

Supplement S1: Details of available software to carry out probabilistic genotyping

Software	Approach	License	Reference
<i>CeesIT</i>	Continuous	(a)	[1]
<i>DNAmixtures</i>	Continuous	Open-source ^(b)	[2]
<i>DNAStatistX</i>	Continuous	Open-source	[3]
<i>DNA-View Mixture Solution</i>	Continuous	Commercial	(c)
<i>eDNA</i>	Continuous	(d)	(d)
<i>EuroForMix</i>	Continuous	Open-source	[4]
<i>Genoproof</i>	Continuous	Commercial	[5]
<i>Kongoh</i>	Continuous	Open-source	[6]
<i>Lab Retriever</i>	Semi-continuous	Open-source	[7]
<i>LikeLTD v.6</i>	Continuous ^(e)	Open-source	[8]
<i>LiRa</i>	Semi-continuous/continuous ^(f)	Commercial	[9]
<i>LoCIM-tool</i>	Empirical	Open-source	[10]
<i>LRmix/LRmix Studio</i>	Semi-continuous	Open-source	[11]
<i>MaSTR</i>	Continuous	Commercial	[12]
<i>STRmixTM</i>	Continuous	Commercial	[13]
<i>TrueAllele</i>	Continuous	Commercial	[14]

Table S1: A list of some of the available software, license conditions and source. Reprinted from Gill, P, Bleka, Ø, Hansson, O, Benschop, C and Haned, H (2020) Forensic Practitioner's Guide to the Interpretation of Complex DNA profiles, London, Academic Press, chapter 7, Copyright (2020) with permission from Elsevier.

- a) *Ceesit* license is: "no cost to local, state and federal forensic DNA laboratories or entities pursuing research, forensic validation or education for non-commercial purposes", but a full license is required for commercial use.
- b) *DNAmixtures* is a free of charge open-source R package, however it requires the HUGIN commercial software to run.

- c) *DNA-View*: <http://dna-view.com/> and <http://dna-view.com/downloads/Mixture%20Solution%20poster.pdf>
- d) *eDNA*: <http://ednalims.com/probabilistic-genotyping/>. Freely available web-based software available to consortium members. *Bullet* uses *LRmix* and *Bulletproof* uses *EuroForMix*. Each program has a custom built graphical user interface.
- e) *LikeLTD* also has a semi-continuous model [15]
- f) *LiRaHT* is available as a continuous model:
https://cdnmedia.euofins.com/europeanwest/media/1418957/lgc_lira_fact_sheet_en_0815_90.pdf

Of the continuous models listed in Table S1, *DNAmixtures*, *DNAView Mixture Solution*, *DNAStatistX*, *LikeLTD*, *LiRaHT* and *EuroForMix* all use the gamma model (Supplement 2) for peak height, but there are differences between software assumptions. *CeesIT* uses a normal distribution. *Kongoh* estimates peak height distributions by using the Monte Carlo simulation based on experimental data to consider allele or locus-specific effects. *STRmixTM* and *TrueAllele* are both commercial solutions, based on a Bayesian approach through specifying prior distributions on the unknown model parameters. They use Markov Chain Monte Carlo (MCMC) methods [16, 17] to calculate marginalised likelihood expressions by simultaneously sampling over the discrete set of genotypes for the unknown contributors specified in the model and the unknown parameters. *MaSTR* and *GenoProof Mixture* also employ MCMC in their calculations.

Supplement S2: Details of the gamma model

The gamma distribution has useful properties: the shape and scale parameters are calculated from M_x , the mixture proportion for contributor 1 and $1-M_x$ for contributor 2; ω is the coefficient of peak height variation and μ is the peak height expectation.

The expected peak height of any given allele is obtained by multiplying together shape and scale parameters:

$$\mu = E[Y] = \alpha \times \beta \quad (1)$$

The coefficient of variance (ω) is the standard deviation divided by the peak height expectation:

$$\omega = CV[Y] = \alpha^{-\frac{1}{2}} \quad (2)$$

Hence the shape and scale parameters are defined as:

$$\alpha = \frac{1}{\omega^2} \quad (3)$$

$$\beta = \mu\omega^2 \quad (4)$$

The per contributor parameters are calculated as follows:

- Contributor 1: $\alpha_1 = M_x/\omega^2$ (5)

- Contributor 2: $\alpha_2 = (1 - M_x)/\omega^2$ (6)

- $\beta = \mu \times \omega^2$ (7)

- $\mu_1 = \alpha_1 \times \beta$ (8)

- $\mu_2 = \alpha_2 \times \beta$ (9)

The gamma model calculates the values of the shape and scale parameters by maximum likelihood estimation. With this method, the value of the evidence under $Pr(O|H_1)$, by optimising the values of M_x , ω , μ . The values are adjusted via a large number of iterative steps, until the log likelihood value is maximised. The process is repeated to calculate the value of the evidence under $Pr(O|H_2)$ to generate a second maximised log likelihood.

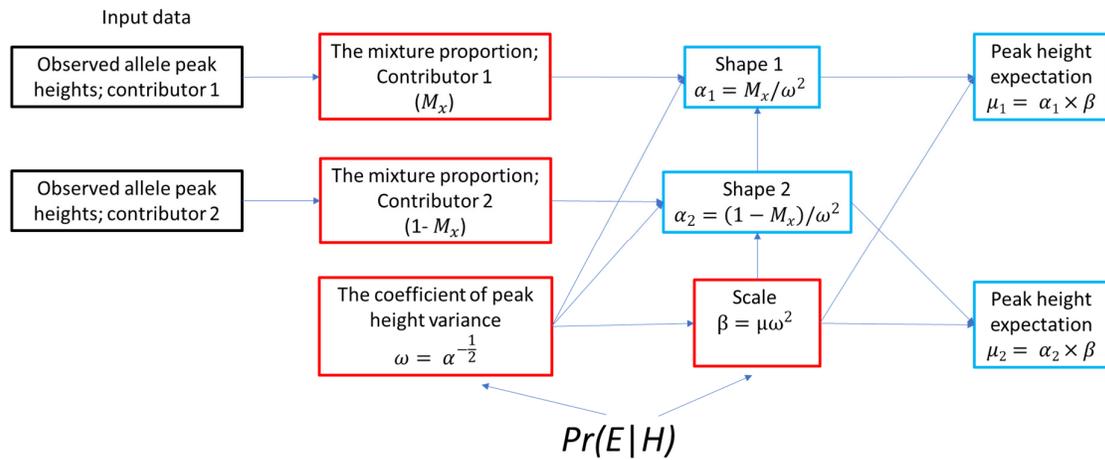


Fig. S1: An outline of the maximum likelihood estimation of the basic gamma model. The input data are the allele peak heights; the M_x , ω , μ parameters are estimated by optimisation (outlined in red): the values are iteratively adjusted to maximise the values of $Pr(O|H_i)$ and

$\Pr(O|H_2)$) respectively, using separate parameter optimisations in order to derive the likelihood ratio.

References

1. Swaminathan, H., et al., *CEESIt: A computational tool for the interpretation of STR mixtures*. Forensic Sci Int Genet, 2016. **22**: p. 149-160.
2. Graversen, T. and S. Lauritzen, *Computational aspects of DNA mixture analysis*. Statistics and Computing, 2015. **25**(3): p. 527-541.
3. Benschop, C.C.G., et al., *DNAXs/DNAstatistX: Development and validation of a software suite for the data management and probabilistic interpretation of DNA profiles*. Forensic Sci Int Genet, 2019. **42**: p. 81-89.
4. Bleka, Ø., G. Storvik, and P. Gill, *EuroForMix: an open source software based on a continuous model to evaluate STR DNA profiles from a mixture of contributors with artefacts*. Forensic Science International: Genetics, 2016. **21**: p. 35-44.
5. Götz, F.M., et al., *GenoProof Mixture 3—New software and process to resolve complex DNA mixtures*. Forensic Science International: Genetics Supplement Series, 2017. **6**: p. e549-e551.
6. Manabe, S., et al., *Development and validation of open-source software for DNA mixture interpretation based on a quantitative continuous model*. PLoS One, 2017. **12**(11): p. e0188183.
7. Inman, K., et al., *Lab Retriever: a software tool for calculating likelihood ratios incorporating a probability of drop-out for forensic DNA profiles*. BMC Bioinformatics, 2015. **16**: p. 298.
8. Steele, C.D., M. Greenhalgh, and D.J. Balding, *Verifying likelihoods for low template DNA profiles using multiple replicates*. Forensic Sci Int Genet, 2014. **13**: p. 82-9.
9. Puch-Solis, R. and T. Clayton, *Evidential evaluation of DNA profiles using a discrete statistical model implemented in the DNA LiRa software*. Forensic Sci Int Genet, 2014. **11**: p. 220-8.
10. Benschop, C.C. and T. Sijen, *LoCIM-tool: An expert's assistant for inferring the major contributor's alleles in mixed consensus DNA profiles*. Forensic Sci Int Genet, 2014. **11**: p. 154-65.
11. Haned, H., K. Slooten, and P. Gill, *Exploratory data analysis for the interpretation of low template DNA mixtures*. Forensic Sci Int Genet, 2012. **6**(6): p. 762-74.
12. Adamowicz, M., et al., *Validation of MaSTR™ software: Extensive study of fully-continuous probabilistic mixture analysis using PowerPlex® Fusion 2–5 contributor mixtures*. Forensic Science International: Genetics Supplement Series, 2019. **7**(1): p. 641-643.
13. Taylor, D., J.A. Bright, and J. Buckleton, *The interpretation of single source and mixed DNA profiles*. Forensic Sci Int Genet, 2013. **7**(5): p. 516-28.
14. Perlin, M.W. and A. Sinelnikov, *An information gap in DNA evidence interpretation*. PLoS One, 2009. **4**(12): p. e8327.
15. Steele, C.D., M. Greenhalgh, and D.J. Balding, *Evaluation of low-template DNA profiles using peak heights*. Stat Appl Genet Mol Biol, 2016. **15**(5): p. 431-445.
16. Hastings, W.K., *Monte Carlo sampling methods using Markov chains and their applications*. 1970.
17. Metropolis, N., et al., *Equation of state calculations by fast computing machines*. The journal of chemical physics, 1953. **21**(6): p. 1087-1092.