* and *EDNRA* genotypes are associated with birth weight, and *PRKAA1* with uterine artery diameter and metabolic homeostasis at high altitude. *Physiol. Genom.* **2014**, *46*, 687–697. [[CrossRef](#)]
103. Roos, S.; Powell, T.L.; Jansson, T. Placental mTOR links maternal nutrient availability to fetal growth. *Biochem. Soc. Trans.* **2009**, *37*, 295–298. [[CrossRef](#)]
104. Yung, H.W.; Calabrese, S.; Hynx, D.; Hemmings, B.A.; Cetin, I.; Charnock-Jones, D.S.; Burton, G.J. Evidence of placental translation inhibition and endoplasmic reticulum stress in the etiology of human intrauterine growth restriction. *Am. J. Pathol.* **2008**, *173*, 451–462. [[CrossRef](#)]
105. Yung, H.W.; Cox, M.; Tissot van Patot, M.; Burton, G.J. Evidence of endoplasmic reticulum stress and protein synthesis inhibition in the placenta of non-native women at high altitude. *FASEB J.* **2012**, *26*, 1970–1981. [[CrossRef](#)]
106. Hill, C.M.; Baya, A.; Gavlak, J.; Carroll, A.; Heathcote, K.; Dimitriou, D.; L'Esperance, V.; Webster, R.; Holloway, J.; Virues-Ortega, J.; et al. Adaptation to life in the High Andes: nocturnal oxyhemoglobin saturation in early development. *Sleep* **2016**, *39*, 1001–1008. [[CrossRef](#)] [[PubMed](#)]
107. Monge, C.; Arregui, A.; Leon-Velarde, F. Pathophysiology and epidemiology of chronic mountain sickness. *Int. J. Sports Med.* **1992**, *13*, S79–S81. [[CrossRef](#)] [[PubMed](#)]
108. Julian, C.G.; Vargas, E.; Gonzales, M.; Davila, R.D.; Ladenburger, A.; Reardon, L.; Schoo, C.; Powers, R.W.; Lee-Chiong, T.; Moore, L.G. Sleep-disordered breathing and oxidative stress in preclinical chronic mountain sickness (excessive erythrocytosis). *Respir. Physiol. Neurobiol.* **2013**, *186*, 188–196. [[CrossRef](#)] [[PubMed](#)]
109. Julian, C.G.; Gonzales, M.; Rodriguez, A.; Bellido, D.; Salmon, C.S.; Ladenburger, A.; Reardon, L.; Vargas, E.; Moore, L.G. Perinatal hypoxia increases susceptibility to high-altitude polycythemia and attendant pulmonary vascular dysfunction. *Am. J. Physiol. Heart Circ. Physiol.* **2015**, *309*, H565–H573. [[CrossRef](#)] [[PubMed](#)]
110. Wu, T.Y.; Li, W.; Li, Y.; Ge, R.L.; Cheng, Q.; Wang, S.; Zhao, G.; Wei, L.; Jin, Y.; Don, G. Epidemiology of chronic mountain sickness: Ten years' study in Qinghai-Tibet. In *Progress in Mountain Medicine and High Altitude Physiology*; Ohno, H., Kobayashi, T., Masuyama, S., Nakashima, M., Eds.; Press Committee of the 3rd World Congress on Mountain Medicine and High Altitude Physiology: Matsumoto, Japan, 1998; pp. 120–125.
111. Claydon, V.E.; Gulli, G.; Slessarev, M.; Appenzeller, O.; Zenebe, G.; Gebremedhin, A.; Hainsworth, R. Cerebrovascular responses to hypoxia and hypocapnia in Ethiopian high altitude dwellers. *Stroke* **2008**, *39*, 336–342. [[CrossRef](#)] [[PubMed](#)]
112. Moore, L.G.; Niermeyer, S.; Vargas, E. Does chronic mountain sickness (CMS) have perinatal origins? *Respir. Physiol. Neurobiol.* **2007**, *158*, 180–189. [[CrossRef](#)]

113. Bailey, D.M.; Rimoldi, S.F.; Rexhaj, E.; Pratali, L.; Salinas Salmon, C.; Villena, M.; McEneny, J.; Young, I.S.; Nicod, P.; Allemann, Y.; et al. Oxidative-nitrosative stress and systemic vascular function in highlanders with and without exaggerated hypoxemia. *Chest* **2013**, *143*, 444–451. [[CrossRef](#)] [[PubMed](#)]
114. Kryger, M.; Glas, R.; Jackson, D.; McCullough, R.E.; Scoggin, C.; Grover, R.F.; Weil, J.V. Impaired oxygenation during sleep in excessive polycythemia of high altitude: Improvement with respiratory stimulation. *Sleep* **1978**, *1*, 3–17. [[CrossRef](#)]
115. Rexhaj, E.; Rimoldi, S.F.; Pratali, L.; Brenner, R.; Andries, D.; Soria, R.; Salinas, C.; Villena, M.; Romero, C.; Allemann, Y.; et al. Sleep-disordered breathing and vascular function in patients with chronic mountain sickness and healthy high-altitude dwellers. *Chest* **2016**, *149*, 991–998. [[CrossRef](#)]
116. Reeves, J.T.; Weil, J.V. Chronic mountain sickness. A view from the crow's nest. *Adv. Exp. Med. Biol.* **2001**, *502*, 419–437. [[PubMed](#)]
117. Meadows, G.E.; O'Driscoll, D.M.; Simonds, A.K.; Morrell, M.J.; Corfield, D.R. Cerebral blood flow response to isocapnic hypoxia during slow-wave sleep and wakefulness. *J. Appl. Physiol.* **2004**, *97*, 1343–1348. [[CrossRef](#)] [[PubMed](#)]
118. Rimoldi, S.F.; Rexhaj, E.; Pratali, L.; Bailey, D.M.; Hutter, D.; Fajta, F.; Salmon, C.S.; Villena, M.; Nicod, P.; Allemann, Y.; et al. Systemic vascular dysfunction in patients with chronic mountain sickness. *Chest* **2012**, *141*, 139–146. [[CrossRef](#)] [[PubMed](#)]
119. Sartori, C.; Allemann, Y.; Trueb, L.; Delabays, A.; Nicod, P.; Scherrer, U. Augmented vasoreactivity in adult life associated with perinatal vascular insult. *Lancet* **1999**, *353*, 2205–2207. [[CrossRef](#)]
120. Bigham, A.W.; Wilson, M.J.; Julian, C.G.; Kiyamu, M.; Vargas, E.; Leon-Velarde, F.; Rivera-Chira, M.; Rodriguez, C.; Browne, V.A.; Parra, E.; et al. Andean and Tibetan patterns of adaptation to high altitude. *Am. J. Hum. Biol.* **2013**, *25*, 190–197. [[CrossRef](#)] [[PubMed](#)]
121. Zhou, D.; Udpa, N.; Ronen, R.; Stobdan, T.; Liang, J.; Appenzeller, O.; Zhao, H.W.; Yin, Y.; Du, Y.; Guo, L.; et al. Whole-genome sequencing uncovers the genetic basis of chronic mountain sickness in Andean highlanders. *Am. J. Hum. Genet.* **2013**, *93*, 452–462. [[CrossRef](#)] [[PubMed](#)]
122. Cole, A.M.; Petousi, N.; Cavalleri, G.L.; Robbins, P.A. Genetic variation in *SEN1* and *ANP32D* as predictors of chronic mountain sickness. *High Alt. Med. Biol.* **2014**, *15*, 497–499. [[CrossRef](#)] [[PubMed](#)]
123. Villafuerte, F.C. New genetic and physiological factors for excessive erythrocytosis and chronic mountain sickness. *J. Appl. Physiol.* **2015**, *119*, 1481–1486. [[CrossRef](#)] [[PubMed](#)]
124. Giuliani, C.; Bacalini, M.G.; Sazzini, M.; Pirazzini, C.; Franceschi, C.; Garagnani, P.; Luiselli, D. The epigenetic side of human adaptation: Hypotheses, evidences and theories. *Ann. Hum. Biol.* **2015**, *42*, 1–9. [[CrossRef](#)] [[PubMed](#)]
125. Jones, P.A.; Baylin, S.B. The fundamental role of epigenetic events in cancer. *Nat. Rev. Genet.* **2002**, *3*, 415–428. [[CrossRef](#)]
126. Robertson, K.D. DNA methylation, methyltransferases, and cancer. *Oncogene* **2001**, *20*, 3139–3155. [[CrossRef](#)] [[PubMed](#)]
127. Illingworth, R.S.; Gruenewald-Schneider, U.; Webb, S.; Kerr, A.R.; James, K.D.; Turner, D.J.; Smith, C.; Harrison, D.J.; Andrews, R.; Bird, A.P. Orphan CpG islands identify numerous conserved promoters in the mammalian genome. *PLoS Genet* **2010**, *6*, e1001134. [[CrossRef](#)] [[PubMed](#)]
128. Saxonov, S.; Berg, P.; Brutlag, D.L. A genome-wide analysis of CpG dinucleotides in the human genome distinguishes two distinct classes of promoters. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 1412–1417. [[CrossRef](#)] [[PubMed](#)]
129. Feinberg, A.P. Phenotypic plasticity and the epigenetics of human disease. *Nature* **2007**, *447*, 433–440. [[CrossRef](#)] [[PubMed](#)]
130. Jones, P.A. Functions of DNA methylation: Islands, start sites, gene bodies and beyond. *Nat. Rev. Genet.* **2012**, *13*, 484–492. [[CrossRef](#)] [[PubMed](#)]
131. Kulis, M.; Heath, S.; Bibikova, M.; Queiros, A.C.; Navarro, A.; Clot, G.; Martinez-Trillos, A.; Castellano, G.; Brun-Heath, I.; Pinyol, M.; et al. Epigenomic analysis detects widespread gene-body DNA hypomethylation in chronic lymphocytic leukemia. *Nat. Genet.* **2012**, *44*, 1236–1242. [[CrossRef](#)]
132. Doi, A.; Park, I.H.; Wen, B.; Murakami, P.; Aryee, M.J.; Irizarry, R.; Herb, B.; Ladd-Acosta, C.; Rho, J.; Loewer, S.; et al. Differential methylation of tissue- and cancer-specific CpG island shores distinguishes human induced pluripotent stem cells, embryonic stem cells and fibroblasts. *Nat. Genet.* **2009**, *41*, 1350–1353. [[CrossRef](#)] [[PubMed](#)]

133. Herman, J.G.; Latif, F.; Weng, Y.; Lerman, M.I.; Zbar, B.; Liu, S.; Samid, D.; Duan, D.S.; Gnarr, J.R.; Linehan, W.M.; et al. Silencing of the VHL tumor-suppressor gene by DNA methylation in renal carcinoma. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 9700–9704. [[CrossRef](#)]
134. Lachance, G.; Uniacke, J.; Audas, T.E.; Holterman, C.E.; Franovic, A.; Payette, J.; Lee, S. DNMT3a epigenetic program regulates the HIF-2 α oxygen-sensing pathway and the cellular response to hypoxia. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 7783–7788. [[CrossRef](#)]
135. Yin, H.; Blanchard, K.L. DNA methylation represses the expression of the human erythropoietin gene by two different mechanisms. *Blood* **2000**, *95*, 111–119.
136. Kallio, P.J.; Okamoto, K.; O'Brien, S.; Carrero, P.; Makino, Y.; Tanaka, H.; Poellinger, L. Signal transduction in hypoxic cells: Inducible nuclear translocation and recruitment of the CBP/p300 coactivator by the hypoxia-inducible factor-1 α . *EMBO J.* **1998**, *17*, 6573–6586. [[CrossRef](#)] [[PubMed](#)]
137. Wellmann, S.; Bettkober, M.; Zelmer, A.; Seeger, K.; Faigle, M.; Eltzhig, H.K.; Buhner, C. Hypoxia upregulates the histone demethylase JMJD1A via HIF-1. *Biochem. Biophys. Res. Commun.* **2008**, *372*, 892–897. [[CrossRef](#)] [[PubMed](#)]
138. Watson, J.A.; Watson, C.J.; McCrohan, A.M.; Woodfine, K.; Tosetto, M.; McDaid, J.; Gallagher, E.; Betts, D.; Baugh, J.; O'Sullivan, J.; et al. Generation of an epigenetic signature by chronic hypoxia in prostate cells. *Hum. Mol. Genet.* **2009**, *18*, 3594–3604. [[CrossRef](#)] [[PubMed](#)]
139. Bonasio, R.; Tu, S.; Reinberg, D. Molecular signals of epigenetic states. *Science* **2010**, *330*, 612–616. [[CrossRef](#)] [[PubMed](#)]
140. Reik, W. Stability and flexibility of epigenetic gene regulation in mammalian development. *Nature* **2007**, *447*, 425–432. [[CrossRef](#)] [[PubMed](#)]
141. Khavari, D.A.; Sen, G.L.; Rinn, J.L. DNA methylation and epigenetic control of cellular differentiation. *Cell Cycle* **2010**, *9*, 3880–3883. [[CrossRef](#)] [[PubMed](#)]
142. Li, G.; Xiao, Y.; Estrella, J.L.; Ducsay, C.A.; Gilbert, R.D.; Zhang, L. Effect of fetal hypoxia on heart susceptibility to ischemia and reperfusion injury in the adult rat. *J. Soc. Gynecol. Investig.* **2003**, *10*, 265–274. [[CrossRef](#)]
143. Xue, Q.; Zhang, L. Prenatal hypoxia causes a sex-dependent increase in heart susceptibility to ischemia and reperfusion injury in adult male offspring: Role of protein kinase C ϵ . *J. Pharmacol. Exp. Ther.* **2009**, *330*, 624–632. [[CrossRef](#)]
144. Patterson, A.J.; Chen, M.; Xue, Q.; Xiao, D.; Zhang, L. Chronic prenatal hypoxia induces epigenetic programming of PKC ϵ gene repression in rat hearts. *Circ. Res.* **2010**, *107*, 365–373. [[CrossRef](#)]
145. Rexhaj, E.; Bloch, J.; Jayet, P.Y.; Rimoldi, S.F.; Dessen, P.; Mathieu, C.; Tolsa, J.F.; Nicod, P.; Scherrer, U.; Sartori, C. Fetal programming of pulmonary vascular dysfunction in mice: Role of epigenetic mechanisms. *Am. J. Physiol. Heart Circ. Physiol.* **2011**, *301*, H247–H252. [[CrossRef](#)]
146. Jayet, P.Y.; Rimoldi, S.F.; Stuber, T.; Salmon, C.S.; Hutter, D.; Rexhaj, E.; Thalmann, S.; Schwab, M.; Turini, P.; Sartori-Cucchia, C.; et al. Pulmonary and systemic vascular dysfunction in young offspring of mothers with preeclampsia. *Circulation* **2010**, *122*, 488–494. [[CrossRef](#)] [[PubMed](#)]
147. Smith, Z.D.; Chan, M.M.; Humm, K.C.; Karnik, R.; Mekhoubad, S.; Regev, A.; Eggan, K.; Meissner, A. DNA methylation dynamics of the human preimplantation embryo. *Nature* **2014**, *511*, 611–615. [[CrossRef](#)] [[PubMed](#)]
148. Roemer, I.; Reik, W.; Dean, W.; Klose, J. Epigenetic inheritance in the mouse. *Curr. Biol.* **1997**, *7*, 277–280. [[CrossRef](#)]
149. Anway, M.D.; Cupp, A.S.; Uzumcu, M.; Skinner, M.K. Epigenetic transgenerational actions of endocrine disruptors and male fertility. *Science* **2005**, *308*, 1466–1469. [[CrossRef](#)] [[PubMed](#)]
150. Hitchins, M.P.; Ward, R.L. Constitutional (germline) MLH1 epimutation as an aetiological mechanism for hereditary non-polyposis colorectal cancer. *J. Med. Genet.* **2009**, *46*, 793–802. [[CrossRef](#)] [[PubMed](#)]
151. Chen, Q.; Yan, W.; Duan, E. Epigenetic inheritance of acquired traits through sperm RNAs and sperm RNA modifications. *Nat. Rev. Genet.* **2016**, *17*, 733–743. [[CrossRef](#)] [[PubMed](#)]
152. Gibbs, J.R.; van der Brug, M.P.; Hernandez, D.G.; Traynor, B.J.; Nalls, M.A.; Lai, S.L.; Arepalli, S.; Dillman, A.; Rafferty, I.P.; Troncoso, J.; et al. Abundant quantitative trait loci exist for DNA methylation and gene expression in human brain. *PLoS Genet* **2010**, *6*, e1000952. [[CrossRef](#)]

153. Bell, J.T.; Pai, A.A.; Pickrell, J.K.; Gaffney, D.J.; Pique-Regi, R.; Degner, J.F.; Gilad, Y.; Pritchard, J.K. DNA methylation patterns associate with genetic and gene expression variation in HapMap cell lines. *Genome Biol.* **2011**, *12*, R10. [[CrossRef](#)]
154. Moen, E.L.; Zhang, X.; Mu, W.; Delaney, S.M.; Wing, C.; McQuade, J.; Myers, J.; Godley, L.A.; Dolan, M.E.; Zhang, W. Genome-wide variation of cytosine modifications between European and African populations and the implications for complex traits. *Genetics* **2013**, *194*, 987–996. [[CrossRef](#)]
155. Zhang, X.; Moen, E.L.; Liu, C.; Mu, W.; Gamazon, E.R.; Delaney, S.M.; Wing, C.; Godley, L.A.; Dolan, M.E.; Zhang, W. Linking the genetic architecture of cytosine modifications with human complex traits. *Hum. Mol. Genet.* **2014**, *23*, 5893–5905. [[CrossRef](#)]
156. McClay, J.L.; Shabalina, A.A.; Dozmorov, M.G.; Adkins, D.E.; Kumar, G.; Nerella, S.; Clark, S.L.; Bergen, S.E.; Swedish Schizophrenia, C.; Hultman, C.M.; et al. High density methylation QTL analysis in human blood via next-generation sequencing of the methylated genomic DNA fraction. *Genome Biol.* **2015**, *16*, 291. [[CrossRef](#)] [[PubMed](#)]
157. Zhi, D.; Aslibekyan, S.; Irvin, M.R.; Claas, S.A.; Borecki, I.B.; Ordovas, J.M.; Absher, D.M.; Arnett, D.K. SNPs located at CpG sites modulate genome–epigenome interaction. *Epigenetics* **2013**, *8*, 802–806. [[CrossRef](#)] [[PubMed](#)]
158. Brown, C.J.; Rupert, J.L. Hypoxia and environmental epigenetics. *High Alt. Med. Biol.* **2014**, *15*, 323–330. [[CrossRef](#)] [[PubMed](#)]
159. Scherrer, U.; Allemann, Y.; Rexhaj, E.; Rimoldi, S.F.; Sartori, C. Mechanisms and drug therapy of pulmonary hypertension at high altitude. *High Alt. Med. Biol.* **2013**, *14*, 126–133. [[CrossRef](#)] [[PubMed](#)]
160. Alkorta-Aranburu, G.; Beall, C.M.; Witonsky, D.B.; Gebremedhin, A.; Pritchard, J.K.; Di Rienzo, A. The genetic architecture of adaptations to high altitude in Ethiopia. *PLoS Genet* **2012**, *8*, e1003110. [[CrossRef](#)] [[PubMed](#)]
161. Julian, C.G.; Pedersen, B.S.; Salmon, C.S.; Yang, I.V.; Gonzales, M.; Vargas, E.; Moore, L.G.; Schwartz, D.A. Unique DNA methylation patterns in offspring of hypertensive pregnancy. *Clin. Transl. Sci.* **2015**, *8*, 740–745. [[CrossRef](#)] [[PubMed](#)]
162. Julian, C.G. Epigenomics and human adaptation to high altitude. *J. Appl. Physiol.* **2017**, *123*, 1362–1370. [[CrossRef](#)]
163. Beall, C.M.; Cavalleri, G.L.; Deng, L.; Elston, R.C.; Gao, Y.; Knight, J.; Li, C.; Li, J.C.; Liang, Y.; McCormack, M.; et al. Natural selection on EPAS1 (HIF2 α) associated with low hemoglobin concentration in Tibetan highlanders. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 11459–11464. [[CrossRef](#)]
164. Minamishima, Y.A.; Moslehi, J.; Bardeesy, N.; Cullen, D.; Bronson, R.T.; Kaelin, W.G., Jr. Somatic inactivation of the PHD2 prolyl hydroxylase causes polycythemia and congestive heart failure. *Blood* **2008**, *111*, 3236–3244. [[CrossRef](#)]
165. Takeda, K.; Aguila, H.L.; Parikh, N.S.; Li, X.; Lamothe, K.; Duan, L.J.; Takeda, H.; Lee, F.S.; Fong, G.H. Regulation of adult erythropoiesis by prolyl hydroxylase domain proteins. *Blood* **2008**, *111*, 3229–3235. [[CrossRef](#)]
166. Percy, M.J.; Zhao, Q.; Flores, A.; Harrison, C.; Lappin, T.R.; Maxwell, P.H.; McMullin, M.F.; Lee, F.S. A family with erythrocytosis establishes a role for prolyl hydroxylase domain protein 2 in oxygen homeostasis. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 654–659. [[CrossRef](#)] [[PubMed](#)]
167. Liu, J.; Liu, Y.; Ren, L.H.; Li, L.; Wang, Z.; Liu, S.S.; Li, S.Z.; Cao, T.S. Effects of race and sex on cerebral hemodynamics, oxygen delivery and blood flow distribution in response to high altitude. *Sci. Rep.* **2016**, *6*, 30500. [[CrossRef](#)] [[PubMed](#)]
168. Bernstein, D.L.; Le Lay, J.E.; Ruano, E.G.; Kaestner, K.H. TALE-mediated epigenetic suppression of CDKN2A increases replication in human fibroblasts. *J. Clin. Investig.* **2015**, *125*, 1998–2006. [[CrossRef](#)] [[PubMed](#)]
169. Maeder, M.L.; Angstman, J.F.; Richardson, M.E.; Linder, S.J.; Cascio, V.M.; Tsai, S.Q.; Ho, Q.H.; Sander, J.D.; Reyon, D.; Bernstein, B.E.; et al. Targeted DNA demethylation and activation of endogenous genes using programmable TALE-TET1 fusion proteins. *Nat. Biotechnol.* **2013**, *31*, 1137–1142. [[CrossRef](#)] [[PubMed](#)]

