

Figure S1. FISH (Fluorescence in situ hybridization) of repetitive DNA probes on metaphase plates of the Siberian sturgeon. **a** Satellites $(ACAT)_n$ (red) and *Arut802A* (green), on the right the same plate with $(ACAT)_n$ only, arrows mark chromosomes with co-localized probes; **b** satellite *Arut19A* (red), on the right DAPI inverted plate, arrows mark chromosomes with signals; **c** satellite *Arut30A* (red) on GTG (G-banding by trypsin using Giemsa) differential stained metaphase plate (left). Scale bar is 10 μ m.

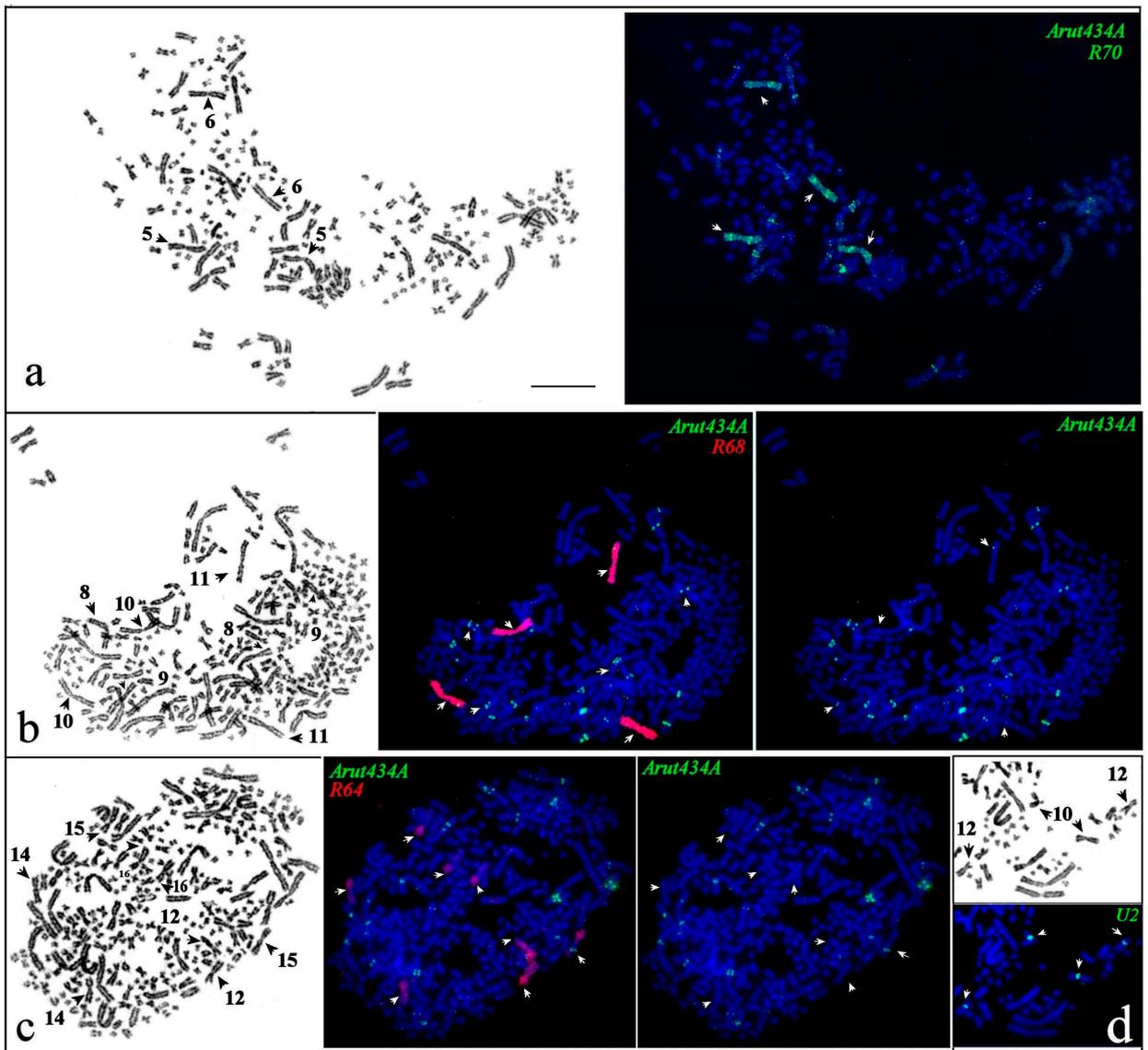


Figure S2. FISH of repetitive and microdissection-derived painting probes (right) on GTG-banded metaphase plates (left) of the Siberian sturgeon (a–c). **a** *Arut434A* (green) and painting probe for sterlet chromosome 3 (R70, green) co-localized on sturgeon chromosomes 5 and 6; **b** *Arut434A* (green) and painting probe for sterlet chromosome 6 (R68, red); **c** *Arut434A* (green) and painting probe for chromosome ARUT8 (R64, red); **d** *Arut802* (red) and U2 snRNA probe (green) on a part of metaphase plate of sterlet. Chromosomes with signals are marked and numbered. Scale bar is 10 μ m.

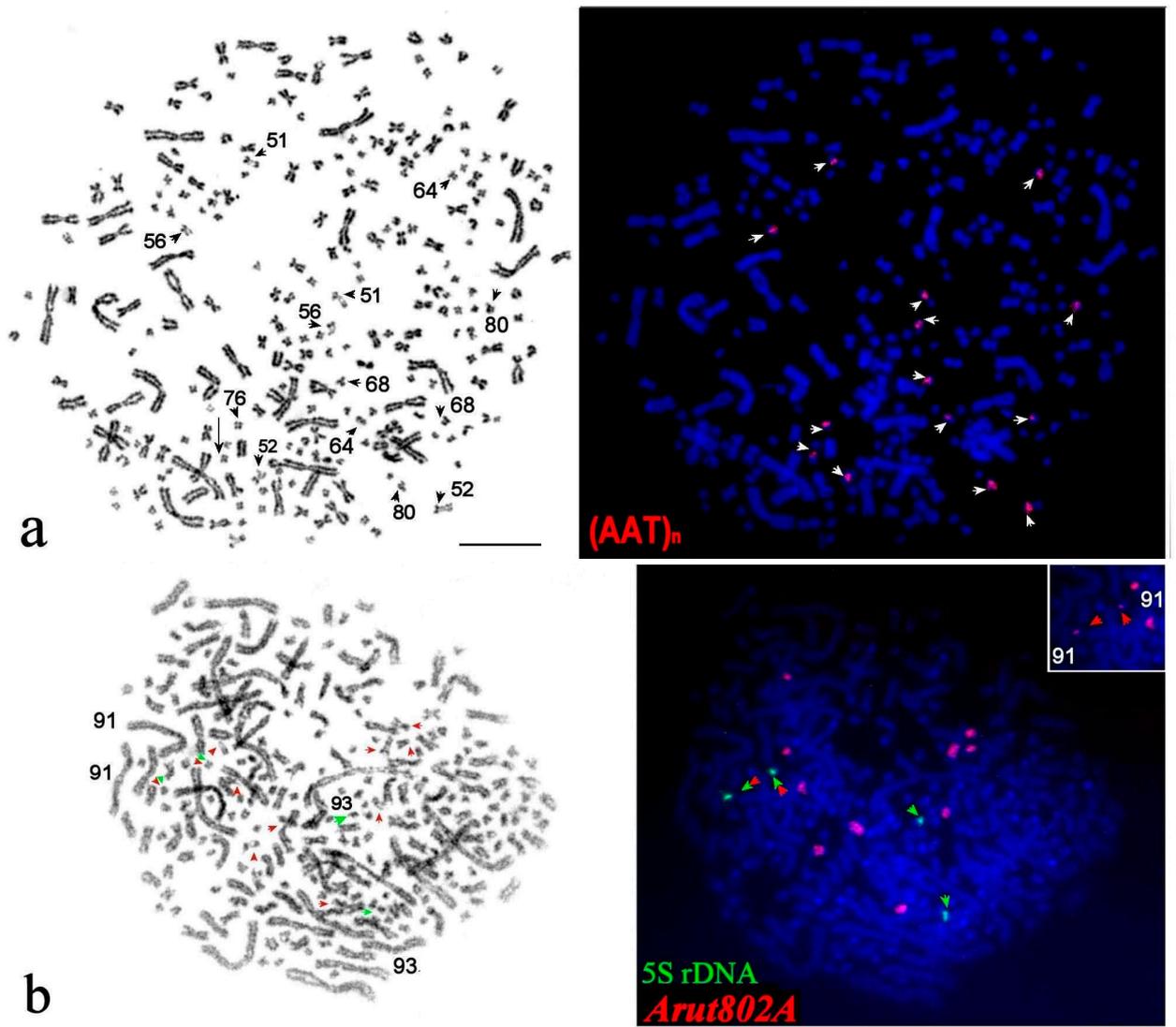


Figure S3. FISH of repetitive probes (right) on GTG-banded metaphase plates (left) of the Siberian sturgeon (a, b). a (AAT)_n (red); b 5S rDNA (green) and *Arut802A* (red), in the right corner is an inset part of the metaphase with overlapped signals. Chromosomes with signals are marked and numbered, the double arrows indicate chromosomes with co-localized probes. Scale bar is 10 μm.

Table S1. Chromosome localization of sterlet-derived chromosome-specific probes on orthologs of *Acipenser ruthenus* (ARUT) and *Acipenser baerii* (ABAE)

#	Molecular markers		Chromosome numbers		Characteristics	
	tandem repeats	ARUT-derived chromosome specific probe	ARUT	ABAE		
1	<i>Arut 434A</i>	1p	1	1	<ul style="list-style-type: none"> - ABAE 1 and 2: strong signals as on ARUT 1; - ABAE 1: a single signal, ABAE 2: two signals 	
				2		
		2p	2	3		<ul style="list-style-type: none"> - ABAE 3 and 4: weak signals as on ARUT 2; - ABAE 4: weak single signal; - ABAE 3: double weak signals
				4		
2	<i>Arut 434A</i>	3	3	5	- ABAE 5 and 6: signals on both arms	
				6		
		4	4	7	<ul style="list-style-type: none"> - one of ARUT 4 ohnologs underwent fission resulted in two acrocentrics; - ABAE 102: strong signals 	
				101		
102						

3	<i>Arut 434A</i>	5	5	8	<ul style="list-style-type: none"> - ABAE 8 and 9: multiple signals on q-arms; - ABAE 9 the distance between the blocks is less than on ABAE 8 	
				9		
		6	6	10		
				11		
4	<i>Arut 434A</i>	7, 14	7	13	<ul style="list-style-type: none"> - one of the orthologs of ARUT 7 divided into 2 acrocentrics: - ABAE 108; - an arm of the submetacentric chromosome ABAE 23; - reduced block on on ABAE 23 and 108 	
				23		
				108		
			14	103		<ul style="list-style-type: none"> - distal end of ABAE 103; - interstitial block on ABAE 104 - ABAE 103 is bigger than ABAE 104, ABAE 104 is close in size and morphology to ARUT 14
5	<i>Arut 434A</i>	8	8	12	<ul style="list-style-type: none"> - ABAE 12 labelled brighter than ABAE 14 by ARUT 8 probe; 	

				14	- missing signals of ARUT434A
			9	15	- p-arms of the both paralogs produce bright signals by ARUT 8 probe; - <i>Arut434A</i> repeat marks one homolog of ABAE 15 brighter
				16	
6	U2	10	10cent	19p cent	- ABAE 19 and 20 are similar
				20p cent	
			12cent	25pq cent	- ABAE 25 and 35 are different in size and G-banding patterns
				35pq cent	
7	<i>28S/18S rDNA</i>	-	30	30	- ABAE 30 and 50 are different in size and G-banding pattern; - ABAE 30 is bigger than ARUT30, and, probably, underwent a fusion with another element; - ABAE 50 are heteromorphic: one of homologs is similar to ARUT 30, while the other is smaller due to reduced p-arm
				50	
			-		
8	<i>28S/18S rDNA,</i> <i>Arut 802A,</i> <i>ArutF 26A,</i> <i>(AAT)n</i>	-	31	51	- ABAE 51 and 52 are similar to ARUT 31 and 32, respectively - ABAE 55 and 56 are smaller than their sterlet orthologs - ABAE 63 probably arose after AcR2;
				56	

				63	- the size of <i>Arut26</i> and $(AAT)_n$ blocks on ABAE 51, 52 and 56 are different
			32		
				52	
				55	
9	<i>Arut F26A</i> , $(AAT)_n$	-	39	68	
				76	
			40	64	
				80	
10	<i>5S rDNA</i> <i>Arut 219A</i>	-	41	91	<p>- <i>5S rDNA</i> and <i>Arut219A</i> colocalized on ABAE91 and 93</p> <p>- ABAE91 and 93 are different in morphology and G-banding pattern;</p> <p>- <i>Arut19A</i>, <i>Arut40A</i> and <i>Arut57A</i> colocalized with <i>Arut219</i>;</p> <p>- <i>5S rDNA</i> and <i>Arut802A</i> co-localized on ABAE 91</p>
				93	
11	<i>Arut F167A</i>	57 (R3)	57	86q	<p>- ABAE 86q and ABAE 100q are orthologs of ARUT 57;</p> <p>- fusion with ABAE 86p;</p> <p>- fusion with ABAE 100p</p>
				100q	

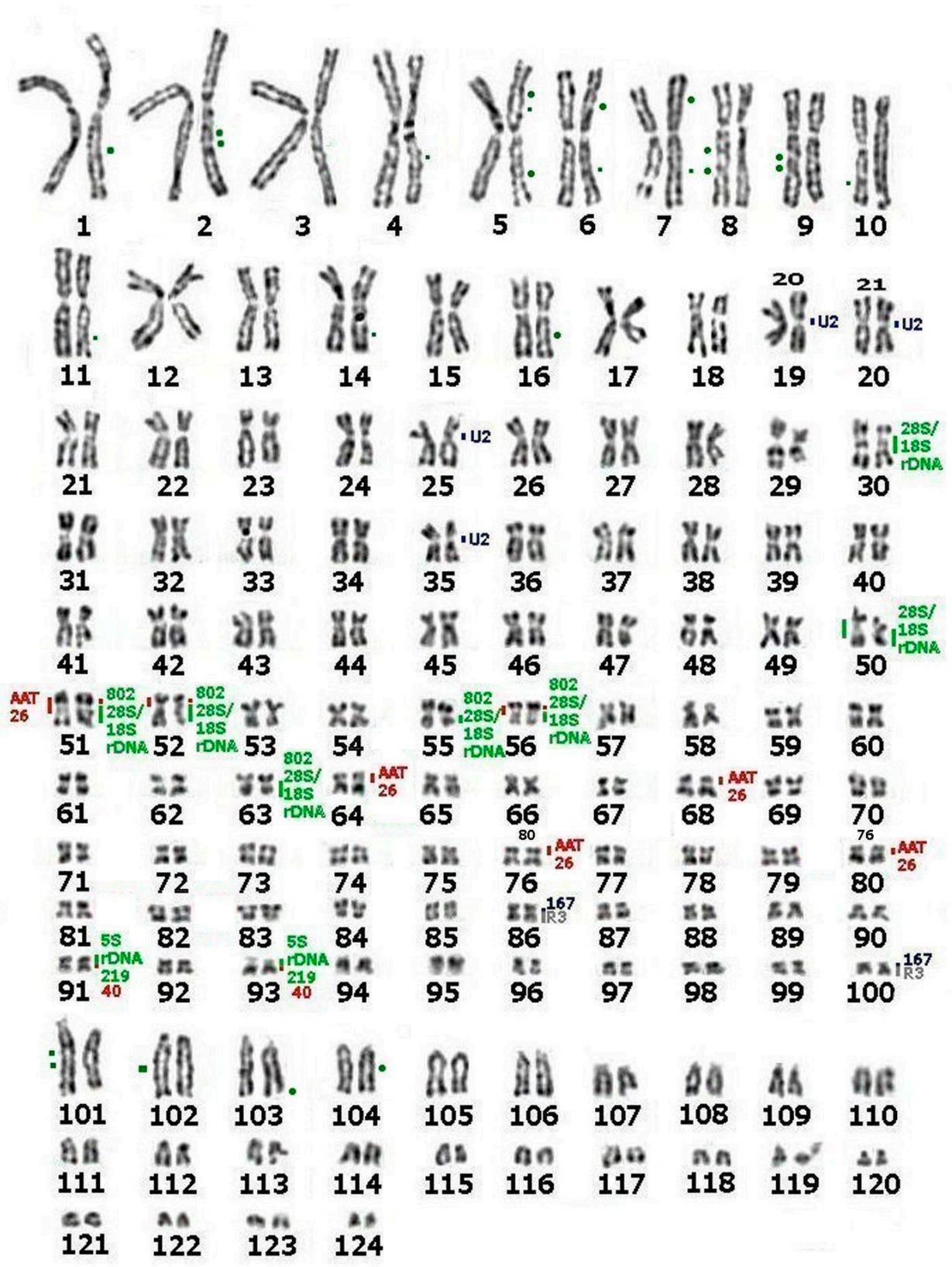


Figure S4. GTG-banded chromosomes Siberian sturgeon with assigned probes: *Arut434A* (434), U2 snRNA genes (U2), ribosomal major cluster (28S/18S rDNA), *Arut26A* (26), *Arut802A* (802), (AAT)_n, 5S ribosomal DNA (5S rDNA), *Arut40A* (40), *Arut167A* (167), *Arut219A* (219), chromosome specific probe ARUT 57 (R3).