

# Particulate Matter in Swine Barns: A Comprehensive Review

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**Abstract:** Particulate matter (PM) represents an air quality management challenge for confined swine production systems. Due to the limited space and ventilation rate, PM can reach relatively high concentrations in swine barns. PM in swine barns possesses different physical, chemical, and biological characteristics than that in the atmosphere and other indoor environments. As a result, it exerts different environmental and health effects and creates some unique challenges regarding PM measurement and mitigation. Numerous research efforts have been made, generating massive data and information. However, relevant review reports are sporadic. This study aims to provide an updated comprehensive review of swine barn PM, focusing on publications since 1990. It covers various topics including PM characteristics, sources, measurement methods, and in-barn mitigation technologies. As PM in swine barns is primarily of biological origins, bioaerosols are reviewed in great detail. Relevant topics include bacterial/fungal counts, viruses, microbial community composition, antibiotic-resistant bacteria, antibiotic resistance genes, endotoxins, and (1→3)-β-D-glucans. For each topic, existing knowledge is summarized and discussed and knowledge gaps are identified. Overall, PM in swine barns is complicated in chemical and biological composition and highly variable in mass concentrations, size, and microbial abundance. Feed, feces, and skins constitute the major PM sources. Regarding in-barn PM mitigation, four technologies (oil/water sprinkling, ionization, alternation of feed and feeders, and recirculating air filtration) are dominant. However, none of them have been widely used in commercial barns. A collective discussion of major knowledge gaps and future research needs is offered at the end of the report.

**Keywords:** swine barn; particulate matter; characteristics; bioaerosols; mitigation; measurement



**Citation:** Yang, X.; Haleem, N.; Osabutey, A.; Cen, Z.; Albert, K.L.; Autenrieth, D. Particulate Matter in Swine Barns: A Comprehensive Review. *Atmosphere* **2022**, *13*, 490. <https://doi.org/10.3390/atmos13030490>

Academic Editor: Carla Viegas

Received: 25 February 2022

Accepted: 13 March 2022

Published: 17 March 2022

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## 1. Introduction

### 1.1. Rationale

The production of pork, as a major source of proteins in many countries, cannot be overemphasized given the growing global population and the pursuit of improved nutrition [1]. The U.S. is the second-largest pork producing country in the world, with 74.8 million pigs in inventory as of 1 March 2021 [2] and exported pork and pork products at a value of USD 7.7 billion in 2020 [3]. The vast majority of pigs are kept in confined swine barns, also known as confinement buildings or indoor systems. Swine barns not only protect pigs from harsh environments, but also improve the management of pigs on the aspects of monitoring, feeding, vaccination, waste management, etc. However, confinement swine production receives public scrutiny especially when it comes to animal welfare and

environmental stewardship [1,4]. Both animal welfare and environmental stewardship are tied to air pollutants generated inside swine barns, such as ammonia, hydrogen sulfide, odors, and particulate matter (PM).

PM refers to a collection of solid particles and liquid droplets suspended in an air environment. PM in the ambient air, also known as atmospheric PM, is one of six criteria air pollutants regulated by the National Ambient Air Quality Standards (NAAQS) [5] due to its adverse impacts on human health and welfare. PM has been known to compromise the human respiratory, cardiovascular, and even reproduction systems [6,7]. The welfare effects of atmospheric PM include impaired visibility, reduced photosynthesis, and the acidification and eutrophication of ecosystems caused by PM deposition [8]. It is important to note that PM in swine barns is different from atmospheric PM in terms of sources, sizes, constituents, etc. As a result, it has certain unique health and welfare implications that justify associated assessment and mitigation efforts. For example, swine barn PM carries a considerable number of microorganisms, and some of them can be pathogenic and responsible for the airborne transmissions of diseases [9]. Another example is that swine barn PM can concentrate and transport odorous chemicals downwind, causing a greater or more persistent odor nuisance [10].

Numerous studies have been conducted on PM in swine barns. They cover a broad range of research topics, including PM sources, characterization, emissions, measurement methods, mitigation technologies, health impacts (on humans and animals), exposure thresholds, etc. A few review efforts were made [11,12]. In a mini-review paper, Ref. [11] summarized the sources, characteristics, measurement methods, dose–response and control standards, impacts on animals, concentration modeling, and mitigation technologies for PM in swine barns. Ref. [12] offered a comprehensive review about PM from livestock production systems, covering various livestock environments besides swine barns. Both papers were published over ten years ago. Since then, numerous publications have become available regarding PM in swine barns. In summary, an updated review specifically of PM in swine barns is needed given the importance of this topic and the lack of similar efforts in recent years. The target readers include environmental engineers, veterinarians, animal scientists, industrial hygienists, agricultural engineers, and government agencies.

### 1.2. Goal and Scope

A preliminary literature search has identified >600 relevant publications, covering a broad range of topics. To make the report manageable, the scope of the review effort is defined as below:

- It focuses on PM inside swine barns. The emissions of PM from swine barns and the transport and fate of PM in the environment are excluded.
- It consists of three major parts: PM characteristics, measurement methodology, and mitigation technologies. For mitigation technologies, only those for in-barn PM mitigation are reviewed.
- It focuses on publications since 1990. Earlier publications are included when necessary (e.g., when discussing technology development).
- No detailed review is included about the health effects of PM in swine barns. As the health effects are closely related to PM components, a brief discussion is sometimes given when reviewing a specific component. For detailed health effect information, several review papers are available [13–18].
- The PM characteristics reviewed include physical characteristics, chemical composition, bioaerosols, and PM sources. The review of bioaerosols covers bacteria, fungi, viruses, antibiotic resistant bacteria, antibiotic resistance genes, and bioaerosol markers in swine barns.

Swine barns are by no means significant PM emitters or contributors to atmospheric PM. In 2005, approximately 2.2 million tons of PM<sub>10</sub> in the U.S. were emitted from anthropogenic sources [19]. Assuming an aggressive PM<sub>10</sub> emission factor for pigs (3 g PM<sub>10</sub> per animal unit per day; compiled from multiple sources), the total PM<sub>10</sub> emission from

the U.S. swine barns would be <10,000 tons in the same year [20]. PM in swine barns is primarily an indoor or local air quality issue.

### 1.3. Terminology and Definitions

PM in swine barns has attracted researchers from various disciplines, backgrounds, or regions. They may use different terms to describe the same entity. To ease the reading experience, standardized terminology is used throughout the report, as clarified below:

- “PM” replaces the following terms: “dust”, “airborne particles” (or “airborne particulates”), “suspended particulate matter”, and “aerosols.” These terms carry slightly different meanings but were interchangeably used with PM in most of the previous studies. “A particle” and “particles” are used in the review when singular and plural expressions are necessary.
- “Total suspended particles (TSP)” replace “total particles.” Although both terms refer to particles of all sizes, many of the previous studies did not use standard TSP samplers to collect total particles. A thorough discussion about the samplers’ efficiency is impossible given the lack of details in the literature. For simplicity, TSP and total particles are treated as the same entity.
- “Bioaerosols” replace “biological aerosols”, “biological particles”, and “biological aerosol particles.” In a broad sense, bioaerosols refer to any PM of biological origins, including bacteria, fungal spores, viruses, pollen grains, insect fragments, and plant detritus [21]. However, this broad-sense definition can hardly apply to PM in swine barns because most in-barn particles are suspended from biomaterials (e.g., feed and feces). To address the dilemma, bioaerosols are defined in this review as PM of microbiological origins.

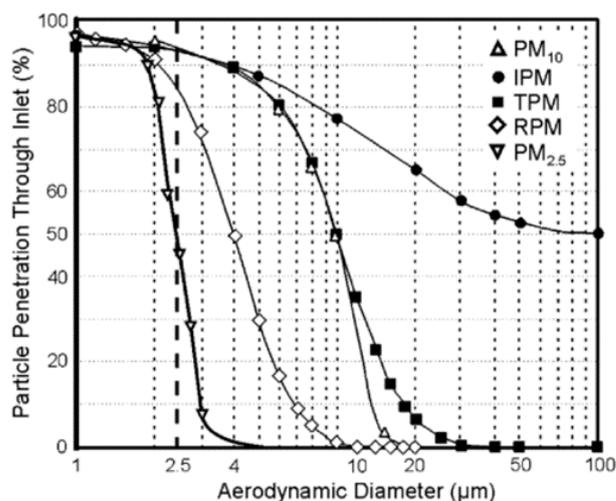
PM concentrations and properties are reported and discussed based on particle size. The following size fractions were often studied in swine barns:

- Inhalable PM refers to the particles that can enter the human respiratory tract through the nose or mouth during normal breathing.
- Respirable PM refers to the particles that can penetrate the human respiratory tract and reach air exchange regions, e.g., alveoli in the lung.
- TSP refers to particles of all sizes.
- PM<sub>10</sub> refers to particles with aerodynamic diameters less than 10 microns (μm).
- PM<sub>2.5</sub>, also known as fine PM, refers to particles with aerodynamic diameters less than 2.5 μm.
- PM<sub>2.5–10</sub>, also known as coarse PM, refers to particles with aerodynamic diameters greater than 2.5 μm but less than 10 μm.

Among these size fractions, inhalable PM and respirable PM are defined by the American Conference of Governmental Industrial Hygienists (ACGIH) for PM exposure assessment in occupational environments [22], based on the compartments where these particles can reach in the human respiratory system. Another ACGIH-defined size fraction is thoracic PM, and it refers to a sub-fraction of inhalable PM that can pass the upper respiratory tract and reach the lower respiratory tract (i.e., thoracic airways). Thoracic PM was rarely measured in swine barns for its similarity to PM<sub>10</sub> (Figure 1). Other size fractions that were occasionally studied include PM<sub>1</sub>, PM<sub>5</sub>, and PM<sub>7</sub>, where PM<sub>x</sub> refers to particles with aerodynamic diameters less than  $x$  μm.

It is noteworthy that the definitions given above are scientific definitions. In reality, PM size fractions are defined by reference sampling methods. Under this definition framework, PM<sub>10</sub> refers to particles collected with a reference PM<sub>10</sub> sampler or any samplers with an equivalent sampling curve (Figure 1). The rationale of the operational definition lies in the fact that no samplers can remove all particles larger than a target size (e.g., 10 μm for PM<sub>10</sub> samplers) while retaining all particles smaller than that size. A key parameter of a sampling curve is 50% cut size (D<sub>50</sub>)—the diameter of particles with 50% sampling efficiency. D<sub>50</sub> is

100 μm for inhalable PM, 4 μm for respirable PM, 10 μm for PM<sub>10</sub> and thoracic PM, and 2.5 μm for PM<sub>2.5</sub>.



**Figure 1.** Sampling curves of the EPA defined reference PM<sub>10</sub> and PM<sub>2.5</sub> samplers, as well as the ACGIH defined inhalable, thoracic, and respirable samplers (courtesy of [23]).

## 2. Methodology

Both peer-reviewed and non-peer-reviewed articles were included. For peer-reviewed articles, six databases (CAB International, AGEICOLA, MEDLINE, EBSCO, Scopus, and Web of Science) were selected as the primary sources. For non-peer-reviewed articles, only scholarly ones (e.g., conference papers, extension factsheets, and theses) were included. Many of them were indexed by the six aforementioned databases. For articles unindexed by any database, the Google search engine was used to search for their records on the websites of universities (e.g., “.edu” and “.ac.uk”) and governmental agencies (e.g., “.gov”). Attempts were also made to include non-scholarly articles (e.g., trade magazine articles and newspaper articles); however, no solid or original data were found.

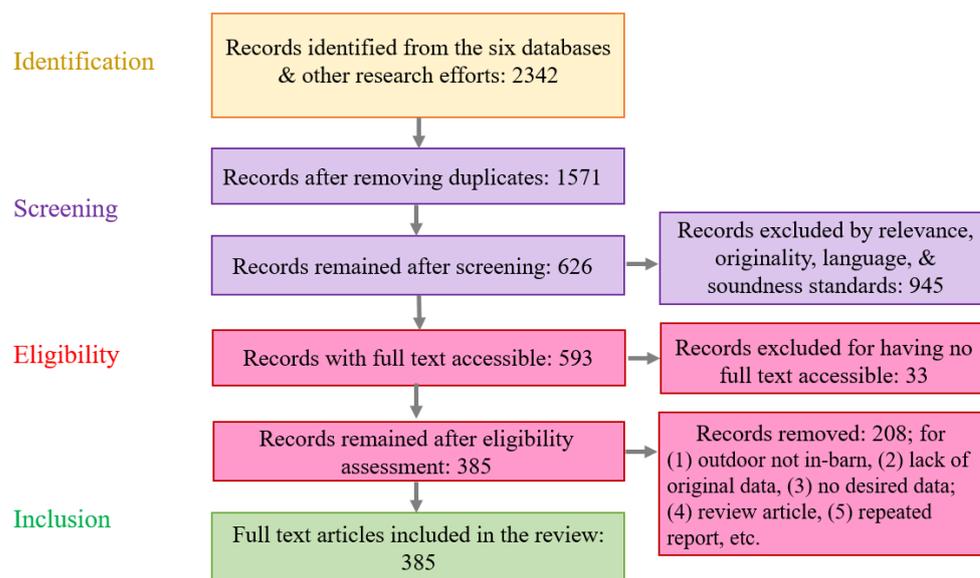
To further increase the number of search records, the references of each identified article were checked to see if any of them would be relevant. The articles citing those identified from the previous steps were searched with Google Scholar. Relevant records were also included. For the broad scope of this study, each section or subsection used a different set of keywords (Table 1). The generic keywords for “overall-swine barn PM” were used in all search efforts. For a specific subject, additional keywords were used to further refine the search results.

**Table 1.** A list of keywords (search terms) in this review project.

Topics	Keywords/Search Terms
Overall swine barn PM	(Swine   pig   hog) and (barn   house   building   farm) and (airborne particle   particulate matter   PM   dust   particulate)
PM characteristics	Concentration   level   count
- concentration	Size distribution   PSD   PM <sub>10</sub>   PM <sub>2.5</sub>   TSP   inhalable   respirable
- size	Source   origin   apportionment
- sources	Composition   elements   ions   odorants   volatile organic compounds   protein   lipid   fatty acid   ash
- chemical composition	(Bioaerosol   biological particles   biological aerosol) and (bacteria   fungi   virus   endotoxin   glucan   antibiotic   DNA   RNA   PCR   sequencing   biomarker)
- bioaerosol	
Measurement methodology	Measurement   sampling   monitoring   sampler   analyzer   monitor   instrument   instrumentation   calibration
Mitigation technologies	Mitigation   abatement   reduction   dedusting   removal

The acquired search records were imported into Zotero (a reference management program), including the title and abstract of each record. With Zotero, the search records were screened based on the following exclusion criteria: (1) Uniqueness. Duplicate records were removed; (2) Relevance. Irrelevant records were removed based on the information in the abstract; (3) Originality. Only original studies were included. The articles citing an original study but providing no further findings were removed; (4) Language. Only articles in English were included. Articles with an English abstract but a non-English full text were removed; and (5) Scientific soundness. Articles with no supporting evidence (observational, experimental, and/or simulative) were removed.

The full texts of the remaining search records were downloaded to further assess their eligibility. Around 6% of the search records had no full texts accessible and were excluded from the review effort. The downloaded full-text articles were read to assess the articles' uniqueness, relevance, originality, and scientific soundness. Articles failing to meet the criteria were removed. Finally, the remaining full-text articles were sorted based on their topics. A total of 385 articles were compiled for review (Figure 2). An additional 98 articles were referenced to support the analysis and discussion of individual topics.



**Figure 2.** A flowchart diagram of the literature search procedure.

### 3. PM Characteristics

#### 3.1. Physical Characteristics

##### 3.1.1. PM Mass Concentrations

PM can be characterized by several concentration measures, including number concentrations, surface area concentrations, and mass concentrations [24]. Among them, PM mass concentrations are most frequently measured because PM regulations are predominantly mass-based. Examples of PM regulations include the NAAQS for PM<sub>10</sub> (daily average concentrations  $\leq 150 \mu\text{g m}^{-3}$ ) and PM<sub>2.5</sub> (daily average concentrations  $\leq 35 \mu\text{g m}^{-3}$ ) [5], and the Occupational Safety and Health Administration (OSHA) Permissible Exposure Limits (PEL) for respirable PM ( $5.0 \text{ mg m}^{-3}$ ) [25]. In swine barns, Ref. [26] recommended an exposure limit of  $2.4 \text{ mg m}^{-3}$  for total dust (TSP) and  $0.23 \text{ mg m}^{-3}$  for respirable PM—both were mass-based. Thus, only PM mass concentrations are reviewed here.

Although PM has been a farm hygiene consideration for over a century [27,28], the first measurement of PM concentrations in swine barns, to our knowledge, was taken in the 1970s [29–32]. These early investigations were largely stimulated by the findings that PM, along with noxious gases (e.g., ammonia), could impact pig health [33,34]. In the 1980s, an increasing number of studies were published about PM concentrations in swine

barns [35–38]. The research focus, however, shifted to occupational exposure assessment. Additionally, in the 1970s and 1980s, various in-barn PM mitigation technologies, such as ionization, oil sprinkling, and feed additives, were initially developed and/or tested. These early measurement and mitigation efforts coincided with the consolidation and concentration of swine production facilities. With a larger number of pigs per barn and a greater stock density than before, swine barns faced a growing challenge regarding indoor air quality management, including PM management.

Moving into the 1990s, the enactment of PM<sub>2.5</sub> standards spurred another round of research interest in swine barn PM. Meanwhile, public awareness of air quality (including PM<sub>10</sub> and PM<sub>2.5</sub>) continued to grow. Table 2 summarizes the monitoring efforts since 1990. It can be seen that PM<sub>10</sub> and PM<sub>2.5</sub> have gradually outpaced inhalable PM and respirable PM as the measurement focus, especially in recent years. Additionally, during this period, several major monitoring campaigns were conducted to determine the baseline emission of air pollutants from animal facilities, e.g., the Four Country study in Northern Europe [39], the Aerial Pollutant Emissions from Confined Animal Buildings (APECAB) study in the U.S. Midwest [40], and the National Air Emissions Monitoring Study (NAEMS) across the U.S. [41]. These campaigns have greatly facilitated the use of highly accurate, real-time instruments and the development of standard operating procedures (SOPs) for PM concentration measurement in swine barns.

**Table 2.** PM mass concentrations in swine barns—A summary of studies since 1990 <sup>a</sup>.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[42] <sup>d</sup>	Mean TSP: 4.13 Mean respirable: 0.48	n/a; n/a	Denmark	n/a	-
[43]	Mean TSP: 3.03–14.05 Mean TSP: 4.10–6.25	Farrowing; n/a Finisher; n/a	Poland	n/a n/a	-
[44]	Control— Mean TSP: 2.1 Sprinkling an oil-water mixture (3 mL oil pig <sup>-1</sup> d <sup>-1</sup> )— Mean TSP: 1.6 Sprinkling an oil-water mixture (11 mL oil pig <sup>-1</sup> d <sup>-1</sup> )— Mean TSP: 0.4	Nursery, finisher; n/a	Denmark	n/a n/a	-
[45] <sup>d</sup>	Geomean inhalable: 2.1 Time 1—	n/a; n/a	Netherlands	All seasons	-
[46] <sup>d</sup>	Geomean TSP: 4.55; 2.62 (GSD) Geomean respirable: 0.23; 2.90 (GSD) Time 2— Geomean TSP: 3.45; 2.39 (GSD) Geomean respirable: 0.26; 2.24 (GSD) Control—	n/a; n/a  n/a; n/a	Iowa	Spring, fall, winter	-
[47]	Mean TSP: 3.59 Mean respirable: 0.27 Adding 4% fat in feed— Mean TSP: 2.26 Mean respirable: 0.122 Control— Mean TSP: 1.99 Mean respirable: 0.124 Adding 4% fat in feed— Mean TSP: 1.16 Mean respirable: 0.075	Nursery; MV   Finisher; MV	Denmark	n/a n/a	-
[48]	Mean TSP: 0.72 (0.12–1.4) Mean respirable: 0.07 (0.01–0.17)	Nursery; MV	Kansas	Warm season (25–34 °C)	Particle size distribution and concentration varied greatly with time.

Table 2. Cont.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[49] <sup>d</sup>	Control— TSP: 3.8 ± 0.2 Oil Sprinkling— TSP: 0.6 ± 0.3	Grower-finisher; MV	Saskatchewan, Canada	Warm season Winter	-
[50]	TSP: 1.0–5.0	Finisher; MV	Germany	n/a	-
[51]	Mean TSP: 8.76 (3.03–14.05) Mean inhalable: 1.87 Mean respirable: 0.24	n/a; n/a Sow, nursery, finisher; n/a	Poland England	n/a Winter, summer	-
[39]	Mean inhalable: 2.43 Mean respirable: 0.25 Mean inhalable: 2.76 Mean respirable: 0.26 Mean inhalable: 1.95 Mean respirable: 0.18	Sow, nursery, finisher; n/a Sow, nursery, finisher; n/a Sow, nursery, finisher; n/a	England Netherlands Denmark	Winter, summer Winter, summer Winter, summer	Swine barns had higher PM concentrations during the day than at night.
[52]	A barn with high-speed air inlet and high exhaust— TSP: 1.29 ± 0.57 Respirable: 0.26 ± 0.095 A barn with a breathing ceiling and low exhaust— TSP: 1.14 ± 0.32 Respirable: 0.15 ± 0.061	Grower-finisher; MV Grower-finisher; MV	Sweden	n/a n/a	Spraying a rapeseed oil-water mixture effectively suppressed PM generation; both automatic spraying and manual spraying were effective.
[53] <sup>d</sup>	Median TSP: 6.71 (0.76–19.09)	n/a; n/a	United Kingdom	All seasons	-
[54]	Mean TSP: 3.54 (2.15–5.60)	Finisher; n/a	Quebec, Canada	Winter, summer	-
[55]	TSP: 0.15 ± 0.04 Respirable: 0.12 ± 0.13 TSP: 0.23 ± 0.12 Respirable: 0.08 ± 0.05 TSP: 0.34 ± 0.13 Respirable: 0.13 ± 0.15 TSP: 0.28 ± 0.28 Respirable: 0.15 ± 0.18 TSP: 0.21 ± 0.07 Respirable: 0.24 ± 0.46	Breeding; NV Farrowing; NV Nursery; NV Grower; NV Finisher; NV	Taiwan	Summer	Finisher barns had the highest respirable PM concentration levels.
[56]	1st feeding period (6–7 a.m.)— TSP: 20.47 ± 8.23 2nd feeding period (6–7 p.m.)— TSP: 20.47 ± 8.14 Non-feeding periods— TSP: 2.32 ± 0.45	Grower; MV	Taiwan	n/a n/a n/a	Feeding caused a surge in PM concentrations.
[57]	Inhalable: 2.13 ± 0.52 Respirable: 0.11 ± 0.02 Inhalable: 2.19 ± 0.61 Respirable: 0.10 ± 0.02	Finisher; MV Finisher; NV	Kansas	All seasons All seasons	PM concentrations were significantly affected by outdoor air temperature and pig weight; no significant difference in PM concentrations was noted between mechanical and natural ventilation barns. The two barns were under the same roof and had the same stock density. Higher ventilation rates were noted for the naturally ventilated barn during the monitoring period.
[58]	PM <sub>10</sub> : 0.46 (0.17–0.91) PM <sub>2.5</sub> : 0.12 (0.08–0.13) PM <sub>10</sub> : 0.17 (0.12–0.25) PM <sub>2.5</sub> : 0.06 (0.05–0.07)	Finisher; MV Finisher; NV	Germany	Fall, winter, spring Fall, winter, spring	
[59] <sup>d</sup>	Median TSP: 3.95 (1.11–13.75) Median TSP: 5.00 (BDL–76.7)	n/a; n/a n/a; n/a	Denmark Germany	n/a n/a	-

Table 2. Cont.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[60]	TSP: 6.86 ± 1.30 PM <sub>10</sub> : 1.63 ± 0.16 Inhalable: 4.56 ± 2.74 Respirable: 0.44 ± 0.15 TSP: 0.42 ± 0.26 PM <sub>10</sub> : 0.24 ± 0.14 Inhalable: 0.64 ± 0.07 Respirable: 0.04 ± 0.02 Control room— TSP: 5.02 ± 0.03 Low ventilation— TSP: 4.56 ± 0.02	Finisher; MV	Minnesota	Winter  Summer	-
[61]	Middle rate ventilation— TSP: 4.05 ± 0.02 High ventilation— TSP: 2.86 ± 0.02 Nighttime— TSP: 4.23 ± 0.02 Daytime— TSP: 7.14 ± 0.04 Air cleaning with dedusters— TSP: 3.82 ± 0.02 Oil sprinkling— TSP: 0.82 ± 0.01	Finisher; MV	Illinois	n/a	A significant diurnal variation in TSP concentrations was noted; oil sprinkling was effective in reducing in-barn dust levels (by 70%).
[62]	Control— Mean respirable: 1.33 Sprinkling soybean oil— Mean respirable: 0.69 Sprinkling canola oil— Mean respirable: 0.60 (1) Control— TSP: 1.02 ± 0.21 (2) Conventional scrapper— TSP: 1.25 ± 0.31 (3) V-shaped scrapper— TSP: 1.12 ± 0.25 (4) Daily V-shape scrapper— TSP: 0.95 ± 0.26 (5) Van Kempen belt— TSP: 1.16 ± 0.24 (6) Cemagref net— TSP: 1.03 ± 0.22	Finisher; n/a Finisher; n/a Finisher; n/a	Iowa	Winter, Spring Winter, Spring Winter, Spring	-
[63]	8:00 to 9:00 am— Mean TSP: 1.04 2:00 to 3:00 pm— Mean TSP: 2.53 8:00 to 9:00 pm— Mean TSP: 1.83 Control (On the first day)— Mean TSP: 1.576 (1.571–1.580) Mean PM <sub>10</sub> : 0.848 (0.752–1.010) Mean PM <sub>2.5</sub> : 0.294 (0.189–0.439)	Grower–finisher; n/a	Quebec, Canada	Summer	Data were acquired from experimental rooms and might not represent the real-world situation.
[64]	Control (On the second day)— Mean TSP: 1.446 (1.431–1.461) Mean PM <sub>10</sub> : 0.911 (0.855–1.005) Mean PM <sub>2.5</sub> : 0.262 (0.136–0.385) Oil-atomization intervention (On the first day)— Mean TSP: 0.627 (0.579–0.676) Mean PM <sub>10</sub> : 0.256 (0.216–0.299) Mean PM <sub>2.5</sub> : 0.085 (0.01–0.161) Oil-atomization intervention (On the second day)— Mean TSP: 0.259 (0.250–0.269) Mean PM <sub>10</sub> : 0.150 (0.066–0.266) Mean PM <sub>2.5</sub> : 0.047 (0.018–0.126)	Grower–finisher; MV	South Korea	Winter	The level of aerial environment risk factors in the building was the highest at 2:00 to 3:00 pm, followed by 8:00 to 9:00 pm and 8:00 to 9:00 am.
[65]	Control (On the second day)— Mean TSP: 1.446 (1.431–1.461) Mean PM <sub>10</sub> : 0.911 (0.855–1.005) Mean PM <sub>2.5</sub> : 0.262 (0.136–0.385) Oil-atomization intervention (On the first day)— Mean TSP: 0.627 (0.579–0.676) Mean PM <sub>10</sub> : 0.256 (0.216–0.299) Mean PM <sub>2.5</sub> : 0.085 (0.01–0.161) Oil-atomization intervention (On the second day)— Mean TSP: 0.259 (0.250–0.269) Mean PM <sub>10</sub> : 0.150 (0.066–0.266) Mean PM <sub>2.5</sub> : 0.047 (0.018–0.126)	Finisher; MV	Mid-Atlantic region, USA	Winter	-

Table 2. Cont.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[66]	Mean TSP: 4.70 (1.58–17.00) Mean TSP: 4.24 (0.00–17.75) Mean TSP: 2.20 (0.00–15.25) (1) With water spray— Mean TSP: 1.98 (2) Without water spray— Mean TSP: 7.94 Mean TSP: 2.18 (0.00–8.00) (1) With water spray— Mean TSP: 3.98 (2) Without water spray— Mean TSP: 3.73 Control—	Gestation; MV	China	Winter (Jan) Spring (Mar)  Summer (Jul)  Summer (Aug)	Feeding resulted in high TSP concentrations.
[67]	TSP: 1.143 ± 0.619 Sprinkling soybean oil— TSP: 0.375 ± 0.185	Finisher; MV	US Midwest	Winter	PM concentrations in swine barns were correlated with animal activity.
[40]	PM <sub>10</sub> : 0.545 ± 0.240 PM <sub>10</sub> : 0.267 ± 0.179 PM <sub>10</sub> : 0.158 ± 0.102	Gestation; MV Farrowing; MV Finisher; MV	US Midwest	All seasons All seasons All seasons	-
[68] <sup>d</sup>	Geomean Inhalable: 2.6 (1.6–5.4); 1.6 (GSD)	n/a; n/a	The Netherlands	All seasons	-
[69]	TSP: 3.18 ± 1.46 Respirable: 0.92 ± 0.81 TSP: 2.93 ± 0.81 Respirable: 1.87 ± 0.62	n/a; n/a n/a; n/a	South Korea	Summer Winter	A significant difference in respirable PM levels was noted between summer and winter.
[70]	Mean PM <sub>10</sub> : 0.31 Mean PM <sub>10</sub> : 0.11–0.40  Mean PM <sub>10</sub> : 0.47  Mean PM <sub>10</sub> : 0.73	Sow; MV Nursery; MV Grower–finisher; MV Grower–finisher; MV	Italy Italy Italy Germany	All seasons All seasons All seasons All seasons	-
[71] <sup>d</sup>	Median inhalable: 4.69 (0.25–7.6); 2.3 (SD) Median respirable: 0.19 (0.03–0.63); 0.19 (SD) Median inhalable: 2.31 (1.9–5.0); 1.16 (SD) Median respirable: 0.17 (0.01–0.3); 0.09 (SD) Median inhalable: 1.49 (0.29–4.4); 1.51 (SD) Median respirable: 0.09 (0.01–3.4); 0.95 (SD) Median inhalable: 1.1 (0.25–3.5); 0.79 (SD) Median respirable: 0.06 (0.01–0.31); 0.11 (SD) Median inhalable: 2.99 (1.1–5.6); 1.49 (SD) Median respirable: 0.19 (0.03–0.63); 0.24 (SD)	Nursery; NV, MV  Finisher; MV  Farrowing; NV  Sow; NV  General farm; MV	Ireland	Spring, summer  Spring, summer  Spring, summer  Spring, summer  Spring, summer	-
[72]	Deep-pit manure storage— Mean TSP: 0.83 Mean respirable: 0.24 Deep-pit manure storage— Mean TSP: 1.52 Mean respirable: 0.51 Manure removal by scrapers— Mean TSP: 1.67 Mean respirable: 0.48 Manure removal by scrapers— Mean TSP: 2.42 Mean respirable: 0.83 Deep-litter bed system— Mean TSP: 2.94 Mean respirable: 1.14	n/a; NV n/a; MV n/a; NV n/a; MV n/a; NV	South Korea	Spring, Fall Spring, Fall Spring, Fall Spring, Fall	-

**Table 2.** *Cont.*

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[73]	PM <sub>10</sub> : 0.316–0.624	Finisher; MV	Italy	April	The highest PM <sub>10</sub> concentration occurred during feeding.
[74]	Mean TSP: 0.24–1.68 Mean TSP: 0.85–3.81 Control— Geomean TSP: 1.03	Wean-to-finish; MV	Illinois	Summer Winter	-
[75]	V-shaped system with the manure scraped daily— Geomean TSP: 0.95 V-shaped scraper with V-shaped concrete gutter— Geomean TSP: 1.12 A net underneath the floor for urine-feces separation— Geomean TSP: 1.03 Conventional flat scraper and stainless gutter— Geomean TSP: 1.25 Feces stayed on an inclined stable rubber belt— Geomean TSP: 1.16 Conventional—	Grower-finisher (lab chambers); MV	Quebec, Canada	Winter, Spring, Summer	Data were acquired from experimental rooms and might not represent the real-world situation.
[76]	TSP: 1.77 ± 0.72 (0.62–2.83) Sawdust bedding— TSP: 1.24 ± 0.94 (0.49–2.49) Source separation system— Mean TSP: 1.02–1.59 Geomean Inhalable: 0.83 (0.50–2.28); 2.34 (GSD)	Finisher; MV	Quebec, Canada	Winter	-
[77] <sup>d</sup>	Geomean Inhalable: 2.53 (1.37–4.11); 1.56 (GSD) Geomean Inhalable: 3.76 (2.19–7.20); 1.91 (GSD) Geomean Inhalable: 1.4; 0.0059 (GSD)	Gestation, farrowing; n/a	US Midwest	Summer Spring Winter	-
[78] <sup>d</sup>	Geomean Inhalable: 1.91; 0.0021 (GSD) TSP: 0.455 ± 0.205 PM <sub>10</sub> : 0.0997 ± 0.0705 PM <sub>7</sub> : 0.0726 ± 0.0269 PM <sub>2.5</sub> : 0.0207 ± 0.0273 PM <sub>1</sub> : 0.0160 ± 0.0163	Grower-finisher, NV (hoop barns) Grower-finisher; MV	Iowa	All seasons	-
[79]	TSP: 0.204 ± 0.169 PM <sub>10</sub> : 0.0335 ± 0.0306 PM <sub>7</sub> : 0.0353 ± 0.0294 PM <sub>2.5</sub> : 0.0112 ± 0.0146 PM <sub>1</sub> : 0.0107 ± 0.0115 TSP: 0.607 ± 0.304 PM <sub>10</sub> : 0.122 ± 0.0946 PM <sub>7</sub> : 0.104 ± 0.0570 PM <sub>2.5</sub> : 0.0179 ± 0.0172 PM <sub>1</sub> : 0.0147 ± 0.0196 TSP: 0.462 ± 0.200 PM <sub>10</sub> : 0.0922 ± 0.0474 PM <sub>7</sub> : 0.0744 ± 0.0235 PM <sub>2.5</sub> : 0.0149 ± 0.0100 PM <sub>1</sub> : 0.0157 ± 0.0123	Nursery; MV	South Korea	Spring Summer Fall Winter	A negative correlation was found between temperature and PM.
[80]	Mean PM <sub>10</sub> : 1.44 Mean PM <sub>10</sub> : 1.27 Mean PM <sub>10</sub> : 0.39	Nursery; MV Grower-finisher; MV Farrowing; MV	Netherlands	Winter Winter Winter	-

Table 2. Cont.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[81]	Mean PM <sub>10</sub> : 0.29	Gestation; MV	China	Summer	-
	Mean PM <sub>2.5</sub> : 0.21			Summer	
	Mean PM <sub>1</sub> : 0.25	Farrowing; MV			
	Mean PM <sub>10</sub> : 0.46	Nursery; MV			
[82]	Mean PM <sub>2.5</sub> : 0.45	Grower–finisher; MV	Illinois	Winter, Spring, Summer	-
	Mean PM <sub>1</sub> : 0.15			Summer	
	Mean PM <sub>10</sub> : 0.83	Finisher; MV		Summer	
	Mean PM <sub>2.5</sub> : 0.29				
[83]	Mean PM <sub>1</sub> : 0.35	Grower–finisher; MV	Saskatchewan, Canada	Winter	-
	Mean PM <sub>10</sub> : 0.98				
[84]	Mean PM <sub>2.5</sub> : 0.22	n/a; MV	Italy	Summer	-
	Mean PM <sub>1</sub> : 0.36				
[85]	PM <sub>10</sub> : 0.014–0.125	Gestation; MV	Illinois	All seasons	Seasons had no significant effect on PM <sub>10</sub> and a significant but weak effect on PM <sub>2.5</sub> inorganic compositions.
	Control—			All seasons	
	Mean TSP: 1.77	All seasons			
	Oil sprinkling—	All seasons			
[86]	Mean TSP: 0.25	Breeding; n/a	Poland	n/a	-
	Mean PM <sub>10</sub> : 0.17			Spring, summer	
	PM <sub>10</sub> : 0.421 ± 0.265	Summer			
	PM <sub>2.5</sub> : 0.156 ± 0.096				
[87]	PM <sub>10</sub> : 0.213 ± 0.114	Sows; MV (chimney)	Denmark	Spring, summer	-
	PM <sub>2.5</sub> : 0.083 ± 0.052			Summer	
	PM <sub>10</sub> : 0.354 ± 0.275	Summer			
	PM <sub>2.5</sub> : 0.075 ± 0.035				
[88]	PM <sub>10</sub> : 0.390 ± 0.303	Finisher; MV	Belgium	Summer	-
	PM <sub>2.5</sub> : 0.177 ± 0.203			Summer	
	Respirable: 0.94 ± 0.93 (0.01–4.69)				
	Inhalable: 2.96 ± 2.69 (0.43–11.8)				
[89]	TSP: 0.8 ± 0.2	n/a; MV	Denmark	Summer	-
	PM <sub>10</sub> : 0.617 ± 0.433 (0.035–1.487)				
[90]	PM <sub>2.5</sub> : 0.033 ± 0.019 (0.006–0.071)	Finisher; MV	Belgium	Summer to winter	Indoor PM <sub>1</sub> , PM <sub>2.5</sub> , and PM <sub>10</sub> concentrations were significantly correlated.
	PM <sub>1</sub> : 0.011 ± 0.007 (0.002–0.03)				
	Mean inhalable: 4.7				
	Geomean inhalable: 3.4				
[91]	Mean inhalable: 5.9	n/a; n/a	Portugal	Winter	-
	Geomean inhalable: 4.8				
	PM <sub>10</sub> : 0.719 ± 0.301				
	PM <sub>2.5</sub> : 0.0389 ± 0.0171				
[92]	PM <sub>1</sub> : 0.015 ± 0.0049	Finisher; MV	Belgium	All seasons	The study compared different pen cleaning techniques and housing systems.
	Mean PM <sub>10</sub> : 2.212				
	Mean PM <sub>5</sub> : 0.439				
	Mean PM <sub>2.5</sub> : 0.046				
[93]	Mean PM <sub>1</sub> : 0.013	Farrowing; MV	Iowa	Winter	-
	Mean PM <sub>0.5</sub> : 0.008				
	Wet protocol—				
	PM <sub>10</sub> : 2.146 ± 0.159				
[93]	PM <sub>2.5</sub> : 0.201 ± 0.012	Farrowing; MV	Iowa	Winter	-
	PM <sub>1</sub> : 0.0287 ± 0.0014				
	Dry protocol—				
	PM <sub>10</sub> : 2.215 ± 0.159				
[93]	PM <sub>2.5</sub> : 0.208 ± 0.012	Farrowing; MV	Iowa	Winter	-
	PM <sub>1</sub> : 0.0263 ± 0.0014				
	Low-ammonia-emission—				
	PM <sub>10</sub> : 2.393 ± 0.159				
[93]	PM <sub>2.5</sub> : 0.219 ± 0.012	Farrowing; MV	Iowa	Winter	-
	PM <sub>1</sub> : 0.0278 ± 0.0014				
	Conventional—				
	PM <sub>10</sub> : 1.968 ± 0.159				
[93]	PM <sub>2.5</sub> : 0.19 ± 0.012	Farrowing; MV	Iowa	Winter	-
	PM <sub>1</sub> : 0.0272 ± 0.0014				
	Respirable: 0.005–0.31				
	Inhalable: 0.17–2.09				

Table 2. Cont.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[94]	Mean PM <sub>10</sub> : 0.76	Grower–finisher, gestation; MV	The Netherlands	n/a	-
[95]	Mean PM <sub>10</sub> : 0.511 (0.159–1.402) TSP: 0.95 ± 0.66 (0.24–2.27)	Finisher; MV	The Netherlands	Fall, winter	-
[96]	TSP: 0.76 ± 0.37 (0.30–1.39) TSP: 1.70 ± 1.41 (0.14–4.59) TSP: 1.76 ± 1.10 (0.32–3.50)	Farrowing; MV Gestation; MV Nursery; MV	Illinois	All seasons	-
[97]	Mean TSP: 1.28	Finisher; MV	Germany	n/a	-
[98]	TSP: 1.40 ± 0.10 PM <sub>10</sub> : 0.78 ± 0.04	Nursery; MV	South Korea	Spring, Fall	-
[99]	Mean PM <sub>10</sub> : 0.96 Mean TSP: 0.59 Mean PM <sub>10</sub> : 0.34 Mean TSP: 2.34 Mean TSP: 3.94	Finisher; NV	China	Summer Fall Winter	A significant seasonality was noted.
[100]	Recirculated air scrubber system (Control)— Mean TSP: 0.93–1.51 Mean PM <sub>10</sub> : 0.37–0.65 Recirculated air scrubber system (Water)— Mean TSP: 0.4 Mean PM <sub>10</sub> : 0.16 Recirculated air scrubber system (Acid)— Mean TSP: 0.42–0.44 Mean PM <sub>10</sub> : 0.18–0.21 Water-oil mixture spraying system (Control)— Mean TSP: 0.80–1.33 Mean PM <sub>10</sub> : 0.26–0.39 Water-oil mixture spraying system (Small nozzle)— Mean TSP: 0.18–0.47 Mean PM <sub>10</sub> : 0.08–0.18 Water-oil mixture spraying system (Large nozzle)— Mean TSP: 0.45–0.65 Mean PM <sub>10</sub> : 0.17–0.27 Air filter modules— Mean TSP: 0.02–0.242	Finisher; MV	Germany	Winter, Spring	The main purpose of the study was to study the PM reduction performance of two mitigation technologies.
[101]	Air filter attic— Mean TSP: 0.003–0.643 Without air filtrations system— Mean TSP: 0.019–0.243 Recirculating air filtration modules— Mean TSP: 0.041–0.280 Mean TSP: 0.2 Mean PM <sub>10</sub> : 0.091	Grower–finisher; MV	Germany	All seasons	-
[102]	Mean PM <sub>4</sub> : 0.06 Mean PM <sub>2.5</sub> : 0.056 Mean PM <sub>1</sub> : 0.053	Finisher; NV	China	All seasons	-
[103]	TSP: 1.56 ± 1.06 (0.164–3.40)	Finisher; MV	Quebec, Canada	Winter	DustTrak DRX yielded much smaller TSP concentrations than gravimetric samplers.
[104]	TSP: 0.635 ± 0.1 (0.228–1.08) PM <sub>10</sub> : 0.388 ± 0.09 (0.152–0.658) PM <sub>2.5</sub> : 0.210 ± 0.09 (0.095–0.415) TSP: 0.777 ± 0.2 (0.307–2.18) PM <sub>10</sub> : 0.338 ± 0.1 (0.116–0.835) PM <sub>2.5</sub> : 0.144 ± 0.06 (0.038–0.374)	Nursery; MV Finisher; MV	China	n/a n/a	Indoor air quality in the rear of the barn was better than in other areas; TSP concentrations in the finisher barn were significantly higher than the nursery barn.
[105]	Inhalable: 0.5 ± 0.35 Respirable: 0.13 ± 0.12	n/a; n/a	South Korea	Summer	-

Table 2. Cont.

Reference	PM Concentration (mg m <sup>-3</sup> ) <sup>b</sup>	Barn and Ventilation Type <sup>c</sup>	Location	Season	Major Findings or Notes
[106]	Mean TSP: 1.14–3.20	Grower-finisher; MV	China	Spring	TSP concentrations were significantly greater in winter than summer.
	Mean PM <sub>10</sub> : 0.24–1.01				
	Mean PM <sub>2.5</sub> : 0.07–0.12				
	Mean TSP: 0.34–0.48			Summer	
	Mean PM <sub>10</sub> : 0.15–0.21				
	Mean PM <sub>2.5</sub> : 0.06–0.10				
	Mean TSP: 1.29–1.81			Fall	
	Mean PM <sub>10</sub> : 0.51–0.69				
	Mean PM <sub>2.5</sub> : 0.09–0.11				
	Mean TSP: 2.22–4.96			Winter	
Mean PM <sub>10</sub> : 0.71–0.88					
Mean PM <sub>2.5</sub> : 0.09–0.20					

<sup>a</sup> References are sorted chronologically in all tables unless otherwise noted. <sup>b</sup> GSD—geometric standard deviation; SD—standard deviation; BDL—below the detection limit. The default format is “mean ± SD” or range. A range of mean values is given when mean concentration data are available from multiple barns. A range of mean or median values are occasionally given when they are available from multiple barns or farms. <sup>c</sup> MV—mechanical ventilation; NV—natural ventilation. <sup>d</sup> Personal exposure samples were collected for PM concentration measurement.

PM mass concentrations in swine barns are affected by many factors. A good understanding of these factors is essential for the development of cost-effective PM mitigation strategies. Ref. [37] reviewed early publications and identified seven factors affecting in-barn PM concentrations. These include: (1) Outside temperature—PM concentrations decrease as the outside temperature goes up. As a ventilation rate increases with the outside temperature, PM in swine barns is diluted by an increased volume of fresh air; (2) Ventilation system—Natural ventilation systems are generally associated with higher in-barn PM concentrations due to their lower ventilation rates than those of mechanical ventilation systems; (3) Air velocity in a barn—An increased air velocity may enhance the suspension and re-suspension of particles, but meanwhile an increased amount of PM may be removed due to enhanced inertial impaction of PM on room surfaces and other objects; (4) Humidity—PM concentrations decrease with relative humidity (RH). The sorption of water enlarges PM size, thereby suppressing PM suspension and re-suspension. However, the effect of moisture becomes significant only under very high RH conditions, e.g., RH ≥ 85%; (5) Animal activity—Increased animal activity leads to elevated PM concentrations. Animal activity is in turn affected by indoor temperature, feeding method, feed type, light, and human activity inside swine barns; (6) Quantity of feed per animal—Feed is a major source of PM. An excessive amount of feed may lead to high PM concentrations; and (7) Barn cleanliness—Dusty floors and wall surfaces are sources of PM and also raise the chance of PM re-suspension. Most of these generalizations still stand today. However, significant changes have occurred to pork production on aspects such as genetics, nutrition, and environmental management. New findings have been reported, including contradictory ones. For example, Gustafsson (1999) found that increasing ventilation rates had a limited effect on in-barn PM concentrations [52].

Upon the analysis of relevant publications since 1990, the following updates are made:

- While North America and Europe lead the effort of PM concentration measurement, a growing interest has been seen in Asian countries (with 12 out of 66 reports from South Korea, China, and Taiwan since 1990), especially in recent years (Table 2). TSP (or total particles) was most frequently measured (Table 3), followed by PM<sub>10</sub> and respirable PM. Since 2010, a rapidly increasing number of reports has been available regarding PM<sub>10</sub> and PM<sub>2.5</sub> concentrations. Ten studies collected personal samples for PM exposure assessment, in which farm workers were asked to wear personal PM samplers during their work shifts. A few studies also used personal samplers but mounted the samplers at a fixed location—a setup known as fixed samplers. In principle, the PM concentrations derived from personal exposure samples cannot be directly compared with those from fixed samples.

**Table 3.** Numbers of measurement efforts for different PM size fractions.

Years	TSP	PM <sub>10</sub>	PM <sub>2.5</sub>	Inhalable	Respirable	Others <sup>a</sup>
1990–1999	7	0	0	2	6	0
2000–2009	16	5	1	6	7	0
2010–Present	12	17	9	4	3	6
Total	35	22	10	12	16	6

<sup>a</sup> Other size fractions include PM<sub>1</sub>, PM<sub>5</sub>, and PM<sub>7</sub>.

- PM concentrations varied greatly in the literature. With no PM mitigation measures implemented, the reported mean TSP concentrations (including + mean, geometric mean (geomean), and median) ranged from 0.15 [55] to >20 mg m<sup>-3</sup> [56]. Sampling or averaging time is highly influential on measurement results. Many studies collected 24 h filter samples or ran PM monitors for 24 h to address diurnal variability and determine daily average PM concentrations. However, a shorter or longer sampling time was occasionally seen. As for the aforementioned TSP concentration range, the lowest value was a daily average [55] while the highest one was an hourly average [56]. Strictly speaking, they cannot be directly compared. This dilemma is further complicated by the lack of sampling or averaging time information in part of the publications. The reported mean concentrations ranged from 0.64 [60] to 5.9 mg m<sup>-3</sup> [89] for inhalable PM, from 0.04 [60] to 1.87 mg m<sup>-3</sup> [69] for respirable PM, from 0.034 [79] to 1.63 mg m<sup>-3</sup> [60] for PM<sub>10</sub>, and from 0.015 [79] to 0.45 mg m<sup>-3</sup> [81] for PM<sub>2.5</sub>. A gradual decrease in TSP concentrations over the past 30 years is noted. The reasons are uncertain but likely related to the improved environmental management of swine barns.
- PM concentrations showed significant seasonality [57,60,66,74,79,89,96,99,106,107], with the highest concentrations generally occurring in winter and the lowest concentrations in summer. Such seasonality is believed to be related to ventilation rates [57,96]. Most modern swine barns use mechanical ventilation or mechanically assisted ventilation (with sidewall curtains dropped in hot weather conditions). In either case, a barn's ventilation rate is maximal in summer—to cool down the barn's temperature—and minimal in winter—to keep the barn warm. An elevated ventilation rate would enhance the dilution of PM by fresh air, thus decreasing in-barn PM concentrations. For a similar reason (i.e., changes in outdoor temperatures), PM concentrations also exhibit significant diurnal variability [39,61]. However, such variability could be caused by a diurnal change in animal activity other than outdoor temperatures.
- No agreement has yet been reached regarding the effect of ventilation systems. Although several studies [37,72] reported that naturally ventilated barns had higher PM concentrations than mechanically ventilated barns, Ref. [57] found no significant difference in inhalable or respirable PM concentrations between naturally and mechanically ventilated finisher barns in Kansas. Another counter finding was reported by Ref. [58] that PM<sub>2.5</sub> and PM<sub>10</sub> concentrations were lower in a naturally ventilated than a mechanically ventilated finisher barn in Germany. A comparison of different ventilation systems is challenging for several reasons. First, many previous publications provide no detailed description of a barn's ventilation system. Even for mechanical ventilation systems, there are various configurations such as tunnel ventilation, cross-flow ventilation, and chimney ventilation. Different configurations could result in different airflow patterns and dilution levels (by fresh air) at the monitoring point. Classifying ventilation systems into natural, mechanical, and mixed types could oversimplify a barn's ventilation conditions. Secondly, it is difficult to find swine barns only differing in ventilation systems. Other environmental and operating parameters (e.g., local climatic conditions and feed types) could substantially affect PM concentrations in swine barns. Thirdly, for naturally ventilation barns, their ventilation rates are highly variable and can be affected by wind speeds and directions, barn locations (e.g., valley or hilltop) and orientations, solar radiation, ground objects, etc.

- PM concentrations increase with animal activity [37,64,67]. Ref. [73] found a significant correlation ( $p < 0.001$ ) between animal activity and PM<sub>10</sub> concentrations inside a finisher barn in Italy. Animal activity was determined through image analysis. Refs. [39,61] observed higher PM concentrations during the day than the night and ascribed this to increased animal activity during the day. Ref. [64] further compared TSP concentrations in three time slots: morning (8–9 a.m.), afternoon (2–3 p.m.), and evening (8–9 p.m.), and found that the overall highest concentrations occurred in the afternoon when pigs were usually most active. Feeding is one of the parameters that regulate animal activity. Elevated in-barn PM concentrations were detected during feeding periods [56,66,73,104], regardless of the feeding methods (hand or automated) [39,108]. However, feeding-resulted high PM concentrations could be related to not only increased animal activity but also the suspension of feed particles by feed delivery systems.
- No agreement has yet been reached regarding the effect of barn types. Refs. [40,85] measured PM<sub>10</sub> concentrations in various barns in the U.S. Midwest and found that the overall lowest concentrations occurred in gestation barns; whereas [81] reported that gestation barns had higher PM<sub>10</sub> concentrations than other barn types in northern China. Another study in China compared PM concentrations in a finisher versus a nursery barn, with higher TSP concentrations detected in the finisher barn [104]. A similar observation was made by Ref. [96] from a field study in Illinois. However, this is contradictory to the findings of Ref. [55] who reported significantly higher TSP concentrations in nursery than finisher barns in Taiwan. Several studies found lower overall PM concentrations in farrowing barns than finisher or nursery barns [55,80,81,85,96]; while higher PM<sub>10</sub> concentrations in farrowing than finisher barns were reported by Ref. [40]. A conclusive comparison among different barn types may require extended monitoring periods and/or additional farm sites to address the uncertainties created by temporal and farm-to-farm variations.
- PM concentrations show a heterogeneous spatial distribution inside swine barns [61,64,107,109–113]. Ref. [109] collected 24 h TSP samples from 16 points inside a partially slatted grower–finisher barn. They found that the TSP mass concentrations ranged from 1.6 to 2.74 mg m<sup>-3</sup>, with the highest concentration at the floor level within non-slatted areas and the lowest concentration at pen divider height over slatted areas. Ref. [61] measured the spatial distribution of TSP concentrations in a partially slatted grower–finisher barn with cross-flow ventilation. The distribution was found to be affected by ventilation rates, diurnal changes in outdoor weather, and dust mitigation efforts such as oil sprinkling. TSP concentrations at the exhaust were lower than the average indoor concentrations, under all tested ventilation rates and outdoor weather conditions. Ref. [112] conducted a similar investigation in a tunnel-ventilated grower–finisher barn. They reported that in December (winter) while air velocities increased longitudinally from the barn’s end-wall intake to tunnel fans (due to air intake from ceiling inlets), TSP concentrations gradually decreased at the heights of 1.6 m and 0.8 m above the floor. However, in June (summer) both TSP concentrations and air velocities increased longitudinally. Ref. [107] mapped inhalable PM mass concentrations inside a tunnel-ventilated gestation barn. In winter, the highest concentrations occurred near the center of the barn; while in spring and summer, the highest concentrations were found close to the tunnel fans. Given the non-uniform spatial distribution of PM, it is important to find sampling/monitoring points representative of average in-barn PM concentrations. However, in reality, it is difficult to do so because PM spatial distribution can be affected by various factors. Caution therefore must be taken when comparing PM concentration data from different studies.
- There has been no consistent conclusion regarding the effect of indoor air humidity. Ref. [58] found that both PM<sub>10</sub> and PM<sub>2.5</sub> concentrations decreased with air humidity levels in two finisher barns in Germany. In Korea, Ref. [64] found that TSP concentrations in a grower–finisher barn significantly decreased with air humidity levels

( $R = -0.52$ ,  $p < 0.05$ ). Ref. [73] measured  $PM_{10}$  concentrations in a chimney-ventilated finisher barn in Italy. The measured  $PM_{10}$  concentrations exhibited a significant but negative correlation with air humidity levels ( $R = -0.929$  and  $p < 0.0001$ ). However, counter findings were also reported. Ref. [48] in Kansas found that in a nursery barn both TSP and respirable PM concentrations increased with air humidity levels and that no significant correlation existed between PM concentrations and air humidity levels. A similar observation was made for  $PM_{10}$  from a barn study in Czech [114]. In the same study,  $PM_{2.5}$  concentrations significantly increased with air humidity levels ( $p < 0.001$ ). As no raw data are available, it remains uncertain whether the air humidity levels in [48,114] exceeded the threshold of  $RH \geq 85\%$  [37] or  $70\%$  [39], above which a significant effect of air humidity could be anticipated.

PM concentrations inside swine barns can additionally be affected by animal age [105,115] and stock density [20]. As few relevant publications are available, no review is provided here. Other influential factors include feed diet and feeders, housekeeping, and mitigation technologies implemented. Detailed information about PM mitigation is available in Section 5.

### 3.1.2. PM Size Distribution

Size is one of the most important physical characteristics of PM. It significantly impacts the PM's health and environmental effects. The smaller a particle is, the more deeply it may penetrate the respiratory tract of humans and animals. Smaller particles can travel a longer distance in the air than larger ones [23]. Particle size is also a key parameter for the design and operation of PM mitigation systems. The PM reduction efficiency of these systems is usually size-dependent and increases with particle size [116]. Thus, from both environmental health and PM mitigation standpoints, the field measurement of PM size distribution in swine barns is of great importance.

For a spherical particle, the particle size is characterized by its geometric diameter. However, most particles collected in polluted environments (including swine barns) are non-spherical and irregular in shape. To apply the concept of particle size to non-spherical particles, several equivalent diameters were defined. Two commonly used equivalent diameters are (1) equivalent volume diameter, defined as the diameter of a sphere with the same volume as the real particle, and (2) aerodynamic diameter, defined as the diameter of a unit density ( $\rho = 1000 \text{ kg m}^{-3}$ ) sphere with the same aerodynamic behaviors (e.g., settling velocity) as the real particle [24,117].

Particles are heterogeneous in size (polydispersed). A particle size distribution (PSD) can be derived by classifying the particles into multiple size channels. The  $y$ -axis of a derived PSD graph represents the quantity or occurrence frequency of particles of a certain size and it can be particle number (count), surface area, volume, or mass, etc. The  $x$ -axis of the graph represents the size of particles (e.g., geometric diameter, or aerodynamic diameter). The central location of a PSD profile can be described by arithmetic mean, geometric mean, median, or mode diameters [24]. Among them, the mass median diameter (MMD)—defined as the diameter that splits the total PM mass by half—is most frequently used, primarily for two reasons. First, PM regulations are predominately mass-based. Secondly, most mass PSD profiles measured in swine barns are approximately lognormal [57,118,119]. A measured PSD profile can therefore be approximated by a lognormal distribution model with MMD representing the central location represented and GSD characterizing the spread (width) of the PSD (Equation (1)):

$$M(d) = \frac{1}{2} \operatorname{erfc} \left( -\frac{\log d - \log MMD}{\sqrt{2} \log GSD} \right) \quad (1)$$

where,  $M(d)$  is a cumulative particle mass fraction at the diameter of  $d$ , as predicted by the lognormal distribution equation, and  $\operatorname{erfc}()$  is the complementary error function.

A measured PSD may consist of tens of size channels. The use of MMD and GSD, thus greatly simplifies data reporting as well as discussions on size-dependent PM properties or

impacts [24]. It is noteworthy that PSDs in livestock barns can be approximated by other statistical models [119,120]. However, the lognormal distribution model has been most commonly used.

To our knowledge, the first PSD measurement in swine barns was reported by Dr. Bundy at Iowa State University [29]. An optical particle counter (OPC; Royco Model 215) was used in a farrowing barn to measure particle numbers in six size channels: 0.5, 1, 2, 3, 5, and 10  $\mu\text{m}$ . The highest particle number concentration was found for particles of 0.5  $\mu\text{m}$  and particle numbers decreased with increased diameters. Ref. [36] measured the size of particles in 21 swine barns in Iowa, with a number median diameter (NMD; the diameter splitting the total PM count by half) of 2.2  $\mu\text{m}$  derived from an optical microscope and an MMD of 9.6  $\mu\text{m}$  determined by a cascade impactor. Ref. [121] collected PM samples from 11 finisher barns, measured the PSD of the collected samples using a Coulter counter, and reported an MMD of 18.5  $\mu\text{m}$  and a GSD of 2.54. Since 1990, additional efforts have been devoted to PSD measurement in swine barns, as summarized in Table 4.

**Table 4.** PM size distribution in swine barns—A summary of studies since 1990.

Reference	Size Measurement Results <sup>a</sup>	Barn and Ventilation Type	Location	Major Findings or Notes
[109]	Mass mean diameter: 14 $\mu\text{m}$	Grower–finisher; MV	Saskatchewan, Canada	-
[122]	NMD: 1.5 $\mu\text{m}$	Grower–finisher; MV	Saskatchewan, Canada	-
[123]	Before oil sprinkling— Number mode diameter: >5.0 $\mu\text{m}$ After oil sprinkling— Number mode diameter: 0.3–0.5 $\mu\text{m}$	Grower–finisher; MV	Saskatchewan, Canada	After oil sprinkling, the particle size distribution inside the swine barn was similar to that in an office.
[48]	MMD: 13 $\mu\text{m}$ (10–19 $\mu\text{m}$ ) GSD: 3 (2–5)	Nursery; MV	Kansas	-
[124]	MMD: 8.31 $\mu\text{m}$ GSD: 1.60	Nursery, lab chamber	Netherlands	-
[57]	Range: 14.0–22.9 $\mu\text{m}$ GMD: 17.9 $\mu\text{m}$ GSD: 2.2	n/a; NV	Kansas	79% of particles by mass were larger than 10 $\mu\text{m}$ .
[57]	Range: 12.1–21.2 $\mu\text{m}$ GMD: 18.1 $\mu\text{m}$ GSD: 2.1	n/a; MV		80% of particles by mass were larger than 10 $\mu\text{m}$ .
[125]	n/a	n/a; n/a	Germany	A bimodal distribution was noted, with one peak in the submicron range and the other in the micron range.
[126]	MMD: 11 (winter) to 14 $\mu\text{m}$ (fall)	Finisher; MV	Iowa	Respirable particles accounted for 25% of the mass of total particles.
[61]	Before oil sprinkling— NMD: 4.82 $\mu\text{m}$ GSD: 1.38 After oil sprinkling— NMD: 1.76 $\mu\text{m}$ GSD: 2.04	Finisher; MV	Illinois	-
[118]	GMD: 16.7 $\mu\text{m}$ GSD: 2.3 GMD: 15.7 $\mu\text{m}$ GSD: 2.2 GMD: 17.4 $\mu\text{m}$ GSD: 2.1	Nursery; MV Experimental finisher; MV Commercial finisher; MV	Kansas	Different barns shared a similar particle size distribution.
[127]	Total particles— NMD: 2.23 $\mu\text{m}$ Viable particles— NMD: 2.79 $\mu\text{m}$ Non-viable particles— NMD: 1.15 $\mu\text{m}$	Grower; NV	Australia	Around 95% of particles by number were smaller than 7 $\mu\text{m}$ ; ~60% of particles by number were smaller than 2.5 $\mu\text{m}$ .
[128]	GMD: 14.0–17.0 $\mu\text{m}$ GSD: 2.0–2.6	Finisher; MV	Kansas	-

Table 4. Cont.

Reference	Size Measurement Results <sup>a</sup>	Barn and Ventilation Type	Location	Major Findings or Notes
[129]	Winter— MMD: 20.33–21.87 $\mu\text{m}$ GSD: 1.46–1.52 Summer— MMD: 30.79–32.21 $\mu\text{m}$ GSD: 1.83–1.89 Horiba LA-300— MMD: 24.3–24.1 $\mu\text{m}$ GSD: 2.8–3.4 Coulter counter— MMD: 9.8–10.1 $\mu\text{m}$ GSD: 2.1–2.4	Wean-to-finish; MV	Illinois	-
[130]	Malvern Mastersizer— MMD: 12.1–13.4 $\mu\text{m}$ GSD: 2.6–3.1 DSP— MMD: 9.1–9.8 $\mu\text{m}$ GSD: 1.8 Horiba LA-300— MMD: 22.2–24.5 $\mu\text{m}$ GSD: 2.2–2.9 Coulter counter— MMD: 9.5–12.8 $\mu\text{m}$ GSD: 2.2–2.3 Malvern Mastersizer— MMD: 11.2–17.3 $\mu\text{m}$ GSD: 2.7–3.2 DSP— MMD: 8.5–12.1 $\mu\text{m}$ GSD: 1.6–1.8 Horiba LA-300— MMD: 19.2–20.7 $\mu\text{m}$ GSD: 2.7–2.9 Coulter counter— MMD: 16.1–16.5 $\mu\text{m}$ GSD: 1.7–1.8 Malvern Mastersizer— MMD: 18.4–19.6 $\mu\text{m}$ GSD: 2.3–2.4 DSP— MMD: 10.7–13.4 $\mu\text{m}$ GSD: 1.4–1.6	Farrowing; MV	Illinois	-
[131]	Mass mode diameter: >10 $\mu\text{m}$ Coulter Counter— MMD: 22.63 $\mu\text{m}$ GSD: 1.58 Horiba LA-300 (all size channels)— MMD: 32.23 $\mu\text{m}$ GSD: 1.89 Horiba LA-300 (only 3–60 $\mu\text{m}$ considered)— MMD: 27.31 $\mu\text{m}$ GSD: 1.92 Before oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$ After oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$	Gestation; MV		-
[131]	Mass mode diameter: >10 $\mu\text{m}$ Coulter Counter— MMD: 22.63 $\mu\text{m}$ GSD: 1.58 Horiba LA-300 (all size channels)— MMD: 32.23 $\mu\text{m}$ GSD: 1.89 Horiba LA-300 (only 3–60 $\mu\text{m}$ considered)— MMD: 27.31 $\mu\text{m}$ GSD: 1.92 Before oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$ After oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$	Wean-to-finish; MV	Iowa	-
[82]	Mass mode diameter: >10 $\mu\text{m}$ Coulter Counter— MMD: 22.63 $\mu\text{m}$ GSD: 1.58 Horiba LA-300 (all size channels)— MMD: 32.23 $\mu\text{m}$ GSD: 1.89 Horiba LA-300 (only 3–60 $\mu\text{m}$ considered)— MMD: 27.31 $\mu\text{m}$ GSD: 1.92 Before oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$ After oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$	Finisher; MV	Illinois	>90% of particles by mass were larger than 10 $\mu\text{m}$ .
[83]	Mass mode diameter: 9.0–10 $\mu\text{m}$ After oil sprinkling— Mass mode diameter: 9.0–10 $\mu\text{m}$	Grower–finisher; MV	Saskatchewan, Canada	-
[132]	MMD: 5.6 $\mu\text{m}$ GSD: 2.2	Finisher; MV	Iowa	Particle size distributions during load-out and power washing were measured. A bimodal distribution was noted, with a minor peak in the submicron range.
[90]	Mass mode diameter: 5.0–6.5 $\mu\text{m}$	Finisher, MV (chimney)	Belgium	

Table 4. Cont.

Reference	Size Measurement Results <sup>a</sup>	Barn and Ventilation Type	Location	Major Findings or Notes
[91]	Mass mode diameter: 2.5–5.0 $\mu\text{m}$	Complex (farrowing, gestation, nursery, finisher); n/a	Portugal	Particle size was the smallest in nursery barns.
[133]	Control— Number mode diameter: 0.3–0.5 $\mu\text{m}$ Treatment (ionization dedusting)— Number mode diameter: 0.3–0.5 $\mu\text{m}$	Nursery; n/a	Kansas	-
[134]	MMD: 10.67–10.86 $\mu\text{m}$ NMD: 0.38–0.36 $\mu\text{m}$ MMD: 9.29 $\mu\text{m}$ NMD: 0.49 $\mu\text{m}$ MMD: 10.26–10.39 $\mu\text{m}$ NMD: 0.38–0.43 $\mu\text{m}$	Sow; MV (chimney) Nursery; MV (chimney) Finisher; MV (chimney)	Netherlands	Particle size was measured for 60 min per barn; no GSD calculation was conducted.
[96]	MMD: 15.7 $\pm$ 1.0 $\mu\text{m}$ GSD: 2.32 $\pm$ 0.34 MMD: 18.0 $\pm$ 1.1 $\mu\text{m}$ GSD: 2.22 $\pm$ 0.25 MMD: 19.3 $\pm$ 2.7 $\mu\text{m}$ GSD: 2.28 $\pm$ 0.22	Wean-to-finish; MV Farrowing; MV Gestation; MV	Illinois	PM <sub>10</sub> accounted for 20–21% of total particles by mass while PM <sub>2.5</sub> accounted for 5–6%; particle size was larger in summer than winter; a bimodal distribution was occasionally seen.
[135]	MMD: 10.73–12.18 $\mu\text{m}$ GSD: >1.22	Finisher; MV	Belgium	No differences were seen between different housing systems (conventional vs. low NH <sub>3</sub> emission) and two cleaning protocols (dry vs. wet).
[136]	No size statistics presented	Nursery; MV and NV	Japan	The highest particle number occurred in the size range of 0.3–0.5 $\mu\text{m}$ .
[102]	No size statistics presented	Nursery; NV	China	>50% of particles by mass were larger than 10 $\mu\text{m}$ .

<sup>a</sup> MMD—mass median diameter; GMD—geometric mean diameter; NMD—number median diameter (50% of particles by number have a diameter smaller than NMD).

Upon the analysis of existing publications, the following generalizations are made:

- A large portion of particles by mass have diameters larger than 10  $\mu\text{m}$  in swine barns. The reported MMD values ranged from 5.6 [126] to 32.23  $\mu\text{m}$  [82]. Among twelve studies with MMD results available, nine of them reported an average MMD value greater than 10  $\mu\text{m}$ . This indicates that PM<sub>10</sub> accounts for only <50% of total PM mass concentrations in many swine barns.
- Although their mass contribution is minor, small particles are predominant in terms of particle counts (numbers). The reported NMD values ranged from 0.33 [134] to 4.82  $\mu\text{m}$  [61], smaller than their corresponding MMD values. Particle counts tend to decrease with size, with the highest counts observed for submicron particles [29,90,123,127,134].
- Particle size tends to be smaller overall in winter than in warm seasons [96,126,129]. This is likely because swine barns run at a minimum ventilation rate in winter [96]. The minimum ventilation rate results in calm air in barns, which encourages the gravitational settling of large particles. Conversely, an elevated ventilation rate results in a high air velocity, thereby enhancing the suspension and resuspension of large particles [96,137].
- Particle size tends to be greater in sow (e.g., farrowing and gestation) barns than other barn types. Ref. [91] surveyed four types of barns (farrowing, gestation, nursery, and finisher) and found that the smallest particle size occurred in nursery barns. A similar observation was reported by [134]. Both studies were conducted in Europe. A field campaign in Illinois revealed that particle size was smaller in wean-to-finish barns than farrowing and gestation barns [96,130]. No dedicated nursery barns were visited in the campaign.
- The mass PSDs in swine barns occasionally show a bimodal distribution, i.e., with a major peak in the large size range (5–20  $\mu\text{m}$ ) and a minor peak in the submicron range (<0.5  $\mu\text{m}$ ) [90,96,125]. The minor peak was considered to originate from the atmospheric PM that entered a barn through the barn's air inlet [90,123]. These ultrafine

particles (with aerodynamic diameters  $<0.5 \mu\text{m}$ ) were designated as “diminutive dust” for air quality management in swine barns [49,123,138,139].

Different PSD analyzers result in different measurement results. The measured MMD and GSD values differed with selected instruments even after a rigorous calibration and validation process [82,119,130]. This is related to the detection principles, assumptions, and experimental procedures adopted by different PSD analyzers (Refer to Section 4.2). The qualitative conclusions however remained largely the same regardless of the selected instruments.

The acquired PSDs were primarily used to estimate the mass fractions or concentrations of PM within a certain size range (e.g.,  $\text{PM}_{10}$ ,  $\text{PM}_{2.5}$ , respirable, and inhalable). Further utilization of the PSD data may include the development and validation of PM mitigation technologies, respiratory health–exposure modeling for pigs and humans [140], and source apportionment of PM in swine barns. PSDs have been used for source apportionment of atmospheric PM through receptor modeling [141,142]. As large and small particles in swine barns originate from different sources [36,121], the same methodology could apply to PM in swine barns as well.

### 3.1.3. Morphology and Density

PM size fractions (e.g.,  $\text{PM}_{10}$  and respirable PM) are defined and regulated based on the aerodynamic diameter of particles. The aerodynamic diameter also dictates the removal of a particle in aerodynamic dedusters such as cyclones and impactors [116]. For a nonspherical particle in the environment (including swine barns), its aerodynamic diameter ( $d_a$ ; m) is related to not only the equivalent volume diameter ( $d_e$ ; m) but also the morphology and density of the particle (Equation (2)) [24].

$$d_a = d_e \left( \frac{C_{ce}\rho_p}{C_{ca}\rho_0\chi} \right)^2 \quad (2)$$

where  $C_{ce}$  is the slip corrector factor for  $d_e$  (dimensionless),  $C_{ca}$  is the slip corrector factor for  $d_a$  (dimensionless),  $\rho_p$  is the particle density ( $\text{kg m}^{-3}$ ),  $\rho_0$  is the unit density ( $1000 \text{ kg m}^{-3}$ ), and  $\chi$  is the shape factor (dimensionless), a measure of PM morphology.

Morphology and density also affect the optical properties of particles and accordingly the response of optical PM monitors [143], as well as the performance of non-aerodynamic dedusters such as baghouses and wet scrubbers [116].

### Morphology

Only a few studies examined the morphology of swine barn PM. Ref. [121] measured the morphology of PM from eleven finisher barns using scanning electron microscopy (SEM). Starch granules, grain meal, and skin particles were identified based on their shape and size. The unidentified particles were classified into irregular, rounded, and cylindrical particles. Ref. [80] used a high-resolution SEM, in coupling with FETEX 2.0 image analysis software, to determine the morphology of PM in four types of swine barns in the Netherlands. They found large, fattened skin particles, and layered manure particles in nursery and grower–finisher barns, and large, folded skin particles in gilt and gestation barns. Ref. [97] calculated the shape factors of PM in a swine production complex (housing sows, piglets, and grower–finishers) in Germany through optical microscope image analysis. The derived shape factors ( $1.28 \pm 0.23$ ; range: 1.07–1.60) increased with particle size. Ref. [104] analyzed the morphology of  $\text{PM}_{2.5}$  in a high-rise nursery barn and a high-rise finisher barn in China, using a field emission SEM. They reported a mixture of roughly spherical and irregularly shaped particles in the nursery barn; while in the finisher barn,  $\text{PM}_{2.5}$  was composed of strip-, rod-, and bar-shaped particles with loose, smooth surfaces. Several additional studies also used microscopes to image PM sampled from the inside of swine barns, e.g., [36,125]. However, no morphology analysis results were reported.

## Density

PM density here refers to the true density of particle material, to distinguish it from the bulk of particles. Density is a key factor affecting the aerodynamic behavior of PM [24]. It is required for conversions between equivalent volume diameters and aerodynamic diameters and between number PSDs and mass PSDs [144].

Only a few publications reported the density of PM in swine barns. All of them but one, [97], used a gas pycnometry method. The method uses settled dust as a surrogate for PM for density determination. Ref. [145] measured PM density in a simulated swine barn and reported an average PM density of  $1600 \text{ kg m}^{-3}$ . Ref. [144] characterized PM sampled from a finisher barn in Kansas and found an average density of  $1840 \text{ kg m}^{-3}$ . Ref. [129] measured the density of settled dust in a mechanically ventilated finisher barn in Illinois and reported an average density of  $1450 \text{ kg m}^{-3}$ . A slightly higher value ( $1580 \text{ kg m}^{-3}$ ) was reported by Ref. [146] in efforts to characterize the  $\text{NH}_3$  emission potency of swine barn PM. Again in Illinois, a PM monitoring campaign was conducted in nine swine barns [96,147]. The measured density values ranged from 1460 to  $2000 \text{ kg m}^{-3}$  (average:  $1640 \text{ kg m}^{-3}$ ), with the highest one ( $1750 \pm 150 \text{ kg m}^{-3}$ ) found in farrowing barns and the lowest ( $1580 \pm 100 \text{ kg m}^{-3}$ ) in gestation barns. The highest PM density in farrowing barns coincided with that farrowing barn PM had the greatest mineral contents [85]. Significant seasonality was noted, with the highest PM density occurring in summer and the lowest in winter [96]. Ref. [97] collected PM samples from the inside of a swine production complex in Germany and measured PM density over six size channels (4.0–5.0, 5.0–7.5, 7.5–10, 10–15, 15–20, and  $>20 \mu\text{m}$ ) using an indirect method. A density value of  $2025 \pm 478 \text{ kg m}^{-3}$  was reported.

Caution should be taken when interpreting the density data derived from gas pycnometry. The measurement assumes that PM and settled dust share the same density and that PM density remains constant regardless of particle size. However, compared to PM, settled dust contained more large particles due to their greater settling velocities [109]. Large and small particles in swine barns originated from different sources [121,148] and, thus, could differ in density. Another counter evidence was given by Ref. [97] who observed PM density to be size-dependent in swine barns; however, only a single data set (with no replicates) was taken in the study. Similar to pycnometry, the indirect method involves several major assumptions. As a result, the acquired size-segregated density data could carry large uncertainties.

The PM density values reported in the literature ranged from 1400 to  $2100 \text{ kg m}^{-3}$ . In comparison, the densities of starch and proteins are  $\sim 1500$  and  $\sim 1350 \text{ kg m}^{-3}$ , respectively; and the density of limestone (a common mineral additive in swine feed) is  $\sim 2700 \text{ kg m}^{-3}$ . This suggests that PM in swine barns is chemically a blend of organics and minerals. For future studies, a PM density value of  $1650 \text{ kg m}^{-3}$  is recommended when no field measurement data is available.

### 3.2. Bioaerosols

#### 3.2.1. Bacterial and Fungal Counts

Bioaerosols in swine barns are highly complex in terms of composition, size, source, and health effect. As a result, no single universal measure of bioaerosols exists. Among various measures, airborne bacterial and fungal counts have been most widely used for assessing bioaerosol contamination levels. The counts can be selective of a specific microbial species (or group) or non-selective of general bacteria or fungi, depending on measurement methodology. Depending on the methodology, the counts can be of viable, culturable, or total bacteria/fungi, with their definitions given below [149]:

- Viable bacteria/fungi refer to living bacterial/fungal cells. These microbes are of particular interest due to their continued growth and reproduction in the environment;
- Culturable bacteria/fungi are a subset of viable bacteria/fungi that can be cultivated with certain growth media under certain environmental conditions;
- Total bacteria/fungi are a collection of viable and non-viable (i.e., dead) bacteria/fungi.



Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[153]	Unit 1— Bacteria: $(4.14 \pm 1.50) \times 10^5$ Fungi: $(1.53 \pm 2.02) \times 10^5$ Unit 2— Bacteria: $(5.10 \pm 1.60) \times 10^5$ Fungi: $(1.83 \pm 0.60) \times 10^5$	Grower; MV	Ontario, Canada	n/a
[154]	Mean microbes: $2 \times 10^5$ – $6 \times 10^6$ Mean fungi: $2 \times 10^3$ – $1 \times 10^5$	Finisher; n/a	Scotland	n/a
[155]	Mean microbes: $1.1 \times 10^5$ Mean Gram-negative bacteria: $7.7 \times 10^3$	n/a, n/a	Netherlands	n/a
[156]	Nuclepore filter— Mean bacteria: $7.78 \times 10^4$ Mean fungi: $5.85 \times 10^3$ Anderson six-stage viable cascade impactor— Mean bacteria: $7.32 \times 10^4$ Mean fungi: $1.97 \times 10^3$ AGI impinge— Mean bacteria: $9.64 \times 10^4$ Mean fungi: $5.38 \times 10^3$	Farrowing, nursery-grower, finisher; n/a	US Midwest	Fall, winter
[43]	Mean fungi: 1.3–7.4 Mean mesophilic bacteria: $1.12 \times 10^3$ – $1.24 \times 10^3$ Mean Gram-negative bacteria: 10.0–36.1 Mean thermophilic <i>Actinomyces</i> : 0.4–7.3 Mean fungi: 1.7–31.2 Mean mesophilic bacteria: 569–1481 Mean Gram-negative bacteria: 13.4–35.8 Mean thermophilic <i>Actinomyces</i> : 0.2–1.4	Farrowing (3 farms); n/a  Finisher (2 farms); n/a	Poland	n/a  n/a
[157]	Control— Bacteria: $1.3 \times 10^5$ – $5.2 \times 10^5$ Recirculating fabric filter— Bacteria: $1.1 \times 10^5$ – $2.7 \times 10^5$ Control— Bacteria: $1.4 \times 10^5$ – $4.5 \times 10^5$ Recirculating electrostatic filter— Bacteria: $0.8 \times 10^5$ – $3.2 \times 10^5$	Grower; MV  Finisher; MV	British Columbia, Canada	All seasons
[51]	Mean microbes: $930.6 \times 10^3$ ( $613.7$ – $1246.7 \times 10^3$ )	n/a; n/a	Poland	n/a
[54]	Mean molds: $883 (557$ – $2.86 \times 10^3)$ Mean bacteria: $4.25 \times 10^5$ ( $1.67 \times 10^5$ – $9.30 \times 10^5$ ) Mean thermophilic <i>Actinomyces</i> : 29 (3–94)	Finisher; n/a	Quebec, Canada	Winter, summer

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[158]	Bacteria: $(4.97 \pm 4.23) \times 10^5$	Breeding; NV	Taiwan	Spring
	Gram-negative bacteria: $50 \pm 93$			
	Fungi: $(3.58 \pm 5.82) \times 10^3$			
	Bacteria: $(1.83 \pm 1.91) \times 10^5$	Farrowing; NV		
	Gram-negative bacteria: $42 \pm 37$			
	Fungi: $(3.01 \pm 1.93) \times 10^3$			
	Bacteria: $(1.03 \pm 1.16) \times 10^5$	Nursery; NV		
	Gram-negative bacteria: $44 \pm 38$			
	Fungi: $(2.30 \pm 1.34) \times 10^3$			
	Bacteria: $(1.27 \pm 1.47) \times 10^5$	Grower; NV		
Gram-negative bacteria: $75 \pm 82$				
Fungi: $(2.47 \pm 2.04) \times 10^3$				
Bacteria: $(0.76 \pm 1.04) \times 10^6$	Finisher; NV			
Gram-negative bacteria: $452 \pm 690$				
Fungi: $(2.69 \pm 2.95) \times 10^3$				
[159]	Filtration—	Grower–finisher; NV	Kansas	Winter, spring, summer
	<i>Staphylococcus</i> : $(5.6 \pm 3.4) \times 10^4$			
	Respirable <i>staphylococcus</i> : $(6.7 \pm 3.7) \times 10^3$			
	<i>Pseudomonas</i> : $(0.98 \pm 1.2) \times 10^3$			
	Respirable <i>Pseudomonas</i> : $(3.4 \pm 4.4) \times 10^2$			
	<i>Bacillus</i> : $(0.50 \pm 1.3) \times 10^4$			
	Respirable <i>Bacillus</i> : $(6.6 \pm 9.1) \times 10^2$			
	<i>Listeria</i> : $(6.3 \pm 5.4) \times 10^2$			
	Respirable <i>Listeria</i> : $(1.1 \pm 1.2) \times 10^2$			
	<i>Enterococcus</i> : $(2.7 \pm 2.3) \times 10^2$			
	Respirable <i>Enterococcus</i> : $(1.8 \pm 4.3) \times 10^1$			
	<i>Nocardia</i> : $(3.3 \pm 7.2) \times 10^2$			
	Respirable <i>Nocardia</i> : $(0.85 \pm 1.7) \times 10^2$			
	<i>Lactobacillus</i> : $(4.3 \pm 3.0) \times 10^2$			
	Respirable <i>Lactobacillus</i> : $(2.3 \pm 1.7) \times 10^2$			
	<i>Penicillium</i> : $(4.8 \pm 5.4) \times 10^2$			
	Respirable <i>Penicillium</i> : $(3.5 \pm 5.7) \times 10^2$			
	Anderson six-stage viable cascade impactor—			
	<i>Staphylococcus</i> : $(7.5 \pm 4.8) \times 10^4$			
	Respirable <i>Staphylococcus</i> : $(2.2 \pm 2.0) \times 10^4$			
	<i>Pseudomonas</i> : $(4.0 \pm 6.9) \times 10^2$			
	Respirable <i>Pseudomonas</i> : $(2.8 \pm 5.8) \times 10^2$			
	<i>Bacillus</i> : $(5.9 \pm 3.8) \times 10^3$			
	Respirable <i>Bacillus</i> : $(2.8 \pm 2.0) \times 10^3$			
	<i>Listeria</i> : $(1.7 \pm 2.7) \times 10^3$			
	Respirable <i>Listeria</i> : $(1.1 \pm 1.9) \times 10^3$			
	<i>Enterococcus</i> : $(4.1 \pm 6.1) \times 10^2$			
	Respirable <i>Enterococcus</i> : $(3.1 \pm 5.4) \times 10^2$			
	<i>Nocardia</i> : $(5.8 \pm 7.7) \times 10^1$			
	Respirable <i>Nocardia</i> : $(2.6 \pm 4.6) \times 10^1$			
	<i>Lactobacillus</i> : $(4.5 \pm 4.5) \times 10^2$			
	Respirable <i>Lactobacillus</i> : $(2.1 \pm 2.0) \times 10^2$			
	<i>Penicillium</i> : $(1.2 \pm 2.1) \times 10^3$			
	Respirable <i>Penicillium</i> : $(7.7 \pm 1.5) \times 10^3$			
	Median fungi: $3.8 \times 10^5$ (BDL- $4.3 \times 10^6$ )	n/a; n/a		
Median bacteria: $5.8 \times 10^6$ (BDL- $1.6 \times 10^8$ )				
[127]	AGI Impinger—	Grower; NV	Australia	n/a
	Bacteria: $(2.89 \pm 1.69) \times 10^5$			
	$(1.12 \times 10^5 - 5.17 \times 10^5)$			
	Fungi: $(1.49 \pm 0.35) \times 10^3$			
	$(9.83 \times 10^2 - 1.85 \times 10^3)$			
	Anderson six-stage viable cascade impactor—			
	Fungi: $(18.2 \pm 6.18) \times 10^2$			
$(1.12 \times 10^3 - 2.79 \times 10^3)$				

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[160]	Site A (tunnel ventilation)— Respirable (<0.8–8 µm) fungi: 81 ± 6.2 Nonrespirable (>8 µm) fungi: 640 ± 320 Respirable bacteria: 7400 ± 1470 Nonrespirable bacteria: 31,000 ± 2680 Site B (chimney ventilation)— Respirable fungi: 90 ± 8.6 Nonrespirable fungi: 160 ± 29 Respirable bacteria: 2100 ± 180 Nonrespirable bacteria: 1500 ± 250 Mean bacteria: 3.02 × 10 <sup>4</sup> –2.58 × 10 <sup>5</sup> Mean fungi: 1.16 × 10 <sup>3</sup> –1.31 × 10 <sup>3</sup>	Grower–finisher; MV        Gestation; n/a	U.S. Midwest	n/a
[161]	Bacteria: (5.38 ± 0.65) × 10 <sup>4</sup> Fungi: (4.89 ± 1.41) × 10 <sup>2</sup> Mean bacteria: 5.95 × 10 <sup>4</sup> –2.21 × 10 <sup>5</sup> Mean fungi: 1.21 × 10 <sup>3</sup> –1.83 × 10 <sup>3</sup> Mean bacteria: 1.24 × 10 <sup>4</sup> –3.45 × 10 <sup>4</sup> Mean fungi: 1.54 × 10 <sup>3</sup> –1.83 × 10 <sup>3</sup> Mean bacteria: 8.05 × 10 <sup>4</sup> –1.76 × 10 <sup>5</sup> Mean fungi: 2.93 × 10 <sup>2</sup> –1.76 × 10 <sup>3</sup>	Farrowing; n/a  Nursery; n/a  Grower; n/a  Finisher; n/a	Taiwan	Winter
[162]	Normal pig activity— Geomean heterotrophic bacteria: 2.2 × 10 <sup>5</sup> (2.8 × 10 <sup>4</sup> –9.0 × 10 <sup>5</sup> ) Geomean <i>E. coli</i> : 21 (3–59) After flushing— Geomean heterotrophic bacteria: 2.2 × 10 <sup>5</sup> Geomean <i>E. coli</i> : 23	Grower; NV	Australia	n/a
[63]	Control— Geomean bacteria: 7.9 × 10 <sup>4</sup> Geomean molds: 930 Conventional scrapper— Geomean bacteria: 3.1 × 10 <sup>4</sup> Geomean molds: 950 V-shaped scraper— Geomean bacteria: 1.3 × 10 <sup>5</sup> Geomean molds: 1190 Daily V-shape scraper— Geomean bacteria: 5.9 × 10 <sup>4</sup> Geomean molds: 1070 Van Kempen belt— Geomean bacteria: 4.9 × 10 <sup>4</sup> Geomean molds: 1020 Cemagref net— Geomean bacteria: 4.3 × 10 <sup>4</sup> Geomean molds: 900	Grower–finisher; MV	Quebec, Canada	Summer
[163]	Mean bacteria: 18,132	Gestation; MV (chimney)	U.S. Midwest	Summer
[164]	Respirable (<0.8–8 µm) bacteria: 9629 ± 2433 Nonrespirable (>8 µm) bacteria: 8556 ± 3737 <i>Staphylococcus aureus</i> ( <i>S. aureus</i> ): (1.40 ± 0.89) × 10 <sup>4</sup> Coliform: (1.3 ± 1.2) × 10 <sup>3</sup>	Grower–finisher; MV	U.S. Midwest	Summer
[165]	Fungi: (11.9 ± 8.87) × 10 <sup>4</sup> Bacteria: (10.4 ± 3.08) × 10 <sup>5</sup>	Grower; MV	South Korea	Summer
[166]	Control— Mean bacteria: 6.7 × 10 <sup>4</sup> Oil spraying system— Mean bacteria: 3.9 × 10 <sup>4</sup> Control— Mean bacteria: 6.6 × 10 <sup>4</sup> Oil spraying system— Mean bacteria: 1.12 × 10 <sup>5</sup>	Nursery; MV   Grower; NV	Australia	n/a   n/a

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[167]	Bacteria: 10 <sup>6.13±2.87</sup> Fungi: 10 <sup>4.26±1.51</sup> Gram-negative bacteria: 10 <sup>4.32±2.66</sup> Bacteria: 10 <sup>5.67±2.09</sup> Fungi: 10 <sup>4.37±1.67</sup> Gram-negative bacteria: 10 <sup>3.98±2.37</sup>	Grower; MV	South Korea	Summer
	Deep pits with slatted floor— Bacteria: 6.76 × 10 <sup>3</sup> (17.4–6.76 × 10 <sup>4</sup> ) Fungi: 56.2 (11–7.08 × 10 <sup>3</sup> ) Manure scraper— Bacteria: 1.10 × 10 <sup>5</sup> (135–2.34 × 10 <sup>8</sup> ) Fungi: 1.38 × 10 <sup>3</sup> (8.1–7.59 × 10 <sup>4</sup> ) Deep pits with slatted floor— Bacteria: 3.31 × 10 <sup>2</sup> (81–1.51 × 10 <sup>4</sup> ) Fungi: 69.2 (3.0–741) Manure scraper— Bacteria: 3.31 × 10 <sup>3</sup> (14.5–2.24 × 10 <sup>5</sup> ) Fungi: 676 (17.4–4.37 × 10 <sup>4</sup> ) Deep litter bed system— Bacteria: 5.75 × 10 <sup>5</sup> (372–1.81 × 10 <sup>10</sup> ) Fungi: 1.38 × 10 <sup>5</sup> (708–7.24 × 10 <sup>6</sup> )	Grower–finisher; MV	South Korea	Summer, fall
[168]	Mean bacteria: 1.26 × 10 <sup>5</sup>	Grower–finisher; n/a	Quebec, Canada	Winter
[169]	Mean bacteria: 4.0 × 10 <sup>4</sup> –1.7 × 10 <sup>6</sup>	n/a; n/a	Belgium	n/a
[170]	Mean fungi: 5.3 × 10 <sup>2</sup> –6.0 × 10 <sup>3</sup>			
[147]	Mean bacteria: 1.2 × 10 <sup>4</sup> –1.6 × 10 <sup>4</sup>	Farrowing, gestation, Finisher; n/a	Illinois	All seasons
[76]	Conventional barns— Mesophilic molds: (8.65 ± 9.36) × 10 <sup>2</sup> Thermotolerant molds: 24 ± 37 Thermotolerant <i>Actinomyces</i> : 5 ± 8 Barns with sawdust beddings— Mesophilic molds: (4.82 ± 9.39) × 10 <sup>4</sup> Thermotolerant molds: (4.07 ± 7.12) × 10 <sup>2</sup> Mesophilic bacteria: (1.54 ± 0.88) × 10 <sup>6</sup> Thermotolerant <i>Actinomyces</i> : (2.66 ± 5.16) × 10 <sup>3</sup> With slatted floors— Mesophilic bacteria: (1.98 ± 2.44) × 10 <sup>5</sup> With source separation— Mesophilic bacteria: 1.37 × 10 <sup>4</sup> –1.44 × 10 <sup>4</sup> Thermotolerant <i>Actinomyces</i> : 2 ± 0	Finisher; MV (17 farms), NV (1 farm)	Quebec, Canada	Winter
[78]	Geomean mesophilic bacteria—tryptic soy agar: 1.57 × 10 <sup>6</sup> (1.48 × 10 <sup>5</sup> –1.8 × 10 <sup>7</sup> ) Reasoner's 2A agar: 8.5 × 10 <sup>5</sup> (4.18 × 10 <sup>4</sup> –1.59 × 10 <sup>7</sup> ) Geomean fungi: 2.83 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –4.28 × 10 <sup>5</sup> ) Geomean mesophilic bacteria—Tryptic soy agar: 6.31 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –8.24 × 10 <sup>5</sup> ) Reasoner's 2A agar: 6.48 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –8.21 × 10 <sup>5</sup> ) Geomean fungi: 2.05 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –2.09 × 10 <sup>5</sup> )	Grower–finisher; NV (hoop barns)	Iowa	All seasons
	Geomean mesophilic bacteria—Tryptic soy agar: 6.31 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –8.24 × 10 <sup>5</sup> ) Reasoner's 2A agar: 6.48 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –8.21 × 10 <sup>5</sup> ) Geomean fungi: 2.05 × 10 <sup>4</sup> (2.1 × 10 <sup>3</sup> –2.09 × 10 <sup>5</sup> )	Grower–finisher, MV		All seasons
[171]	Mean bacteria: 34,399 (161–29.4 × 10 <sup>5</sup> ) Mean fungi: 1882 (136–5.06 × 10 <sup>3</sup> )	Finisher, farrowing, nursery (17 farms); n/a	North Carolina	n/a
[172] <sup>b</sup>	<i>Enterococcus</i> spp.: (1.1 ± 2.6) × 10 <sup>4</sup>	Grower–finisher; n/a	Quebec, Canada	Winter

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[79]	Bacteria: 10 <sup>4.15±0.31</sup> Coliforms: 10 <sup>2.43±0.58</sup> <i>E. coli</i> : 10 <sup>1.36±0.33</sup>	Nursery; HV	South Korea	Spring
	Bacteria: 10 <sup>4.30±0.35</sup> Coliforms: 10 <sup>3.13±0.61</sup> <i>E. coli</i> : 10 <sup>3.04±0.74</sup>			Summer
	Bacteria: 10 <sup>4.01±0.25</sup> Coliforms: 10 <sup>2.08±0.90</sup> <i>E. coli</i> : 10 <sup>1.78±0.87</sup>			Fall
	Bacteria: 10 <sup>2.13±0.35</sup> Coliforms: 10 <sup>2.40±0.42</sup> <i>E. coli</i> : 10 <sup>1.64±0.28</sup>			Winter
[173]	Farm A— Median <i>E. coli</i> : 35 (13–76)	n/a, NV	China	All seasons
	Farm B— Median <i>E. coli</i> : 23 (19–58)	n/a, MV		
	Farm C— Median <i>E. coli</i> : 27 (10–67)			
	Farm D— Median <i>E. coli</i> : 21 (9–47)			
[174]	In all barns— <i>Clostridium difficile</i> : 2–625	Farrowing, gilt, boar; n/a	The Netherlands	
	In farrowing pen 1— <i>Clostridium difficile</i> : 135–575			
	In farrowing pen 2&3— <i>Clostridium difficile</i> : 0–480			
[175]	Impinger— Geomean bacteria: 3.1 × 10 <sup>5</sup> Geomean MRSA <sup>c</sup> : 257 Geomean <i>Staphylococcus</i> spp.: 1.4 × 10 <sup>4</sup> IOM dust sampler— Geomean bacteria: 2.4 × 10 <sup>5</sup> Geomean MRSA: 802 Geomean <i>Staphylococcus</i> spp.: 2.6 × 10 <sup>4</sup>	Finisher, gestation, farrowing, nursery; n/a	Germany	n/a
	[176] Median MRSA: 151 (6–3619)			
	[177] Bacteria: (47.85 ± 33.12) × 10 <sup>4</sup> Respirable bacteria: (23.07 ± 20.65) × 10 <sup>4</sup> Fungi: (1.55 ± 3.03) × 10 <sup>4</sup> Respirable fungi: (1.07 ± 2.01) × 10 <sup>4</sup>			
	[178] Pretreatment— MRSA: 14 (control) and 23 (treatment) During treatment with Stalosan <sup>®</sup> F (a disinfectant)— MRSA: 32–274 (control) and 0–215 (treatment) Post-treatment— MRSA: 17–21 (control) and 11–30 (treatment)			
[179]	Mean fungi: 5.70 × 10 <sup>3</sup> (20–5.26 × 10 <sup>4</sup> ) MSSA <sup>e</sup> : 1.56 × 10 <sup>3</sup> (100–4 × 10 <sup>3</sup> ) Mean MRSA: 300	Nursery, farrowing, finisher; n/a	Switzerland	Summer, winter
	Mean <i>S. aureus</i> : 1.61 × 10 <sup>3</sup> (100–4 × 10 <sup>3</sup> )			
[180]	Mean bacteria: 1 × 10 <sup>5</sup>	Grower–finisher; n/a	Quebec, Canada	n/a
[181]	Mean fungi— >1.8 µm: 3.0 × 10 <sup>3</sup> (1.4 × 10 <sup>3</sup> –9.0 × 10 <sup>3</sup> ) 1–1.8 µm: 2.4 × 10 <sup>2</sup> (BDL–4.8 × 10 <sup>2</sup> ) <1 µm: BDL (BDL–1.6 × 10 <sup>2</sup> ) total: 3.1 × 10 <sup>3</sup> (1.7 × 10 <sup>3</sup> –9.5 × 10 <sup>3</sup> )	n/a; n/a	Taiwan	Summer

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[182]	Farm A—	Grower–finisher; HV	Romania	Summer
	Mean mesophilic bacteria: 5.21 × 10 <sup>4</sup> –7.48 × 10 <sup>4</sup>			
	Mean fungi: 0.21 × 10 <sup>4</sup> –2.0 × 10 <sup>4</sup>			
	Mean Gram-negative bacteria: 3.12 × 10 <sup>2</sup> –3.75 × 10 <sup>3</sup>			
	Mean <i>Staphylococci</i> : 3.36 × 10 <sup>3</sup> –6.06 × 10 <sup>3</sup>			
	Mean <i>Streptococci</i> : 3.93 × 10 <sup>4</sup> –6.65 × 10 <sup>4</sup>			
	Farm B—	Grower–finisher; MV		
	Mean mesophilic bacteria: 1.79 × 10 <sup>5</sup> –1.86 × 10 <sup>5</sup>			
	Mean fungi: 1.18 × 10 <sup>4</sup> –1.53 × 10 <sup>4</sup>			
	Mean Gram-negative bacteria: 2.62 × 10 <sup>2</sup> –4.50 × 10 <sup>2</sup>			
	Mean <i>Staphylococci</i> : 4.21 × 10 <sup>4</sup> –5.03 × 10 <sup>4</sup>			
	Mean <i>Streptococci</i> : 1.50 × 10 <sup>5</sup> –1.60 × 10 <sup>5</sup>			
Farm A—	Grower–finisher; HV		Winter	
Mean mesophilic bacteria: 3.51 × 10 <sup>5</sup> –9.25 × 10 <sup>5</sup>				
Mean fungi: 0.85 × 10 <sup>4</sup> –7.23 × 10 <sup>4</sup>				
Mean Gram-negative bacteria: 7.62 × 10 <sup>2</sup> –8.12 × 10 <sup>2</sup>				
Mean <i>Staphylococci</i> : 4.43 × 10 <sup>4</sup> –4.82 × 10 <sup>4</sup>				
Mean <i>Streptococci</i> : 1.91 × 10 <sup>5</sup> –7.63 × 10 <sup>5</sup>				
Farm B—	Grower–finisher; MV			
Mean mesophilic bacteria: 1.32 × 10 <sup>6</sup> –1.56 × 10 <sup>6</sup>				
Mean fungi: 5.50 × 10 <sup>2</sup> –7.12 × 10 <sup>2</sup>				
Mean Gram-negative bacteria: 7.50 × 10 <sup>2</sup> –8.50 × 10 <sup>2</sup>				
Mean <i>Staphylococci</i> : 2.42 × 10 <sup>5</sup> –2.92 × 10 <sup>5</sup>				
Mean <i>Streptococci</i> : 8.80 × 10 <sup>5</sup> –9.18 × 10 <sup>5</sup>				
[183] <sup>f</sup>	For particles > 5 µm— Mean bacteria: 363–2.32 × 10 <sup>4</sup> Mean MRSA: 0–825 For particles < 5 µm— Mean bacteria: 255–1.38 × 10 <sup>4</sup> Mean MRSA: 24–471	Nursery-grower; n/a	U.S. Midwest	n/a
[184]	Farm A: Median bacteria: 18,688 Mean Gram-negative bacteria: 72 Median fungi: 124 (malt extract agar) Median fungi: 160 (DG18 agar) Farm B: Median bacteria: 13,660 Mean Gram-negative bacteria: 72 Median fungi: 140 (malt extract agar) Median fungi: 400 (DG18 agar) Farm C: Median bacteria: 11,944 Mean Gram-negative bacteria: 4 Median fungi: 604 (malt extract agar) Median fungi: 604 (DG18 agar) Farm D: Median bacteria: 14,720 Mean Gram-negative bacteria: 24 Median fungi: 104 (malt extract agar) Median fungi: 356 (DG18 agar) Farm E: Median bacteria: 28,210 Mean Gram-negative bacteria: 60 Median fungi: 2500 (malt extract agar) Median fungi: 2680 (DG18 agar)	Complex (gestation, farrowing, finisher); HV	Portugal	Summer

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[185]	Respicon <sup>g</sup> —	Finisher; n/a	Denmark	Fall
	Mean inhalable <i>S. aureus</i> : 227			
	Mean thoracic <i>S. aureus</i> : 113			
	Mean respirable <i>S. aureus</i> : 91			
	Mean inhalable MRSA: 219			
	Mean thoracic MRSA: 95			
	Mean respirable MRSA: 61			
	Anderson six-stage viable cascade impactor—	Farrowing; n/a		Winter
	Mean inhalable <i>S. aureus</i> : 246			
	Mean respirable <i>S. aureus</i> : 80			
	Mean inhalable MRSA: 266			
	Mean respirable MRSA: 88			
	Respicon—			
	Mean inhalable <i>S. aureus</i> : $2.6 \times 10^3$			
	Mean thoracic <i>S. aureus</i> : 850			
	Mean respirable <i>S. aureus</i> : 732			
	Mean inhalable MRSA: 100			
	Mean thoracic MRSA: 34			
	Mean respirable MRSA: 29			
	Anderson six-stage viable cascade impactor—	Nursery; n/a		Winter
Mean inhalable <i>S. aureus</i> : $2.9 \times 10^3$				
Mean respirable <i>S. aureus</i> : 990				
Respicon—				
Mean inhalable <i>S. aureus</i> : 327				
Mean thoracic <i>S. aureus</i> : 194				
Mean respirable <i>S. aureus</i> : 138				
Mean inhalable MRSA: 202				
Mean thoracic MRSA: 113				
Mean respirable MRSA: 93				
Anderson six-stage viable cascade impactor—	Sick pigs; n/a		Winter	
Mean inhalable <i>S. aureus</i> : 441				
Mean respirable <i>S. aureus</i> : 172				
Mean inhalable MRSA: 151				
Mean respirable MRSA: 85				
Respicon—				
Mean inhalable <i>S. aureus</i> : 211				
Mean thoracic <i>S. aureus</i> : 62				
Mean respirable <i>S. aureus</i> : 32				
Mean inhalable MRSA: 16				
Mean thoracic MRSA: 5.6				
Mean respirable MRSA: 3.6				
Anderson six-stage viable cascade impactor—	Nursery; MV and NV	Japan	Winter	
Mean inhalable <i>S. aureus</i> : 188				
Mean respirable <i>S. aureus</i> : 43				
Mena aerobic microbes: $10^{4.50}$ – $10^{5.42}$				
<i>Staphylococcus aureus</i> : $10^{3.90}$ – $10^{5.06}$				
<i>E. coli</i> : $10^{0.91}$ – $10^{2.97}$			Summer	
Mena aerobic microbes: $10^{3.93}$ – $10^{5.53}$				
<i>Staphylococcus aureus</i> : $10^{1.69}$ – $10^{5.14}$				
<i>E. coli</i> : BDL– $10^{2.01}$				

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[101]	Barn 1 (supply air filter modules)— Mean bacteria: $1.78 \times 10^2$ – $2.07 \times 10^6$ Mean MRSA: $2$ – $5.99 \times 10^4$ Coliforms: 0–95 <i>E. coli</i> : 0–34 Barn 2 (supply air filter attic)— Mean bacteria: $2.07 \times 10^2$ – $4.36 \times 10^6$ Mean MRSA: $3$ – $6.65 \times 10^4$ Mean Coliforms: 0–47 Mean <i>E. coli</i> : 0–125 Barn 3 (without air filtration system)— Mean bacteria: $1.36 \times 10^3$ – $2.56 \times 10^6$ Mean MRSA: $36$ – $7.76 \times 10^4$ Mean Coliforms: 0–45μ Mean <i>E. coli</i> : 0–79 Barn 4 (recirculating air filtration)— Mean bacteria: $60$ – $2.03 \times 10^6$ Mean MRSA: $0$ – $5.31 \times 10^4$ Mean Coliforms: 0–292 Mean <i>E. coli</i> : 0–361	Finisher; MV	Germany	All seasons
[186]	Geomean bacteria: 21,777 Geomean tetracycline-resistant bacteria: 2011 Geomean erythromycin-resistant bacteria: 5876	n/a; n/a	China	Fall
[187]	UVC-recirculating air filtration <sup>h</sup> — Mean bacteria: $3.8 \times 10^4$ – $1.3 \times 10^6$ No UVC-recirculating air filtration— Mean bacteria: $1.1 \times 10^5$ – $1.07 \times 10^6$	Nursery; MV	Germany	All seasons
[188]	Bacteria: 3428 ± 1244 (spring) 9824 ± 2157 (summer) 1707 ± 957 (fall) 2322 ± 1352 (winter) Bacteria: 8325 ± 3209 (spring) 18,254 ± 5166 (summer) 4258 ± 1438 (fall) 6124 ± 1527 (winter) Bacteria: 13,254 ± 6108 (spring) 24,088 ± 9274 (summer) 8254 ± 2416 (fall) 12,470 ± 4869 (winter)	Gestation-farrowing; MV  Nursery; MV  Grower-finisher; MV	Korea	All seasons
[189]	Group housing system— Bacteria: $10^{4.22}$ – $10^{5.67}$ <i>Haemolytic streptococci</i> : up to $10^{4.78}$ Fungi: up to $10^{3.43}$ Loose housing system— Bacteria: $10^{4.03}$ – $10^{5.37}$ <i>Haemolytic streptococci</i> : up to $10^{4.64}$ Fungi: up to $10^{3.50}$ Farrowing crates— Bacteria: $10^{3.43}$ – $10^{5.58}$ <i>Haemolytic streptococci</i> : up to $10^{4.92}$ Fungi: up to $10^{3.64}$	Farrowing; n/a	Germany	All seasons
[103]	Total bacteria: $1.55 \times 10^4$ – $1.55 \times 10^6$ <i>Staphylococcus aureus</i> : $4.19 \times 10^2$ – $9.05 \times 10^4$ MRSA: $<14$ – $7.91 \times 10^2$ <i>Salmonella</i> spp.: $<14$ <i>Clostridium difficile</i> : $<14$ – $1.75 \times 10^3$ <i>Mycobacterium avium</i> : $<14$ – $2.12 \times 10^3$ <i>Listeria monocytogenes</i> : $<14$	Finisher; MV	Quebec, Canada	Winter

Table 5. Cont.

Reference	Culturable Counts (CFU m <sup>-3</sup> )	Barn and Ventilation Type <sup>a</sup>	Location	Season
[190]	Bacteria: $(1.96 \pm 1.15) \times 10^4$ ( $5.37 \times 10^3$ – $5.19 \times 10^4$ ) <i>E. coli</i> : $(2.29 \pm 1.89) \times 10^3$ ( $1.41 \times 10^2$ – $6.82 \times 10^3$ ) <i>Streptococcus aureus</i> : $(1.44 \pm 0.59) \times 10^4$ ( $7.70 \times 10^3$ – $2.68 \times 10^4$ ) <i>Staphylococcus</i> : $(9.16 \pm 4.15) \times 10^3$ ( $2.65 \times 10^3$ – $1.52 \times 10^4$ )	Grower; MV	China	Fall
[191]	<i>Actinobacillus pleuropneumoniae</i> : BDL– $1.26 \times 10^6$	Wean-to-finish; MV	Australia	n/a
[192]	Fungi: 1281 ( $377$ – $2.17 \times 10^3$ )	Finisher; n/a	Denmark	Summer, winter
[193]	Mean mesophilic bacteria: $2.6 \times 10^5$ – $7.3 \times 10^5$ <i>Staphylococcus</i> spp.: $9.4 \times 10^3$ – $8.8 \times 10^4$ <i>Aerococcus</i> spp.: $4.1 \times 10^4$ – $4.2 \times 10^5$ Mean mesophilic bacteria: $2.7 \times 10^4$ – $1.1 \times 10^5$ <i>Staphylococcus</i> spp.: $1.2 \times 10^3$ – $1.2 \times 10^4$ <i>Aerococcus</i> spp.: $9.3 \times 10^3$ – $1.1 \times 10^5$	Finisher; n/a  Grower–finisher; n/a	Austria	Winter  Spring

<sup>a</sup> HV—hybrid (mixed) ventilation (mechanical + natural). <sup>b</sup> *Clostridium perfringens*, *E. coli*, and *Yersinia enterocolitica* were also quantified. However, no concentration numbers were available. <sup>c</sup> MRSA—Methicillin-resistant *Staphylococcus aureus* (*S. aureus*). <sup>d</sup> Experimental rooms, not real farms. <sup>e</sup> MSSA—Methicillin-susceptible *Staphylococcus aureus*. <sup>f</sup> Microbial counts were compiled from the Supplementary Materials of the paper. <sup>g</sup> Respicon is a three-stage cascade impactor manufactured by TSI Inc. (Shoreview, MN) and it uses filters for bioaerosol collection. <sup>h</sup> UVC-recirculating air filtration—two ultraviolet C light (UVC) tubes combined with air filters.

To our knowledge, the earliest study of culturable bacterial/fungal counts in swine barns was conducted by Dr. Fiser from the University of Veterinary Brno, Czech [194,195]. In the U.S., the first of such efforts were made by Dr. Curtis and his colleagues from the University of Illinois at Urbana-Champaign [31,32]. They collected bacterial-colony-forming particles (BCFP; i.e., particles carrying culturable bacteria) on tryptose agar using an Andersen viable cascade impactor and reported that culturable bacterial counts ranged from  $2.1 \times 10^4$  to  $1.5 \times 10^5$  colony forming units (CFU) m<sup>-3</sup>. They further studied the size distribution of BCFPs in university research barns and commercial barns. These efforts advanced our understanding of the impact of swine barn aerial environments on pig performance [196] and occupational health [197] and, in turn, stimulated a growing interest in the measurement of culturable bacterial and fungal counts in swine barns in the 1970s and 1980s [35,36,38,108,198–204]. From these early studies, a correlation was found of culturable bacterial/fungal counts with the incidence of respiratory diseases among farmers [38] and with reduced pig performance [196,201].

The interest in culturable bacterial and fungal counts continued into the 1990s and 2000s. Table 5 summarizes relevant publications since 1990. A few clarifications are provided below for readers with no relevant experience:

- Total culturable bacteria and fungi are abbreviated as bacteria and fungi, respectively, in the table. In the literature, “total” refers to the total culturable counts of all size stages as multiple-stage cascade impactors are widely used for bioaerosol sampling. It should not be confused with total bacteria/fungi where “total” refer to a collection of viable and non-viable ones.
- Molds and yeasts are two subcategories of fungi. They form colonies differing in size, shape, and texture and they were counted separately in a few studies, e.g., [152]. On a few occasions, molds and fungi were interchangeably used, e.g., [54,63].
- Different publications could use different statistical measures to summarize counting results. The commonly used measures include mean, median, geometric mean (geomean), standard deviation, and range. A few studies use logarithmic numbers, e.g., [31,167]. Caution must be taken when comparing the data from different publications.
- No official occupational exposure limits exist for culturable airborne bacterial or fungal counts. Ref. [205] proposed a threshold limit value of 5000 CFU m<sup>-3</sup> for culturable

airborne bacteria and a more stringent value ( $1000 \text{ CFU m}^{-3}$ ) was later proposed by [206]. For culturable fungi, a concentration of  $\leq 100 \text{ CFU m}^{-3}$  could detriment immunosuppressed people [207]. No threshold limits exist regarding pig exposure to culturable bacteria or fungi.

It is a daunting challenge to compile, compare, and analyze existing data in the literature because, as aforementioned, culturable bacterial/fungi counts vary with measurement methods and are presented in various forms. This challenge is further complicated by size-segregated sampling, i.e., many publications offered culturable counts in multiple size ranges. Upon the analysis of the publications since 1990, the following observations are made:

- Among the 53 publications compiled, 21 were conducted in Europe, followed by the U.S. (9), Canada (8), and South Korea (5). This is consistent with the geographical distribution of PM studies in Table 2. Thirty-three publications reported general bacterial counts, and 21 presented general fungal/mold counts. The counts of a specific species (or group) were reported by 20 publications. The typical species (or group) of interest include Gram-negative bacteria, *Staphylococcus*, methicillin-resistant *Staphylococcus aureus* (MRSA), *Actinomyces*, and *E. coli*. Many studies were limited to short-term farm surveys (which is partly related to the methodology constraints). Only a few studies involved long-term monitoring efforts, e.g., [101,157,187].
- The total culturable bacterial count varied substantially in the literature, ranging from 14.5 [168] to  $1.82 \times 10^{10} \text{ CFU m}^{-3}$  [168]. Another high concentration ( $1.6 \times 10^8 \text{ CFU m}^{-3}$ ) was reported by [59]. Caution should be taken when interpreting extremely high count values. Assuming that bacteria weigh at  $1 \times 10^{-12} \text{ g}$  per cell [208], the count of  $1.82 \times 10^{10} \text{ CFU m}^{-3}$  would indicate an airborne bacterial mass concentration of  $18.2 \text{ mg m}^{-3}$ , which is unlikely in reality. Excluding the extremes, most of the total culturable bacterial counts fall into the orders of magnitude of  $10^3$ – $10^6 \text{ CPU m}^{-3}$ .
- The total culturable fungal count also varied greatly, ranging from  $1.3 \text{ CFU m}^{-3}$  in a farrowing barn in Poland [43] to  $7.24 \times 10^6 \text{ CFU m}^{-3}$  in a grower–finisher barn in South Korea [168]. Another high concentration level ( $4.3 \times 10^6 \text{ CFU m}^{-3}$ ) was observed in Denmark [59]. Most of the total culturable fungal counts in the literature fall into the orders of magnitude of  $10^2$ – $10^4 \text{ CPU m}^{-3}$ .
- Gram-negative bacteria accounted for only a small portion of culturable bacterial counts (0.1% to 1% of the latter). This is consistent with the findings derived from the molecular biology analysis of airborne bacterial communities in swine barns [169,209,210]. However, molecular biology methods such as polymerase chain reaction (PCR) and DNA sequencing may detect both living and dead microorganisms (with DNA molecules yet to be degraded) [209]. Gram-negative bacteria are of particular concern when it comes to human and animal health as they include pathogens such as *E. coli* and *Salmonella* spp. When dead, Gram-negative bacteria also release endotoxins, a cell wall component with well-established health implications [211].
- No consistent seasonality was observed. Ref. [54] reported that culturable bacterial counts were significantly lower in summer than in winter. This contradicts an early finding [212] that a significantly lower bacterial count occurred in winter/spring than in summer/fall. A significant seasonal variation was also noted by Refs. [78,147]. Different microbial species or groups may exhibit different seasonality. In the same study by Ref. [54], no significant difference was seen between summer and winter in terms of fungi, thermophilic actinomycetes, or *Saccharopolyspora rectivirgula*. The lack of significant seasonality was also reported by Ref. [167] for fungi, bacteria, and Gram-negative bacteria. While there was no significant seasonality in bacteria, significant seasonal changes in coliforms and *E. coli* were reported by Ref. [79]. Similarly, Ref. [182] found a significant seasonality in mesophilic bacteria, but no significant seasonal effect on fungi, Gram-negative bacteria, *Streptococci*, or *Staphylococci*.
- Limited information is known about the effect of barn types. Only a few studies compared culturable bacterial or fungal counts between different barn types—a sharp

contrast to PM concentration assessment. Even worse, many of the existing studies [43,166,185] provided no conclusive comparison due to the lack of statistical analysis. Ref. [158] compared five types of swine barns in Taiwan: breeding, farrowing, nursery, grower, and finisher. A significantly higher bacterial count was detected in finisher barns than farrowing, nursery, and grower barns, but no significant difference in culturable fungal counts was noted among these barns. Ref. [78] compared two types of grower–finisher barns: hoop barns (with natural ventilation) versus regular barns (with mechanical ventilation); and found a significant effect of barn types on total microbes but no significant effect on mesophilic bacteria. Ref. [147] compared farrowing and finisher barns in Illinois and reported no significant difference in culturable bacterial counts.

- Limited information is known about the effect of waste management systems. Ref. [63] compared six manure separation systems and observed the highest bacterial and mold counts in the barn with V-shaped scrapers. However, no significant difference was found among different separation systems. The study by Ref. [78] also involved a comparison of waste management systems: deep manure pits in regular barns versus bedded floors in hoop barns. The overall higher bacterial and fungal counts in hoop barns could be related to manure build-up and microbial growth on bedding materials. In an extensive farm survey, Ref. [171] compared 17 different manure treatment technologies and found a significant effect on culturable bacterial counts.
- Culturable bacterial and fungal counts could be affected by animal (e.g., age/weight and the number of pigs) and environmental factors (e.g., air temperature, humidity, and air speed). Ref. [54] found that airborne culturable bacterial counts correlated negatively with barn dustiness and positively with the number of pigs, while culturable mold counts correlated positively with dustiness. Ref. [78] found significant effects on mesophilic bacteria by air temperature, humidity, wind speed, and the number of pigs, and significant effects on total microbes by air temperature and humidity. Ref. [79] investigated the influence of barn microclimate on airborne bacteria and found a significant effect of air temperature and air speed on coliform and *E. coli*; however, no significant effect of air humidity was noted. Similar observations were made by [167,177]. It is noteworthy that air temperature in swine barns varies with outdoor temperature and ventilation conditions. Thus, the effect of air temperature may confound with seasonality. Two studies [101,187] measured culturable bacterial counts over multiple production cycles and noted that the bacterial counts generally increased with animal age/weight.
- The effectiveness of mitigation technologies remains uncertain for culturable bacteria/fungi. Ref. [166] tested an oil sprinkling system for its effectiveness in dust, total bacteria, and ammonia removal and reported a 42% reduction in culturable bacterial counts in a nursery barn. However, in a grower barn, oil sprinkling increased culturable bacterial counts by 70%. The reason remains unknown. Ref. [157] tested two recirculating filtration technologies (fabric filters and electrostatic filters) and found culturable bacteria reduction efficiencies of 10–50% for the fabric filter in a grower barn and 20–52% for the electrostatic filter in a finisher barn. Ref. [101] compared four air filtration options (no filtration, supply air filter modules, supply air filter attic, and recirculating air filtration modules) in finisher barns. Pig performance was improved with recirculating air filtration modules. However, no significant reduction in culturable bacterial counts was observed. Ref. [187] investigated the combination of ultraviolet C (UVC) light and air filtration for its mitigation of culturable bacteria and reported a reduction efficiency of 31.6% in trial 1 and 63% in trial 2.
- Culturable bacteria and fungi show different size distribution patterns. Culturable bacterial counts tend to increase with particle size [147,177,185,188]. This, consistent with early findings [201], suggests that the bacteria may not exist as individual particles but bind to large particles in the air. Comparatively, culturable fungal counts peaked at approximately 2–5  $\mu\text{m}$  [177,192], suggesting that airborne fungi in swine barns

may exist as individual spores/particles. Different size distribution patterns suggest that airborne bacteria and fungi may undergo different aerodynamic transport and removal processes in swine barns [24] and different strategies may be required for the mitigation of airborne bacteria than fungi.

- No agreement has been reached regarding the relationship between culturable bacterial counts and PM concentrations. A significant and positive correlation was observed by [54,147]. However, a later study [79] found no significant correlation of PM concentrations with culturable bacterial, coliform, or *E. coli* counts. The inconsistency in the literature might be caused by sampling duration. Due to the high bioaerosol concentrations inside swine barns, most bioaerosol samplers must be run for a short sampling duration (typically a few minutes) compared to PM samplers (typically 24 h). Future research is needed to further investigate the relationship and alike. If a correlation exists, it will simplify the assessment and mitigation of airborne bacteria in swine barns.

### Total Bacterial and Fungal Counts

To our knowledge, the first measurement of total bacterial/fungal counts in swine barns was conducted by Ref. [213], immediately after the invention of the Nuclepore filter method (also known as CAMNEA method) by the same authors [214]. An average count of  $2 \times 10^9$  cells  $m^{-3}$  was reported for total microbes (fungi + bacteria). The method involved the use of acridine orange (AO) to stain PM collected on a Nuclepore (i.e., polycarbonate) filter and the identification and enumeration of microbes with epifluorescence microscopy (EPM). Using the same method, Ref. [38] measured bioaerosol concentrations in 30 swine barns in Sweden and found an average count of  $1.4 \times 10^6$  cells  $m^{-3}$  for bacteria,  $2 \times 10^4$  cells  $m^{-3}$  for fungi, and  $1.8 \times 10^7$  cells  $m^{-3}$  for total microbes. Additional measurement results have been available since 1990, as summarized in Table 6.

**Table 6.** Total bacterial and fungal counts in swine barns—A summary of studies since 1990.

Reference	Total Counts (Cell $m^{-3}$ ; Unless Otherwise Noted) <sup>a,b</sup>	Building and Ventilation Type	Location	Season	Analytical Method <sup>c</sup>
[215]	Bacteria with AGI impingers— Cytometry/DAPI: $\sim 3 \times 10^7$ EFM/DAPI: $\sim 1.5 \times 10^7$ Bacteria with May impingers— Cytometry/DAPI: $\sim 1.3 \times 10^7$ EFM/DAPI: $\sim 0.9 \times 10^7$ (Read from Figure 4 of [215])	n/a; n/a	Iowa	n/a	DAPI staining and flow cytometry; DAPI staining and EPM
[59]	Mean bacteria: $4.2 \times 10^8$ (BDL- $1.6 \times 10^{10}$ ) Mean fungi: $3.8 \times 10^5$ (BDL- $4.3 \times 10^6$ ) Mean total microbes— EPM/AO: $6.05 \times 10^6$ – $6.63 \times 10^6$ EPM/DAPI: $5.03 \times 10^6$ – $5.82 \times 10^6$ FISH/DAPI: $7.31 \times 10^6$ – $1.05 \times 10^7$	n/a; n/a Gestation; n/a	Denmark	n/a	AO staining and EPM
[161]	Total microbes— EPM/AO: $(1.08 \pm 0.03) \times 10^7$ EPM/DAPI: $(9.43 \pm 0.25) \times 10^6$ FISH/DAPI: $(8.20 \pm 0.84) \times 10^6$ Mean total microbes— EPM/AO: $8.82 \times 10^6$ – $2.18 \times 10^7$ EPM/DAPI: $5.69 \times 10^6$ – $2.17 \times 10^7$ FISH/DAPI: $9.87 \times 10^6$ – $1.20 \times 10^7$ Mean total microbes— EPM/AO: $5.48 \times 10^6$ – $7.53 \times 10^6$ EPM/DAPI: $5.39 \times 10^6$ – $7.14 \times 10^6$ FISH/DAPI: $5.66 \times 10^6$ – $2.20 \times 10^7$ Mean total microbes— EPM/AO: $6.91 \times 10^6$ – $1.12 \times 10^7$ EPM/DAPI: $5.93 \times 10^6$ – $1.07 \times 10^7$ FISH/DAPI: $7.65 \times 10^6$ – $1.32 \times 10^7$	Farrowing; n/a Nursery; n/a Grower; n/a Finisher; n/a	Taiwan	Winter	AO or DAPI staining and EPM; DAPI staining and FISH

Table 6. Cont.

Reference	Total Counts (Cell m <sup>-3</sup> ; Unless Otherwise Noted) <sup>a,b</sup>	Building and Ventilation Type	Location	Season	Analytical Method <sup>c</sup>
[170]	Mean total microbes: 7.9 × 10 <sup>4</sup> –9.4 × 10 <sup>5</sup> Mean bacteria: 7.5 × 10 <sup>4</sup> –9.3 × 10 <sup>5</sup> Mean fungi: 1.01 × 10 <sup>3</sup> –1.2 × 10 <sup>4</sup>	n/a; n/a	Belgium	n/a	Staining and flow cytometry: ChemChrome V6/ChemSol B16 for total microbes, and ChemChrome V6/ChemSol B2 for fungi 16S rRNA qPCR
[76]	Bacteria: ~1 × 10 <sup>6</sup> –1 × 10 <sup>8</sup> (read from Figure 2 of [76])	Finisher; MV (17 farms), NV (1 farm)	Quebec, Canada	Winter	
[78]	Geomean total microbes: 3.01 × 10 <sup>6</sup> (2 × 10 <sup>4</sup> –1.8 × 10 <sup>7</sup> ) Geomean total microbes: 1.49 × 10 <sup>6</sup> (1.8 × 10 <sup>4</sup> –1.22 × 10 <sup>7</sup> )	Grower–finisher; NV (hoop barns) Grower–finisher; MV	Iowa	All seasons	AO staining and EPM
[216]	Bacteria: 5 × 10 <sup>6</sup> –4 × 10 <sup>8</sup> copies m <sup>-3</sup>	Farrowing, nursery, grower–finisher; n/a	Quebec, Canada	All seasons	16S rRNA qPCR
[87]	Total microbes: (2.7 ± 0.7) × 10 <sup>7</sup> Fungi: (1.2 ± 0.3) × 10 <sup>6</sup>	Sows; MV (chimney)	Denmark	Spring, summer	Staining and EPM: AO for total microbes, and PAS for fungi 16S rRNA qPCR for bacteria; qPCR with a special primer pair for <i>Staphylococcus</i> spp.
[179]	Mean bacteria: 3.6 × 10 <sup>9</sup> copies m <sup>-3</sup> (1.6 × 10 <sup>5</sup> –6.1 × 10 <sup>10</sup> ) <i>Staphylococcus</i> spp.: 1.6 × 10 <sup>7</sup> copies m <sup>-3</sup> (2 × 10 <sup>3</sup> –4 × 10 <sup>8</sup> )	Nursery, farrowing, finisher; n/a	Switzerland	Summer, winter	16S rRNA qPCR for bacteria; qPCR with a special primer pair for <i>Staphylococcus</i> spp.
[217]	Mean total microbes: 1.1 × 10 <sup>6</sup> Mean total microbes: 2 × 10 <sup>7</sup> Mean total microbes: 1.1 × 10 <sup>6</sup> (Read from Figure 1 of [217])	Nursery; n/a Grower; n/a Finisher; n/a	Colorado	Summer, fall	DAPI staining and EPM
[180]	Bacteria: 1 × 10 <sup>8</sup> –2 × 10 <sup>8</sup> <i>Streptococcus suis</i> : 4 × 10 <sup>5</sup> –1 × 10 <sup>6</sup> <i>Streptococcus suis</i> serotypes 2 and 1/2: 1 × 10 <sup>3</sup> –3 × 10 <sup>4</sup>	Grower–finisher; n/a	Quebec, Canada	n/a	16S rRNA qPCR for bacteria; qPCR with a special primer pair for <i>S. suis</i> and its serotypes
[218]	Mean bacteria: 1.4 × 10 <sup>8</sup> copies m <sup>-3</sup> Mean bacteria: 1.2 × 10 <sup>7</sup> copies m <sup>-3</sup>	Grower–finisher; n/a	South Korean	Winter Summer	16S rRNA qPCR
[181]	Mean fungi: 4.1 × 10 <sup>5</sup> (3.6 × 10 <sup>5</sup> –5.6 × 10 <sup>5</sup> )	n/a; n/a	Taiwan	Summer	AO or PI staining and EPM
[219]	Deep pit with slatted floor— Mean bacteria: ~10 <sup>8.2</sup> copies m <sup>-3</sup> Manure scraper— Mean bacteria: ~10 <sup>7.6</sup> copies m <sup>-3</sup> Litter bedding— Mean bacteria: ~10 <sup>7.2</sup> copies m <sup>-3</sup> (Read from Figure 2 of [219])	Grower–finisher; n/a	South Korean	Winter	16S rRNA qPCR
[220]	Mean fungi: ~10 <sup>5.3</sup> –10 <sup>5.8</sup> copies m <sup>-3</sup> Mean fungi: ~10 <sup>4.5</sup> –10 <sup>5.7</sup> copies m <sup>-3</sup> (Read from Figure 2 of [220])	Grower–finisher; n/a	South Korean	Winter Summer	ITS qPCR
[221]	Mean bacteria: ~10 <sup>20</sup> copies m <sup>-3</sup> Mean bacteria: ~10 <sup>16</sup> copies m <sup>-3</sup> Mean bacteria: ~10 <sup>10.5</sup> copies m <sup>-3</sup> Mean bacteria: ~10 <sup>14</sup> copies m <sup>-3</sup> (Read from Figure 1 of [221])	n/a; n/a	Switzerland	Winter Spring Summer Fall	16S rRNA qPCR
[103]	Bacteria: 8.06 × 10 <sup>7</sup> –3.34 × 10 <sup>9</sup> copies m <sup>-3</sup> <i>Staphylococcus aureus</i> : 5.04 × 10 <sup>4</sup> –7.43 × 10 <sup>5</sup> copies m <sup>-3</sup> MRSA: <8–1.19 × 10 <sup>4</sup> copies m <sup>-3</sup> <i>Salmonella</i> spp.: 7.59 × 10 <sup>2</sup> –1.07 × 10 <sup>6</sup> copies m <sup>-3</sup> <i>Clostridium difficile</i> : <8–4.21 × 10 <sup>4</sup> copies m <sup>-3</sup> <i>Mycobacterium avium</i> : <8 copies m <sup>-3</sup> <i>Listeria monocytogenes</i> : <8–3.16 × 10 <sup>3</sup> copies m <sup>-3</sup>	Finisher; MV	Quebec, Canada	Winter	16S rRNA qPCR for bacteria; qPCR with special primer pairs for individual species

Table 6. Cont.

Reference	Total Counts (Cell m <sup>-3</sup> ; Unless Otherwise Noted) <sup>a,b</sup>	Building and Ventilation Type	Location	Season	Analytical Method <sup>c</sup>
[191]	<i>Actinobacillus pleuropneumoniae</i> : BDL- $5.3 \times 10^3$	Wean-to-finish; MV	Australia	n/a	<i>apxIV</i> qPCR
[222]	Mean bacteria: $10^{12}$ copies m <sup>-3</sup> (Read from Chap 5, Figure 1 of [222])	n/a; n/a	Nine European countries	n/a	16S rRNA qPCR

<sup>a</sup> Some studies did not distinguish bacteria from fungi. Instead, they presented the measurement results as the counts were for total microbes, i.e., fungi plus bacteria. <sup>b</sup> AO—acridine Orange; DAPI—4',6-diamidino-2-phenylindole; EPM—epifluorescence microscope; FISH—fluorescence in situ hybridization; BDL—below the detection limit. <sup>c</sup> PAS—Periodic Acid–Schiff; PI—propidium iodide; ITS—internal transcribed spacer, an indicator gene for fungi; 16S rRNA—16S ribosome RNA gene, an indicator gene for bacteria; *apxIV*—a species-specific gene for *Actinobacillus pleuropneumoniae*.

Besides AO, fluorochromes such as DAPI and PI were also used as staining agents for microscopic or cytometric counting of total bacteria and/or fungi. With technological advances in molecular biology, real-time quantitative polymerase chain reaction (qPCR) became a prevalent, alternative method in recent years. The results derived from qPCR were often presented in the number of gene copies per m<sup>3</sup> of air (copies m<sup>-3</sup>). Here, the gene refers to an indicator gene selected for general bacteria, general fungi, or specific bacterial/fungal groups. A single microbial cell may have multiple copies of a target gene. Thus, the results from qPCR cannot be directly compared with those from fluorochrome-assisted counting. Moreover, different fluorochromes may result in different counting results [161,181]. In short, total bacterial/fungal counting is highly method dependent. Further information about measurement methodology can be found in Section 4.4.1.

Several publications also discussed the measurement of total bacterial/fungal counts in swine barns [156,223–226]. However, they focused on methodology development with no measurement data provided. Upon the analysis of existing publications, the following observations are made:

- Among the 19 reports since 1990, six were from Europe, followed by Canada (four). Only three studies were conducted in the U.S., all over ten years ago. Eleven studies measured total bacterial counts, five monitored total fungal counts, five examined total microbial (bacterial + fungal) counts, and four reported the count of specific microbes. Among various barn types, finisher and grower–finisher barns were most studied.
- Total bacterial counts (in cells m<sup>-3</sup>) varied greatly in swine barns. Both the lowest (BDL) and the highest counts ( $1.6 \times 10^{10}$ ) were reported by Ref. [59]. Again, special attention should be paid to exceptionally high count values. The total bacterial counts typically fell into the range of  $10^6$ – $10^8$  cell m<sup>-3</sup>, two-to-three orders of magnitude greater than that for culturable bacteria ( $10^3$ – $10^6$  CPU m<sup>-3</sup>). The total bacterial counts derived from 16S rRNA qPCR exhibited even greater variability, ranging from  $1.6 \times 10^5$  [179] to  $\sim 10^{20}$  copies m<sup>-3</sup> [221]. A single bacterial cell can contain multiple copies of 16S rRNA genes and the number of copies varies with bacterial species [227]. Thus, there is no well-established relationship that can readily translate bacterial counts from gene copies to cells.
- Total fungal counts (in cells m<sup>-3</sup>) also varied substantially in the literature and typically stayed in the range of  $10^3$ – $10^6$  cells m<sup>-3</sup>. This is one-to-two orders of magnitude greater than that of culturable fungi ( $10^2$ – $10^4$  CPU m<sup>-3</sup>). Only one study reported the fungal counts derived from ITS qPCR [220], spanning from  $\sim 10^4$  to  $10^7$  copies m<sup>-3</sup>.
- A direct comparison of culturable versus total counts revealed that only a small portion of airborne bacteria/fungi in swine barns are culturable. Ref. [215] reported that culturable bacterial counts were about two orders of magnitude lower than total bacterial counts derived from DAPI staining coupled with cytometric or EFM counting. Ref. [59] found that culturable bacterial counts accounted for on average only 1.4% of total bacteria counts while culturable fungi counts were 4.4% of total fungal counts in the same barns. Ref. [161] compared culturable bacterial counts versus

total bacterial counts derived from three different methods (EFM/AO, EFM/DAPI, and FISH/DAPI) and found that, regardless of the methods, culturable counts were one to three orders-of-magnitude lower than total counts. Similar observations were made by Refs. [76,78,170,180,181].

- Total bacterial/fungal counts showed significant seasonal variability. In nearly all reports, total bacterial/fungal counts were highest in winter and lowest in summer [218–221]. The only exception is Ref. [78] in which the highest counts of total microbes occurred in summer in regular swine barns but fall in hoop barns. A significant seasonal effect was found through reduced ANOVA modeling. No winter samples were analyzed in the same study.
- Little is known about the effect of barn types or waste management systems. Only one study compared different barn types [161], with no significant effect found. However, multiple types of barns are often collocated to form a farm complex in Taiwan, which is different from the setting in the U.S. Thus, it is uncertain whether the finding would apply to the U.S. pork production systems. Only one study compared different waste management systems [219], with significantly greater total bacterial counts detected in barns with slatted floors and deep pits than those with beddings. No discussion about possible reasons was offered.
- Challenges remain regarding the interpretation of qPCR-derived total count data. As qPCR becomes increasingly accessible and affordable, numerous qPCR data were generated in the past decade. To date, the data (in copies  $m^{-3}$ ) have only been used to compare the relative abundance of bacteria/fungi in different environments or barn conditions. How to relate them to the health and welfare of pigs or farm workers remains a technical challenge.

#### Viable Bacterial and Fungal Counts

Viability is critical for assessing the health effects of airborne microbes. Despite the importance of viable bacteria and fungi, the direct measurement of their airborne counts is technically challenging. Only a few studies reported the measurement results from swine barns. The measurement usually involved various assumptions and combined use of multiple technologies. As only a few reports are available, both the measurement results and methodology are summarized in this section.

Ref. [127] measured the counts of viable particles in swine barns using an ultraviolet aerodynamic particle sizer (UVAPS). Viable particles refer to airborne particles carrying viable microbes. The detection of viable particles by UVAPS relies on the autofluorescence of nicotinamide adenine dinucleotide phosphate (NADPH), a fluorophore and an essential molecule for all living organisms [228]. An assumption behind the measurement is that only viable microbes contain NADPH and that no interference from other fluorophores occurs at the exciting light wavelength (335 nm) in the UVAPS. An average concentration of  $(10.71 \pm 4.41) \times 10^6$  particles  $m^{-3}$  was reported.

Ref. [161] studied the viability of airborne microbes in swine barns with three methods: (1) PI staining followed by EPM counting—as PI can stain completely damaged cells, viable counts can be acquired by subtracting damaged dead cells from total counts; (2) YOPRO-1 staining followed by EPM counting—YOPRO-1 can stain both partially and completely damaged cells so viable counts can be estimated by subtracting damaged cells from total counts; and (3) FISH coupled with EPM counting—assuming that only viable cells have enough DNA to be hybridized with fl-Univ FISH probes, viable counts can be acquired from EPM counting of FISH-labeled cells. Approximately 55% of total microbes were found to be viable with the PI method, 37% with the YOPRO-1 method, and 66% with the FISH method. Ref. [181] used a similar method (PI staining followed by EPM counting) to study the viability of swine barn aerosols. An average viable fungal count of  $4.1 \times 10^5$  cells  $m^{-3}$  (range:  $3.6 \times 10^5$ – $5.6 \times 10^5$ ) was reported, accounting for on average 27.9% (range: 22.2–62.5%) of total fungi.

Ref. [226] used viable-qPCR (vPCR) to measure viable airborne bacteria in multiple indoor environments, including swine barns. The method combined propidium monoazide (a DNA stain) and 16S rRNA qPCR to construct a calibration curve (of 16rRNA gene copies versus viable cells) for viability assessment. However, no measurement data was given for swine barns.

### 3.2.2. Viruses

Airborne transmission has been long recognized as a pathway for the spread of infective zoonotic viruses, including porcine viruses [229]. For example, Donaldson and Ferris studied the survivability of airborne African swine fever viruses—which remain as one of the most critical swine pathogens today—at different relative humidity levels [230]. Other investigated airborne porcine viruses include influenza A virus (IAV) and foot-and-mouth disease virus (FMDV), as summarized in [231]. Early investigations focused on the survivability of porcine viruses in the air, a key factor for viral spread and infection; and they stimulated the development of relevant measurement technologies [232,233]. Nearly all these investigations were conducted in the lab, likely due to biosecurity considerations and viral detection being then complicated and labor-intensive.

A milestone paper about airborne porcine viruses is Ref. [234]; it provided the first experimental evidence for airborne viral transmission in swine barns. The experiment spatially separated healthy pigs from pigs inoculated with porcine reproductive and respiratory syndrome viruses (PRRSV) and the initially healthy group was found to be PRRSV-positive after 16 days. This artificial inoculation method was adopted by many follow-up studies. For example, Ref. [235] reported the detection of PRRSV-positive bioaerosol samples 4.7 km away from artificially infected barns, backing a longstanding viewpoint that airborne transmission can attribute to a regional outbreak of porcine reproductive and respiratory syndromes (PRRS) [236]. Despite intense interest in airborne porcine viruses, field measurement reports remain sporadic.

To our knowledge, the first field measurement of airborne viruses in swine barns was conducted by Ref. [216]. Since then, 15 field studies have been reported (Table 7). A brief overview of part of these studies can be found in Ref. [9]. This section summarizes each study in greater detail and includes the latest efforts (2017–present). Only the direct measurement of airborne viruses in production barns is reviewed here. Indirect measurement (e.g., using infected pigs as an indication of airborne transmission [237,238]) or artificially infected barns (e.g., [234,235,239]) is excluded. These studies are important in understanding the airborne transmission of pathogenic viruses in and around swine farms, but do not represent actual barn conditions.

**Table 7.** Field measurement of airborne viruses in swine barns.

Reference	Barn and Ventilation Type	Location	Season	Analytical Method <sup>a</sup>	Key Findings and/or Notes <sup>b</sup>
[216]	Farrowing, nursery, grower–finisher; n/a	Quebec, Canada	All seasons	qPCR	Most TSP samples were PCV2 positive. Airborne PCV2 concentrations ranged from $2 \times 10^3$ to $10^7$ copies $m^{-3}$ . A significant but weak correlation was found between airborne PCV2 and PM concentrations.

Table 7. Cont.

Reference	Barn and Ventilation Type	Location	Season	Analytical Method <sup>a</sup>	Key Findings and/or Notes <sup>b</sup>
[240]	Nursery (1 farm), wean-to-finish (3 farms); n/a	Iowa, Minnesota	Spring, fall	Real-time RT-PCR	Presumptively infected farms were visited. All farms but one (wean-to-finish) were found to be IAV positive with collected bioaerosol samples. The average IAV concentration was $3.20 \pm 4.01 \times 10^5$ copies $m^{-3}$ . The detected IAV subtypes included H1N2, H1N1, and H3N2. The air at the exhaust and that downwind (up to 1.9 km) from the infected barns also contained IAV.
[217]	Nursery, grower, finisher, Wean-to-finish; n/a	Colorado	Spring, summer, fall	qPCR	PCV2 was detected in all but nursery barns, with airborne concentrations ranging from $10^3$ – $10^5$ copies $m^{-3}$ . The PCV2 genotypes identified through DNA sequencing included PCV2, PCV2a, PCV2b, PCV1/2a, and PCV2e.
[241]	Farrowing, gestation, gilts; n/a	Iowa, Minnesota, South Dakota	Fall, winter	Real-time RT-PCR	Bioaerosol samples were collected 30 m downwind from farms during an outbreak, with 37% of the samples being PRRSV positive. Phylogenetic analysis was conducted through sequencing, with 19 viral clades identified. Viral concentrations were presented in median tissue culture infectious dose per mL of sample (TCID <sub>50</sub> mL <sup>-1</sup> ).
[242]	Wean-to-finish; n/a	Denmark	n/a	Real-time RT-PCR	Bioaerosol samples from four infected herds were analyzed. The majority of in-barn samples were PRRSV positive. Outdoor samples (~30 m downwind) were also tested, with four out of 20 being positive.
[243]	Nursery, grower, finisher, sow; n/a	China	Summer, fall, winter	Real-time RT-PCR	None of the summer bioaerosol samples were IAV positive; while nine fall/winter bioaerosol samples (out of 95) were positive. The positive rate of bioaerosol samples was lower than that of pig oral secretion samples or environmental swab samples.
[244]	n/a; n/a	Iowa, Minnesota	Fall, winter, spring	Real-time RT-PCR	A total of 43% of in-barn bioaerosol samples were IAV positive, slightly lower than the positive rate (48%) of pig oral secretion samples. Bioaerosol samples were also collected 25 m upwind and downwind from barns but none of them was IAV positive.
[245]	n/a; n/a	Iowa	n/a	Real-time RT-PCR	Two samplers (NIOSH bioaerosol sampler BC251 and PHISH) were used for assessing the personal exposure of veterinarians in infected barns. The geometric mean IAV concentration was 2094 copies $m^{-3}$ using the NIOSH sampler and 545 copies $m^{-3}$ using the PHISH sampler. Three IAV subtypes were identified: H1N1, H3N2, and H1N2.

Table 7. Cont.

Reference	Barn and Ventilation Type	Location	Season	Analytical Method <sup>a</sup>	Key Findings and/or Notes <sup>b</sup>
[246]	Nursery; n/a	Minnesota	Spring	Real-time RT-PCR	Presumptively infected farms were visited to compare two bioaerosol samplers (Andersen eight-stage non-viable cascade impactor and Tisch four-stage non-viable cascade impactor) for their performance regarding PRRSV and PEDV sampling. Seven out of 16 samples were PRRSV positive, and 12 out of 12 samples were PEDV positive. The PRRSV positive rate derived from Andersen. The airborne virus concentration was $10^{6.04 \pm 0.72}$ copies $m^{-3}$ for PEDV and $10^{3.79 \pm 1.35}$ copies $m^{-3}$ for PRRSV. Outdoor bioaerosol samples were also tested.
[136]	Nursery; MV and NV	Japan	Winter, summer	Real-time RT-PCR	Barn-average airborne PSV concentrations ranged from $10^{3.57}$ to $10^{4.03}$ copies $m^{-3}$ in summer and from $10^{5.22}$ to $10^{5.85}$ copies $m^{-3}$ in winter. No correlation between airborne PSV and PM concentrations was found.
[247]	Nursery, finisher; n/a	Austria	n/a	Real-time RT-PCR	The study collected bioaerosol samples from three PRRSV-2 infected barns with three different samplers (Coriolis <sup>®</sup> $\mu$ , MD8 Airscan, and IOM Multidust). None of the field samples were PRRSV-2 positive.
[101]	Finisher; MV	Germany	All seasons	Real-time RT-PCR	No PRRSV was detected in any barns. The reason was ascribed to no infection or blow the qPCR's detection limit.
[248]	Farrowing-to-wean; n/a	U.S. Midwest	n/a	Real-time RT-PCR	Deposition samples in an infected barn were collected (until 23 weeks after an outbreak) for PRRSV analysis. For newborns (3–5 days), 24% of aerosol samples were PRRSV positive. For piglets of ~21 days, 33.3% of aerosol samples were positive.
[249]	Farrowing, gestation; MV	Ontario, Canada	All seasons	Real-time RT-PCR	Four samplers were used: a PTFE filter in an open cassette, a NIOSH cyclone, a Coriolis <sup>®</sup> $\mu$ sampler, and a SASS 3100 bioaerosol sampler. IAV-positive samples accounted for 28.9%, 30.3%, 11%, and 0% of those from the four samplers, respectively. Viral phenotyping was conducted through high-throughput sequencing.
[250]	Wean-to-finish; NV	Spain	n/a	qPCR	Three batches were monitored: one unvaccinated and two vaccinated. Bioaerosol samples from the unvaccinated batch showed greater PCV2 concentrations ( $1.19 \times 10^3$ – $2.42 \times 10^7$ copies $m^{-3}$ ; Note: positive samples only) than those from vaccinated batches ( $5.39 \times 10^2$ – $5.93 \times 10^4$ copies $m^{-3}$ ). Positive bioaerosol samples occurred earlier than positive blood samples.

<sup>a</sup> RT-PCR—reverse transcript polymerase chain reaction. <sup>b</sup> PCV2—porcine circovirus type 2; IAV—influenza A virus; PRRSV—porcine reproductive and respiratory syndrome virus; PEDV—porcine epidemic diarrhea virus; PSV—porcine sapelovirus; PHISH—personal high-flow inhalable sampler head.

Among the 15 field measurement reports, seven came from the U.S. (in particular Upper Midwest), four from Europe, and two each from Canada and Asia. Five common porcine viruses were studied: PCV2, IAV, PRRSV, PEDV, and PSV. PCV2 is a DNA virus. The rest are RNA viruses, demanding a different measurement method. IAV has been the most measured virus, followed by PRRSV. The presence of IAV, PCV2, and PSV is common in commercial production barns because they present relatively low hazards (except for high-risk subtypes); whereas PRRSV and PEDV are highly hazardous, with stringent surveillance and elimination programs implemented to prevent their presence. Accordingly, airborne PRRSV and PEDV measurement was primarily conducted in infected or presumptively infected swine barns [241,242,246,248]. Three studies further examined the subtypes of viruses through sequencing [217,241,249].

The measurement results affirmed the presence of IAV, PCV2, and PSV in the air of swine barns. For IAV, the occurrence frequency of positive samples ranged from 0% (summer samples of Ref. [243]) to 68% (calculated from Ref. [240]) and the measured airborne concentration was up to  $1.25 \times 10^6$  copies  $m^{-3}$  [244]. For PCV2, positive samples ranged from 25% [250] to 91% [216] in occurrence frequency and from  $10^3$  [217] to  $2.42 \times 10^7$  copies  $m^{-3}$  [250] in airborne concentrations. For PSV, only one study was available [136] in which all bioaerosol samples were found PSV-positive and the airborne concentration ranged from  $5.25 \times 10^4$  to  $1.78 \times 10^8$  copies  $m^{-3}$ .

The measurement results affirmed the presence of PRRSV and PEDV in the air of swine barns during and after an outbreak. Ref. [101] reported the absence of airborne PRRSV in uninfected finisher barns. In contrast, 24% [248] to 44% [246] of bioaerosol samples in infected barns were PRRSV-positive. Only one report is available about PEDV measurement [246]. All collected bioaerosol samples were found PEDV-positive. Airborne PRRSV and PEDV concentration data are sporadic, with a single report available [246]. Bioaerosol samples persisted to be PRRSV-positive 14 weeks after an outbreak [248]. No reference (threshold) concentration or dose values were available regarding inhalation exposure to the two viruses.

Five studies measured porcine viruses in outdoor bioaerosol samples. Four of them identified the presence of viruses but at lower concentrations or occurrence frequency than in-barn samples [240–242,246]. Only one study reported the absence of viruses (IAV) in outdoor samples [244]. In particular, Ref. [240] reported an average IAV concentration of  $6.83 \times 10^3$  copies  $m^{-3}$  1.9 km downwind from an IAV-infected commercial farm. Positive outdoor samples suggest the possibility of farm-to-farm airborne viral transmission and the necessity of having filtered air inlets to protect high-value pig herds from airborne porcine viruses.

Little is known regarding the effect of barn types and seasons. Due to the limited data, no statistical analysis was conducted to compare different barn types or seasons. Ref. [216] noted a farrowing barn to be the only PCV2-negative barn during their field survey. Ref. [217] reported that the highest airborne PCV2 concentrations occurred in a grower barn. Ref. [136] compared airborne PSV concentrations between winter and summer samples and found an overall higher concentration in winter. However, Ref. [216] reported that no correlation existed between airborne PCV2 concentrations and outdoor temperatures.

Sampling methods affect measurement results. Ref. [245] reported that higher IAV concentrations were derived from a NIOSH BC251 bioaerosol sampler than from a PHISH sampler. Ref. [246] compared an Andersen eight-stage non-viable cascade impactor with a Tisch four-stage non-viable cascade impactor and found the Tisch yield higher airborne viral concentrations than the Andersen impactor. Ref. [249] compared four different bioaerosol samplers and found that the highest percent of IAV-positive samples came from a NIOSH cyclone sampler and the lowest percent (0%) from a SASS 3100 dry bioaerosol sampler.

Limited information is available regarding the size distribution of airborne porcine viruses. Only two studies conducted size-segregated viral measurement [245,246]. Both studies reported that the majority of airborne viruses (IAV, PRRSV, and PEDV) were associated with coarse PM ( $>4 \mu m$ ). This suggests that these viruses tend not to exist as individual

particles in the air but rather attach to large particles during airborne transmission. For IAV and PEDV, a minor size distribution peak occurred in the submicron range (i.e.,  $<1 \mu\text{m}$ ), which may be attributed to individual viral particles.

Bioaerosol monitoring may serve as a useful tool for field surveillance of porcine viruses. In reality, many field studies took this (tool development) as a goal. In addition to bioaerosol samples, pig nasal swab, oral fluid, and environmental swab samples were often collected. Due to the small volume of bioaerosol samples (and accordingly fewer viral genomic materials), airborne viral detection was less sensitive and required a greater cycle threshold (Ct) than the analysis of other samples (Refer to Section 4.4.2). However, bioaerosol measurement still showed its great potential as it yielded only slightly lower positive counts than pig oral secretion or environmental swab samples [243,244,250], enabled earlier viral detection than blood sample analysis [250], and was able to reflect the onset, peak, and disappearance of a viral infection episode [248,250].

Extensive research has been conducted to mitigate airborne porcine viruses, including mitigation technology development and performance assessment. The former research was predominantly conducted in the lab with artificial viral suspensions or artificially inoculated pigs. For example, [251,252] examined the effectiveness of spraying functional water in reducing airborne PSV, PEDV, and picornaviruses. The test was conducted in an experimental room using a nebulizer to aerosolize viral particles. Other mitigation technology development studies include Refs. [253–256], to name a few. None of them were conducted in actual production barns. The performance of viral mitigation technologies was assessed through lab experiments, epidemiological (cohort) studies [257], and ventilation modeling [258]. The latter two types of studies have been, to date, restricted to filtered air inlet systems (that are increasingly popular in the U.S. Upper Midwest). No field measurement of airborne viruses has been conducted to validate the performance of viral mitigation technologies.

### 3.2.3. Microbial Composition

Different from cell or viral counting, microbial composition (also known as microbial community composition) analysis focuses on the identification of microbial species in PM samples. Although counting can be selective of target microbial species (e.g., *E. coli* or *Salmonella* spp.), it lacks the capability of generating a taxonomic spectrum. Two types of methods are used in microbial composition analysis: culture-dependent and culture-independent methods. Correspondingly, this section is structured into two parts. A review of the relevant methods is available in Section 4.4.3. Culture-dependent methods were commonly used in early investigations. With the advances in molecular biology technologies, most of the recent studies selected culture-independent methods for composition analysis.

#### Culture-Dependent Methods

The interest in PM microbial composition is largely driven by health considerations. Pathogenic bacteria (e.g., *E. coli* and *Salmonella*) and fungi (e.g., mucormycetes (black fungus)) can be transmitted in the air and cause a variety of adverse health effects on humans and animals [13,21]. These pathogens can colonize and infect organs, produce toxins, reproduce, and hijack nutrients only when they are alive. Thus, it is important to identify viable microbes in swine barn PM. However, this is technically difficult, and for decades, culturable microbes have been studied as an alternative.

The effort to identify microbial species in swine barn PM can date back to the 1970s. Ref. [198] measured culturable bacterial counts in a grower–finisher barn in Nebraska. They further identified *Staphylococcus* and *Salmonella* on formed bacterial colonies using selective agars and biochemical testing. In a follow-up study in Nebraska, [200] found that culturable bacteria in the air of swine barns were primarily micrococci. Ref. [35] identified eight bacterial species or genera in the air of swine barns (*Acinetobacter calcoaceticus*, *Alcaligenes odorans*, *Enterobacter agglomerans*, *Enterococcus*, *E. coli*, *Klebsiella*, *Pseudomonas*, and *Pseudomonas syringae*) and found that a large percentage of bacterial colonies comprised of

Gram-positive *Enterococcus*. A fungal species *Aspergillus fumigatus* was also identified using a selective agar. Ref. [36] reported six predominant fungal genera (*Penicillium*, *Alternaria*, *Aspergillus*, *Fucarium*, *Verticillium*, and *Scopulariopsis*) detected in swine barns in Iowa and six predominant fungal genera (*Penicillium*, *Alternaria*, *Aspergillus*, *Rhizopus*, *Cladosporium*, and *Homodendrum*) in Swedish swine barns. Ref. [204] measured bacterial and fungal counts in 79 farms in Sweden including swine barns, and found *Aspergillus* and *Penicillium* to be the predominant fungal genera.

Additional bacterial and fungal species have been identified since 1990. To make a summary manageable, the identification results are organized based on barn types and classified into multiple taxonomic levels (Table 8). No references are given for each identified microbial species (because it would otherwise make the table too wordy). Instead, all the relevant references are listed below the table. Bacterial and fungal species were also detected in settled dust, e.g., [259]. However, it is uncertain whether settled dust and PM share a similar microbial composition because many factors can affect the viability of microbes. Thus, no relevant studies are included in this review effort.

**Table 8.** Microorganisms identified in swine barn PM using culture-dependent methods—A summary of studies since 1990.

Barn Type	Microorganisms Identified <sup>a,b</sup>
Farrowing	<p>Bacteria—                      Phyla: Proteobacteria, Actinobacteria, Firmicutes                      Classes: Gammaproteobacteria, Actinobacteria, Bacilli                      Orders: Pseudomonadales, Enterobacterales, Pasteurellales, Micrococcales, Mycobacteriales, Bacillales, Streptomycetales, Caryophanales, Pseudonocardiales, Streptosporangiales                      Families: Moraxellaceae, Enterobacteriaceae, Pasteurellaceae, Micrococcaceae, Corynebacteriaceae, Staphylococcaceae, Streptomycetaceae, Nocardiaceae, Erwiniaceae, Thermoactinomycetaceae, Pseudonocardiaceae, Thermomonosporaceae,                      Genera: <i>Acinetobacter</i>, <i>Enterobacter</i>, <i>Escherichia</i>, <i>Moraxella</i>, <i>Pasteurella</i>, <i>Pseudomonas</i>, <i>Arthrobacter</i>, <i>Corynebacterium</i>, <i>Staphylococcus</i>, <i>Streptomyces</i>, <i>Rhodococcus</i>, <i>Klebsiella</i>, <i>Erwinia</i>, <i>Thermoactinomycetes</i>, <i>Saccharopolyspora</i>, <i>Saccharomonospora</i>, <i>Thermomonospora</i>                      Species: <i>Acinetobacter calcoaceticus</i>, <i>Enterobacter agglomerans</i>, <i>E. coli</i>, <i>Moraxella</i> spp., <i>Pasteurella</i> spp., <i>Pseudomonas</i> spp., <i>Klebsiella pneumoniae</i>, <i>Klebsiella oxytoca</i>, <i>Erwinia herbicola</i>, <i>Enterobacter cloacae</i>, <i>Thermoactinomycetes vulgaris</i>, <i>Thermoactinomycetes thalophilus</i>, <i>Saccharopolyspora rectivirgula</i>, <i>Saccharomonospora viridis</i>, <i>Thermomonospora fusca</i></p> <p>Fungi—                      Phyla: Ascomycota, Mucoromycota, Basidiomycota                      Classes: Eurotiomycetes, Mucorales, Sordariomycetes, Saccharomycetes, Tremellomycetes, Dothideomycetes                      Orders: Eurotiales, Mucorales, Hypocreales, Saccharomycetales, Microascales, Tremellales, Pleosporales                      Families: Trichocomaceae, Syncephalastraceae, Dipodascaceae, Mucoraceae, Microasceae, Saccharomycetaceae, Trichosporonaceae, Pleosporaceae                      Genera: <i>Aspergillus</i>, <i>Circinella</i>, <i>Fusarium</i>, <i>Geotrichum</i>, <i>Mucor</i>, <i>Penicillium</i>, <i>Scopulariopsis</i>, <i>Candida</i>, <i>Trichosporon</i>, <i>Eurotium</i>, <i>Alternaria</i>, <i>Monilinia</i>                      Species: <i>Aspergillus</i> spp., <i>Circinella</i> spp., <i>Fusarium</i> spp., <i>Geotrichum</i> spp., <i>Mucor</i> spp., <i>Penicillium</i> spp., <i>Scopulariopsis</i> spp., <i>Candida</i> spp., <i>Torulopsis candida</i>, <i>Trichosporon beigelli</i>, <i>Aspergillus fumigatus</i>, <i>Eurotium repens</i>, <i>Alternaria alternata</i>, <i>Monilinia</i> spp.</p>
Gestation	<p>Bacteria—                      Phyla: Firmicutes, Actinobacteria, Proteobacteria                      Classes: Bacilli, Actinobacteria, Gammaproteobacteria,                      Orders: Bacillales, Micrococcales, Enterobacterales, Pseudomonadales                      Families: Bacillaceae, Micrococcaceae, Erwiniaceae, Pseudomonadaceae, Moraxellaceae                      Genera: <i>Staphylococcus</i>, <i>Arthrobacter</i>, <i>Erwinia</i>, <i>Pseudomonas</i>, <i>Acinetobacter</i>                      Species: <i>Staphylococcus equorum</i>, <i>Arthrobacter arilaitensis</i>, <i>Erwinia persicina</i>, <i>Pseudomonas poae</i>, <i>Acinetobacter lwoffii</i></p> <p>Fungi—                      Phyla: Ascomycota                      Classes: Eurotiomycetes, Sordariomycetes                      Orders: Eurotiales, Hypocreales, Onygenales                      Families: Trichocomaceae, Stachybotryaceae, Onygenaceae                      Genera: <i>Penicillium</i>, <i>Stachybotrys</i>, <i>Chrysosporium</i>                      Species: <i>Penicillium</i> spp., <i>Stachybotrys chartarum</i>, <i>Chrysosporium</i> spp.</p>

Table 8. Cont.

Barn Type	Microorganisms Identified <sup>a,b</sup>
Nursery	<p>Bacteria—            Phyla: Proteobacteria, Firmicutes            Classes: Gammaproteobacteria, Bacilli            Orders: Caryophanales, Pasteurellales            Families: Staphylococcaceae, Pasteurellaceae            Genera: <i>Staphylococcus</i>, <i>Actinobacillus</i>            Species: <i>Staphylococcus aureus</i>, <i>Actinobacillus pleuropneumoniae</i></p>
Finisher <sup>c</sup>	<p>Bacteria—            Phyla: Proteobacteria, Actinobacteria, Firmicutes            Classes: Gammaproteobacteria, Actinobacteria, Bacilli, Clostridia            Orders: Pseudomonadales, Enterobacterales, Pasteurellales, Micrococcales, Mycobacteriales, Bacillales, Streptomycetales, Caryophanales, Pseudonocardiales, Streptosporangiales, Bacillales, Lactobacillales, Clostridiales            Families: Moraxellaceae, Enterobacteriaceae, Pasteurellaceae, Micrococcaceae, Corynebacteriaceae, Staphylococcaceae, Streptomycetaceae, Nocardiaceae, Erwiniaceae, Thermoactinomycetaceae, Pseudonocardaceae, Thermomonosporaceae, Bacillaceae, Listeriaceae, Enterococcaceae, Lactobacillaceae, Streptococcaceae, Clostridiaceae, Mycobacteriaceae            Genera: <i>Acinetobacter</i>, <i>Enterobacter</i>, <i>Escherichia</i>, <i>Moraxella</i>, <i>Pasteurella</i>, <i>Pseudomonas</i>, <i>Arthrobacter</i>, <i>Corynebacterium</i>, <i>Staphylococcus</i>, <i>Streptomyces</i>, <i>Rhodococcus</i>, <i>Klebsiella</i>, <i>Erwinia</i>, <i>Thermoactinomycetes</i>, <i>Saccharopolyspora</i>, <i>Saccharomonospora</i>, <i>Thermomonospora</i>, <i>Bacillus</i>, <i>Listeria</i>, <i>Enterococcus</i>, <i>Nocardia</i>, <i>Lactobacillus</i>, <i>Aerococcus</i>, <i>Streptococcus</i>, <i>Clostridium</i>, <i>Mycobacterium</i>, <i>Salmonella</i>            Species: <i>Acinetobacter calcoaceticus</i>, <i>Enterobacter agglomerans</i>, <i>E. coli</i>, <i>Moraxella</i> spp., <i>Pasteurella</i> spp., <i>Pseudomonas</i> spp., <i>Klebsiella pneumoniae</i>, <i>Klebsiella oxytoca</i>, <i>Erwinia herbicola</i>, <i>Enterobacter cloacae</i>, <i>Thermoactinomyces vulgaris</i>, <i>Thermoactinomyces thalophilus</i>, <i>Saccharopolyspora rectivirgula</i>, <i>Saccharomonospora viridis</i>, <i>Thermomonospora fusca</i>, <i>Aerococcus</i> spp., <i>Clostridium difficile</i>, <i>Salmonella</i> spp., <i>Mycobacterium avium</i>, <i>Listeria monocytogenes</i>, <i>Staphylococcus aureus</i></p> <p>Fungi—            Phyla: Ascomycota, Mucoromycota, Basidiomycota            Classes: Eurotiomycetes, Mucorales, Sordariomycetes, Saccharomycetes, Tremellomycetes, Dothideomycetes, Leotiomycetes            Orders: Eurotiales, Mucorales, Hypocreales, Saccharomycetales, Microascales, Tremellales, Pleosporales, Helotiales, Capnodiales            Families: Trichocomaceae, Syncephalastraceae, Dipodascaceae, Mucoraceae, Microascaceae, Saccharomycetaceae, Trichosporonaceae, Pleosporaceae, Sclerotiniaceae, Hypocreaceae, Cordycipitaceae, Davidiellaceae            Genera: <i>Aspergillus</i>, <i>Circinella</i>, <i>Fusarium</i>, <i>Geotrichum</i>, <i>Mucor</i>, <i>Penicillium</i>, <i>Scopulariopsis</i>, <i>Candida</i>, <i>Trichosporon</i>, <i>Eurotium</i>, <i>Alternaria</i>, <i>Monilia</i>, <i>Acremonium</i>, <i>Beauveria</i>, <i>Cladosporium</i>, <i>Scedosporium</i>, <i>Sepedonium</i>, <i>Paecilomyces</i>, <i>Petriella</i>            Species: <i>Aspergillus</i> spp., <i>Circinella</i> spp., <i>Fusarium</i> spp., <i>Geotrichum</i> spp., <i>Mucor</i> spp., <i>Penicillium</i> spp., <i>Scopulariopsis</i> spp., <i>Candida</i> spp., <i>Torulopsis candida</i>, <i>Trichosporon beigelli</i>, <i>Aspergillus fumigatus</i>, <i>Eurotium repens</i>, <i>Alternaria alternata</i>, <i>Monilia</i> spp., <i>Aspergillus flavus</i>, <i>Aspergillus terreus</i>, <i>Aspergillus versicolor</i>, <i>Scedosporium apiospermum</i>, <i>Candida dubliniensis</i></p>
Unspecified <sup>d</sup>	<p>Bacteria—            Phyla: Proteobacteria, Actinobacteria, Firmicutes            Classes: Gammaproteobacteria, Actinobacteria, Bacilli            Orders: Pseudomonadales, Micrococcales, Mycobacteriales, Streptomycetales, Bacillales, Actinomycetales            Families: Moraxellaceae, Micrococcaceae, Corynebacteriaceae, Streptomycetaceae, Bacillaceae, Actinomycetaceae            Genera: <i>Acinetobacter</i>, <i>Arthrobacter</i>, <i>Corynebacterium</i>, <i>Endomycopsis</i>, <i>Streptomyces</i>, <i>Bacillus</i>, <i>Actinomyces</i>            Species: <i>Acinetobacter calcoaceticus</i>, <i>Arthrobacter globiformis</i>, <i>Corynebacterium xerosis</i>, <i>Endomycopsis capsularis</i>, <i>Streptomyces</i> spp.</p> <p>Fungi—            Phyla: Ascomycota, Zygomycota, Mucoromycota, Basidiomycota            Classes: Saccharomycetes, Dothideomycetes, Zygomycetes, Eurotiomycetes, Leotiomycetes, Saccharomycetes, Pleosporomycetidae, Ascomycetes, Sordariomycetes, Agaricomycetes, Dothideomycetes            Orders: Endomycetales, Capnodiales, Mucorales, Pleosporales, Eurotiales, Helotiales, Eurotiales, Saccharomycetales, Mucorales, Pleosporales, Incertae sedis, Hypocreales, Agaricales, Erysiphales, Dothideales            Families: Endomycetaceae, Davidiellaceae, Cunninghamellaceae, Pleosporaceae, Trichocomaceae, Sclerotiniaceae, Trichocomaceae, Saccharomycetaceae, Mucoraceae, Pleosporaceae, Incertae sedis, Nectriaceae, Typhulaceae, Dipodascaceae, Erysiphaceae, Dothioraceae            Genera: <i>Endomycopsis</i>, <i>Cladosporium</i>, <i>Absidia</i>, <i>Alternaria</i>, <i>Aspergillus</i>, <i>Botrytis</i>, <i>Cladosporium</i>, <i>Eurotium</i>, <i>Candida</i>, <i>Mucor</i>, <i>Penicillium</i>, <i>Trichoderma</i>, <i>Ulocladium</i>, <i>Cephalosporium</i>, <i>Fusarium</i>, <i>Curvularia</i>, <i>Sclerotium</i>, <i>Geotrichum</i>, <i>Drechslera</i>, <i>Oidium</i>, <i>Aureobasidium</i>, <i>Stemphylium</i>, <i>Monilia</i>, <i>Paecilomyces</i>            Species: <i>Endomycopsis capsularis</i>, <i>Aspergillus niger</i>, <i>Aspergillus flavus</i>, <i>Aspergillus circumdatus</i>, <i>Aspergillus aspergillus</i>, <i>Aspergillus versicolor</i>, <i>Aspergillus ustus</i>, <i>Aspergillus candidus</i>, <i>Aspergillus nidulan</i>, <i>Cladosporium</i> spp., <i>Penicillium</i> spp., <i>Fusarium culmorum</i></p>

<sup>a</sup> The lists were compiled primarily from [43,51,54,59,76,87,152,158,159,184]. Several publications in Table 5 (e.g., [174,178]) used selective agar media to identify and quantify certain microorganisms. Although they were not truly microbial composition analyses, the identified microorganisms are also included in the table. <sup>b</sup> A microorganism unreported in a barn type does not exclude the possibility of its existence. This could simply be because no relevant analysis was conducted in that barn type. <sup>c</sup> For simplicity, finisher barns here include grower–finisher and wean-to-finish barns. <sup>d</sup> The category also includes the studies with multiple barn types (including gestation barns) but providing no barn type-specific microbial composition information.

The following findings are made upon the analysis of the previous publications:

- Although most of the identified microorganisms are unharmed, pathogenic or potentially pathogenic bacteria and fungi were occasionally identified in swine barns. These include bacterial species *Acinetobacter lwoffii*, *E. coli*, *Enterobacter cloacae*, *Klebsiella*

*pneumonia*, *Klebsiella oxytoca*, *Listeria monocytogenes*, *Pseudomonas* spp., *Staphylococcus aureus*, and *Salmonella* spp., bacterial genera *Acinetobacter*, *Actinomyces*, *Aerococcus*, *Bacillus*, *Clostridium*, *Corynebacterium*, *Enterobacter*, *Enterococcus*, *Escherichia*, *Klebsiella*, *Listeria*, *Moraxella*, *Mycobacterium*, *Pasteurella*, *Pseudomonas*, *Staphylococcus*, *Streptococcus*, and *Salmonella*, fungal species *Aspergillus* spp., *Aspergillus niger*, *Aspergillus flavus*, *Aspergillus circumdatus*, and *Aspergillus versicolor*, and fungal genera *Aspergillus*, *Candida*, and *Trichosporon*. The information of pathogens here is acquired from Hartmann Science Center, Pathogen Search A-to-Z (<https://www.bode-science-center.com/center/relevant-pathogens-from-a-z.html>) (accessed on 30 August 2021).

- The majority of bacteria identified in swine barn PM are Gram-positive bacteria. The Gram-negative bacteria observed include genera *Acinetobacter*, *Enterobacter*, *Escherichia*, *Erwinia*, *Klebsiella*, *Moraxella*, *Pasteurella*, *Pseudomonas*, and *Salmonella*. Although Gram-negative bacteria account for only a small fraction of the bacterial community, they include many known pathogenic bacterial strains, such as *Pseudomonas aeruginosa*, *E. coli*, and *Salmonella* spp. Thus, they were often counted separately (Table 5) and identified at genus or species levels [43].
- Quantitative analysis was occasionally conducted by counting the colonies assigned to be a specific species or group. Ref. [43] examined three farrowing and two finisher barns in Poland for the relative abundance of different mesophilic bacteria and found *Corynebacteria* to be the most abundant. A similar comparison was conducted for Gram-negative bacteria, thermophilic actinomycetes, and fungi. Ref. [51] compared the colony counts of six microbial groups among farms and found a significant effect of geographic locations. At two locations, the microbial community was dominated by *Corynebacteria*, while at one location, Gram-positive micrococci were predominant. Gram-negative bacteria accounted for <3% of culturable microbial counts. Ref. [158] studied airborne fungal composition in various swine barns in Taiwan and found the dominance of genus *Cladosporium* (>92%) in all visited barn types. Ref. [159] reported higher counts of *Staphylococcus* than other identified bacterial genera (*Pseudomonas*, *Bacillus*, *Listeria*, *Enterococcus*, *Nocardia*, and *Lactobacillus*) in grower–finisher barns in Kansas. Ref. [87] reported the dominance of bacterial phylum Firmicutes and fungal phylum Ascomycota in the air of a Danish gestation barn. Again only a portion of viable microbes is culturable. Thus, the quantitative information derived from culture-dependent methods may not represent the actual composition of viable microbes.
- Little is known regarding the effect of barn types. Only a few studies compared PM microbial composition in different types of barns, likely because microbial counting and identification require intensive time and labor input. Ref. [43] compared farrowing barns with finisher barns in Poland. While no considerable difference was seen in terms of mesophilic bacteria or thermophilic actinomycetes, a substantial difference in Gram-negative bacteria and fungi was observed between the two barn types. Specifically, the visited finisher barns had a lower abundance of *Acinetobacter calcoaceticus*, *Erwinia Herbicola*s, *E. coli*, *Klebsiella* spp., and *Pseudomonas* spp., but a higher abundance of yeast than farrowing barns. Ref. [158] measured airborne fungal composition in five types of barns. The measured fungal composition differed with barn types. However, due to limited data points, it is uncertain whether the difference was statistically significant.

Although numerous efforts have been made to measure PM microbial composition with culture-dependent methods, several questions remain unanswered. For example, no studies have been conducted regarding the effects of seasons, ventilation systems, and waste management systems, even though their effects on PM microbial composition are anticipated. From a disease transmission standpoint, the PM size of bacterial or fungal pathogens is of critical importance. However, no size-segregated analysis of PM microbial composition (e.g., microflora within 1.1–2.1  $\mu\text{m}$ ) has been conducted. Again, this is likely due to the intensive time and labor required for culture-dependent methods. Some of these questions have been addressed by studies using culture-independent methods.

### Culture-Independent Methods

Culture-independent methods can be classified into two categories: genomics and proteomics. The genomic methods derive microbial species information from the analysis of genetic materials extracted from swine barn PM samples; while the proteomic methods achieve it through fingerprinting proteins and peptides extracted from the PM samples. Most previous studies used genomic methods to analyze microbial composition in swine barn PM. Only a few recent studies used proteomic methods. Under the category of genomics methods, various technologies have been adopted, such as clone library and next-generation pyrosequencing (NGS). For simplicity, the findings acquired from different technologies are discussed collectively in this section.

It is a daunting challenge to summarize microbial composition analysis results from the previous studies because culture-independent methods often generate massive amounts of data. For example, Ref. [209] (a study we participated in) used 16S rRNA NGS to analyze bacterial composition in TSP samples from six poultry barns and 14 swine barns. The study generated 214,795 sequences. Even after bioinformatics analysis, the amount of taxonomic data it created was still formidable. To make the review manageable, no exhaustive list of identified microbial species is provided. Instead, key findings from each study were summarized in Table 9. Following that, a summary and discussion of the existing findings are provided. Microbial species were also identified in settled dust using culture-independent methods, e.g., [260] (Note: The study is academically important as it was the first study using shotgun metagenomic pyrosequencing for the analysis of swine barn dust). However, for a potential difference in microbiota between PM and settled dust, no relevant studies are included. Quantitative information, often presented as relative abundance, was available in many publications. The relative abundance of a microbe was measured by the percentage of its counts, sequences, or clones in total identified bacteria or fungi.

**Table 9.** Microbial composition in swine barn PM—A summary of studies using culture-independent methods.

Reference	Barn Type	Location	Methods <sup>a</sup> and Target Microbes	Major Findings and/or Notes
Proteomic methods [261]	Finisher	Germany	MALDI-TOF MS for bacteria	A reference database was created by comparing proteomics results against 16S rRNA sequencing results. Using the database, 18 bacterial species were identified, including five risk group 2 pathogens ( <i>Aerococcus viridians</i> , <i>Corynebacterium striatum</i> , <i>Staphylococcus epidermidis</i> , <i>Staphylococcus pasteurii</i> , and <i>Staphylococcus saprophyticus</i> ). Two PM size fractions (PM <sub>20</sub> and PM <sub>5</sub> ) were examined.
[210] <sup>b,c</sup>	Farrowing, nursing <sup>d</sup> , nursery, finisher	Denmark	MALDI-TOF MS for bacteria (aerobes and anaerobes) and fungi	The study used MALDI-TOF MS to identify colony isolates. With that, 120 (96 + 24) bacterial species and 27 fungal species were identified. Many of them were classified into species levels. The identified bacteria and fungi included 28 risk group 2 pathogens.
[192] <sup>c</sup>	Finisher	Denmark	MALDI-TOF MS for fungi	The study used MALDI-TOF MS to identify colony isolates. With that, 40 fungal species were identified, including 16 allergens and five risk group 2 pathogens. Size distribution of these species was also available. The majority of fungi occurred in the size range of 1.1–3.3 µm. However, it differed with species.

Table 9. Cont.

Reference	Barn Type	Location	Methods <sup>a</sup> and Target Microbes	Major Findings and/or Notes
Genomic methods [169]	Grower–finisher	Quebec, Canada	16S rRNA DGGE and clone library for bacteria	A total of 245 sequences (clones) were generated from selected DGGE bands. They, along with blast analysis of DGGE bands, indicated the existence of <i>Aerococcus</i> spp., <i>Lactobacillus</i> spp., <i>Streptococcus</i> spp., <i>Bacillus</i> spp., <i>Anaerococcus</i> spp., <i>Clostridium chauvoei</i> , and <i>Clostridium litusburense</i> , among many others. Bacterial composition was dominated by three classes: Clostridia, Bacteroidetes, and Lactobacillales.
[262]	Grower–finisher	Quebec, Canada	16S rRNA DGGE and clone library for archaea	A total of 566 sequences were generated from selected DGGE bands. They suggested the dominance (>94.5%) of <i>Methanosphaera stadtmanea</i> . <i>Methanosarcina siciliae</i> were also detected.
[209] <sup>e</sup>	Farrowing, gestation, nursery, finisher	Illinois	16S DGGE and rRNA NGS for bacteria	Phylum Firmicutes were dominant, followed by Bacteroidetes. Actinobacter were the third most abundant phylum in farrowing and gestation barns. PM samples from nursery and finisher barns contained a significantly higher abundance of <i>Prevotella</i> , <i>Roseburia</i> , <i>Faecalibacterium</i> , <i>Megaspaera</i> , and <i>Subdoligranulum</i> spp. than those from farrowing and gestation barns. Twelve identified genera were potentially associated with pathogens. Swine barn PM samples showed overall a significantly different bacterial composition than those from poultry barns and non-farm office environments.
[87] <sup>e</sup>	Gestation	Denmark	16S rRNA clone library and FISH for bacteria 18S rRNA clone library for fungi FISH for archaea	Archaea were identified by FISH but only accounted for 0.3% of total microbial counts. Sixty-two sequences were generated for bacteria. They were classified into 15 operational taxonomic units (OTUs) and were dominated by phylum Firmicutes and genus <i>Clostridium</i> . The 73 fungal sequences generated were grouped into six OTUs and were dominated by genera <i>Aspergillus</i> and <i>Eurotium</i> .
[217] <sup>e</sup>	Nursery, grower, finisher, Wean-to-finish	Colorado	16S rRNA clone library for bacteria	A total of 810 sequences were generated for in-barn PM samples. Phylum Firmicutes were predominant, followed by Bacteroidetes. In-barn samples showed a significantly different bacterial composition than outdoor samples. Within Firmicutes, the dominant species included <i>Bacillus</i> spp., <i>Lachnospiraceae</i> spp., and <i>Clostridium</i> spp. Some identified species were also found in pig gastrointestinal tracts.
[218]	Grower–finisher	South Korea	16S rRNA NGS for bacteria	A total of 497,607 sequences were generated, classed into 13,597 OTUs. Firmicutes were the most abundant phylum, followed by Bacteroidetes and Proteobacteria. At the genus level, <i>Lactobacillus</i> and <i>Prevotella</i> were dominant. A significantly different bacterial composition was found between winter and summer samples, with winter samples having more OTUs (i.e., greater richness) and a higher abundance of <i>Prevotella</i> .
[263]	n/a	North Carolina	16S rRNA NGS for bacteria	Note: PM samples were collected 5 m away from a barn. The 2364 sequences generated were classified into 441 OTUs. Phylum Proteobacteria were predominant, followed by Actinobacteria and Bacteroidetes. At the genus level, <i>Sphingomonas</i> and <i>Hymenobacter</i> were most abundant.

Table 9. Cont.

Reference	Barn Type	Location	Methods <sup>a</sup> and Target Microbes	Major Findings and/or Notes
[219]	Grower–finisher	South Korea	16S rRNA NGS for bacteria	A total of 14,315 sequences were generated, classified into 976 OTUs. Firmicutes were the predominant phylum. The relative abundances of Firmicutes, Actinobacteria and Proteobacteria differed with waste management systems (deep pits, beddings, and scraper). Bedding systems showed a significantly higher abundance of <i>Corynebacterium</i> than other systems. Manure scraper systems had over the lowest OTUs.
[220]	Grower–finisher	South Korea	ITS NGS for fungi	A total of 22,399 OTUs were identified. Winter samples had different fungal compositions and greater diversity than summer samples. At the phylum level, Ascomycota were the most abundant, followed by Basidiomycota and Zygomycota. The most abundant genera were <i>Clavaria</i> and <i>Fusarium</i> . Twenty-nine potential pathogens or allergens were found. Significant barn-to-barn and within-barn variations were found in fungal composition.
[261]	Finisher	Germany	16S rRNA clone library for bacteria	The study used 16S rRNA clone library to identify colony isolates. <sup>p</sup> With that, 65 sequences were generated and classified to 21 bacterial OTUs.
[264]	n/a	Switzerland	16S rRNA NGS for bacteria	The richness and diversity of bacteria in swine barn PM were greater than those in pigs' nasal swabs but lower than those in farmer's nasal swabs. Regarding bacterial composition, PM sat somewhere in-between pigs' and pig farmers' nasal swabs. Farmers from the same barn tended to share similar nasal microbiota, suggesting a significant influence of in-barn PM.
[265]	n/a	Denmark	16S rRNA NGS for bacteria	16S rRNA genes in PM were more abundant but less diverse in swine barns than farmer's homes. Bacterial community was dominated by phylum Firmicutes, followed by Actinobacteria and Proteobacteria. The most abundant bacteria were Clostridiales at the order level and Clostridiaceae and Peptostreptococcaceae at the family level. No significant seasonality was noted in bacterial composition or diversity.
[221] <sup>e</sup>	n/a	Switzerland	16S rRNA NGS for bacteria	The richness and diversity of bacteria varied significantly with seasons and they were greater in winter. Phylum Firmicutes were predominant, followed by Actinobacteria and Proteobacteria. Dominant genera included <i>Veillonellaceae</i> , <i>Clostridiaceae</i> , <i>Lactobacillaceae</i> , and <i>Peptostreptococcaceae</i> . About 45–65% of pig farmers' nasal microbiome was from PM and the percentage was greater in winter and lower in summer.
[266]	Finisher	Quebec, Canada	16S rRNA NGS for bacteria	Phylum Firmicutes were the most abundant bacterial group, followed by Bacteroidetes and Actinobacteria. At the class level, Clostridia Bacilli, and Bacteroidia were dominant. Microbial composition in PM samples resembled that in farm workers' nasal swabs.
[210] <sup>b</sup>	Farrowing, nursing <sup>d</sup> , nursery, finisher	Denmark	16S rRNA NGS for bacteria ITS NGS for fungi	A total of $4.0 \times 10^5$ bacterial $1.5 \times 10^5$ fungal sequences were generated. The bacterial community was dominated by genera <i>Clostridium sensu stricto</i> , <i>Lactobacillus</i> , <i>Terrisporobacter</i> , <i>Turicibacter</i> , <i>Romboutsia</i> , <i>Methanobrevibacter</i> , <i>Aerococcus</i> , and <i>Weissella</i> . Dominant fungal genera were <i>Filobasidium</i> , <i>Apiotrichum</i> , <i>Wallemia</i> , and <i>Candida</i> . A significant effect of barn type was observed on fungal and bacterial compositions.

Table 9. Cont.

Reference	Barn Type	Location	Methods <sup>a</sup> and Target Microbes	Major Findings and/or Notes
[267] <sup>e</sup>	Farrowing, gestation, nursery, grower–finisher, boar	China	Shotgun metagenomic sequencing for microbiota	Most genes (>88.8%) were assigned to bacteria. Firmicutes were the most abundant bacterial phylum, followed by Bacteroidetes, Actinobacteria, and Proteobacteria. At the genus level, <i>Prevotella</i> , <i>Clostridium</i> , <i>Corynebacterium</i> , <i>Bacteroides</i> , and <i>Ruminococcus</i> are abundant. Bacterial composition differed with barn types. Bacteria were also classified based on functional characteristics (functional genes). Archaea, viruses, and eukaryotes were detected but not discussed. Only five composite samples were analyzed.
[268]	n/a	Netherlands	16S rRNA NGS for bacteria ITS NGS for fungi	PM <sub>2.5–10</sub> in two pig barns was sampled for analysis. For bacteria, <i>Lactobacillus</i> and <i>Clostridium</i> were the most abundant at the genus level. For fungi, genus <i>Emmericella</i> was the most abundant, followed by genera <i>Penicillium</i> and <i>Candidia</i> . Swine barns showed significantly different PM bacterial or fungal compositions than chicken and goat barns.
[269] <sup>f</sup>	n/a	Nine European countries	Shotgun metagenomic sequencing for bacteria	At the class level, Clostridia were the most abundant, followed by Bacilli, Bacteroidia, Actinobacteria, Betaproteobacteria, and Erysipelotrichia.
[270] <sup>e</sup>	Nursery	China	16S rRNA NGS for bacteria ITS NGS for fungi	PM <sub>2.5</sub> was sampled for analysis. Thirty-three bacterial phyla and 460 genera were identified. At the phylum level, Firmicutes were the most abundant, followed by Bacteroidetes. At the genus level, <i>Lactobacillus</i> was the most abundant. Eleven fungal phyla and 317 genera were identified, with Basidiomycota being the predominant phylum. A significant seasonality was seen in both bacterial and fungal compositions, as well as microbial diversity.
[192]	Finisher	Denmark	ITS NGS for fungi	A total of 334,862 fungal sequences were generated. They were classified into 59 genera (Note: Relevant information is available in the Supplementary Materials of the paper).
[271]	Nursery, finisher	South Korea	16S rRNA NGS for bacteria	A total of 351,016 sequences were generated. Firmicutes were the most abundant phylum, followed by Proteobacteria and Bacteroidetes. <i>Clostridium</i> was the most abundant genus, followed by <i>Terrisporobacter</i> and <i>Turcibacter</i> . Four genera ( <i>Curvibacter</i> , <i>Sediminibacterium</i> , <i>Bradyrhizobium</i> , and <i>Pelomonas</i> ) were associated with potential pathogens.
[272]	n/a	China	16S rRNA NGS for bacteria	At the phylum level, Firmicutes were predominant, followed by Bacteroidetes and Proteobacteria. At the order level, Clostridia was predominant, followed by Lactobacillales and Bacteriales. At the genus level, <i>Clostridium</i> and <i>Streptococcus</i> were the most abundant. Summer samples contained a high abundance of opportunistic pathogens.
[273]	Nursery	China	16S rRNA NGS for bacteria	A total of 300 bacterial genera were identified. Bacterial composition differed with size fractions (six fractions). The dominant phylum was Bacteroidetes in >7 and 4.7–7.0 µm, Firmicutes in 3.3–4.7 µm, and Fusobacteria in 1.1–2.1 and 2.1–3.3 µm. Note: Only one set of PM samples was sequenced.

Table 9. Cont.

Reference	Barn Type	Location	Methods <sup>a</sup> and Target Microbes	Major Findings and/or Notes
[274] <sup>e</sup>	Farrowing, gestation, nursery, grower–finisher	China	16S rRNA NGS for bacteria	A total of 70,763 bacterial sequences were generated, classified into 2643 OTUs. Bacterial composition showed a significant effect by barn types. Proteobacteria was the most abundant phylum, followed by Actinobacteria, Bacteroidetes, and Firmicutes. <i>Acinetobacter</i> was the most abundant genus in nursery and grower–finisher barns, while <i>Psychrobacter</i> and <i>Rothia</i> were dominant genera in farrowing and gestation barns. Three potentially pathogenic genera were detected.

<sup>a</sup> MALDI-TOF MS—matrix-assisted laser desorption/ionization time-of-flight mass spectrometry; DGGE—denaturing gradient gel electrophoresis. <sup>b</sup> Although the title of the paper said settled dust, PM (dust) samples were collected using an electrostatic precipitator (ESP). <sup>c</sup> Strictly speaking, the method was culture-dependent because microbial identification was conducted on colony isolates. As the method generated substantially more information than traditional culture-dependent identification methods (colony appearance-based identification, biochemical testing, and selective/differential media), the research results are included in the table for comparison. <sup>d</sup> Nursing barns are a barn type in certain western European countries. The barns separate nursing from farrowing activity. <sup>e</sup> Sequencing data were uploaded on Genbank, NCBI, and alike, allowing for post-hoc analysis by others. Other studies could also have archived microbial composition data online; however, no relevant information or data link was provided. <sup>f</sup> Relevant information was retrieved from the publication’s Supplementary Materials.

Upon the analysis of the above publications, the following summary and observations are made:

- Microbial composition analysis of swine barn PM with culture-independent methods has received increasing attention since the first report by Ref. [169]. Out of the 24 papers compiled in the table, 14 were published in the past three years (2018 to present). This has not included non-English papers. Besides the pioneering work of Ref. [169], three milestone papers are worth mentioning from the methodology standpoint: (1) Ref. [209], for the first report of using NGS to study microbial composition in swine barn PM; (2) Ref. [220], for the first use of ITS sequencing for fungal composition analysis in swine barns; and (3) Ref. [267], for the first attempt of using shotgun metagenomic pyrosequencing for analysis of microbiota in swine barn PM (Note: Ref. [260] used the same technology for settled dust). A brief overview of these methods is available in Section 4.4.3.
- Regarding the geographic distribution of relevant studies, nine papers were from Asia (China and South Korea), nine from Europe, three from the U.S., and three from Canada. Twenty papers studied bacterial composition, six studied fungi, and one studied archaea. Seventeen papers used NGS for microbial identification, five used clone library, and three used MALDI-TOF MS. Nine studies each analyzed  $\geq 20$  PM samples [192,209,210,220,221,264,265,269,274]. Others had relatively small sample sets. Finisher barns were the most studied, followed by nursery barns. Regardless of barn types, identification methods, and farm locations, many studies reached similar observations about PM microbial composition in swine barns.
- Bacteria and fungi in swine barn PM are highly diverse. Numerous species have been identified, especially with NGS and shotgun metagenomic sequencing—both technologies generated a large number of reads. For bacteria, up to 13,597 OTUs [218] and 460 genera [270] were identified; while for fungi, the identified OTUs and genera were up to 22,399 OTUs [220] and 317 [270], respectively. An even larger number of OTUs or genera could exist in the literature, but it requires a thorough inspection of raw data. It is noteworthy that many factors could affect the number of identified species, such as volume of PM samples, selection of primers, depth of sequencing, and classification criteria. Several studies used the number of OTUs predicted from rarefaction curves (OTUs versus sequences) to enable a reasonable comparison among samples [192,209,210].

- Firmicutes, the majority of which are Gram-positive, were reported by many to be the predominant (typically  $\geq 60\%$ ) bacterial phylum in swine barn PM. Other abundant bacterial phyla included Actinobacteria, Bacteroidetes, and Proteobacteria. One exception is Ref. [274], in which Proteobacteria was found to be the most abundant. However, in an earlier report by the same authors, Firmicutes were dominant [267]. At the genus level, *Clostridium* and *Lactobacillus* often accounted for significant fractions. Other abundant genera included *Bacteroides*, *Corynebacterium*, *Peptostreptococcaceae*, *Prevotella*, and *Terrisporobacter*; however, their abundance lacks the same degree of universality as that of *Clostridium* and *Lactobacillus*. This could be attributed to differences in factors such as geographical locations, barn types, waste management, and sampling seasons. It is noteworthy that *Clostridium*, a genus in phylum Firmicutes, contains several important pathogens. *Clostridium*, *Lactobacillus*, *Prevotella*, and *Bacteroides* were usually associated with pig gastrointestinal tracts and fecal microbiota, suggesting that a significant portion of airborne bacteria in swine barns are of fecal origins [169,209,219,267,270].
- Many bacterial pathogens or potential pathogens were identified. A complete list was usually unavailable in the literature due to the large number of identified species. A partial list, including opportunistic and potential pathogens, is as follows: genera *Aerococcus*, *Acinetobacteria*, *Arcobacter*, *Bacillus*, *Campylobacter*, *Clostridium*, *Erysipelothrix*, *Escherichia*, *Fusobacterium*, *Helicobacter*, *Leptotrichia*, *Moraxella*, *Pseudomonas*, *Salmonella*, *Staphylococcus*, *Streptococcus*, and *Treponema*; and species *Aerococcus viridans*, *Bacillus cereus*, *Clostridium perfringens*, *Clostridium bifermentans*, *Clostridium cadaveris*, *Clostridium baratii*, *Enterococcus avium*, *Enterococcus casseliflavus*, *Enterococcus durans*, *Enterococcus faecalis*, *Enterococcus villorum*, *Enterococcus gallinarum*, *Enterococcus hirae*, *Enterococcus faecium*, *Filobasidium untiguttulatum*, *Leuconostoc mesenteroides*, *Serratia marcescens*, *Staphylococcus aureus*, *Staphylococcus saprophyticus*, *Staphylococcus paseuri*, *Staphylococcus haemolyticus*, *Staphylococcus hominis*, *Staphylococcus hyicus*, *Staphylococcus pettenkoferi*, *Staphylococcus epidermidis*, *Streptococcus parauberis*, *Streptococcus lutetiensis*, and *Vagococcus fluvialis* [209,210,217,261,263,270–272,274].
- Out of the six fungal studies, five reported Ascomycota and one found Basidiomycota to be the predominant phylum (division). The abundant fungal genera included *Aspergillus*, *Eurotium*, *Clavaria*, *Fusarium*, *Filobasidium*, *Apiotrichum*, *Wallemia*, and *Candida*; and their presence and/or relative abundance differed among publications. Several potential fungal pathogens or allergens were identified, including genera *Fusarium*, *Aspergillus*, *Penicillium*, *Schizophyllum*, *Trichoderma*, *Wallemia*, *Humicola*, and *Sporobolomyces*; and species *Aspergillus fumigatus*, *Aspergillus niger*, *Aspergillus terreus*, *Trichosporon asahii*, *Trichosporon cutaneum*, *Alternaria alternata*, *Aureobasidium pullulans*, *Cladosporium cladosporoides*, *Cladosporium herbarum*, and *Schizophyllum commune* [192,210,220,270].
- Barn types affect bacterial and fungal compositions. Ref. [209] compared the bacterial composition of TSP sampled from four types of swine barns. Two clusters were identified: (1) farrowing and gestation and (2) nursery and finisher. The latter had higher abundances of *Prevotella*, *Roseburia*, *Faecalibacterium*, *Megasphaera*, and *Subdoligranulum* spp. than the former barns. Ref. [210] found that both bacterial and fungal compositions varied with barn types; however, farm locations exhibited an even greater influence. For barns of different types but situated at the same farm sites, they shared a similar PM microbial composition. A significant influence of barn types on bacterial composition was reported by Refs. [267,274]. PM samples from finisher barns contained more Firmicutes but lesser Actinobacteria than those from farrowing and gestation barns. At the genus level, finisher barn samples had a higher abundance of *Aerococcus* but lower *Kocuria* than gestation barns. Within the same barn type, PM bacterial composition differed with selected waste management systems [219].
- No agreement has yet been reached regarding the effect of seasons. A significant seasonal variation in bacterial or fungal composition was reported by Refs. [218,220,221,270];

however, Ref. [265] found no significant seasonality in PM bacterial composition. Temperature is a major shaping factor for airborne microbial communities [275]. In-barn air temperature varies with seasons even with ventilation, heating, and/or cooling systems. The variation, however, is relatively small as compared with outdoor environments. Other biotic and abiotic factors (e.g., air humidity and animal age) than temperature could affect microbial communities, causing uncertainties in observed seasonal patterns. Regarding the specific effects of seasons, Ref. [272] reported that bacterial phyla Proteobacteria and Actinobacteria were more abundant in winter than summer samples. Refs. [218,220] found that winter samples had a greater level of microbial richness (i.e., more OTUs) than summer samples. An opposite finding was reported by Refs. [270,272]. It should be noted that PM<sub>2.5</sub> (rather than TSP) was sampled in Ref. [270]. A large portion of PM<sub>2.5</sub> in swine barns could originate from ambient PM [24], with different seasonal patterns than PM generated in barns.

- Limited information is available regarding the size distribution of microbial communities. Only two recent studies analyzed the microbial composition of size-segregated PM samples. Ref. [192] collected PM samples of different size ranges using Andersen multistage cascade impactors and examined the fungal composition of these samples. Most *Wallemia* spp. was found in PM of 0.65–2.1 µm; while, the majority of *Candida catenulate* occurred in PM of 1.1–7 µm. Fungal genera *Cladosporium*, *Malassezia*, and *Kazachstania* were detected in all size ranges. Ref. [273] reported that genus *Mycoides* dominated the bacterial community in PM of >7.0 µm and 4.7–7.0 µm and genus *Escherichia-Shigella* was most abundant in PM of 1.1–2.1 and 2.1–3.3 µm. In comparison, PM of 3.3–4.7 µm showed the greatest bacterial diversity, with four abundant bacterial genera in it: *Burkholderia-Caballeronia-Paraburkholderia*, *Streptococcus*, *Actinobacillus*, and *Veillonella*.

Culture-independent methods are becoming increasingly affordable and accessible. Ten years ago, the analysis of bacterial composition with NGS cost USD ~100 per sample [209]. Today it costs only USD ~18 per sample, according to Illumina, Inc., an industrial leader of NGS technology [276]. This has been less expensive than many regular physical and chemical tests of PM samples. In addition, many universities now offer low-cost or free bioinformatics services. Reduced cost, along with a growing interest in bioaerosols, leads to the booming of relevant publications in the past few years. Massive amounts of data have been generated (Table 9), which, however, raises a great challenge to data analysis and utilization (e.g., translating the data into the knowledge that average pork producers can utilize). Meanwhile, some research questions remain unanswered or unclarified, e.g., how do PM microbial compositions vary before, during, and after a swine disease outbreak, and how do pathogenic viruses correlate with bacterial and/or fungal linkages? Microbial composition could also be related to other air quality parameters. For example, our previous study found a significant correlation between PM-borne odorants and microbial composition (unpublished data) [209,277]. Additional efforts, thus, are needed to examine and interpret PM microbial composition in swine barns.

#### 3.2.4. Antibiotic Resistant Bacteria (ARB) and Antibiotic Resistance Genes (ARGs)

The use of antibiotics is critical for successful pork production. It also raises public concerns about the spread of ARB and ARGs from farms to the environment which could compromise the ability to curb bacterial illnesses. A renowned example of ARB is MRSA, touted by some as “superbugs”. It has been detected and quantified in the air of swine barns (Table 5). Antibiotic resistance can be acquired by other bacteria in the environment through horizontal gene transfer, a reason for the importance of ARG analysis [278]. In the U.S., ~80% of antibiotics are used by animal agriculture, including swine barns [279]. Although pork producers have made tremendous efforts to restrict the selection and dosage of swine antibiotics, public concerns and pressures still linger. In the past 20 years, numerous studies have been conducted to examine ARB and ARGs in and around swine barns.

Swine manure is believed to be a main source for the propagation of ARB and ARGs in the environment. Their presence in swine manure, waste management systems, manure-fertilized soils, and downstream waters has been extensively studied [280,281]. As PM in swine barns originates partially from manure, it can serve as a carrier of ARB and ARGs and disperse them into the environment. It is important to recognize that airborne transmission is different from water- or soil-borne transmission in that it is not restricted by watersheds or manure sheds (defined as the land area where the manure is applied) and, thus, may disperse ARB and ARGs further away from farms. To our knowledge, the first observation of airborne ARB in swine barns was reported by Ref. [282]. Since then, 24 publications have been available regarding ARB and ARGs in swine barn bioaerosols.

A relevant finding is the occurrence of PM-borne antibiotics in swine barns. In a 20-year study, fourteen antibiotics were detected in swine barn PM samples and six of them (tetracycline, oxytetracycline, chlortetracycline, tylosin, chloramphenicol, and sulfamethazine) were quantified using high-performance liquid chromatography (HPLC) [283,284]. However, it remains unclear whether and how these PM-borne antibiotics would correlate with airborne ARB and ARGs.

#### Antibiotic Resistant Bacteria

ARB were usually measured with culture-dependent methods through the use of selective or differential cultivation media. Commercial selective or differential agars are available, e.g., a CHROMagar MRSA chromogenic agar for MRSA detection in swine barns [101,183]. Self-prepared cultivation media were used in four studies to examine bacterial colonies for their resistance to multiple antibiotics, following the minimal inhibitory concentration (MIC) agar dilution method [285] or the Kirby–Bauer diffusion disk method [160,163,286]. The measurement results include qualitative (presence or absence) and quantitative ones (CFU m<sup>-3</sup>) (Table 10). Settled dust samples were also analyzed [287,288] but relevant studies were excluded for possible differences in microbiota between PM and settled dust.

**Table 10.** Airborne antibiotic resistant bacteria in swine barns.

Reference	Barn Type	Location	Target ARB	Major Findings and/or Notes
[282]	Finisher	Iowa	Tylosin-resistant bacteria	The average concentration ranged from 49,400 to 16,700 CFU m <sup>-3</sup> , accounting for ~80% of culturable bacterial counts. The vast majority of bacterial isolates showed resistance to the examined antibiotics except for Penicillin. While most <i>S. aureus</i> isolates were Penicillin-resistant, only a small portion of the other three bacteria was resistant to Penicillin. Upwind samples contained a much lower percentage of ARBs than downwind samples.
[160] <sup>a</sup>	Grower–finisher	U.S. Midwest	Six antibiotics (ampicillin, erythromycin, oxytetracycline, penicillin, tetracycline, and tylosin) × four bacteria ( <i>S. aureus</i> , <i>Salmonella</i> spp., fecal coliforms, and coliforms)	No vancomycin-resistant <i>Enterococcus</i> was detected. The percentage of antibiotic resistant isolates differed with bacterial genera or species, as well as antibiotics. Around 98% of isolates showed resistance to ≥2 antibiotics.
[285]	Finisher	U.S. Mid-Atlantic	Five antibiotics (clindamycin, erythromycin, tetracycline, vancomycin, and virginiamycin,) × three genera ( <i>Enterococcus</i> , <i>Staphylococcus</i> , and <i>Streptococcus</i> ) and five <i>Enterococcus</i> species ( <i>E. dispar</i> , <i>E. Durans</i> , <i>E. faecalis</i> , <i>E. faecium</i> , and <i>E. hirae</i> )	Among bacterial isolates from the inside of the barns, 45% were resistant to all six antibiotics and 95% were resistant to ≥2 antibiotics. These numbers were much greater than those for upwind samples. The percentage of ABR in culturable bacteria gradually decreased downwind away from the barn.
[163] <sup>a</sup>	Gestation	U.S. Midwest	Six antibiotics (ampicillin, erythromycin, lincomycin, oxytetracycline, penicillin, and tetracycline) × four bacteria ( <i>S. aureus</i> , <i>Streptococci</i> , fecal coliforms, and coliforms)	

Table 10. Cont.

Reference	Barn Type	Location	Target ARB	Major Findings and/or Notes
[175] <sup>a</sup>	Farrowing, gestation, nursery, finisher	Germany	MRSA	IOM dust samplers (a filter-based method) yielded a lower MRSA occurrence frequency and a lower average airborne MRSA count than impingers. The occurrence of MRSA was confirmed through coagulase reactions and real-time PCR. Settled dust, pig nasal swab, boot swab, and fecal samples from gestation and finisher barns were also tested and found positive for MRSA. Quantitative information is available in Table 5. MRSA was found on soil surface downwind from barns. Selected MRSA colonies were cultivated with sheep blood agar and confirmed for their existence via coagulase reactions and real-time PCR.
[176] <sup>a</sup>	Gestation, finisher	Germany	MRSA	The study focused on the MRSA mitigation performance of a disinfectant. In addition to PM samples, bedding and pig nasal swab samples were also analyzed. The disinfectant showed limited effectiveness in MRSA reduction.
[178] <sup>a</sup>	Grower	Denmark	MRSA	A genotype MRSA CC398 was targeted. MSSA was also measured. The occurrence of MRSA was confirmed through an agglutination kit diagnostic test and molecular identification.
[179] <sup>a</sup>	Nursery, farrowing, finisher	Switzerland	MRSA	Sedimentation agar plates were used. Both kanamycin and oxacillin plates had over 300 colonies, among which 37 randomly selected colonies were subjected to 16S rRNA sequencing analysis. Results showed the dominance of phyla Firmicutes, Bacteroidetes, and Proteobacteria.
[263]	n/a	North Carolina	Kanamycin- and oxacillin-resistant bacteria	Multiple genera and species were identified. In-barn samples were collected with three different sampling durations. Downwind samples were also analyzed. Molecular typing of suggestive MRSA isolates was conducted through antimicrobial susceptibility testing, <i>mecA</i> PCR, <i>spa</i> typing, <sup>b</sup> and Pantón–Valentine leucocidin PCR. Several isolates were also resistant to tetracycline, clindamycin, and erythromycin.
[183] <sup>a</sup>	Nursery-grower	U.S. Midwest	MRSA	The study compared three industrial swine barns versus an antibiotic-free swine operation (open pasture). No MRSA or MDRSA was detected at the latter site. In swine barns, two samples were positive for MDRSA but at low concentrations (7 and 9 CFU m <sup>-3</sup> ).
[289]	n/a	North Carolina	MRSA and multi-drug resistant <i>S. aureus</i> (MDRSA)	Two samplers (Andersen six-stage viable cascade impactor and Respicon) were used but showed no significant differences. The highest airborne MRSA counts were found in nursery barns. The geometric mean diameter of MRSA was 7.2 µm (slightly greater than that of <i>S. aureus</i> ) and it differed with sampling locations.
[185] <sup>a</sup>	Farrowing, nursery, finisher, sick pigs	Denmark	MRSA	

Table 10. Cont.

Reference	Barn Type	Location	Target ARB	Major Findings and/or Notes
[101] <sup>a</sup>	Finisher	Germany	MRSA	The study compared four air filtration systems regarding their effectiveness in reducing MRSA and other bioaerosols. Suggestive MRSA isolates were further studied through <i>spa</i> and <i>mecA</i> PCR. High MRSA concentrations were ascribed to the use of Coriolis <sup>®</sup> $\mu$ samplers (a wet cyclone) for MRSA sampling.
[286]	n/a	China	Resistance of <i>E. coli</i> to ampicillin, piperacillin, amoxicillin/clavulanic acid, ampicillin/sulbactam, piperacillin/sulbactam, cephalothin, cefuroxime, aztreonam, gentamicin, kanamycin, streptomycin, amikacin, tetracycline, ciprofloxacin, ofloxacin, nalidixic acid, sulfamethoxazole, chloramphenicol, and nitrofurantoin	Twenty-six <i>E. coli</i> isolates were tested for antibiotic susceptibility. No resistance to amoxicillin/clavulanic acid, ampicillin/sulbactam, nitrofurantoin, or piperacillin/sulbactam was seen. Resistance to other antibiotics was found in part of <i>E. coli</i> isolates.
[186]	n/a	China	Tetracycline and erythromycin resistant bacteria	Self-prepared selective agar plates were used to screen tetracycline and erythromycin-resistant bacteria. DNA was extracted from bacterial isolates and subjected to 16S rRNA sequencing analysis. Genera <i>Staphylococcus</i> and <i>Rothia</i> were found to be abundant. The majority of ARB species occurred ubiquitously in both fine and coarse PM fractions. Note: Cattle, broiler, and layer barns were also studied and discussed collectively. In-barn airborne MRSA counts ranged from $\sim 0.3$ to $1.95 \text{ CFU m}^{-3}$ and reached the maximum in week 2 of weaning and the minimum in week 6. A slight increase was noted in weeks 7 and 8. The concentrations were much greater than those during the finisher stage. A significant correlation between in-barn and outdoor MRSA counts was noted.
[290]	Nursery	Denmark	MRSA	

<sup>a</sup> The publication has been summarized in Table 5 for culturable microbial counts. It is listed here for the convenience of readers, with minimal repeated information. <sup>b</sup> *spa* refers to *Staphylococcal* protein A, a critical factor affecting the virulence of *S. aureus*. *spa* typing is a method to distinguish *S. aureus* genotypes through PCR detection of *spa* genes.

The existing publications were from the U.S. (7), Europe (7), and China (2). Nine of them focused on MRSA and seven tested bacteria other than *S. aureus* for antibiotic resistance. Methicillin resistance was most extensively investigated, followed by tetracycline and erythromycin resistance (Note: The resistance of MRSA to tetracycline and erythromycin was examined by several MRSA studies). Finisher barns were the most studied barn type, followed by nursery barns.

Airborne MRSA counts varied greatly in the literature, from undetected (zero) to  $>10^4 \text{ CFU m}^{-3}$ . Most of the reported MRSA counts fell into the range of 10 to  $10^3 \text{ CFU m}^{-3}$ . MRSA accounted for only a small portion of cultural bacteria [101,175,183]. Regarding the percentage of *S. aureus* being methicillin resistant, no agreement has been reached. Ref. [185] found that in finisher and nursery barns, the majority of airborne *S. aureus* strains were MRSA; while in farrowing and sick pig barns, the percentage was down to  $<10\%$ . Comparatively, Ref. [179] reported that  $\sim 20\%$  of *S. aureus* strains in visited nursery, finisher, and farrowing barns were MRSA.

Two studies measured the percentage of other ARBs in total culturable bacteria. Ref. [282] reported that ~80% of airborne culturable bacteria in two finisher barns in Iowa were tylosin resistant. Ref. [186] found that, on average, 9% of airborne culturable bacteria in four swine barns in China were tetracycline resistant and 27% were erythromycin resistant. An overall higher percentage of ARB was found in broiler barns and a lower percentage in cattle barns in the same study.

PM samples in swine barns were frequently MRSA positive, especially those sampled from the inside of pig pens. The detected MRSA strains often carried resistance to other swine antibiotics (e.g., tetracycline) [183,289]. *Spa* typing was performed to determine the genotype of MRSA isolates [101,175,183,289,290]. The identified *spa* genotypes include t011, t034, t108, and t337. Among them, t011 and t034 are known to be livestock associated [183].

The exhaust air from swine barns can be a major source of ARB in surrounding areas. Several studies [160,163,176,183,290] analyzed downwind and/or upwind bioaerosol samples. Results revealed a substantial increase in ARB counts or occurrence frequency downwind from swine barns. Another supporting evidence is a significant correlation between in-barn and outdoor airborne MRSA counts [290].

Two studies investigated the control of airborne ARB in swine barns. Ref. [178] tested a commercial disinfectant (Stalosan<sup>®</sup>F) but did not see a significant reduction in MRSA counts after disinfection treatment. Ref. [101] compared four air filtration systems. However, no significant difference in in-barn airborne MRSA counts was reported. Ref. [185] examined the size distribution of MRSA and found its geometric mean diameter to be 7.2  $\mu\text{m}$ , suggesting that the majority of MRSA could be attached to large particles. Although it is not an MRSA mitigation study, the size data derived are valuable for the development of MRSA mitigation technologies, as many PM removal processes are size-dependent (Refer to Section 5).

In summary, multiple studies detected the presence of airborne ARB (including MRSA) in swine barns. The emissions of ARB via the barns' exhaust were found to elevate downwind ARB concentrations in the air and soil. Although efforts were made, no effective mitigation technology has been identified. ARB mitigation, from source reduction to end-of-pipe abatement, should receive considerable attention in future research.

#### Antibiotic Resistance Genes (ARGs)

Bacteria including human pathogens can acquire antibiotic resistance from other bacterial cells through horizontal gene transfer. An investigation of ARGs in various media (e.g., air, water, food, and soil), therefore, enables an improved understanding of the dissemination of antibiotic resistance in the environment. Molecular biology technologies, such as PCR and qPCR, are extensively used for the detection of ARGs in swine barn bioaerosols. These technologies can be part of culture-dependent or culture-independent methods. Examples of culture-dependent applications include the confirmation of MRSA colonies through *mecA* PCR (Table 10) where *mecA* (also written as *mec(A)*) is a gene that grants methicillin resistance to bacteria. This subsection focuses on ARG analysis results derived from culture-independent methods which constitute the vast majority of existing findings.

To our knowledge, the first analysis of ARGs in swine barn bioaerosols was reported by Ref. [291], as an add-on effort to culture-dependent ARB monitoring in the same barns [285]. Both were conducted by Dr. Schwab and his colleagues at Johns Hopkins University. Table 11 summarizes ARG studies since then. From a methodology standpoint, the findings acquired from PCR/qPCR of ARGs cannot be directly compared with those from metagenomic sequencing. For simplicity, they are listed here in the same table; a brief discussion of the methods is available in Section 4.4.4.

**Table 11.** Airborne antibiotic resistance genes in swine barn PM.

Reference	Barn Type	Location	Target ARGs <sup>a</sup>	Analytical Method	Major Findings and/or Notes <sup>b</sup>
[291]	Finisher	Iowa	MLS resistance: <i>erm</i> (A), <i>erm</i> (B), <i>erm</i> (C), <i>erm</i> (F), <i>mef</i> (A)Tetracycline resistance: <i>tet</i> (M), <i>tet</i> (O), <i>tet</i> (S), <i>tet</i> (K), <i>tet</i> (L)	DNA-DNA hybridization, PCR	Sixteen <i>Enterococcus</i> spp. and 16 <i>Streptococcus</i> spp. isolates selected from ARB screening were subjected to ARG analysis. All colonies carried multiple MLS and tetracycline resistance genes.
[172]	Grower–finisher	Quebec, Canada	Tetracycline resistance: <i>tet</i> (A), <i>tet</i> (C), <i>tet</i> (G), <i>tet</i> (M), <i>tet</i> (O), <i>tet</i> (P), <i>tet</i> (Q), <i>tet</i> (S), <i>tet</i> (T), <i>tet</i> (W)	qPCR for <i>tet</i> (G)PCR for others	The concentration of <i>tet</i> (G) was $2.5 \pm 6.6 \times 10^6$ copies $m^{-3}$ . All 18 visited barns were positive for tetracycline resistance genes analyzed. Tetracycline resistance was also found among culturable pathogens at various occurrence frequencies. These tetracycline resistance genes were abundant in DNA extracts from swine barn PM ( $9.55 \times 10^2$ – $1.69 \times 10^6$ copies $ng^{-1}$ ). <i>tet</i> (O) and <i>tet</i> (Q) were the most abundant; while <i>tet</i> (B) was the least abundant. Swine barn PM had much greater <i>tet</i> (H) and <i>tet</i> (W) contents than poultry barn PM but lower <i>tet</i> (Z) contents than turkey barn PM.
[209]	Farrowing, gestation, nursery, finisher	Illinois	Tetracycline resistance: <i>tet</i> (B), <i>tet</i> (H), <i>tet</i> (Z), <i>tet</i> (O), <i>tet</i> (Q), <i>tet</i> (W)	qPCR	<i>tet</i> (Q) was the most abundant ( $8.89 \times 10^5 \pm 1.45 \times 10^6$ copies $m^{-3}$ ) while <i>tet</i> (B) was the least abundant. A significant difference in ABG abundance was found for <i>tet</i> (H), <i>tet</i> (O), <i>tet</i> (Q), and <i>tet</i> (W) between winter and summer samples. All tested ARGs but <i>tet</i> (B) were more abundant in barns with deep manure pits. Barns with bedded litter floors had the lowest ARGs abundance in the air.
[218]	Grower–finisher	South Korea	Tetracycline resistance: <i>tet</i> (B), <i>tet</i> (H), <i>tet</i> (Z), <i>tet</i> (O), <i>tet</i> (Q), <i>tet</i> (W)	qPCR	Twenty-six <i>E. coli</i> isolates were analyzed. Among them, 19 carried at least one and 4 carried two or more quinolone resistance genes. The detected subtypes included <i>qnr</i> (A), <i>qnr</i> (S1), <i>qnr</i> (S2), <i>qnr</i> (B2), <i>aac</i> (6′)- <i>lb-cr</i> , <i>qep</i> (A), and <i>oqx</i> (AB). Other environmental samples were also analyzed.
[219]	Grower–finisher	South Korea	Tetracycline resistance: <i>tet</i> (B), <i>tet</i> (H), <i>tet</i> (Z), <i>tet</i> (O), <i>tet</i> (Q), <i>tet</i> (W)	qPCR	All PM samples carried <i>czt</i> (C) and 60% of samples carried <i>bla</i> CTX-M-1 and <i>mcr</i> -1. Their concentrations were $1.73 \times 10^2$ – $1.78 \times 10^5$ , $<8$ – $9.89 \times 10^2$ , and $<8$ – $9.87 \times 10^2$ copies $m^{-3}$ , respectively.
[286] <sup>c</sup>	n/a	China	Quinolone resistance in <i>E. coli</i> : <i>qnr</i> (A), <i>qnr</i> (B), <i>qnr</i> (S), <i>qnr</i> (S1), <i>qnr</i> (S2), <i>qnr</i> (A), <i>aac</i> (6′)- <i>lb-cr</i> , <i>aac</i> (6′)- <i>lb-cr</i> , <i>qep</i> (A), <i>oqx</i> (AB)	PCR	A total of 304, 300, 300, 277, and 304 ARG subtypes were identified in boar, farrowing, gestation, nursery, and finisher barns, respectively. In terms of ARG composition, two clusters were found: (1) boar, farrowing, and gestation, and (2) nursery and finisher.
[103]	Finisher	Quebec, Canada	Zinc resistance: <i>czt</i> (C)Cephalosporin resistance: <i>bla</i> CTX-M-1 Colistin resistance: <i>mcr</i> -1	qPCR	The top 10 ARG subtypes were resistant to aminoglycosides, aminocoumarin, mupirocin, elfamycin, fluoroquinolone, pleuromutilin, rifampin, and lincosamide. Bacterial phyla Firmicutes and Bacteroidetes carried the majority of ARGs.
[267]	Farrowing, gestation, nursery, grower–finisher, boar	China	Various resistance	Shotgun metagenomic sequencing	

Table 11. Cont.

Reference	Barn Type	Location	Target ARGs <sup>a</sup>	Analytical Method	Major Findings and/or Notes <sup>b</sup>
[269]	n/a	Nine European countries	Various resistance	Shotgun metagenomic sequencing	The most abundant ARG was tetracycline resistance genes, followed by macrolide and aminoglycoside. Nine other major ARG types were reported: beta-lactam, colistin, nitroimidazole, oxazolidinone phenicol, phenicol, quinolone, sulphonamide, trimethoprim, and vancomycin.
[272]	n/a	China	Tetracycline resistance: <i>tet(M)</i> , <i>tet(G)</i> , <i>tet(O)</i> Sulfonamide resistance: <i>sul1</i> , <i>sul2</i> Quinolones resistance: <i>qnr(A)</i> Macrolides resistance: <i>erm(A)</i> , <i>erm(B)</i>	qPCR	ARGs were more abundant (in copies $m^{-3}$ ) in winter than summer samples. The most dominant ARG was <i>tet(M)</i> . Mobile genetic elements (MGEs), a carrier of ARGs in bacteria, were also quantified. Genera <i>Lactobacillus</i> , <i>Prevotella</i> , <i>Prevotellaceae</i> , <i>Balutia</i> , and <i>Muribaculaceae</i> correlated significantly with ARGs in winter. Genera <i>Prevotellaceae</i> and <i>Ochrobactrum</i> correlated significantly with tetracycline resistance genes.
[274] <sup>d</sup>	Farrowing, gestation, nursery, grower–finisher	China	Various resistance	Metagenomic sequencing <sup>e</sup> for ARGs and bacteria	Twenty-two ARG types were identified, with 12 shared by all barn types. Nursery barns had the fewest ARG types and finisher barns had the lowest abundance of ARGs. The top three most abundant ARGs were <i>aph(3'')-III</i> , <i>aad(E)</i> , and <i>tet(W)</i> . Genera <i>Leadbetterella</i> and <i>Methylobacterium</i> were hosts of most ARGs; while 11 other genera carried $\geq 3$ ARGs.

<sup>a</sup> MLS—macrolide, lincosamide, and streptogramin. <sup>b</sup> Several papers presented qPCR quantitative results in figures only. It is difficult to derive accurate readings from those figures. Accordingly, no quantitative results were provided in this table. <sup>c</sup> The study did not use PCR to confirm the existence of ARG. Instead, it focused on the determination of ARG types and subtypes. Thus, it is also included in this table. <sup>d</sup> The same paper used 16S rRNA NGS for bacterial taxonomic classification. <sup>e</sup> Although no direct information was given, the selected technology appeared to be shotgun metagenomic sequencing based on method descriptions.

A rapidly growing interest in ARGs is witnessed. Out of the eleven publications in the table, six were published in the past two years. Regarding the geographic distribution of relevant studies, six were from Asia (China and South Korea) and only one from Europe. Although the U.S. pioneered in this subject, no relevant research has been reported since 2012.

Seven studies used qPCR for ARG detection and quantification. The abundance of ARGs was presented as the number of ARG copies per ng of DNA extracts (copies  $ng^{-1}$ ) or the number of ARG copies per  $m^{-3}$  of barn air (copies  $m^{-3}$ ). Three studies used PCR for qualitative (presence or absence of an ARG) or quantitative analysis (occurrence frequency of an ARG in bacterial colony isolates). Three studies used metagenomic analysis to survey the existence of various ARGs and to determine host bacterial groups.

Tetracycline resistance genes (TRGs) were the most studied ARGs (with eight publications) due to the extensive use of tetracycline for pork production [292]. Multiple TRG subtypes were investigated in the literature, including *tet(A)*, *tet(B)*, *tet(C)*, *tet(G)*, *tet(H)*, *tet(K)*, *tet(L)*, *tet(M)*, *tet(O)*, *tet(P)*, *tet(Q)*, *tet(S)*, *tet(T)*, *tet(W)*, and *tet(Z)*. They confer tetracycline resistance to bacteria through several mechanisms including efflux pump, ribosomal protection, and enzymatic inactivation [293]. TRGs encoding ribosomal protection proteins were commonly found in the gastrointestinal tracts of pigs [294]. Other ARGs that were reported by multiple publications include quinolones resistance genes and macrolides (or MLS) resistance genes.

Little information is known regarding the effect of seasons and barn types. Ref. [209] compared four barn types but found no significant difference in TRG profiles. Ref. [219]

compared three waste management systems for finisher barns and revealed a significant effect of waste management on TRG concentrations. Ref. [272] examined four types of ARGs in winter and summer PM samples and found that ARGs were more abundant in winter. This is understandable, as in-barn PM concentrations are typically the highest in winter.

Metagenomic analysis revealed that various airborne bacteria in swine barns could carry ARGs. The major ARG hosts identified include phyla Firmicutes and Bacteroidetes and genera *Lactobacillus*, *Prevotella*, *Prevotellaceae*, *Balutia*, *Leadbetterella*, and *Methylobacterium*. For TRGs, the major hosts include genera *Prevotellaceae* and *Ochrobactrum*. It is noteworthy that phyla Firmicutes and Bacteroidetes are two dominant bacterial groups in swine barn bioaerosols, as well as genera *Lactobacillus* and *Prevotella* (Refer to Section 3.2.3). However, it is uncertain whether other abundant bacterial groups in swine barn bioaerosols, such as genera *Clostridium*, *Bacteroides*, and *Corynebacterium*, are significant hosts of ARGs due to limited data in the literature.

Although significant progress has been made, several fundamental questions remain unanswered regarding ARGs in swine barn bioaerosols. For example, the air is an unideal environment for many gastrointestinal or fecal bacteria. Thus, it is uncertain how long airborne bacteria carrying ARGs can survive and whether they can propagate ARGs through horizontal or vertical gene transfer after a long travel in the air. No exposure model has been well established to describe the potential impact of airborne ARB and ARGs on animal and community health.

### 3.2.5. Endotoxins and other Bioaerosol Markers

Bioaerosols can be characterized by their fingerprint components or metabolites [21]. These components and metabolites are known as bioaerosols markers, or simply biomarkers. Some of them are etiological agents of diseases or disorders in animals and humans; while others have had no known health implications. In a previous review paper, etiological agents were listed as a separate category from markers [295]. For simplicity, this review tags the agents in both categories (etiological and non-etiological) as bioaerosol markers.

Various markers have been used for bioaerosols assessment. In principle, any organism(s)-specific substance can potentially be selected as a bioaerosol marker. The specificity can be at the taxonomic level of domain, kingdom, phylum, etc., or even simply distinguish organisms from non-organisms. Table 12 lists several commonly selected markers. Among them, endotoxin is most frequently used for bioaerosols assessment in swine barns, followed by (1→3)-β-D-glucan. Others are very occasionally measured. Accordingly, this section is structured into three subsections: endotoxin, (1→3)-β-D-glucan, and other bioaerosol markers.

**Table 12.** Bioaerosol markers for different organisms.

Organism	Marker <sup>a</sup>	Reference
General organisms	Total proteins	[296]
	ATP	[297]
	(1→3)-β-D-glucan	[298]
	Ergosterol	[299]
Fungi	EPS	[300]
	Mannitol	[301]
	Arabitol	[302]
Gram-negative bacteria	Mycotoxins (e.g., aflatoxin B1)	[303]
	Endotoxin (3-hydroxy fatty acids)	[304,305]
Gram-positive bacteria	Peptidoglycan (muramic acid)	[306,307]

<sup>a</sup> ATP—Adenosine triphosphate; EPS—Extracellular polysaccharides.

## Endotoxin

An endotoxin (lipopolysaccharide, LPS) is a cell envelop component of Gram-negative bacteria and is released when the bacteria are lysed or at the multiplication stage [308]. An LPS molecule consists of three parts: O-antigen, core polysaccharide, and lipid A. Among them, lipid A is believed to be responsible for the toxicity of endotoxins. Lipid A comprises two glucosamine groups with one phosphate and multiple acyl chains attached to each group. Different Gram-negative bacteria may differ in the number, length, and attachment site of acyl chains, resulting in different degrees of toxicity [309]. Even for the same species, the structure of lipid A may change when the bacteria grow at different lifetime periods or under different environmental conditions [211].

Endotoxins are mildly toxic to mammals. Upon injection into the blood, endotoxins immediately trigger a series of immunoreactions, leading immune cells to release pro-inflammatory cytokines. For human, a dose of 2 ng of *Salmonella abortus-equi* endotoxin per kg body weight could increase a body temperature by 1.9 °C [310]. Symptoms such as “fever, change in white blood cell counts, disseminated intravascular coagulation, hypotension, shock, and death” can be observed [311]. Inhalation of endotoxins is associated with various acute and chronic symptoms in humans, including fever, shivering, pulmonary inflammation, non-allergenic asthma, airway obstruction, and impaired lung functions [14]. Acute lung function impairment and acute bronchial obstruction in swine farm workers were found to be highly dependent on endotoxin levels [13,38,312]. Chronic bronchial obstruction and hyperactive airways in swine farm workers could result from long-term exposure to airborne endotoxins [155,313]. Efforts were made to develop endotoxin exposure limits through dose–response studies [38,314]. Ref. [26] recommended an exposure limit of 100 endotoxin units (EU) m<sup>-3</sup> for swine barns. Inhalation of endotoxins could also compromise pig performance and health, causing decreased growth rates [315], bronchial hyperreactivity, lung inflammation [316], and even respiratory failure [317].

Endotoxins are extensively used as a marker of Gram-negative bacteria [311]. Previous studies have revealed the ubiquitous occurrence of Gram-negative bacteria in swine barn PM samples. For example, Ref. [159] measured culturable bacterial counts in a finisher barn in Kansas and reported the presence of genera *Pseudomonas*, *Vibro*, and Gram-negative *Bacilli*. Similar findings were acquired using culture-independent methods. Ref. [169] examined the bacterial diversity of PM sampled from eight swine barns in Quebec, Canada, and found that most Gram-negative bacteria belonged to genera *Moraxella*, *Bacteroides*, and *Pseudomonas*. Using 16S rRNA NGS, Ref. [209] found four Gram-negative bacterial phyla (Bacteroidetes, Proteobacteria, Fusobacteria, and Cyanobacteria) in TSP samples from 12 swine barns in Illinois. They further identified Gram-negative bacterial genera, such as *Acinetobacter*, *Moraxella*, and *Fusobacterium*.

To our knowledge, the first measurement of airborne endotoxins in swine barns was conducted by Dr. Thedell and his colleagues at the National Institute of Occupational Safety and Health (NIOSH) [318]. Using a gel clot *Limulus* amoebocyte lysate (LAL) assay, they reported the endotoxin levels of 4.77–47.74 µg per gram of PM in two swine barns in the U.S. Midwest. Following their work, multiple monitoring efforts were made in the 1980s [35,36,38,108,312,313,319,320]. These early studies were often coupled with epidemiological surveys to further unravel or affirm the health effects of airborne endotoxins. Additional studies have been reported since 1990 (Table 13). However, the focus areas of the studies shifted to (1) comparing and further refining endotoxin sampling and testing protocols, and (2) determining baseline airborne endotoxin levels and their effects by environmental and operating parameters in swine barns.

**Table 13.** Airborne endotoxin concentrations in swine barns—a summary of studies since 1990.

Reference	Endotoxin Concentration (EU m <sup>-3</sup> ) <sup>a,b,c</sup>	Barn and Ventilation Type	Location	Season	Analytical Method
[42] <sup>d</sup>	TSP: 64 ± 34 (9–120) ng m <sup>-3</sup> Respirable: 5 ± 3 (1–13) ng m <sup>-3</sup> During tending— TSP: 37 ng m <sup>-3</sup>	Breeding; n/a	Denmark	n/a	Chromogenic LAL assay
[321] <sup>d</sup>	Respirable: 8 ng m <sup>-3</sup> During feeding— TSP: 315 ng m <sup>-3</sup> Respirable: 17 ng m <sup>-3</sup>	n/a; n/a	Sweden	n/a	LAL assay
[322] <sup>d</sup>	TSP: 702	Breeding; n/a	Denmark	n/a	LAL assay
[323] <sup>d</sup>	TSP: 0.21–0.40 µg m <sup>-3</sup>	n/a; n/a	Sweden	n/a	Chromogenic LAL assay
[43]	TSP: 1.88–31.25 µg m <sup>-3</sup> TSP: 31.25–75.00 µg m <sup>-3</sup>	Farrowing; n/a Finisher; n/a	Poland	n/a n/a	Clot LAL assay
[324] <sup>d</sup>	TSP: 202.4 Respirable: 16.59	n/a; n/a	Iowa	All seasons	Chromogenic LAL assay
[325] <sup>d</sup>	TSP: 111 (5.6–825) ng m <sup>-3</sup> TSP: 150 (10.6–1503) ng m <sup>-3</sup>	n/a; n/a n/a; n/a	Netherlands	Summer Winter	Kinetic LAL assay
[46] <sup>d</sup>	Time 1— TSP: 202.7 ng m <sup>-3</sup> (GM); 4.33 (GSD) Respirable: 17.0 ng m <sup>-3</sup> (GM); 2.30 (GSD) Time 2— TSP: 176.1 ng m <sup>-3</sup> (GM); 3.16 (GSD) Respirable: 11.9 ng m <sup>-3</sup> (GM); 2.88 (GSD)	n/a; n/a	Iowa	Spring, fall, winter	Endpoint chromogenic LAL assay
[49]	Area samples— TSP: 7.03 × 10 <sup>3</sup> without oil sprinkling TSP: 566 with oil sprinkling Personal sample— TSP: 1.60 × 10 <sup>3</sup> without oil sprinkling TSP: 3.44 × 10 <sup>3</sup> with oil sprinkling	Grower–finisher; MV	Saskatchewan, Canada	Winter	Kinetic chromogenic LAL assay
[326] <sup>d</sup>	TSP: 2040–24,100; 8290 (GM)	n/a; n/a	Iowa	n/a	KLARE <sup>e</sup> & endpoint chromogenic LAL assay
[51] <sup>d</sup>	TSP: 22.8 µg m <sup>-3</sup> Daytime— Inhalable: 114.6 ng m <sup>-3</sup> Respirable: 8.3 ng m <sup>-3</sup>	n/a; n/a	Poland	n/a	LAL assay
[327]	Nighttime— Inhalable: 52.3 ng m <sup>-3</sup> Respirable: 7.4 ng m <sup>-3</sup> Daytime— Inhalable: 186.5 ng m <sup>-3</sup> Respirable: 17.7 ng m <sup>-3</sup> Nighttime— Inhalable: 157.4 ng m <sup>-3</sup> Respirable: 18.9 ng m <sup>-3</sup> Daytime— Inhalable: 135.1 ng m <sup>-3</sup> Respirable: 13.0 ng m <sup>-3</sup> Nighttime— Inhalable: 109.1 ng m <sup>-3</sup> Respirable: 11.4 ng m <sup>-3</sup>	Sow; n/a  Nursery; n/a  Finisher; n/a	England, Netherlands, Denmark, Germany	n/a	LAL clot assay
[328] <sup>d</sup>	Control— TSP: 3984; 498 (SE) Oil sprinkling— TSP: 452; 66 (SE)	Grower–finisher; MV	Saskatchewan, Canada	n/a n/a	Chromogenic LAL assay
[53] <sup>d</sup>	TSP: 60–1.49 × 10 <sup>4</sup> ng m <sup>-3</sup> ; 631 ng m <sup>-3</sup> (median); 660 ng m <sup>-3</sup> (GM)	n/a; n/a	United Kingdom	All seasons	Kinetic turbidimetric LAL assay
[54] <sup>d</sup>	TSP: 4.9 × 10 <sup>3</sup>	Finisher; MV	Quebec, Canada	Winter	Endpoint chromogenic LAL assay

Table 13. Cont.

Reference	Endotoxin Concentration (EU m <sup>-3</sup> ) <sup>a,b,c</sup>	Barn and Ventilation Type	Location	Season	Analytical Method
[55]	TSP: 36.8 ± 18.1 (15.8–73.2)	Breeding; NV	Taiwan	Summer	KLARE assay
	Respirable: 14.1 ± 16.0 (3.4–56.6)	Farrowing; NV		Summer	
	TSP: 82.1 ± 85.0 (14.4–277)			Summer	
	Respirable: 48.6 ± 166 (3.5–837)	Nursery; NV		Summer	
	TSP: 298 ± 249 (32.0–818)	Grower; NV		Summer	
[59] <sup>d</sup>	Respirable: 20.9 ± 31.9 (1.6–155)	Finisher; NV	Denmark	Summer	Kinetic turbidimetric LAL assay
	TSP: 145 ± 81.4 (40.1–298)	n/a; n/a		n/a	
	Respirable: 21.8 ± 45.6 (0.02–217)	n/a; n/a		n/a	
	TSP: 136 ± 105 (30.8–418)				
	Respirable: 129 ± 396 (5.6–1643)				
[63]	TSP: 1.30–1101.7 ng m <sup>-3</sup> ; 58.01 ng m <sup>-3</sup> (median)	Grower–finisher; n/a	Quebec, Canada	Summer	LAL assay
	TSP: 0.01–2090.1 ng m <sup>-3</sup> ; 76.3 ng m <sup>-3</sup> (median)				
	Control—				
	TSP: 1.72 × 10 <sup>3</sup> (GM); 1.5 (GSD)				
	Conventional scrapper—				
[68] <sup>d</sup>	TSP: 1.85 × 10 <sup>3</sup> (GM); 2.0 (GSD)	n/a; n/a	The Netherlands	All seasons	Kinetic chromogenic LAL assay
	V-shaped scraper—				
	TSP: 2.14 × 10 <sup>3</sup> (GM); 2.3 (GSD)				
	Daily V-shape scraper—				
	TSP: 1.53 × 10 <sup>3</sup> (GM); 2.1 (GSD)				
[329] <sup>d</sup>	Van Kempen belt—	Finisher; n/a	Germany	All seasons	Kinetic chromogenic LAL assay
	TSP: 1.85 × 10 <sup>3</sup> (GM); 2.5 (GSD)				
	Cemagref net—				
	TSP: 1.43 × 10 <sup>3</sup> (GM); 2.1 (GSD)				
	TSP: 992–6970; 1510 (GM); 2.1 (GSD)				
[71] <sup>d</sup>	Inhalable: 43.2–7.47 × 10 <sup>3</sup> ; 668.7 (median)	Nursery, finisher; n/a	Ireland	Spring, summer	Endpoint LAL assay
	Respirable: 1.9–236; 23.1 (median)				
	TSP: 1.67 × 10 <sup>5</sup> (maximum)				
	Inhalable: 3400 (GM); 6.9 (GSD)				
	TSP: 6.55 × 10 <sup>3</sup> (2.22 × 10 <sup>3</sup> –2.59 × 10 <sup>4</sup> )				
[330] <sup>d</sup>	TSP: 2.57 × 10 <sup>4</sup> (1.80 × 10 <sup>3</sup> –6.91 × 10 <sup>4</sup> )	Finisher; n/a	Quebec, Canada	Summer	Endpoint chromogenic LAL assay
	With slatted floors—				
	TSP: (2.67 ± 1.44) × 10 <sup>4</sup>				
	With sawdust beddings—				
	TSP: (5.19 ± 3.04) × 10 <sup>4</sup>				
[76] <sup>d</sup>	With source separation—	Finisher; MV	Quebec, Canada	Winter	chromogenic LAL assay
	TSP: 3.17 × 10 <sup>3</sup> –7.25 × 10 <sup>3</sup>				
	Inhalable: ~450				
	Inhalable: ~1400				
	Inhalable: ~2500				
[77] <sup>d</sup>	Inhalable: 48–3.77 × 10 <sup>4</sup> ; 3.25 × 10 <sup>3</sup> (GM); 4.9 (GSD)	Gestation, farrowing; n/a	US Midwest	Summer Spring Winter	Kinetic chromogenic LAL assay
	Inhalable: 59–5.78 × 10 <sup>4</sup> ; 3.10 × 10 <sup>3</sup> (GM); 5.8 (GSD)				
	TSP: 384.9 (2.6–4.15 × 10 <sup>3</sup> ); 120 (median); 109.0 (GM)				
	Inhalable: 5.2 × 10 <sup>3</sup> (160–3.7 × 10 <sup>5</sup> ); 1.4 × 10 <sup>3</sup> (GM); 3.2 (GSD)				
	Inhalable: 5.3 × 10 <sup>3</sup> (BDL–1.10 × 10 <sup>5</sup> ); 2.4 × 10 <sup>3</sup> (GM); 3.1 (GSD)				
[78] <sup>d</sup>	TSP: 1.29 × 10 <sup>3</sup> (17–6.15 × 10 <sup>3</sup> ); 636 (GM)	Grower–finisher; NV (hoop barns)	Iowa	All seasons	Kinetic chromogenic LAL assay
	TSP: 510 ± 317 (164–991); 419 (GM); 1.98 (GSD)				
	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)				
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				
	TSP: 1508 ± 978, (693–3.59 × 10 <sup>3</sup> ); 1285 (GM); 1.51 (GSD)				
[171]	TSP: 384.9 (2.6–4.15 × 10 <sup>3</sup> ); 120 (median); 109.0 (GM)	Finisher, farrowing, nursery; MV	North Carolina	n/a	Kinetic chromogenic LAL assay
	Inhalable: 5.2 × 10 <sup>3</sup> (160–3.7 × 10 <sup>5</sup> ); 1.4 × 10 <sup>3</sup> (GM); 3.2 (GSD)				
	Inhalable: 5.3 × 10 <sup>3</sup> (BDL–1.10 × 10 <sup>5</sup> ); 2.4 × 10 <sup>3</sup> (GM); 3.1 (GSD)				
	TSP: 1.29 × 10 <sup>3</sup> (17–6.15 × 10 <sup>3</sup> ); 636 (GM)				
	TSP: 510 ± 317 (164–991); 419 (GM); 1.98 (GSD)				
[89] <sup>d</sup>	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)	Gestation; MV	Switzerland	Winter, summer	Chromogenic kinetic LAL assay
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				
	TSP: 1508 ± 978, (693–3.59 × 10 <sup>3</sup> ); 1285 (GM); 1.51 (GSD)				
	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)				
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				
[179]	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)	Farrowing; MV	Illinois	All seasons	Chromogenic kinetic LAL assay
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				
	TSP: 1508 ± 978, (693–3.59 × 10 <sup>3</sup> ); 1285 (GM); 1.51 (GSD)				
	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)				
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				
[332]	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)	Nursery; MV			
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				
	TSP: 1508 ± 978, (693–3.59 × 10 <sup>3</sup> ); 1285 (GM); 1.51 (GSD)				
	TSP: 508 ± 617 (98–2.10 × 10 <sup>3</sup> ); 334 (GM); 2.47 (GSD)				
	TSP: 1971 ± 2816 (217–8.7 × 10 <sup>3</sup> ); 1017 (GM); 3.30 (GSD)				

Table 13. Cont.

Reference	Endotoxin Concentration (EU m <sup>-3</sup> ) <sup>a,b,c</sup>	Barn and Ventilation Type	Location	Season	Analytical Method
[333]	PM <sub>10</sub> : 74.7 ± 54.2; 63.9 (GM); 1.72 (GSD) PM <sub>2.5</sub> : 23.3 ± 11.2; 20.5 (GM); 1.78 (GSD) PM <sub>10</sub> : 313 ± 321; 201 (GM); 2.77 (GSD) PM <sub>2.5</sub> : 66.1 ± 56.4; 46.8 (GM); 2.52 (GSD) PM <sub>10</sub> : 173 ± 104; 148 (GM); 1.82 (GSD) PM <sub>2.5</sub> : 26.2 ± 14.0; 22.8 (GM); 1.79 (GSD) PM <sub>10</sub> : 198 ± 164; 163 (GM); 1.83 (GSD) PM <sub>2.5</sub> : 84.2 ± 97.0; 64.1 (GM); 2.50 (GSD)	Gestation; MV Farrowing; MV Nursery; MV Finisher; MV	Illinois	All seasons	Chromogenic kinetic LAL assay
[103]	TSP: (9.03 ± 9.52) × 10 <sup>3</sup> (6.02 × 10 <sup>2</sup> –3.40 × 10 <sup>4</sup> )	Finisher; MV	Quebec, Canada	Winter	Chromogenic kinetic LAL assay
[334] <sup>d</sup>	Inhalable: 2026 (GM); 5.3 (GSD)	n/a; n/a	Iowa	Spring, fall	Chromogenic kinetic LAL assay

<sup>a</sup> A unit of EU m<sup>-3</sup> is used unless otherwise stated. <sup>b</sup> Airborne endotoxins are measured as a component of particles (e.g., TSP, PM<sub>10</sub>, and PM<sub>2.5</sub>). <sup>c</sup> Concentrations are expressed in the default forms of arithmetic mean, arithmetic mean ± standard deviation, or concentration range. Other statistics, including median, geometric mean (GM), GSD, and standard error (SE), are reported when relevant data are available. A note in the parenthesis specifies the type of reported data. <sup>d</sup> Personal exposure samples were collected and analyzed for endotoxins. <sup>e</sup> KLARE—Kinetic Limulus assay with resistant-parallel-line estimation.

It is noteworthy that in some previous studies (especially those before 2000), airborne endotoxin concentrations were presented in the unit of ng m<sup>-3</sup> or µg m<sup>-3</sup>. This is primarily due to the concentration unit of available endotoxin standards. These standards are extracted from pure Gram-negative bacterial strains. They can be quantitated by mass (e.g., ng of endotoxins) or endotoxin potency (i.e., endotoxin unit (EU)). Again, different bacterial strains, or the same strain but different batches, could possess different endotoxin potency (toxicity). In recent years, nearly all endotoxin standards come with a predetermined potency value (e.g., 1 ng *E. coli* O55:B85 endotoxin = 12 EU). For studies with no potency information available, an approximate relationship of 1 ng endotoxin = 10 EU may be used [326,335].

Taking the conversion factor (1 ng = 10 EU), the following observations were derived from the analysis of existing publications:

- All but one endotoxin study was conducted in North America and Europe. In major pork-producing Asian countries like South Korea and China, no endotoxin measurement has been reported—a sharp contrast to a growing interest in in-barn bioaerosol composition in these countries. The majority of existing studies measured the endotoxin concentrations associated with large particles (TSP and inhalable). Only a few studies (7 out of 32 since 1990) measured the endotoxin concentrations associated with small particles (e.g., PM<sub>2.5</sub> and respirable). Many studies used personal PM samplers for endotoxin exposure assessment. As personal samplers are carried around by farm workers during their work shifts, the measured concentrations cannot be directly compared with those derived from fixed samplers, e.g., area samples [38].
- Airborne endotoxin concentrations varied greatly in the literature. For endotoxins associated with TSP and inhalable particles, their concentrations ranged from tens [55,78] to hundreds of thousands EU m<sup>-3</sup> [43,89]. Such variability (over four orders of magnitude) is more pronounced than that in TSP or inhalable PM concentrations. However, the majority of measured concentrations associated with TSP or inhalable PM fall into the range of 10<sup>2</sup>–10<sup>4</sup> EU m<sup>-3</sup>. In comparison, the endotoxin concentrations associated with respirable PM were much smaller, with a typical range of a few to hundreds of EU m<sup>-3</sup>. Only one study measured the endotoxin concentrations associated with PM<sub>10</sub> and PM<sub>2.5</sub> [333]. No conclusion is drawn due to limited data.
- Airborne endotoxin concentrations differed with barn types. For endotoxins associated with large particles (inhalable and TSP), the overall highest concentrations were found in nursery barns, followed by grower or finisher barns; while the lowest concentrations occurred in sow (farrowing and gestation) barns [55,327,332]. It is noteworthy that the

three studies on the effects of barn types were each conducted in Asia, North America, and Europe, respectively. Thus, the observed variability could be representative. For endotoxins associated with small particles, no agreement was reached. Ref. [327] reported that, for respirable PM, the overall highest endotoxin concentrations occurred in nursery barns. However, a later study [55] found that the highest respirable endotoxin concentrations occurred in finisher barns; and nursery barns had even lower respirable endotoxin concentrations than farrowing barns. A similar observation was reported by [333] from the endotoxin measurement of PM<sub>2.5</sub> samples.

- Airborne endotoxin concentrations were overall higher in winter than in summer [77,89,325,331–333]. This is consistent with seasonality in in-barn PM concentrations. Elevated ventilation rates in summer enhance the dilution of endotoxin-laden PM by fresh air, thereby resulting in lower airborne endotoxin concentrations. However, it is noteworthy that the in-barn thermal environment, including temperature and humidity, could affect endotoxin loadings in PM samples. The loading is normally measured as the number of endotoxin units per mass of PM (e.g., EU mg<sup>-1</sup> PM). Ref. [77] found that winter inhalable particle samples had overall higher endotoxin loadings (on average 713 EU mg<sup>-1</sup>) than summer samples (on average 550 EU mg<sup>-1</sup>). This contradicts the study [332] in which endotoxin loadings in TSP increased with outdoor temperatures. They ascribed the higher endotoxin loadings in summer (on average 1308 EU mg<sup>-1</sup>; versus on average 484 EU mg<sup>-1</sup> in winter) to enhanced bacterial growth as a result of elevated temperatures in swine barns and feed storages.
- Airborne endotoxin concentrations varied with manure collection systems. Ref. [63] compared six different manure collection systems in experimental swine barns and found that the barns with V-shaped manure scrapers had the highest endotoxin concentrations. However, no significant difference was noted. Ref. [76] observed overall higher endotoxin concentrations in swine barns with sawdust beddings than those with slatted floors. They further reported that by implementing a source separation (i.e., solid-liquid separation) system, the airborne endotoxin concentrations were reduced by nearly an order of magnitude. Ref. [78] compared inhalable endotoxin concentrations in hoop barns (with beddings and natural ventilation) versus regular barns (with slatted floors, manure pits, and mechanical ventilation). Slightly higher endotoxin concentrations were found in hoop barns. However, no significant difference was noted.
- Airborne endotoxin concentrations were affected by animal activity. Ref. [327] found that airborne endotoxin concentrations were higher during the day than the night. Pigs are usually more active during the daytime, with their activity peaked in the afternoon [336]. Animal activity is related to feeding. Ref. [321] reported significantly higher airborne endotoxin concentrations during feeding than tending. In addition to its regulation of animal activity, feeding could directly result in the suspension of endotoxin-laden feed particles, thereby raising airborne endotoxin concentrations.
- Airborne endotoxin concentrations could be reduced by oil sprinkling [49,328]. However, both studies targeted TSP-associated endotoxins. It remains unknown whether oil sprinkling would be similarly effective in reducing airborne endotoxins associated with small particles (respirable, PM<sub>10</sub>, or PM<sub>2.5</sub>) and whether other in-barn PM mitigation technologies (Refer to Section 5) would be effective. It is noteworthy that endotoxins are released from lysed Gram-negative bacteria. In-barn PM mitigation technologies that kill bacteria could elevate free endotoxin loadings in PM samples.

#### (1→3)-β-D-glucan

(1→3)-β-D-glucan is a fungal cell wall component, and it also occurs in the cell wall of certain bacteria and high plants [337]. (1→3)-β-D-glucan is a water insoluble D-glucose polysaccharide linked by β (1→3) glycosidic bonds. In fungal cell walls, a (1→3)-β-D-glucan molecule comprises of a (1→3) β-D-glucose backbone and numerous branches attached to the backbone at (1→6) positions. For its ubiquitous existence in fungi, (1→3)-

$\beta$ -D-glucan is selected as an indicator of mold contamination as direct, accurate mold measurement can be difficult [338].

The health implications of (1→3)- $\beta$ -D-glucans vary with molecular weight, shape, structure, and source, and are not always detrimental. As they can activate immune systems, (1→3)- $\beta$ -D-glucans were occasionally used as medicines or supplements for cancer therapy and infection prevention [337]. Inhalation of (1→3)- $\beta$ -D-glucans, however, is believed to exert adverse human health effects [21]. Similar to endotoxins, (1→3)- $\beta$ -D-glucans are non-allergenic but strongly inflammatory [339]. Airborne (1→3)- $\beta$ -D-glucans have been associated with atopy and respiratory symptoms in humans, including non-allergic asthma, airway inflammation, and deteriorated pulmonary functions [340,341]. (1→3)- $\beta$ -D-glucans and endotoxins may have synergistic health effects in promoting the secretion of cytokines [342]. No (1→3)- $\beta$ -D-glucan exposure limits have been available for the lack of essential dose-response data.

Airborne (1→3)- $\beta$ -D-glucans primarily originate from fungi. Relatively high temperature and humidity levels in swine barns are suitable for the growth of many fungal species. Fungal genera *Aspergillus*, *Penicillium*, and *Mucor* were observed by Ref. [327] in the air of swine barns in four European countries. Ref. [159] reported fungal genus *Penicillium* in PM sampled from a finisher barn in Kansas. Both studies measured culturable fungi only. The majority of airborne fungal spores can be non-culturable [343]. Using 18S rRNA clone library technology, [87] found the dominance of fungal genera *Aspergillus* and *Eurotium* in TSP sampled from a gestation barn in Denmark. Ref. [220] studied airborne fungal composition in eight barns in South Korea with ITS NGS and found that phyla Ascomycota, Basidiomycota, Zygomycota, and Glomeromycota were dominant. They further identified the top 20 most abundant fungal genera.

Only six existing studies measured airborne (1→3)- $\beta$ -D-glucans concentrations in swine barns (Table 14). Due to limited data availability, it remains uncertain whether the findings derived from these studies are representative of average barns. The measured concentrations associated with large PM (inhalable and TSP) ranged from 0.5 [181] to 38,490 ng m<sup>-3</sup> [45] and tended to decrease in recent publications. Ref. [86] analyzed inhalable and respirable (1→3)- $\beta$ -D-glucan samples from 30 swine farms in Poland. Through the statistical analysis, they found that inhalable and respirable (1→3)- $\beta$ -D-glucan concentrations were significantly affected by manure disposal, feeding method, and flooring type (with versus without beddings). Beddings, hand feeding, and manure scraping were found to significantly increase the concentrations of both inhalable and respirable (1→3)- $\beta$ -D-glucans. The use of manure pits and slatted floors significantly reduced airborne (1→3)- $\beta$ -D-glucan concentrations. Ref. [332] collected TSP samples from 12 swine barns in Illinois. The overall highest (1→3)- $\beta$ -D-glucan concentrations occurred in finisher barns while the lowest concentrations occurred in farrowing barns. Similar to endotoxins, (1→3)- $\beta$ -D-glucans had higher airborne concentrations in winter than summer. (1→3)- $\beta$ -D-glucan loadings in TSP (ng mg<sup>-1</sup>) were lower in winter possibly because fungal growth was suppressed by cold temperatures.

**Table 14.** Airborne (1→3)- $\beta$ -D-glucan concentrations in swine barns—a summary of relevant studies since 1990.

Reference	(1→3)- $\beta$ -D-glucan Concentration (ng m <sup>-3</sup> ) <sup>a,b</sup>	Barn and Ventilation Type	Location	Season	Analytical Method
[45] <sup>c</sup>	Inhalable: BDL-38,490; 4340 (GM); 3.4 (GSD)	n/a; n/a	Netherlands	All seasons	Inhibition enzyme immunoassay
[344]	Inhalable: 33–410	n/a; n/a	Germany	n/a	Monoclonal antibody-based two-site enzyme immunoassay
[86]	Inhalable: 446 ± 724 (14–3594); 190 (GM); 3.90 (GSD) Respirable: 124 ± 183 (1–703); 37.0 (GM); 6.80 (GSD)	n/a; MV and NV	Poland	n/a	Chromogenic kinetic LAL assay

Table 14. Cont.

Reference	(1→3)-β-D-glucan Concentration (ng m <sup>-3</sup> ) <sup>a,b</sup>	Barn and Ventilation Type	Location	Season	Analytical Method
[332]	TSP: 25.2 ± 20.2 (3.7–50.8); 17.2 (GM); 2.71 (GSD)	Gestation; MV	Illinois	All seasons	Chromogenic kinetic LAL assay
	TSP: 21.2 ± 20.2 (2.4–50.7); 12.5 (GM); 3.50 (GSD)	Farrowing; MV		All seasons	
	TSP: 34.9 ± 48.4 (3.8–140.1); 16.3 (GM); 3.58 (GSD)	Nursery; MV		All seasons	
[181]	TSP: 32.7 ± 22.0 (3.9–65.0); 24.3 (GM); 2.62 (GSD)	Finisher; MV	Taiwan	All seasons	Chromogenic kinetic LAL assay
	>1.8 μm: 9.5 (3.6–14) 1–1.8 μm: 3.0 (0.5–53) <1 μm: 2 (8.0–9.0) Total: 12 (10–71)	n/a, n/a		Summer	
[334] <sup>c</sup>	Inhalable: 33.5 (GM); 11.0 (GSD)	n/a, n/a	Iowa	Spring, fall	Chromogenic kinetic LAL assay

<sup>a</sup> (1→3)-β-D-glucan is measured as a component of particles (e.g., inhalable and respirable). <sup>b</sup> Concentrations are expressed in the default forms of arithmetic mean, arithmetic mean ± standard deviation, or concentration range. Other statistics, including GM and GSD, are given when relevant data are available. A note in the parenthesis specifies the type of reported data. <sup>c</sup> Personal exposure samples.

### Other Bioaerosol Markers

Proteins are essential constituents of organisms and, thus, are occasionally selected as a general marker for bioaerosols [345]. Ref. [31] measured crude-protein contents in finisher barn PM samples and reported an average content of 28.7 ± 2.7%. Ref. [36] collected TSP samples from 21 swine barns in Iowa and found that total proteins accounted for 23% of dry mass in collected TSP samples. However, due to the high protein contents in the feed, it remains questionable whether total proteins are a valid bioaerosol marker. Total proteins were also used as a measure of organics during swine barn organic dust extract (ODE) preparation [346,347]. A swine barn ODE was prepared primarily for toxicity research (through cell or tissue experiments). In addition to total proteins, markers in Table 12, such as peptidoglycan, muramic acids, 3-hydroxy fatty acids, and ergosterol, were also analyzed for their presence and concentrations in swine barn ODEs [348]. However, no translation of the analysis results to airborne bioaerosol concentrations was reported. Ref. [349] analyzed muramic acid in swine barn PM samples with gas chromatograph-mass spectrometry (GC-MS) to estimate the airborne concentration of peptidoglycans. An average concentration of 6.5 μg m<sup>-3</sup> (range: 2.7–13 μg m<sup>-3</sup>) was reported.

### 3.3. Chemical Characteristics

#### 3.3.1. Elemental Composition

Elemental composition analysis is extensively being conducted for ambient PM. Examples of such efforts in the U.S. include Chemical Speciation Network (CSN) and Interagency Monitoring of Protected Visual Environments (IMPROVE), each with hundreds of monitoring stations. Elemental composition analysis has also been conducted for various PM sources and the acquired source chemical profiles are included in SPECIATE, a USEPA's database of source profiles.

The effort to analyze chemical elements in swine barn PM can date back to Ref. [350]. PM samples were collected on a glass fiber filter for spectrometric analysis. Nine elements were identified and quantified: Ca, Mg, P, Al, Cu, Na, Fe, Si, and B, with Ca being the most abundant. For swine barn PM, the analysis of elemental composition can serve multiple purposes. First, it improves our understanding of PM's health and environmental implications. Secondly, elemental composition can be used as the fingerprint of swine barn PM, thereby facilitating PM source apportionment in areas with intensive pork production. Thirdly, elemental composition, along with other physical and chemical information, enables the identification of PM origins in swine barns, thereby promoting in-barn PM management. Despite its scientific importance, only a few studies analyzed the elemental composition of PM in swine barns (Table 15). Differing in measurement methods, they are

summarized into two categories: (1) elemental composition of individual particles and (2) elemental composition of bulk PM. Other than the listed, two studies also examined the elemental composition of swine barn PM [131,351]. However, both collected PM samples from the outside of barns and, thus, are excluded in the table.

**Table 15.** Elemental composition of swine barn PM—a summary of studies since 1990.

Reference	Elements Detected or Quantitated	Barn and Ventilation Type	Location	Major Findings or Notes
Composition of individual particles				
[125]	PM <sub>0.18–0.35</sub> , PM <sub>0.35–0.65</sub> , PM <sub>0.65–1.2</sub> , PM <sub>1.2–3.5</sub> and PM <sub>3.5–10</sub> : C, O, Al, S, N, Na, Mg, P, K, Ca	n/a; n/a	Germany	-
[80]	PM <sub>2.5–10</sub> and PM <sub>2.5</sub> : N, Na, Mg, Al, Si, P, S, Cl, K, Ca, Fe, Ni, Cu, Zn, Ag, Pb, Sn, Cr, Co, Ba, Br, Ti, V, Sb, Au	Nursery, grower–finisher, gestation; MV	Netherlands	The study focused on source apportionment; PM was not the true in-barn PM but suspended from in-barn materials.
[148]	PM <sub>2.5–10</sub> : P, N, K, S, Cl, Al, Ca, Cr, Na, Mg, Ba, Fe PM <sub>2.5</sub> : P, N, Cl, S, K, Si, Na, Al, Ca, Mg, Sn	Nursery, grower–finisher, farrowing, gestation; MV	Netherlands	PM sources varied in elemental composition; PM was suspended from in-barn materials.
[104]	PM <sub>2.5</sub> : C, N, O, Na, Mg, Si, P, K, Ca, Fe, Zn	Nursery, finisher; MV	China	Only four scans were conducted.
Composition of bulk PM				
[124]	TSP: N, P, K, Cl, Na	Grower–finisher; MV	Netherlands	The elemental composition of PM was close to that of settled dust; PM was suspended from in-barn materials.
[85]	PM <sub>10</sub> : Al, B, Ba, Ca, Cr, Cu, Fe, K, Mg, Mn, Na, Ni, P, S, Si, Sr, Ti, Zn PM <sub>2.5</sub> : Al, B, Ba, Ca, Co, Cr, Cu, Fe, K, Mg, Mn, Na, P, S, Si, Sr, Ti, Zn	Farrowing; MV	Illinois	The elements accounted for on average 13.0% in PM <sub>10</sub> and 10.3% in PM <sub>2.5</sub> by mass.
	PM <sub>10</sub> : Al, B, Ba, Ca, Co, Cr, Cu, Fe, K, Mg, Mn, Na, P, S, Si, Sr, Ti, Zn PM <sub>2.5</sub> : Al, B, Ba, Ca, Co, Cr, Cu, Fe, K, Mg, Mn, Na, P, S, Si, Sr, Ti, Zn	Gestation; MV		The elements accounted for on average 9.98% in PM <sub>10</sub> and 8.92% in PM <sub>2.5</sub> by mass.
	PM <sub>10</sub> : Al, B, Ba, Ca, Cr, Cu, Fe, K, Mg, Mn, Na, P, S, Si, Sr, Ti, Zn PM <sub>2.5</sub> : Al, B, Ba, Ca, Co, Cr, Cu, Fe, K, Mg, Mn, Na, P, S, Si, Sr, Ti, Zn	Nursery; MV		The elements accounted for on average 7.83% in PM <sub>10</sub> and 7.81% in PM <sub>2.5</sub> by mass.
	PM <sub>10</sub> : Al, B, Ba, Ca, Co, Cr, Cu, Fe, K, Mg, Mn, Na, Ni, P, S, Si, Sr, Ti, Zn PM <sub>2.5</sub> : Al, B, Ba, Ca, Cr, Cu, Fe, K, Mg, Mn, Na, Ni, P, S, Si, Sr, Ti, Zn	Finisher; MV		The elements accounted for on average 8.09% in PM <sub>10</sub> and 7.84% in PM <sub>2.5</sub> by mass.

It is hard to generalize the findings from the previous studies due to differences in barn conditions, sampling setup, and analytical methods. They are individually discussed as follows:

- Ref. [125] was an exploratory study, with the purpose to test and showcase two PM characterization technologies (electron probe X-ray microanalysis (EPXMA) and laser ablation microprobe mass analysis (LAMMA)) for their applicability to swine barn PM. Both technologies target single individual particles. No quantitative results were presented.
- Refs. [80,148] focused on the development of source profiles for several major PM sources (feed, skin, and feces) in swine barns, with a combination of size, shape, and elemental composition information derived from scanning electron microscope-energy dispersive X-ray spectroscopy (SEM-EDX) analysis. The developed source profiles were then used to determine the contribution of different sources.
- Ref. [104] used PM elemental composition, along with morphology information acquired from SEM-EDX, to assign individual particles to different origins. Carbon (C) and oxygen (O) were found to be the most abundant elements, indicating the PM’s organic origins.

- Ref. [124] compared the element composition of PM with those of settle dust, feed, skins, and feces, with aims to identify the major sources of PM in swine barns. The average N, phosphorus (P), and potassium (K) content in PM were 6.703%, 1.47%, and 2.78%, respectively. No information about analytical methods was presented.
- Ref. [85] examined the elemental composition of multi-season PM samples from 12 swine barns using inductively coupled plasma-atomic emission spectroscopy (ICP-AES; also known as ICP-OES). The objectives were to study the effect of barn types and seasons on PM elemental composition and to develop PM source chemical profiles for typical barn types in the U.S. Midwest. While no significant seasonality was noted, PM<sub>10</sub>/PM<sub>2.5</sub> compositions differed significantly with barn types. The lower mineral (inorganic) content in PM<sub>2.5</sub> than PM<sub>10</sub> samples suggests that PM<sub>2.5</sub> in swine barns was of more organic origins than PM<sub>10</sub>. No toxic heavy metals (e.g., lead, cadmium, or arsenic) were detected. Among all the elements quantified, Ca was the most abundant. This is consistent with the early finding by Ref. [350].

It is noteworthy that several studies did not analyze true PM samples [80,124,148]. Instead, the samples analyzed were suspended from pre-collected in-barn dust or powder materials (e.g., feed, feces, and skin). It remains uncertain whether and to what degree this indirect method would affect the analysis results. Additionally, an SEM requires a high vacuum in its testing chamber unless it is an environmental SEM (Note: No previous studies appear to use it). A high vacuum environment would result in the volatilization loss of volatiles and semivolatiles in PM [352], creating a measurement bias.

It is important to note that swine barn PM holds unique physical and chemical properties. Thus, the sampling and analysis protocol for ambient PM may not necessarily apply [85]. Future efforts should be made to develop a SOP for swine barn PM, produce more and complete data sets, and interface the acquired measurement data with the databases for ambient PM and other PM sources.

### 3.3.2. PM-Borne Odors

PM is considered to play a critical role in the propagation of odors from animal barns [10]. As particles are much larger than gas molecules ( $\sim 10^{-4}$   $\mu\text{m}$ ), their generation, transport, and transformation inside and outside of swine barns are substantially different. For example, PM may deposit on hairs, clothes, tools, and vehicles, be carried around by these objects, and create persistent odor nuisance [10]. When reaching human and animal receptors, PM and gases also behave differently regarding their transport and fate in respiratory systems [353]. As a result, the management and mitigation of PM-borne odors demand different strategies.

The first study of PM-borne odorants in swine barns was conducted by Dr. Day and his colleagues at the University of Illinois at Urbana-Champaign [350]. Besides air samples, they also collected PM samples on glass fiber filters for odor assessment. Although the further chemical analysis was unsuccessful due to then methodology constraints, they found a strong odor from heated PM samples and PM extracts in methanol. This pioneering work spurred several follow-up studies in the 1970s and 1980s [36,354–358].

Ref. [354] extracted settled dust from swine barns in water, concentrated the extract through distillation, and analyzed the dust's odorant composition using thin-layer chromatography and gas-liquid chromatography. A total of 19 odorous compounds were identified, including acids, phenols, aldehydes, and ketones. In the authors' later study [355], two swine barn PM samples were collected using an electrostatic precipitator (ESP) and extracted in diethyl ether. The extracts were further derivated, purified, and then analyzed using gas chromatography (GC). Thirty-four odorants were identified and quantified, including 15 acids, 16 carbonyls (ketones and aldehydes), and three phenols. Ref. [356] tested a wet scrubber for its performance of odor reduction in a finisher barn and found that odor reduction highly correlated with PM removal. Ref. [357] collected settled dust from a finisher barn in Germany and extracted the dust in ethanol. The extract was alkalized with NaOH for volatile fatty acid (VFA) separation and then acidified to recover phenols.

Using GC, six VFAs and five phenolic/indolic compounds were identified and quantified. Ref. [36] reported that settled dust from swine barns in Iowa contained 3.9 mg NH<sub>3</sub> per gram of dust. Ref. [358] summarized past publications and concluded that >60 odorants could exist in livestock barn PM samples.

Additional measurement efforts have been reported since 1990, as summarized in Table 16. Besides field monitoring, modeling of odors carried by swine barn PM was also attempted, including odor adsorption dynamics, PM deposition, and inhalation dose simulation [56,359–361]. All of the modeling work was conducted by Dr. Liao and his group at the National Taiwan University in Taiwan. However, no model calibration or validation has been conducted, likely due to the lack of essential experimental data.

**Table 16.** Measurement of PM-borne odors in swine barns since 1990.

Reference	Odorant(s) Identified and/or Qualified	Analytical Method <sup>a</sup>	Barn and Ventilation Type	Location	Major Findings and/or Notes
[362]	Settled dust: >100 VOCs	Solvent extraction followed by GC-FID	Finisher; MV	Illinois	No compound identification was conducted. VOC diversity was estimated from the number of peaks.
[363] <sup>b</sup>	Settled dust: 10 acids, 2 phenols, indole, and skatole	Solvent extraction followed by GC-FID	n/a; n/a	North Carolina	Ozonation and manure pit additives significantly reduced the concentrations of many odorants.
[364] <sup>b</sup>	Settled dust: 5 aldehydes, 1-octane, and H <sub>2</sub> S	Solvent extraction followed by GC-MS	n/a; n/a	Georgia	Settled dust was separated into several size ranges before analysis.
[365] <sup>b,c</sup>	TSP (qualitative): 13 acids, 11 ketones, 16 aldehydes, 9 esters, 3 phenols, 5 nitrogen-containing compounds, 13 hydrocarbons, 3 ethers, methylene chloride, dimethyl disulfide, and 3 others TSP (quantitative): 3 acids and 2 aldehydes TSP: 4 alkanes, 4 alcohols, 8 aldehydes, 7 ketones, 8 acids, 6 amines and nitrogen heterocycles, 3 sulfides and thiols, 7 aromatics, and 2-pentylfuran PM <sub>10</sub> : 3 alkanes, 4 alcohols, 8 aldehydes, 7 ketones, 7 acids, 3 amines and nitrogen heterocycles, 3 sulfides and thiols, 7 aromatics, and 2-pentylfuran	Qualitative: solvent extraction, SPME <sup>2</sup> and P&T, followed by GC-MS Quantitative: P&T followed by GC-MS	Grower–finisher; MV	Kansas	Solvent extraction, SPME, and P&T yielded different identification results. Among the quantitated odorants, acetic acid was the most abundant.
[366]	PM <sub>2.5</sub> : 1 alkanes, 2 alcohols, 6 aldehydes, 2 ketones, 2 acid, 3 amines and nitrogen heterocycles, 2 sulfides and thiols, and 7 aromatics PM <sub>1</sub> : 3 alkanes, 2 alcohols, 3 aldehydes, 3 ketones, 8 acids, 3 amines and nitrogen heterocycles, 2 sulfides and thiols, and 7 aromatics	Headspace SPME followed by GC-MS for odorants	Grower–finisher; MV	Iowa	PM <sub>1</sub> contained higher odorant contents (by %) than PM <sub>10</sub> and then TSP; Carboxen/PDMS was the most effective SPME fiber; no quantitation was conducted and odorant content comparisons were based on peak areas.
[146] <sup>b</sup>	Settled dust: NH <sub>3</sub> and odor	TD followed by NH <sub>3</sub> analyzer for NH <sub>3</sub> ; TD followed by olfactometry for odor	Farrowing, nursery, grower, finisher; MV	Illinois	An average odor emission rate was 1.43 ± 0.37 OU min <sup>-1</sup> g <sup>-1</sup> dust; no quantitative information about NH <sub>3</sub> was available; odor-carrying capacity was related to barn type.

Table 16. Cont.

Reference	Odorant(s) Identified and/or Qualified	Analytical Method <sup>a</sup>	Barn and Ventilation Type	Location	Major Findings and/or Notes
[367] <sup>b</sup>	PM of unknown size: 8 acids, 5 phenols, 2 ketones, and 2 sulfides	TD-GC-MS	Boars, gilts, finisher; n/a	Denmark	A dual filter (front plus backup filters) setup and denuders were used to study gas-particle partitioning of odorants and their adsorption on filters.
[277] <sup>b</sup>	TSP: 18 aldehydes, 1 ketone, 8 alcohols, 20 acids, 7 phenols, and 3 nitrogen-containing compounds PM <sub>10</sub> : 18 aldehydes, 1 ketone, 8 alcohols, 20 acids, 7 phenols, and 3 nitrogen-containing compounds	Solvent extraction followed by stable isotope dilution GC-MS	Farrowing, gestation, nursery, finisher; MV	Illinois	PM <sub>10</sub> contained higher odorant contents (by %) than TSP; For both TSP and PM <sub>10</sub> , their odorant composition varied significantly with barn types and seasons; Among the odorant quantitated, acetic acid is the most abundant. A column packed with PM was challenged with acetic acid, butanoic acid, phenol, and dimethyl disulfide to study their gas-particle partitioning using SIFT-MS; <sup>e</sup> these odorants were concentrated in PM but occurred at much lower fractions in the particle than the gas phase.
[368] <sup>d</sup>	PM <sub>10</sub> (qualitative): 7 acids, 7 aldehydes, 3 alcohols, 2 phenols, 2 esters, 4 heterocycles, 2 sulfur-containing compounds, 1 amine, 1 ketone, and 1 terpene	SPME followed by GC-MS	n/a; n/a	Belgium	

<sup>a</sup> GC-FID—gas chromatography-flame ionization detector; GC-MS—gas chromatography-mass spectrometry; SPME—solid-phase micro-extraction; P&T—purge and trap; TD—thermal desorption; TD-GC-MS—thermal desorption gas chromatography-mass spectrometry. <sup>b</sup> Quantitative information was available in the publication. <sup>c</sup> The authors reported a similar if not the same effort on a conference [369]. However, the sampling method was slightly different and involved the use of backup glass fiber filters. <sup>d</sup> Quantitative analysis was conducted for amino acids and triacylglycerols. <sup>e</sup> SIFT-MS: selective ion flow tube mass spectrometry.

Numerous odorants have been detected in swine barn PM, including NH<sub>3</sub>, H<sub>2</sub>S, organic acids, alcohols, aldehydes, ketones, phenols, nitrogen-containing organic compounds, and sulfur-containing organic compounds. Table S1 lists all the detected odorants (159 in total) and relevant publications. Many of these odorants were also found in the air of swine barns and/or in swine manure [370].

Upon the analysis of the previous publications, the following observations are made:

- All but three measurement efforts (including those before 1990) were conducted in the U.S. The only three non-U.S. ones were from Europe. Several simulation papers were published by researchers from Taiwan but no experimental data were available. The existing publications covered a broad range of focus subjects, including testing of concepts or methodology development [146,354,355,357,364,366], regular field monitoring [36,277,365], gas-particle partitioning [367,368], transport and exposure modeling [56,359–361], and mitigation technologies [356,363].
- Nearly half of the publications, especially early ones, provide no quantitative data. Semi-quantitative analysis was occasionally conducted based on the chromatography peak areas of individual odorants, e.g., [366]. Quantitative data were available from ten studies, including seven studies since 1990. However, most of the data were from one or a few farm visits and extensive field sampling is largely lacking. The only multi-farm, multi-season monitoring effort was [277].
- PM samples of various size fractions, including TSP [277,355,365,366], PM<sub>10</sub> [277,366,368], PM<sub>2.5</sub> [366], and PM<sub>1</sub> [366], were collected for odorant analysis. In many early studies, settled dust was selected as a surrogate for PM [36,146,354,357,362–364]. However, settled dust and PM could differ in size and origins. Thus, it is uncertain whether

the findings derived from settled dust also apply to actual PM samples. No direct comparison between PM and settled dust has been reported.

- Aldehydes, acids, phenols, and nitrogen-containing compounds were often detected in collected PM and settled dust samples. The most frequently detected odorants ( $\geq 7$  out of 12 publications in Table S1) were hexanal, acetic acid, propanoic acid, butanoic acid, pentanoic acid, phenol, *p*-cresol, indole, and skatole. Among the odorants quantified, acetic acid was the most abundant [277,355,357,365,367]. The second most abundant odorant differed among publications, including propanoic acid [355,365], *p*-cresol [357], nonanoic acid [363], pentanoic acid [367], and ethanol [277]. Known for their malodors, sulfur-containing compounds were found to occur at very low concentrations in PM. Many of these compounds are highly volatile and, thus, would exist predominantly in gaseous forms [277,367].
- The detected PM-borne odorants vary considerably in their sensory characteristics and odor thresholds [277,366]. It is noteworthy that the odor thresholds compiled in the literature are for gaseous (odor thresholds in the air (OTA)) or waterborne odorants (odor thresholds in water (OTW)) only. They may not apply to PM-borne odorants, for two reasons. First, the gas-particle partition coefficient ( $K_p$ ; defined as the ratio of the mass fraction of a compound in PM to its concentration in the gas phase) of an odorant governs the equilibrium concentration of the odorant in the gaseous form. A PM-borne odorant with a low OTA (meaning that it carries a strong smell as a gas) could have a high  $K_p$  and therefore occur at low gaseous concentrations. Secondly, PM can be trapped and accumulated in the human's nasal cavity before being sensed by olfactory cells. Odor perception, in this case, could be substantially different from that for gaseous odor molecules [277,355]. Odor activity values (OAVs) were occasionally calculated in the literature by normalizing PM-borne odorant concentrations with OTAs [277,366]. However, they do not represent actual PM-associated odor levels, for the reasons discussed above.
- The detected odorants vary substantially in their sources [277]. For example, fatty acids can be generated from the bacterial conversion of simple carbohydrates, whereas phenols can be degraded from tyrosine [371,372]. Indole and skatole, two malodorants of great concern, were believed to be of fecal origins and produced from the degradation of tryptophan or indole-3-carboxylic acid by gastrointestinal microbes [371,372]. Thus, their occurrence in swine barn PM suggests that PM could partly originate from swine feces, which is consistent with the findings from PM source studies. Some of the PM-borne odorants were also found in swine feed but at lower mass fractions [373]. However, it remains uncertain whether these odorants were of feed origins or came from other sources but becoming sorbed or deposited to feed particles.
- Little is known about the total concentrations or fractions of odorants in swine barn PM. This is primarily due to methodology constraints. Various methods have been adopted, but none of them can detect and quantify all possible odorants. From the existing data—though they are incomplete as explained—it appears that odorants account for only a small fraction of PM mass. Ref. [355] reported that ~2% of PM mass was attributed to the odorants quantified. Ref. [357] analyzed 11 odorants, and they accounted for ~0.09% of settle dust by mass. The five odorants quantified by Ref. [365] accounted for ~0.01% of collected TSP samples. Ref. [277] quantified 57 odorants in PM sampled from 12 swine barns. The total mass fractions of these odorants were on average 2.86% in farrowing TSP, 5.55% in farrowing PM<sub>10</sub>, 3.24% in gestation TSP, 6.24% in gestation PM<sub>10</sub>, 2.53% in nursery TSP, 4.28% in nursery PM<sub>10</sub>, 2.65% in finisher TSP, and 6.58% in finisher PM<sub>10</sub>. PM<sub>10</sub> contained significantly higher odorant fractions than TSP. A similar observation (PM<sub>1</sub> > PM<sub>10</sub> > TSP) was made by Ref. [366] through a comparison of GC-MS peak areas. The finding is consistent with that smaller particles in swine barns were of more fecal origins (Refer to Section 3.4).
- Many odorants occurred at a lower concentration (e.g.,  $\mu\text{g}$  odorant per  $\text{m}^3$  of barn air) in PM than in gaseous forms. Direct comparisons of PM-borne with gaseous odorants

showed that most detected odorants existed primarily in the gas phase [367,368]. The comparison result was further assessed through thermodynamic simulation. Using experimentally determined particle-to-gas partition coefficients ( $K_{PM/air}$ ; a dimensionless version of  $K_p$ ), it was calculated that the total mass fraction of PM-sorbed acetic acid, butanoic acid, dimethyl disulfide, and phenol would not exceed 0.11% of PM mass at  $1 \text{ mg m}^{-3} \text{ PM}_{10}$  [368]. The calculated odorant fractions were lower than some field measurement results [277,355]. Both  $K_p$  and  $K_{PM/air}$  are equilibrium constants. For PM inside swine barns (which is freshly generated and close to its sources), it is unclear whether gas-particle partitioning would have reached equilibriums. According to Ref. [367], odorants with higher  $K_p$  values (e.g., hexanoic acid) would be enriched in PM and those with lower  $K_p$  values (e.g., 3-hydroxybutanone) would stay predominately in the gas phase. Indirect comparisons between PM-borne and gaseous odorants, in which part of the data was from the literature, also led to the same conclusion that many odorants occurred primarily in the gas phase rather than in PM [277,355]. For  $\text{NH}_3$ , assuming a TSP concentration of  $2 \text{ mg m}^{-3}$ , the  $3.9 \text{ mg NH}_3 \text{ g}^{-1}$  dust value reported by Ref. [36] can be translated into a PM-borne  $\text{NH}_3$  concentration of  $7.8 \times 10^{-3} \text{ mg m}^{-3}$ , which is  $\sim 3$  orders of magnitude smaller than typical  $\text{NH}_3$  gas concentrations in swine barns.

- PM can concentrate and amplify odors. PM carried substantially larger amounts of odorants than the same volume of air [355,368]. Thus, PM deposited on the olfactory region in the human's nasal cavity will trigger an odor perception equivalent to a much larger volume of air. The enrichment ratio of an odorant in PM can be experimentally determined [355] or calculated from its gas-particle partition coefficient ( $K_p$  or  $K_{PM/air}$ ; [368]). Furthermore, the nasal cavity serves as a physical barrier for PM. The turns and curves inside the cavity lead to the aerodynamic separation of PM from inhaled air and the deposition of PM in the olfactory region. Compared to gaseous odorants, PM-borne odorants, especially those carried by large particles, are more likely perceived by olfactory cells, thereby amplifying swine odors [355]. However, no quantitative information about the deposition of PM-borne odors versus gaseous odors in the olfactory region has been available.
- Little is known about the effect of barn types and seasons. Only one publication investigated PM-borne odorants sampled from multiple barn types and multiple seasons [277]. Results showed a large influence of barn types on odorant composition. Specifically, TSP and  $\text{PM}_{10}$  samples from nursery barns were significantly different from those from farrowing and gestation barns; while the samples from finisher barns sit somewhere in-midst. Overall, PM samples from swine barns differed significantly from those from turkey and layer hen barns. Seasons also exhibited a significant influence. The total mass odorant fraction in both TSP and  $\text{PM}_{10}$  increased as the weather warmed up. Summer PM samples contained more hexanal, heptanal, nonanal, ethanol, 1-octanol, and 2-ethyl-1-hexanol but lesser acetic acid, propanoic acid, and 3-methyl-butanoic acid than winter samples. Similar seasonality was observed for odorants in swine feeds [373], a major PM source in swine barns.
- The modeling work has yet to be validated. As aforementioned, modeling papers were available regarding the sorption and transport of PM-borne odors in swine barns, as well as human exposure to the odors [56,359–361]. No calibration or validation has been conducted. An in-depth analysis of the models is beyond our expertise and the scope of this review effort. Various assumptions were taken during model development. For example, it was assumed that the occurrence of odorants in PM was attributed to gas adsorption only and that the adsorption equilibrium and kinetics could be described by the Freundlich isotherm and the Langmuir–Hinshelwood equation, respectively. A revisit to these assumptions might be necessary given the research advances in the past 20 years. How to effectively utilize the modeling framework and interface it with other management models or tools remains an immense challenge.

Odors are a top air quality challenge facing pork producers. PM-borne odors have intrigued academia and industry for decades. After 55 years' research (since the pioneering work by Ref. [350]), our understanding of PM-borne odors and odorants has substantially improved. However, many fundamental questions remain unanswered, e.g., to what degree does PM contribute to a downwind odor nuisance? How are PM-borne odors transported and decayed in the environment? What is the most cost-effective way to de-odor the smelly PM that sticks on cloth, vehicles, or walls? Answers to these questions will enable improved management of swine odors and therefore benefit pork producers in the long run.

### 3.3.3. Ions and Others

Ambient PM composition is often broken down into trace elements (excluding C, O, and H), soluble ions, and carbonaceous material [23]. The corresponding analyses are being conducted for ambient PM samples gathered from hundreds of air monitoring stations in the world. Carbonaceous material consists of organic carbon (OC) and elemental carbon (EC), where OC refers to carbon in organic matter and EC refers to carbon in elemental forms. No OC/EC analysis has been performed for swine barn PM. As PM in swine barns primarily originates from feeds and animal feces, a high OC content and a low EC content are anticipated.

#### Soluble Ions

Soluble ions include  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$ , and  $\text{NH}_4^+$ . Other anions (e.g.,  $\text{PO}_4^{3-}$ ) and cations (e.g.,  $\text{K}^+$  and  $\text{Na}^+$ ) are often not analyzed because they can be quantified through elemental analysis. Only two studies analyzed soluble ions in swine barn PM. Ref. [85] collected  $\text{PM}_{10}$  and  $\text{PM}_{2.5}$  samples from 12 swine barns in Illinois and examined their  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$ , and  $\text{NH}_4^+$  concentrations using ion chromatography (IC). A significant effect of barn types was observed. These ions accounted for on average 3.02% of  $\text{PM}_{10}$  and 3.42% of  $\text{PM}_{2.5}$  in farrowing barns, 3.93% of  $\text{PM}_{10}$  and 4.94% of  $\text{PM}_{2.5}$  in gestation barns, 3.89% of  $\text{PM}_{10}$  and 4.90% of  $\text{PM}_{2.5}$  in nursery barns, and 3.46% of  $\text{PM}_{10}$  and 3.98% of  $\text{PM}_{2.5}$  in finisher barns by mass. No significant seasonality was found for  $\text{PM}_{10}$ ; while summer  $\text{PM}_{2.5}$  samples had overall higher ion contents than winter samples. Ref. [99] analyzed  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$ , and  $\text{NH}_4^+$  in TSP and  $\text{PM}_{10}$  samples from a finisher barn in China. A significant seasonality was observed, with summer TSP and  $\text{PM}_{10}$  samples containing substantially higher  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$  and  $\text{NH}_4^+$  contents than fall and winter samples. The authors assigned the sum of  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$  and  $\text{NH}_4^+$  as secondary inorganic aerosols (SIAs) and found that they accounted for on average 12.0% of TSP mass and 13.9% of  $\text{PM}_{10}$  mass—which are greater than those reported by [85]. Caution should be taken when discussing SIAs in swine barns. It is an important topic, as  $\text{NH}_3$  emitted from animal barns is an essential precursor for SIA formation and SIAs account for a significant fraction of atmospheric  $\text{PM}_{2.5}$ . However, the IC quantification data from both studies [85,99] showed that  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  were not fully neutralized by  $\text{NH}_4^+$  in PM. Thus, part of  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  could originate from minerals or nitrification and should not be included in the calculation of SIA mass.

The analysis of elements, soluble ions, and OC/EC in swine barns is important from the ambient air quality management standpoint. It will enable the inclusion of swine barns as a PM source in the EPA's SPECIATE database and PM receptor modeling as part of State Implementation Plans (SIPs). However, how to utilize the composition information for the benefit of pork production remains a question.

#### Others

Other tested swine barn PM components include moisture, ashes, lipids, proteins, and amino acids. Ref. [121] quantified ash contents (reported as inorganic contents) in TSP samples from 11 swine barns in Kansas using AOAC method 7.009 and found an average ash content of 13.1% (range: 7.3–18.2%), slightly lower than that in settled dust (13.5%, range: 8.7–33.2%). Ref. [124] reported that PM sampled from swine barns contained

7.90 ± 0.35% of moisture and 14.95 ± 0.35% of ash contents by mass; and their fractions were similar to those in settled dust, feces dust, and skin particle dust. No information about analytical methods was provided. Similar ash content levels (reported as minerals) were reported by [85] by summing all quantified elements and soluble ions. Total protein contents in swine barn PM were also measured [31,36], with the average content ranging from 23% to 28.7% (Refer to Section 3.2.5). The Kjeldahl method was selected for crude protein quantification by [31]. Commercial assays, such as Bradford assay and NanoOrange protein quantification kit, have been used to analyze total proteins in ambient PM. They should also work for swine barn PM characterization. Ref. [87] conducted a series of analyses to determine the composition of swine barn TSP samples (drying at 105 °C for moisture, ignition at 550 °C for volatile organic solids, CBQCA protein quantification kit for protein, anthrone carbohydrate method for carbohydrate, and GC-FID for fatty acids) and found an average moisture content of 14% and a volatile organic solid content of 83%. The volatile organic solids consisted of 82% carbohydrate, 21% protein, and 1% fatty acids. Ref. [368] analyzed lipids (triacylglycerols) in swine barn PM<sub>10</sub> samples by saponifying the lipids and converting produced fatty acids into their corresponding methyl esters (fatty acid methyl esters (FAME)). Using GC-FID, nine saturated and eleven unsaturated fatty acids (C6 to C20) were quantified, together accounting for 4.5% of PM<sub>10</sub> mass. The same study also hydrolyzed proteins in the PM<sub>10</sub> samples into amino acids and quantified amino acid derivatives using HPLC. Seventeen amino acids were quantified, attributing to 33.2% of PM<sub>10</sub> mass. Considering mass gains of proteins during hydrolysis, the value was consistent with early reports by [31,36].

#### 3.4. Sources

PM in animal barns is highly complex in composition and originates from various sources including feed, feces, animal skin and hair (feathers for poultry), beddings, construction materials, insect fragments, and microorganisms [24]. PM can be classified into inorganic, organic, and biological particles (often referred to as bioaerosols) according to their composition and origins. However, an individual particle can be a mixture of inorganic, organic, and biological components, or an agglomerate of small particles of inorganic, organic, and biological origins. In fact, many particles in animal barns are formed through the agglomeration of small particles possibly from different sources [374].

Source identification and apportionment constitute two consecutive steps for the research of particle origins. The former aims to identify the sources of particles while the latter aims to quantitate the respective contributions of identified sources. The results of source apportionment are often presented in the unit of particle number or mass percentage (%). Both source identification and apportionment can be conducted on a bulk scale (i.e., a collection of particles) or individual particles. Although an individual particle can be of multiple origins, many previous studies examined the shape, size, and/or composition of individual particles with optical or electron microscopy [36,121] and assigned their origins (e.g., as feed or fecal particles) accordingly.

Early studies found feed as a primary PM source in swine barns [31,36,121,375]. Ref. [31] compared the crude-protein (CP) contents in feed, settled dust, and PM under different swine diet conditions and found the CP contents in PM were consistently close to those in the feed and settled dust. Based on this bulk scale analysis, they concluded that the majority of PM in swine barns was feed particles. Ref. [375] collected PM samples from four pig pens in an experimental room and used an optical microscope to count and identify particles. They found that hair and skin accounted for a small portion of particles (by number) and assumed the remaining cubical or spherical particles to be of feed origins. Ref. [36] studied the sources of PM in 21 swine barns in Iowa using an optical microscope. They reported that PM was primarily from feed (starch, grain meal, plant trichomes, and corn silk; by particle number) and fecal materials (microorganisms, animal cells, and undigested feed), and the feed content increased with the size of pigs. The existence of dander, molds, pollens, insect fragments, and minerals in PM was also observed. Ref. [121]

collected PM samples from 11 swine barns in Kansas with eight visits to each barn and used an SEM and an optical microscope to identify and classify particles. They found that the majority of particles were from the feed. Around 65% (by number) of particles were grain meal particles and 13.5% were starch granules. Most of the starch granules were larger than 6.7  $\mu\text{m}$ . Grain meal particles were, on average, smaller, but most of them were still larger than 3.0  $\mu\text{m}$ . The study also revealed that PM composition varied with particle size and, in particular, the feed contents were lower for small particles than large particles. Ref. [124] collected PM, settled dust, feed dust, manure dust, and skin particle samples from a grower–finisher barn in the Netherlands and compared their contents of N, P, K, Cl, Na, ash, and dry matter. Through the analysis, they found that PM and settled dust shared nearly the same chemical composition and suggested that PM mass in swine barns primarily originated from feed and skin particles.

However, a later study in the Netherlands indicated that manure and skin could be greater PM sources than animal feeds in swine barns [80]. The study combined the morphology and elemental composition information of individual particles for source identification and apportionment. A high-resolution SEM-EDX was used for relevant analyses. Results showed that, by mass, most particles in swine barns originated from the skin (0–79% in  $\text{PM}_{2.5}$  and 0–71% in  $\text{PM}_{2.5-10}$ ) and manure (14–95% in  $\text{PM}_{2.5}$  and 23–92% in  $\text{PM}_{2.5-10}$ ). It should be noted that the findings derived from the study do not fully contradict those from earlier studies. First, Ref. [80] only investigated particles smaller than 10  $\mu\text{m}$  ( $\text{PM}_{2.5}$  and  $\text{PM}_{2.5-10}$ ) while many earlier studies considered particles of all sizes. According to Refs. [36,121], feed particles are usually large, e.g., >10  $\mu\text{m}$ . Secondly, the source contributions reported by Ref. [80] were in mass percentage, while many of the earlier studies presented their results in number percentage [36,121,375]. As PM regulations are based on mass concentrations, mass source apportionment results would be more useful for air quality management.

Source identification and apportionment provide valuable information for the development of control strategies. Once a major source is determined, measures can be implemented to suppress the source's PM generation, thereby reducing in-barn PM concentrations. For example, adding fat to animal feed was found effective in reducing the aerosolization of feed particles [376]. From the emission control standpoint, future source studies should focus on small particles, because once released from swine barns, large particles will quickly settle down, but the small particles will travel a relatively long distance in the atmosphere, reaching neighboring communities and creating potential air pollution problems.

It is noteworthy that different source apportionment methods are used for atmospheric PM. Microscopic morphology analysis of individual particles is used, but less often than receptor models. Receptor models rely on PM characteristics (e.g., chemical composition, size, and morphology of atmospheric PM) and source profiles (e.g., chemical composition, size, and morphology of source-emitted PM) to identify the contributing sources and quantitate their PM mass contributions. The commonly used receptor models include chemical mass balance (CMB), positive matrix factorization (PMF), and UNMIX. They are all statistical models and require no meteorology input. Receptor models are potentially useful for the study of PM sources in swine barns [12]. A discussion of receptor models is beyond the scope of this review. Further information can be found in [377,378].

## 4. Measurement Methodology

### 4.1. Mass Concentration Determination

In general, PM mass concentrations can be determined with two types of methods: gravimetric and real-time. The gravimetric method, also known as the time-average or integrated method, involves the collection of PM samples on a filter at a known sampling airflow rate over a known period (e.g., 24 h), and the weighing of the filter before and after the sample collection. Thus, the measurement result represents the average PM mass concentration over the entire sampling period. The real-time method, also known as the

online method, generates continuous or semi-continuous mass concentration readings by feeding PM-laden air to a sensing element or elements. It is noteworthy that “real-time” is an inaccurate generalization. In reality, many PM monitors are not truly real-time due to the time latency required for PM sensing.

#### 4.1.1. Gravimetric Methods

Various PM samplers were used for PM collection in swine barns (Table 17). Most of them are size-selective, i.e., they collect PM of a certain size range (e.g., PM<sub>10</sub>). This is achieved by using aerodynamic size separators to remove undesired PM, i.e., PM beyond the target size range. Two types of aerodynamic size separators were commonly used: impactors and cyclones. Both separate particles of different sizes based on their inertia (which is size-dependent) [24]. The PM samplers varied greatly in portability, cost, and sampling airflow rate. Among them, personal samplers were developed for PM exposure assessment in occupational environments. They are battery-powered and run at a relatively low sampling airflow rate. They are the most portable and usually the least expensive units, and can be worn by farm workers to assess their average PM exposure during a work shift. Other samplers must be installed at a fixed location, and many of them rely on AC power to run sampling pumps.

**Table 17.** Gravimetric PM samplers used in the past swine barn PM studies.

Gravimetric PM Sampler <sup>a,b</sup>	Size Separator	Available PM Size	Sampling Airflow Rate (LPM)	Manufacturer, City, Country	References
Ambient air particle sampler	Impactor, cyclone	PM <sub>10</sub> and PM <sub>2.5</sub>	16.7	Dandong Baite Instrument Co., Ltd., Dandong, China	[104]
BGI personal sampler	Cyclone	PM <sub>10</sub> , PM <sub>2.5</sub> , PM <sub>1</sub> , respirable, PM <sub>4</sub> , and PM <sub>0.8</sub>	Up to 5	Mesa Labs, Lakewood, CO, USA	[48,57]
Cassette only <sup>c</sup>	n/a <sup>d</sup>	TSP	n/a	Many vendors have filter cassettes available	[54,61,69,75,103,112,167,168]
Conical inhalable sampler	impactor	PM <sub>10</sub> , PM <sub>2.5</sub> , inhalable, and respirable	3.5	JS Holdings, Stevenage, UK	[89]
Harvard impactor (discontinued)	Impactor	PM <sub>10</sub> , PM <sub>2.5</sub> , and PM <sub>1</sub>	5–20	Air Diagnostics and Engineering Inc., Harrison, ME, USA	[85]
Isokinetic sampler	n/a <sup>d</sup>	TSP	n/a <sup>e</sup>	University of Illinois, Urbana, IL, USA	[96,332]
Low-volume PM <sub>10</sub> reference sampler <sup>f</sup>	Impactor	PM <sub>10</sub>	38.3	n/a	[95,379]
Medium-volume air sampler	Impactor, cyclone	PM <sub>10</sub> and PM <sub>2.5</sub>	16.7	Tianhong Co., Wuhan, China	[99]
MiniVol PM sampler	Impactor	TSP, PM <sub>10</sub> , and PM <sub>2.5</sub>	5	Airmetrics, Springfield, OR, USA	[60,380]
SKC Personal sampler	Impactor, cyclone	TSP, PM <sub>10</sub> , PM <sub>2.5</sub> , inhalable, and respirable	Up to 5	SKC Ltd., Blandford, UK	[39,52,57,60,62,65,73,77,78,105,167]
URG cyclone sampler	Cyclone	PM <sub>10</sub> and PM <sub>2.5</sub>	3–92	URG Corp., Chapel Hill, NC, USA	[94,95,379]
Virtual cascade impactor	Impactor	PM <sub>10</sub> and PM <sub>2.5–10</sub>	3.11	Helmut Und GmbH, Wetzlar, Germany	[80,148]
Zefon personal sampler	Cyclone	Respirable	1.7–2.5	Zefon International, Ocala, FL, USA	[55,64,72]

<sup>a</sup> Some publications contain no sampler information. They might use different samplers than the listed. <sup>b</sup> No federal reference method (FRM) gravimetric samplers were used in swine barns, likely due to their poor portability and high equipment costs. <sup>c</sup> No size separator is used. A closed-face or open-face filter cassette is connected to a timer-controlled vacuum pump for TSP sampling. <sup>d</sup> No size separator is required for TSP (total particle) sampling. <sup>e</sup> The sampling airflow rate is adjustable to achieve isokinetic TSP sampling (Refer to [24]). <sup>f</sup> A reference PM<sub>10</sub> size separator per European standards (EN 12341).

Table 18 summarizes the types of filters selected for PM mass concentration measurement in swine barns. Among them, ringed Teflon filters (i.e., PTFE filters with a PMP supporting ring) are recommended by the U.S. EPA for ambient PM monitoring due to the filters’ superior chemical stability, lightweight, and minimal gas and water adsorption [381]. However, glass fiber filters were most frequently used, for two reasons. First, glass fiber filters are more affordable than Teflon filters. Secondly, PM concentrations in swine barns are normally much greater than those in the ambient air. Thus, even though the gas and water adsorption by glass fiber filters causes mass measurement uncertainties, its influence would be relatively minor. To minimize the uncertainties, filters must be conditioned before mass measurement (including both pre-sampling and post-sampling weighing). In many swine barn PM studies, filters were conditioned in a desiccator at room temperature for 24–48 h. No specific humidity and temperature values were given; however, a relative humidity (RH) level of <20% and a temperature level of 20–25 °C (68–77 °F) could be assumed when the lab and the desiccator were managed properly. This is in contrast to 30–45% RH and 20–22 °C that the U.S. EPA recommends for ambient PM monitoring [377]. The filter mass measurement was normally conducted on a microbalance with a readability of 1 µg or 10 µg. The former (1 µg) is preferred, as the PM mass concentration can be as low as tens of µg m<sup>-3</sup> in swine barns.

**Table 18.** PM filters used in the past swine barn PM studies.

Filter Type	References
Ringed Teflon filter (e.g., Teflo™)	[65,85,99]
Teflon membrane filter (e.g., Zefluor™)	[73,74,82,130]
Cellulose filter	[46,52,61,63,75]
Glass fiber filter	[44,45,47–49,53,57,59,62,66,68,69,71,72,74,83,84,89,94,96,106]
Quartz fiber filter	[104]
Polycarbonate filter	[80,148,158]
PVC filter	[43,54,62,76,77,103,105]

The mass concentration of a PM sample ( $C_p$ ; mg m<sup>-3</sup>) can be calculated with Equation (3):

$$C_p = \frac{M_p}{V_a} = \frac{M_t - M_0}{Q \times t} \tag{3}$$

where  $M_p$  is the mass (weight) of collected particles (mg),  $V_a$  is the volume of sampled air (m<sup>3</sup>),  $M_t$  is mass of a filter after sampling (with collected particles) (mg),  $M_0$  is the mass of the filter before sampling (mg),  $Q$  is the volumetric sampling airflow rate (m<sup>3</sup> h<sup>-1</sup>), and  $t$  is the sampling period (h). The volume of sampled air varies with temperature and pressure. For PM concentrations measured under different temperature or pressure conditions, a correction is required to convert the volume of sampled air to a standard volume, i.e., the volume of sampled air under a standard condition (Equation (4)):

$$C_{p,std} = C_p \times \frac{T_{std} + 273.15}{T + 273.15} \times \frac{P}{P_{std}} \tag{4}$$

where  $C_{p,std}$  is a corrected PM mass concentration under the standard condition (mg m<sup>-3</sup>),  $T_{std}$  is the standard air temperature (°C),  $T$  is the actual air temperature during PM sampling (°C),  $P$  is the actual air pressure (atm), and  $P_{std}$  is the standard air pressure (atm). Different regulations may define different sets of standard conditions. For example, in the U.S. NAAQS, the standard condition refers to a temperature of 25 °C and a pressure of 1 atm; while in the U.S. Standards of Performance for New Sources, the standard condition is defined as 20 °C and 1 atm. The relevant information, however, was often lacking in the previous swine barn PM studies, making it difficult to compare the measurement results from different studies and against air quality standards.

Many PM samplers and filters were developed for sampling in the ambient air where PM is dominated by fine particles and its mass concentrations are substantially lower. However, these conditions do not stand for swine barns, causing potential issues. The

most prevalent is overloading [96,379]. Overloading could occur to size separators and filters. For size separators, overloading could shift a separator's penetration curve beyond its acceptable range and cause the reentry of captured large particles to the sampling airflow. Both could bias measurement results. For example, the reentry of large particles leads to overestimates of PM concentrations. For filters, overloading could dislodge captured particles off the filter surface, resulting in underestimates of PM concentrations. No systematic investigation has been conducted on the overloading issue in swine barns. When it occurs, a possible solution is to reduce the sampling period, e.g., using a digital timer to turn on and off a sampling pump periodically [85].

Given the challenges facing size-selective PM sampling, an indirect method was occasionally used. The method features the collection of TSP on a hydrophobic filter and the analysis of the weight and PSD of the TSP sample. The TSP mass concentration ( $C_{TSP}$ ) is determined with the regular gravimetric method. The mass fraction of  $PM_{10}$  or  $PM_{2.5}$  in the TSP ( $C_{PM10}/C_{TSP}$  or  $C_{PM2.5}/C_{TSP}$ ) is determined from the TSP's PSD profile. The  $PM_{10}$  or  $PM_{2.5}$  mass concentration can, thus, be estimated [82,96,382]. The indirect method is easy to implement and circumvents possible issues with size separators. However, it relies on multiple assumptions, e.g., particles of all sizes having the same density and refractive index [96]. These assumptions have yet to be fully validated.

The gravimetric method is widely used in various air environments, including swine barns. Even though real-time PM monitors are becoming increasingly available, the gravimetric method is anticipated to continue its popularity given its ease to operate and maintain. Compared to the real-time method, the gravimetric method is relatively simple, but it involves many technical details or considerations (e.g., gas-particle partitioning). An in-depth discussion about the gravimetric method can be found in Refs. [117,381].

#### 4.1.2. Real-Time Methods

With the advancement in sensor technologies, real-time methods are becoming increasingly powerful, user-friendly, and prevalent. In the past decade, numerous new PM instruments have entered the market at an affordable price, and many of them fall into the category of optical PM monitors. Only the instruments that have been used for swine barn PM monitoring are reviewed here (Table 19).

**Table 19.** Real-time PM monitors used in the past swine barn PM studies.

Instrument	Manufacturer, City, Country	References
TEOM	ThermoFisher Scientific, Waltham, MA, USA	[40,67,95,106]
BAM	ThermoFisher Scientific, Waltham, MA, USA; Met One Instruments, Inc., Grants Pass, OR, USA	[95]
DustTrak	Optical PM monitors	
DustTrak DRX	TSI Inc., Shoreview, MN, USA	[58,80,81,94,95,104,383]
Dylos DC1100	TSI Inc., Shoreview, MN, USA	[93,101–103]
EPAM 5000 particulate monitor	Dylos Corp., Riverside, CA, USA	[383]
Grimm aerosol spectrometer	SKC Ltd., Blandford, UK	[70,73]
GT-331 handheld particle counter	Grimm Aerosol Technik, Ainring, Germany	[56,88,90,92,95,97,98,100]
Handheld 3016 particle counter	Met One Instruments, Inc., Grants Pass, OR, USA	[79]
pDR-1200	Lighthouse Worldwide Solutions, Fremont, CA, USA	[91]
	ThermoFisher Scientific, Waltham, MA, USA	[383]

#### Tapered Element Oscillating Microbalance (TEOM)

TEOM uses a tapered element, which is a hollow quartz cantilever with a Teflon filter mounted on its tip, to measure a change in the mass of particles collected on the filter. The mass measurement is based on a simple physical principle that the resonant vibration frequency of a cantilever decreases when additional mass is added to the cantilever. The mass–frequency relationship can be presented as:

$$\Delta m = K_0 \left( \frac{1}{f_1^2} - \frac{1}{f_0^2} \right) \quad (5)$$

where  $\Delta m$  is the mass change of a cantilever-filter assembly (attributed to collected particles),  $K_0$  is the spring constant (determined by the mechanical property of the cantilever),  $f_1$  is the final resonant vibration frequency (Hz), and  $f_0$  is the initial resonant vibration frequency (Hz).

Thus, TEOM by nature is a gravimetric method; but, different from gravimetric samplers, TEOM tracks the mass change of collected particles periodically, thereby enabling semi-continuous monitoring of PM mass concentrations. TEOM can be configured for TSP, PM<sub>10</sub>, or PM<sub>2.5</sub> monitoring. Each requires a different sampler inlet and/or size separator. For TSP monitoring, a TSP inlet is used. It is noteworthy that the TSP inlet for TEOM (and BAM) does not truly collect total particles of all sizes. The inlet has a 50% cut size between 25 and 40  $\mu\text{m}$ , depending on wind speeds [23]; that is, only 50% of particles larger than 25–40  $\mu\text{m}$  would be measured. For PM<sub>10</sub> monitoring, a low-volume (16.67 LPM, i.e., 1  $\text{m}^3 \text{h}^{-1}$ ) PM<sub>10</sub> inlet is used. The inlet has an impactor cup inside to remove particles larger than 10  $\mu\text{m}$ . For PM<sub>2.5</sub> monitoring, a PM<sub>2.5</sub> size separator is mounted downstream of a PM<sub>10</sub> inlet to further remove particles larger than 2.5  $\mu\text{m}$ . The separator can be a Well Impactor Ninety-Six (WINS) or a BGI PM<sub>2.5</sub> Very Sharp Cut Cyclone (VSCC). For PM<sub>2.5</sub> monitoring in swine barns, VSCC is a better option for its superior performance under heavy PM loading conditions [384]. TEOM could also be coupled with other low-volume inlets and size separators, e.g., URG cyclones and dichotomous virtual impactors. The latter combination (TEOM 1405D) was used in a recent swine barn PM study [106].

To minimize the interference of varying air density and water vapor condensation with PM mass measurement, the cantilever-filter assembly of TEOM is maintained at an elevated temperature (usually 50 °C). However, this could cause the volatilization loss of volatile and semi-volatile substances from particles. A large portion of fine particles in swine barns originate from feces, and they are rich in volatiles and semivolatiles. Thus, using TEOM for PM<sub>2.5</sub> monitoring in swine barns could carry biases. Although no relevant studies were conducted in swine barns, the measurement biases were affirmed by Ref. [385] from the field assessment of TEOM in a layer hen house. TEOM is listed by the U.S. EPA as a Federal Equivalent Method (FEM) for PM<sub>10</sub> monitoring [386]. However, it is not a FEM for PM<sub>2.5</sub>, for a similar reason (volatilization-induced biases). To apply TEOM for PM<sub>2.5</sub> monitoring, one could decrease the temperature or add a filter dynamic measurement system (FDMS) to condition the PM-laden air before it being directed to the sensing element. TEOM-FDMS is a U.S. EPA-certified FEM for PM<sub>2.5</sub> and PM<sub>2.5-10</sub> [386].

#### Beta Attenuation Monitor (BAM)

BAM, also known as a beta gauge or beta-radiation attenuation monitor, derives PM mass from the PM's attenuation (absorption) of beta-rays. The instrument collects particles on a moving glass fiber filter tape. Before a clean spot of the filter tape is subjected to PM collection, it is irradiated with beta-rays emitted from a carbon-14 (<sup>14</sup>C) radiation source. A radiation counter on the other side of the filter determines the baseline beta radiation level, i.e., a zero reading. The clean spot then advances to a filter cassette where PM-laden air is pulled through the spot. After a certain period (e.g., 1 h), the PM-laden spot moves back in-between the beta-ray source and the radiation counter to measure the beta radiation level after PM collection. According to Beer–Lambert's Law, the mass of PM on the filter spot can be calculated as:

$$\frac{m}{A} = \frac{1}{\mu} \ln \left( \frac{I_0}{I_1} \right) \quad (6)$$

where  $m$  is the mass of particles on the filter spot ( $\mu\text{g}$ ),  $A$  is the area of the filter spot ( $\text{cm}^2$ ),  $\mu$  is the absorption cross-section of particles<sup>1</sup> ( $\text{cm}^2 \mu\text{g}^{-1}$ ; Note: it varies little with the composition of PM substances),  $I_0$  is the beta radiation level before PM collection (i.e., a

clean filter spot), and  $I_1$  is the beta radiation level after PM collection (i.e., a PM-laden filter spot).

BAM is listed by the U.S. EPA as FEMs for PM<sub>2.5</sub> and PM<sub>10</sub> monitoring. It can be coupled with various low-volume (16.67 LPM) sampler inlets and/or size separators. The glass fiber filter used by BAM is more water-absorbing than the Teflon filter used by TEOM. Thus, for BAM, the relative humidity (RH) of the sampled air must be controlled before it reaches the filter spot. The RH control is conducted using a fixed-temperature inlet heater, or a smart inlet heater with its temperature adjusted based on the temperature and humidity of the sampled air. Similar to TEOM, BAM should be housed in an environmental enclosure during field deployment. The enclosure provides a relatively constant temperature and prevents the instrument from dust, water, and other hazards.

Both BAM and TEOM provide semi-continuous PM mass concentration readings and can be deployed in swine barns for months without substantial maintenance. Regular maintenance work includes replacing the filter or filter tape, cleaning the sampler inlet or size separator, and performing leak checks and flow audits [67]. BAM, by nature, is an indirect method. Thus, a calibration against the gravimetric method is recommended before using BAM for PM monitoring in swine barns. For BAM, the minimal time interval is 1 h for updating its PM concentration reading. In contrast, TEOM can offer an updated PM reading every 10 s and, thus, would be preferred when short-term PM concentration changes are of interest. However, as an FEM for PM<sub>10</sub> and PM<sub>2.5</sub>, BAM is more versatile; and a recent BAM model (Thermo Scientific Model 5028i) has overcome the time-interval limitation and is capable of measuring PM<sub>10</sub> and PM<sub>2.5</sub> simultaneously with a single instrument.

#### Optical PM Monitors

Interactions between light and PM are complicated. When a light beam irradiates airborne particles, part of the light passes by, part of the light is absorbed by the particles, and part of light changes its direction due to processes such as refraction and reflection. In physics, these three interactions are defined as light transmission, absorption, and scattering, respectively. To our knowledge, all the optical PM monitors used in swine barns are based on light scattering. Thus, only light scattering PM monitors are reviewed.

Light is an electromagnetic wave. Its interactions with PM are therefore described by Maxwell's equations. For PM within the micron size range, Maxwell's equations can be approximated by the Mie scattering theory. The theory assumes that all particles are spherical, solid, and homogenous. According to the theory, the size and, accordingly, the volume of particles are related to and can be computed from the angle and intensity of scattered light. By further assuming particle density, PM mass concentration can be derived. Differing in functionality, light scattering PM monitors can be grouped into two categories: light scattering photometer and Optical Particle Counter (OPC).

A light scattering photometer (e.g., EPAM 5000, DataRam pDR-1200, TSI DustTrak 8520 and 8530) measures the mass concentration of a PM sample. It uses an optical detector to measure the intensity of scattered light at a single fixed angle. A photometer is usually coupled with a size-selective sampler inlet and/or separator to measure the PM of the desired size, e.g., PM<sub>2.5</sub>; and PM mass concentration ( $C_p$ ,  $\text{g m}^{-3}$ ) can be computed as:

$$C_p = \frac{b_{sp}}{\alpha_M} \quad (7)$$

where  $b_{sp}$  = intensity of scattered light ( $\text{m}^{-1}$ ) and  $\alpha_M$  = mass scattering efficiency ( $\text{m}^2 \text{g}^{-1}$ ).

The mass scattering efficiency  $\alpha_M$  characterizes the light scattering per unit mass of particles and it varies with measured PM substances. For example, assuming that a PM sample follows the lognormal size distribution, for a given scattered angle,  $\alpha_M$  can be calculated based on the Mie scattering theory [143].

$$\alpha_M = \frac{1.5}{\rho_p} \int \frac{Q_{scat}(n, k, D, \lambda) f(D, D_g, \sigma_g)}{D} dD \quad (8)$$

where  $\rho_p$  is particle density ( $\text{g m}^{-3}$ ),  $D$  is particle diameter (m),  $n$  is the real part of particle refractive index ( $n \pm ik$ ),  $k$  is the imaginary part of particle refractive index,  $\lambda$  is the light beam wavelength (m),  $Q_{\text{scat}}(n, k, D, \lambda)$  is the Mie scattering efficiency (dimensionless),  $D_g$  is the MMD of particles (m),  $\zeta_g$  is the GSD of particles (dimensionless), and  $f(D, D_g, \zeta_g)$  is the lognormal density function of particle size distribution.

Thus, light scattering photometers do not directly measure PM mass concentrations. They must be calibrated with the measured PM to ensure their measurement accuracy. The calibration can be conducted through a field comparison of the photometer with a co-located gravimetric PM sampler or samplers [95,373]. It is noteworthy that some instrument vendors provide calibration services; however, their calibration standard could be substantially different from swine barn PM, resulting in an improper calibration factor. Taking TSI DustTrak 8520 as an example, it is calibrated in the factory using Arizona road dust with different properties than PM in swine barns. As a result, TSI DustTrak was found to significantly underestimate TSP and  $\text{PM}_{10}$  mass concentrations in swine barns when a factory calibration factor was used [95,103].

An OPC measures the size distribution of a TSP sample and optionally derives size-segregated PM mass concentrations from the measured PSD profile. While a photometer measures the light scattered by a cloud of particles, an OPC measures the pulse signal of scattered light created by individual particles. A light pulse is detected by an optical detector when a particle quickly passes through a light beam. Thus, the signal can be used for particle counting. According to the Mie scattering theory, the height of the pulse signal is proportional to the particle size. By compiling the sizing results of many individual particles over a time period, a PSD profile can therefore be generated. To achieve the desired function, an OPC has a specially designed airflow and optical system. As a result, an OPC is often more complicated and expensive than a light scattering photometer of similar measurement accuracy. The derivation of PM mass concentrations from PSD profiles involves several assumptions with such as particle morphology and density. These assumptions are often addressed by comparing an OPC with co-located gravimetric samplers or certified monitors (to derive PM-specific calibration factors). One of the most prevalent OPCs for swine barn PM studies is Grimm aerosol spectrometers (Models 1.100 and 11-X series). The instrument can classify particles into 31 size channels and provide the mass concentration readings of  $\text{PM}_1$ ,  $\text{PM}_{2.5}$ ,  $\text{PM}_{10}$ , respirable, thoracic, and inhalable particles simultaneously. Other OPCs for real-time PM concentration measurement include GT-331 handheld particle counter, Lighthouse handheld 3016 particle counter, and Dylon DC1100 and DC1700.

A unique design combining a light-scattering photometer with a simple OPC was proposed by Ref. [387]. The design has been implemented in TSI DustTrak DRX series products (Models 8533, 8533EP, and 8534). The instruments can simultaneously measure  $\text{PM}_1$ ,  $\text{PM}_{2.5}$ ,  $\text{PM}_{10}$ , respirable, and total PM (TPM) mass concentrations.

#### 4.2. Size Distribution Measurement

Various techniques/instruments have been used to determine the PSD of swine barn PM samples (Table 20). They can be classified into three categories: aerodynamic, light scattering, and microscopic methods. Different categories of methods target different diameters. An aerodynamic sizer measures aerodynamic diameters, a light scattering sizer measures optical diameters, and a microscope measures geometric diameters. These diameters can be converted to one another. A conversion often involves numerous assumptions. A detailed discussion about PM diameters and their inter-conversions is beyond the scope of this review and can be found in Ref. [24].

**Table 20.** Measurement methods for PM size distribution in swine barns.

Method Category	PM Sizer <sup>a</sup>	Manufacturer, Country	References
Aerodynamic, gravimetric	Berner low-pressure impactor	Hauke-MP GmbH, Hodonín, Czechia	[351]
	Marple cascade impactor	ThermoFisher Scientific, Waltham, MA, USA	[132]
	Non-viable Andersen cascade impactor (eight stages)	Westech Scientific Instruments, Essex, UK; Tisch Environmental, Cleves, OH, USA; ThermoFisher Scientific, Waltham, MA, USA (Graseby Andersen Inc. until 2005)	[36,48,57,83,118,124,128,388]
Aerodynamic, real-time	APS or UVAPS	TSI Inc., Shoreview, MN, USA	[109,120,122,127]
	Aerosizer DSP particle sizer	TSI Inc., Shoreview, MN, USA	[119,130]
Light scattering, real-time	Aerosol Particle Size Spectrometer	Topas GmbH, Dresden, Germany	[254]
	CLIMET laser particle counter	Climet Instrument Company, Redlands, CA, USA	[61,65,389]
	DustTrak DRX	TSI Inc., Shoreview, MN, USA	[102]
	Grimm aerosol spectrometer	Grimm Aerosol Technik, Ainring, Germany	[90,125,134,135]
	Lighthouse laser particle counter	Lighthouse Worldwide Solutions, Fremont, CA, USA	[91,133]
	Met One laser particle counter	MetOne Instrument, Grants Pass, OR, USA	[49,138,390]
Light scattering, filter-based	Horiba particle size analyzer	Horiba Ltd., Kyoto, Japan	[82,96,129,130]
	Malvern Mastersizer	Malvern Panalytical Ltd., Malvern, UK	[130]
Electrical resistivity, filter-based	Coulter counter	Beckman Coulter Life Sciences, Indianapolis, IN, USA	[82,121,129,130]
Microscopy, filter-based	Optical microscopy	n/a	[36]
	SEM	n/a	[80,121,148,391]

<sup>a</sup> APS—Aerodynamic particle sizer.

#### 4.2.1. Aerodynamic Sizers

##### Gravimetric Methods

The concurrent concentration measurement of PM<sub>2.5</sub>, PM<sub>10</sub>, and PM of other size ranges can provide PSD information. The measurement can be conducted by co-locating multiple gravimetric samplers each equipped with a different aerodynamic inlet or size separator (differing in 50% cut size ( $D_{50}$ )). However, this is costly and labor-intensive. Due to the limited cut size options, the derived PSD profile is of low resolution. To address the limitations, cascade impactors are often used instead.

A cascade impactor, also called a multi-stage impactor, is a stack of impactors each holding a different cut size. From the inlet to the outlet of the stack, the cut size of a stage/impactor decreases. A stage collects particles larger than its cut size on a collection medium (e.g., a filter disc or an aluminum foil) and transfers particles smaller than the cut size to the next stage. The collection medium can be submitted for gravimetric analysis. With a known sampling period and a known sampling airflow rate, the PM mass concentration of each size stage can, thus, be determined. The greater the number of stages is, the better size resolution a cascade impactor could offer.

Among various impactors, non-viable Andersen cascade impactors have been most commonly selected for swine barn PM studies (Table 20). A non-viable Andersen cascade impactor consists of eight stages (0 to 7), with cut sizes of 9.0, 5.8, 4.7, 3.3, 2.1, 1.1, 0.7, and 0.4  $\mu\text{m}$ . A pre-separator is installed before Stage 0 to remove particles larger than 10  $\mu\text{m}$ . Therefore, the eight stages collect particles with aerodynamic diameters of 9.0–10, 5.8–9.0, 4.7–5.8, 3.3–4.7, 2.1–3.3, 1.1–2.1, 0.7–1.1, and 0.4–0.7  $\mu\text{m}$ . It is noteworthy that Andersen cascade impactors have viable versions. A viable Andersen cascade impactor is used to collect bioaerosol samples on agar plates. Due to its different design specifications, a viable Andersen cascade impactor is unsuitable for gravimetric PSD analysis.

A Berner low-pressure impactor (BLPI) consists of six stages, with cut sizes of 0.25, 0.43, 0.86, 1.73, 3.42, and 6.61  $\mu\text{m}$ . BLPI is also available in eight or ten stages, with additional stages stacked to determine the size distribution of submicron particles. A Maple cascade impactor is a personal sampler compact in size and can be carried by farm workers for occupational exposure assessment.

Other than the aforementioned cascade impactors, one may use a Micro-Orifice Uniform Deposit Impactor (MOUDI; TSI Inc., Shoreview, MN, USA) or a Dekati Low-Pressure Impactor (DLPI; Dekati Ltd., Kangasala, Finland) for size distribution measurement. Both instruments have been extensively used for atmospheric and indoor (including livestock barns) PM studies and they are available in different size-stage configurations. For MOUDI, five configurations are available, including three-stage, six-stage, eight-stage, ten-stage, and thirteen-stage impactors. For DLPI, three size-stage configurations are offered (four, five, and fourteen).

#### Real-Time Methods

PM size distribution can be derived in real-time by continuously measuring the mass of particles collected on each stage of a cascade impactor. Examples of such instruments include Quartz Crystal Microbalance (QCM) MOUDI (TSI Inc., USA) and Electrical Low-Pressure Impactor (ELPI; Dekati Ltd., Finland). A QCM MOUDI uses QCM, a highly sensitive mass transducer, to measure the mass of particles deposited on each stage. An ELPI charges the sampled particles before feeding them into a cascade impactor and estimates the mass of particles collected on each impactor stage by quantifying the total charge carried by the stage. Neither QCM MOUDI nor ELPI has been used in swine barns. The ELPI was used in a poultry barn PM study in Iowa [392].

Another real-time method is the use of time-of-flight particle sizers. In a time-of-flight particle sizer, a particle is accelerated by pulling a highly diluted PM-laden air sample through an orifice. Immediately after the orifice is two closely located, parallel laser beams. When the particle passes through the beams, its scattering of the laser light results in two pulses on an optical detector. The time difference between the two pulses, i.e., the time required for a particle to travel the small distance between the two laser beams, is related to the particle's aerodynamic diameter. Due to their greater inertia, larger particles have lower accelerating rates and, thus, take a longer time to travel a given distance, whereas smaller particles accelerate faster and take a shorter time to travel the same distance. Based on the time-of-flight principle, the aerodynamic diameter of a particle can be determined. A PSD profile can, thus, be derived by summarizing the sizing results of many particles in the air sample.

Two time-of-flight particle sizers have been used in swine barn PM studies: Aerodynamic Particle Sizer (APS) and Aerosizer DSP. Both are patented products of TSI Inc. Aerosizer DSP was discontinued in 2002 but is still available in some research groups. It can classify particles from 0.3 to 700  $\mu\text{m}$  into 44 size channels. APS has undergone several upgrades since the 1980s. The current model is model 3321 and it can classify particles from 0.5 to 20  $\mu\text{m}$  into 52 size channels based on their aerodynamic diameters. Different from previous models, the APS model 3321 is additionally equipped with an OPC that enables the classification of particles from 0.37 to 20  $\mu\text{m}$  into 16 size channels based on their optical diameters.

#### 4.2.2. Light Scattering Sizers

##### Real-Time Methods

The size distribution of a PM sample can be analyzed in real-time using an OPC. As described in Section 4.1.2, an OPC estimates the size of a single particle from the pulse height of the scattered light that the particle creates when traveling through a light beam. By sizing numerous particles over a certain time interval, a PSD profile can be generated. In theory, a mathematical relation between particle size and pulse height is given by Mie scattering. However, in reality, the relationship is experimentally determined by calibrating an OPC with monodispersed particle (i.e., particles uniform in size) standards of different sizes. Instrument design and calibration affect the size classification resolution of an OPC.

Although some OPCs read PM mass concentrations, the majority of OPCs measure PSD only and present their measurement results in the form of number PSD, i.e., the number (count) of particles within each size channel. Again, an OPC measures the optical

diameter of particles. To convert a number PSD derived from an OPC to a volume PSD, i.e., the volume of particles within each size channel, two assumptions are required. First, particles are solid and spherical so that the optical diameter of a particle is equal to its geometric diameter. The geometric diameter can then be used to calculate a particle's volume. Second, all particles within a size channel share the same diameter (which is usually the geometric or arithmetic mean diameter of the channel). To further convert a volume PSD to a mass PSD (i.e., the mass of particles within each size channel), a density value must be assumed. For simplicity, particles of all sizes are often assumed to share the same density. It should be noted that the mass PSD here is based on geometric diameters (assumed to be equal to optical diameters); whereas, PM<sub>10</sub>, PM<sub>2.5</sub>, respirable PM, and alike are defined based on aerodynamic diameters. Thus, to derive PM mass concentrations from the mass PSD, a further conversion from geometric diameters to aerodynamic diameters is required [24].

#### Filter-Based Methods

In several previous studies, a TSP sample was collected on a hydrophobic filter medium (e.g., a Teflon membrane filter) and extracted into an aqueous solution for PSD analysis. Light scattering particle sizers for liquid samples were used, including Horiba LA-300, Malvern Mastersizer, Beckman LS 13 320, and LS230 [382]. These analyzers have a different optical system than OPCs. Instead of using a single optical detector to target individual particles, they use multiple detectors to measure the light scattered by a collection of particles over a broad angle. To enhance the extraction efficiency and preserve particles during extraction, chemical stabilizers such as sodium polymetaphosphate (NaPMP) and lithium chloride (LiCl) were often added to the aqueous solution [96,147]. In addition to light scattering particle sizers, a Coulter counter (Beckman Coulter Life Sciences, USA) was also used to determine the size distribution of filter-collected PM samples [82,129,130]. The Coulter counter determines the size of particles based on their induced changes in the electrical resistance of sample extracts.

The filter-based methods exempt the field deployment of real-time particle sizers. They can be combined with gravimetric PM sampling and analysis, thereby simplifying the field monitoring setup. However, particles could change their size during PM collection, extraction, and size analysis. For example, the dissolution of soluble particles or soluble parts of particles could significantly bias the size measurement results. The accuracy of the filter-based methods has yet to be fully assessed.

#### 4.2.3. Microscopic Methods

The size of individual particles can be determined using a microscope. This requires the field collection of a PM sample on a filter medium. Polycarbonate filters (e.g., Whatman Nuclepore) are recommended for sample collection because their smooth surface and round, uniformly-sized pores provide a good contrast for PM identification and size measurement [393]. However, other filter types (e.g., glass filter filters) were still occasionally used [104]. Size measurement can be conducted manually, automatically, or semi-automatically. All of the previous studies in swine barns used the manual method, i.e., manually measuring the shape and size of individual particles on acquired images. The semi-automated method uses image analysis software programs (e.g., ImageJ) to automatically detect particles on a processed image and measure their PSD. The method, however, requires the manual adjustment of image processing parameters such as thresholds and backgrounds. The automated method uses an automated microscope (e.g., Phenom desktop SEM (ThermoFisher Scientific, USA)) for high-throughput, automatic measurement of particle size. To our knowledge, neither semi-automated nor automated methods have been employed for PSD measurement in swine barns. The semi-automated method was previously used for counting bioaerosols in a Danish swine barn [87].

Both optical microscopes and SEMs were used. As electron beams (adopted by SEMs for object illumination) have smaller wavelengths than visible light, SEMs typically provide

greater amplification than optical microscopes. Thus, SEMs are particularly suitable for the size measurement of fine particles. Some SEMs are equipped with additional detectors such as energy-dispersive X-ray spectroscopy (EDX or EDS). It enables the concurrent analysis of the chemical composition of individual particles and, thus, benefits the identification and classification of particles (Refer to Section 3.4).

### 4.3. Morphology and Density

#### 4.3.1. Morphology

PM morphology is determined through microscopic image analysis. Modern particle imaging systems can provide automated, high-throughput morphology measurement. Examples of such systems include Morphologi G4 (Malvern Instruments Ltd., Worcestershire, UK) and Horiba PSA300 (Horiba Group, Edison, NJ, USA). However, they have not been used in any swine barn PM studies.

Most previous studies used SEMs to visually measure the shape, size, and texture of PM samples from swine barns. These included a regular SEM [121,394], a field emission SEM [104], and a field emission high-resolution SEM [80], differing in electron sources and resolution. An SEM uses a focused electron beam (typically 0.2–40 k eV) to scan the surface of a test sample. The interaction of the electron beam with the sample produces various signals such as secondary electrons and back-scattered electrons. Among them, the secondary electrons are commonly used for morphology analysis and they are generated from electron beam-induced ionization at the sample surface. In an SEM, these secondary electrons are detected by an Everhart–Thornley detector. The intensity of detected signals is related to material composition and surface morphology. A digital image can, thus, be created when the beam finishes the scanning of a sample. A major advantage of an SEM over an optical microscope (Note: used by [97]) lies in its superior magnification (SEM: up to 300,000 $\times$  versus optical microscope: up to 1500 $\times$ ) and resolution. The magnification and resolution can be further improved with field emission or high-resolution SEMs. This allows one to easily identify and measure fine particles. The manual inspection and measurement of PM morphology are highly labor-intensive [302]. Image analysis software programs (e.g., FETEX 2.0 [80]) were often used to facilitate the process. With advancements in machine learning-assisted image analysis, it is expected that PM morphology will gain increasing attention. Many unresolved questions (e.g., the size and mixing state of airborne viral, bacterial, and fungal particles) will possibly be addressed through morphology analysis.

A high vacuum environment in an SEM's specimen chamber could result in the volatilization loss of volatiles and semivolatiles in PM. This, along with electron beam irradiation, could cause the deformation of particles [352]. Therefore, the morphology (including PM size) observed from an SEM specimen does not necessarily represent actual PM morphology.

#### 4.3.2. Density

Particles in swine barns are highly variable in sources, composition, mixing states, hygroscopicity, etc. Accordingly, their density values vary greatly and the reported PM density is an average of a collection of particles. PM density measurement is a longstanding challenge for aerosol science [395,396]. For PM in swine barns, its density can be measured or estimated with two different methods:

- Gas pycnometry. A gas pycnometer is an instrument specifically for density measurement. It utilizes a gas replacement principle to measure the net volume of a test sample (excluding external and internal pores). Along with precise mass measurement, the density of the sample can be calculated. However, the instrument has several limitations when applied to PM. Taking the AccuPyc II 1340 pycnometer (Micromeritics Instrument Corp., Norcross, Georgia) as an example, the bulk volume of test samples must be  $\geq 0.5$  cm<sup>3</sup>. This requires at least  $\sim 200$  mg of PM samples to be collected, which is impossible in reality. To address this limitation, settled dust was selected as a

surrogate for density measurement in the literature [96,129,146,147]. The dust samples were usually collected from exhaust fans, pen dividers, and feed lines, and they were conditioned in desiccators before pycnometry analysis to prevent measurement biases induced by absorbed water. A detailed analysis procedure can be found from [20,147].

- Indirect method. PM density can be calculated from Equation (2) when the PM's aerodynamic diameter, equivalent volume diameter, and shape factor are known [397]. However, this requires online, simultaneous measurement of both diameters and was rarely used due to the complexity of required measurement systems [395,398]. An alternative method was used by Ref. [97] in which a sedimentation cylinder was built to measure the settling velocity of particles ( $v_s$ ) and the PM density was calculated as:

$$\frac{\rho_p}{\chi} = 18 \frac{v_s \eta}{g d_e^2 C_{ce}} \quad (9)$$

where  $\eta$  is the dynamic viscosity of air (Pa sec). Specifically, a pulse release of PM was conducted at the top of the sedimentation cylinder. The time for particles of different sizes to arrive at the cylinder bottom, i.e., settling time was tracked using a Grimm aerosol spectrometer. A settling velocity was then calculated by dividing the cylinder height by the measured settling time.

Both methods involve many assumptions. Thus, the measurement results may carry large uncertainties. A discussion of the assumptions and associated uncertainties requires advanced knowledge of aerosol mechanics and instrumentation and it is beyond the scope of this review. Further information can be found from Refs. [24,117].

#### 4.4. Bioaerosol Characterization

##### 4.4.1. Bacterial and Fungal Counts

##### Culturable Bacterial and Fungal Counts

The measurement of culturable bacterial/fungal counts typically consists of three steps: sampling, cultivation, and enumeration. Sampling represents the collection of airborne microbes (more strictly speaking, microbe-laden particles), cultivation involves the growth of target microbes on a growth medium or media to form visible colonies, and enumeration refers to the counting of formed bacterial or fungal colonies [295]. An optional step is identification, with aims to determine microbial species based on the appearance of the colonies formed [43,51,158,159,193]. Identification can also be conducted by taking individual colonies for further investigation including microscopic analysis, biochemical testing, DNA sequencing, and proteomic analysis [59,101,173,183,192].

Various sampling methods are available (Table 21). In general, they can be classified into two categories (dry and wet) based on collection media. The dry methods collect airborne microbes on filters or agar plates. For filter methods, the same setup for PM sampling can be used. However, it is noteworthy that filters pose stress to collected airborne microbes due to impaction and desiccation, thereby reducing the culturability of the microbes [399]. After collection, the filter is extracted in a surfactant solution (e.g., Tween-20 [63,175] and Tween-80 [59,185]) and the extract is diluted and transferred to an agar plate or plates for cultivation.

For agar plate methods, airborne microbes are directly collected onto agar plates. Before they settle on the agar plate surface, airborne microbes are usually segregated into multiple size ranges using cascade impactors. Among various impactors, a viable Andersen six-stage impactor is most frequently used. The impactor was invented by Mr. Ariel Andersen in the 1950s [400] and it quickly became one of the most prevalent bioaerosol samplers. The working principle of cascade impactors can be found in Section 4.2.1. Again, Andersen cascade impactors have viable and non-viable versions. They cannot be interchangeably used due to their different design specifications. A viable Andersen six-stage impactor separates airborne microbes into six size ranges: >7, 4.7–7, 3.3–4.7, 2.1–3.3, 1.1–2.1, and 0.65–1.1  $\mu\text{m}$ . An agar plate is placed in each impactor stage for bioaerosol collection. A viable Andersen six-stage impactor and its brother products (single- or two-

stage) are commercially available from Thermo Fisher Scientific Inc. (Waltham, MA, USA) or Tisch Environmental (Clevs, OH, USA).

**Table 21.** Sampling methods adopted in swine barn culturable airborne microbe studies since 1990.

Method Category	Collection Medium	Specific Method and the Studies Adopting the Method
Dry	Filter	Cellulose nitrate membrane filter: [159] Glass fiber filter: [63] Polycarbonate filter: [59,175,181] Teflon filters: [185] Viable Andersen six-stage impactor: [43,54,76,127,147,152,153,156,158,159,162,166,173,183,185,188,190,192] Viable Andersen two-stage impactor: [160,163,164] Viable Andersen single-stage impactor: [165,168]
	Agar plate	MB1 MICROBIO Air Sampler: [174] Microbiological air sampler MAS-100: [79,170,179,182] RCS centrifugal air sampler: [157] Slit sampler: [43,51]
Wet	Liquid	AGI impinger: [54,78,127,156,158,161,162,167,171,172,175,176,193] Coriolis <sup>®</sup> $\mu$ sampler: [101,103,180,184,187,189]

The wet methods collect airborne microbes in a liquid medium, usually a sterile phosphate-buffered saline (PBS) or peptone solution amended with a non-ionic surfactant (e.g., Tween-80 [54,101] and TritonX-100 [184]) and/or an antiform agent [127,156]. After sample collection, the medium is transferred to an agar plate or plates for cultivation. Two devices have been used for swine barn bioaerosol studies. A Coriolis<sup>®</sup>  $\mu$  sampler uses a wet cyclone design in which airborne microbes, driven by a centrifugal force, are separated from air streams and scrubbed into a liquid medium in the cyclone chamber. An AGI impinger is another prevalent bioaerosol sampling device besides the Andersen impactor. It utilizes an impaction mechanism, that is, airborne microbes, after acceleration at a nozzle, impact into a liquid medium due to their greater inertia than gas molecules. Particle diffusion is also believed to play a role in bioaerosol collection by the AGI impinger [401]. Other impingers include SKC Biosamplers (with multiple models). Their applications in swine barn bioaerosol research can be found in Section 4.4.3.

Sampling methods affect the final counting results of culturable bacteria or fungi. Higher culturable counts from AGI impingers than Andersen six-stage impactors were observed in two previous swine barn studies [127,156]. Liquid media are considered to be less stressful (Note: desiccation stress) to airborne microbes than solid media including agar plates. In addition, Andersen six-stage impactors were designed for bioaerosol sampling in relatively clean environments. They can easily be overloaded in swine barns with typically high bioaerosol concentrations. Specifically, each impactor stage has 400 nozzles so in principle the number of colonies formed on an agar plate is no greater than 400. Overloading occurs when multiple microbe-laden particles deposit and form colonies beneath the same nozzle and nearly all nozzles are saturated in this manner. The overloading, if occurring, will result in an underestimate of culturable bacterial/fungal counts. To address this issue, Andersen six-stage impactors were often operated for only tens of seconds to a few minutes in swine barns and a positive-hole conversion was conducted to correct for the underestimation caused by the “carpooling” of multiple particles [402]. For further information regarding sampler selection and sampling time determination, readers may refer to Ref. [401].

Nearly all the previous studies used agar plates to cultivate collected bacteria or fungi. Depending on target microbes and subsequent analyses, different cultivation media and cultivation conditions were selected (Table 22). For Andersen six-stage impactors and alike, the same agar plates were used for both sampling and cultivation. Thus, they are summarized in the same table.

**Table 22.** Cultivation media and conditions used in swine barn bioaerosol studies since 1990.

Target Microbe(s)	Cultivation Medium	Cultivation Condition
Bacteria <sup>a</sup>		Room temperature for 5–7 d: [171] 20 °C for 4 d: [170] 25 °C for 1–3 d: [78] 25 °C for 5 d: [127]
	Blood-based agar: [175]	30 °C for 2 d: [162,177]
	Brain heart infusion agar: [103,180]	30 °C for 2–5 d: [158]
	Columbia agar: [101,182]	30 °C for 5 d: [54]
	Horse blood agar: [166]	30 °C for 5–7 d: [156]
	Nutrient agar: [177,190]	30 °C for 7 d: [184]
	R2A agar: [78,127,162,171]	30–35 °C for 2 d: [157]
	Sheep blood agar: [43]	35 °C for 40–120 hr: [152]
	Trypticase soy agar (TSA): [54,63,69,78,79,147,152,156– 158,160,161,163–165,168,170,184,187– 189,193]	35 °C for 1–2 d: [69,160,163,164] 37 °C for 1 d: [161,182,189] 37 °C for 1–2 d: [188]
	Tryptone glucose extract agar: [59]	37 °C for 2 d: [63,79,103,147,165,166,168,175,190,193] 37 °C for 2 d, w/5% O <sub>2</sub> on day 2: [101] 37 °C for 1 d, 22 °C for 3 d and 4 °C for 3 d: [43] 30 °C for up to 10 d: [158] 35 °C for 1–2 d: [69,173]
Gram-negative bacteria	Endo agar: [182,183]	35 °C for 40–120 hr: [152]
	Esosin methylene blue (EMB) agar: [43]	37 °C for 1 d: [182]
	MacConkey agar: [152,158,173,193]	37 °C for 2 d: [193]
	Modified Conradi agar: [69]	37 °C for 1 d, 22 °C for 3 d and 4 °C for 3 d: [43]
Coliform	Brilliance Coliform selective agar: [101]	30–35 °C for 2 d: [157]
	Chromocult Coliform agar: [79]	35 °C for 5–7 d: [184]
	MacConkey agar: [157]	37 °C for 2 d: [79,101]
	Violet read bile agar: [184]	
<i>E. coli</i>	Chromocult Coliform agar: [79,162]	
	Brilliance <i>E. coli</i> selective agar: [101]	
	EMB agar: [162]	
	MacConkey agar: [164,190]	35 °C for 1 d: [162] for Chromocult 37 °C for 1 d: [162] for EMB and MMGM 37 °C for 2 d: [79,101,190] 44.5 °C for 18–24 h: [172]
<i>Salmonella</i> spp.	mFC basal medium supplemented with 3-bromo-4-chloro-5-indolyl-β-D- glucuronide: [174]	
	Mineral modified glutamate medium (MMGM) agar: [162]	
<i>Clostridium difficile</i>	Bismuth sulfite agar: [103]	37 °C for 2 d: [103]
<i>Clostridium perfringens</i>	CLO-agar: [174]	
	<i>Clostridium difficile</i> agar: [103]	37 °C for 2 d w/o O <sub>2</sub> : [103,174]
<i>Staphylococcus</i> spp.	mCP agar: [172]	44.5 °C for 1 d: [172]
	Baird–Parker agar: [190]	37 °C for 1 d: [182]
<i>Staphylococcus aureus</i>	Chapman agar: [182]	37 °C for 2 d: [175,190]
	Mannitol salt agar: [164,175]	
	ChromoID Saureus agar: [179,193]	
	CHROMagar Staph aureus agar: [103]	37 °C for 2 d: [103,193] 37 °C for 3 d: [179]
MRSA	<i>S. aureus</i> selective agar: [185]	
	Brilliance MRSA agar: [185]	
	CHROMagar MRSA agar: [101,103,176,183]	37 °C for 1 d: [178] 37 °C for (24 + 17) hr: [176] 37 °C for 2 d: [101,103,175] 37 °C for 3 d: [179]
	MRSA chromogenic agar: [179]	
	MRSA screen agar: [175]	
<i>Streptococcus aureus</i> <i>Haemolytic streptococci</i> ( <i>Streptococcus pyogenes</i> )	MRSA selective agar: [178]	
	Fresh blood agar: [190]	37 °C for 2 d: [190]
	Sheep blood azide agar: [189]	35 °C for 1–2 d with 5% CO <sub>2</sub> : [189]
<i>Campylobacter</i>	Charcola cefoperazone desoxycholate agar: [172]	42 °C for 2 d w/o O <sub>2</sub> : [172]

Table 22. Cont.

Target Microbe(s)	Cultivation Medium	Cultivation Condition
<i>Enterococcus</i>	mEnterococcus agar: [172]	37 °C for 2 d: [172]
<i>Listeria monocytogenes</i>	PALCAM agar: [103]	37 °C for 2 d w/5% CO <sub>2</sub> : [103]
<i>Mycobacterium avium</i>	Middlebrook 7H10 Agar + OADC growth supplement: [103]	37 °C for 2 d w/5% CO <sub>2</sub> : [103]
<i>Yersinia enterocolitica</i>	Cefsulodin-Irgasan-Novobiocin agar: [172]	30 °C for 18 h: [172]
Thermophilic <i>Actinomycetes</i>	TSA: [43,76]	52 °C for 5 d: [76] 55 °C for 5 d: [43]
		Room temperature for 5–7 d: [156,171]
		20 °C for 4 d: [170]
		20–25 °C for 3–5 d: [165,168]
		22 °C for 5 d: [182]
		25 °C for 2 d: [161]
	DG-18 agar: [59,184,189,192]	25 °C for 2–5 d: [78]
	Malt extract agar (MEA): [43,59,69,76,78,156,158,161,165,168,171,177,181,184]	25 °C for 3 d: [179]
Fungi <sup>b</sup>	Rose Bengal agar: [54,76,127,170]	25 °C for 5 d: [76,127]
	Sabourand dextrose agar: [152,179,182]	25 °C for 5–7 d: [63,158]
	Sterile water: [63]	25 °C for 7 d: [181,189]
		27 °C for 5–7 d: [184]
		30 °C for 4 d and 22 °C for 4 d: [43]
		30 °C for 5 d: [54,177]
		35 °C for 1–2 d: [69]
		35 °C for 40–120 h: [152]
		52 °C for 5 d: [76] <sup>3</sup>
<i>Aspergillus</i> spp.	Czapek solution agar: [152]	35 °C for 40–120 hr: [152]

<sup>a</sup> Bacteria here refer to general bacteria, including mesophilic bacteria presented by [43,78,182,193]. <sup>b</sup> Fungi here refer to general fungi and are interchangeably termed with molds. They include mesophilic molds (25 °C for five days) and thermotolerant molds (52 °C for five days) presented by [76].

Various cultivation media and conditions were used. For general bacteria, TSA was the most frequently selected medium. To suppress fungal growth, bacterial agar was sometimes amended with cycloheximide, a natural fungicide [54,165,193]. The cultivation temperature ranged from room temperature (20–25 °C) to 37 °C and the cultivation time varied from 1 to 7 days. The most commonly selected cultivation condition is 37 °C for 2 days, which applies to not only general bacteria but most individual bacterial species or groups. Several studies presented their results as mesophilic bacterial counts (Table 5). They can be considered the same as bacterial counts as there was no difference in cultivation media or conditions (Table 22). In principle, mesophilic bacteria are bacteria that grow between 20 °C and 45 °C [403]. All the reported cultivation temperatures for bacteria fall into this range.

For general fungi, MEA was most frequently selected. To inhibit bacterial growth, antibacterial agents such as chloramphenicol and streptomycin were usually added [76,127,152,184]. The cultivation temperature ranged from room temperature to 35 °C and the cultivation time varied from 1–7 days. The only exception is Ref. [76] in which agar plates were cultivated at 52 °C to measure thermotolerant molds. The most prevalent cultivation temperature and time were 25 °C and 5 days, respectively, for fungi. Cultivation time must be sufficient for bacteria or fungi to grow into colonies. However, it cannot be too long to avoid the overgrowth of colonies causing difficulty in counting and isolating.

After cultivation, the colonies formed can be enumerated visually or with the aid of equipment (e.g., Quebec colony counters [127,156,157] or software (e.g., ImageJ)). Again, for bacterial/fungal counts derived from Andersen impactors, a multi-jet impactor positive-hole correction is needed [402]. Impingers are less prone to overloading. However, the liquid medium must be serially diluted (up to a dilution ratio of 10<sup>5</sup>) to ensure the number of colonies formed on an agar plate stays within a manageable range for visual enumeration [54,69,78].

### Total Bacterial and Fungal Counts

Various methods are available for the measurement of total bacteria/fungi in indoor air environments [295]. The methods that have been used for swine barns can be classified into two categories: (1) fluorochrome-assisted counting and (2) qPCR. They are discussed in separate paragraphs.

The fluorochrome-assisted counting methods involve the use of fluorescent cell stains (e.g., DAPI and AO) or fluorescent nucleic acid hybridization probes (e.g., a fluorochrome-labeled DNA probe). Fluorescent cell stains are normally coupled with EPM or flow cytometry for microbial enumeration. Fluorescent nucleic acid probes constitute a core part of FISH technology and cell enumeration is also normally conducted with EPM. Numerous fluorochromes are available, e.g., Ref. [404]. However, only a few have been used for staining airborne microbes sampled from swine barns, as listed below:

- AO is a nucleic acid stain with a maximum excitation wavelength at 502 nm and a maximum emission wavelength at 526 nm when binding to DNA and 460 nm when binding to RNA. It is cell-permeable, meaning that it can penetrate the cell membranes and organelles of both viable and non-viable microbes to interact with DNA or RNA.
- DAPI is also a nucleic acid stain and binds strongly to DNA regions rich in A (adenine) and T (thymine). It has a maximum excitation wavelength at 350 nm and a maximum emission wavelength at 470 nm. Similar to AO, DAPI can pass through cell membranes and, thus, can stain both viable and non-viable microbes.
- PI is a nucleic acid stain with enhanced fluorescence after binding to DNA. It shows a maximum excitation wavelength of 536 nm and a maximum emission wavelength of 617 nm. Different than DAPI and AO, PI is not cell-permeable and can only stain damaged cells. Thus, it is commonly used to study the viability of microbes.
- PAS is a staining method targeting polysaccharides such as glucans. It uses periodic acid to oxidize these sugars into aldehydes that then react with the Schiff reagent to create a purple-magenta color. PAS can be a regular or a fluorescent staining method. When conjugating the Schiff reagent with aniline blue, a fluorochrome, it becomes a fluorescent stain. PAS is often used to label fungal spores as fungal cell walls are abundant in glucans (Refer to Section 3.2.5).
- ChemChrome V6 is an ester precursor of fluorescein (a prevalent nucleic acid stain) and it is converted to fluorescein in the presence of esterase. Fluorescein can penetrate cell membranes and bind to DNA or RNA and has a maximum excitation wavelength of 494 nm and a maximum emission wavelength of 518 nm. ChemChrome V6 was used together with ChemSol B16 and B2 solutions [170], but no detailed information about these combinations is given.

For filter samples [59,87,181], bioaerosols must be first extracted from a PM-laden filter, usually in a surfactant (e.g., Tween 80) amended PBS solution. For liquid samples from impingers [78,161,215], no extraction is needed, but serial dilution may be necessary to adjust cell concentrations within a detectable range. The liquid acquired from extraction or dilution is then stained with a selected fluorochrome. When flow cytometry is used, the stained liquid is submitted for cell counting without further treatment. When EPM is used, the liquid is usually filtered with a black polycarbonate filter (offering a great contrast), and the cells retained on the filter are counted at a magnification of 400–1000 $\times$ . EPM is typically equipped with a tungsten lamp and multiple excitation optical filters to achieve a desired excitation wavelength and multiple emission optical filters to selectively detect the target emission. Flow cytometry adopts a similar detection principle to EPM; however, it has a specially designed flow chamber to allow microbial cells to pass the excitation light beam on an individual basis and, thus, counts the cells from emission light pulses. Microbial identification (e.g., distinguishing fungal spores from bacteria) can be conducted with EPM through the morphology analysis of observed microbes and/or combined use of other stains such as FISH probes [161]. Some flow cytometers can simultaneously measure the size of cells, which may also help with microbial identification or classification. Only

one study adopted FISH [161], and further information about the technology is available in Section 4.4.3.

The qPCR method quantifies the copy number of a target gene in a bioaerosol sample. It amplifies a target gene (or a segment of the gene) over multiple cycles and the amplification selectivity is achieved using a pair of primers. A primer is a short, single-strand DNA or RNA sequence that complements the starting or end sequence of a target gene. Thus, a primer pair defines the bacterial or fungal DNA region to be amplified. The gene region selected must serve as a fingerprint (indicator) of the microbes of interest, e.g., 16S rRNA for bacteria and ITS for fungi. 16S rRNA is a hypervariable region in bacterial DNA and it has been extensively used for bacterial identification [405]. Similarly, ITS genes are highly variable among fungal species. Different from regular PCR, qPCR uses fluorescent nucleic acid stains or fluorescent DNA probes to track the amplification efficiency of every cycle, thereby allowing one to back-calculate the number of gene copies in an original sample. Further information about qPCR can be found in Refs. [406,407]. The total bacteria/fungi measurement with qPCR typically consists of three steps: bioaerosol sampling, DNA extraction, and qPCR.

- The following bioaerosol samplers were used in the literature: filter samplers [179,218–220], Coriolis<sup>®</sup>  $\mu$  wet cyclones [103,180,191,221], impingers [76], and electrostatic dustfall collectors [222]. While solid samples were acquired from filter samplers and electrostatic dust fall collectors, liquid samples were derived from wet cyclones and impingers.
- Multiple DNA extraction kits were used, including QIAamp DNA Minikit [76,180], Fast Spin Kit [179], Power Soil Kit [218–220], Qiagen DNA Minikit [221], Ultra Clean GelSpin DNA Extraction Kit [191], and Nucleospin 8 Plant II Kit [222].
- All existing studies performed qPCR with commercial qPCR systems. *E. coli* genomes [76,103,180] or synthetic bacterial DNA [222] with known 16S rRNA gene copies were diluted to build a calibration curve for the quantification of total bacteria. For fungi, synthetic fungal amplicons were used as the calibration standard [220]. For specific bacterial species, plasmid vectors ligated with species-specific genes were used to build calibration curves [179]. Information about the selected primers can be found in corresponding publications.

A single microbial cell can have multiple gene copies and the number of gene copies per cell differs with microbial species. Therefore, for total bacteria or fungi, the qPCR results can hardly be translated into actual cell counts. However, for a known species, the number of gene copies per cell is relatively constant, e.g., seven 16S rRNA gene copies per *E. coli* cell [408]. Ref. [180] presented the counts of total bacteria and *Streptococcus suis* in cells  $m^{-3}$ ; however, no conversion factors were offered.

The first use of PCR and FISH for examining swine barn bioaerosols was reported by Ref. [409] (no quantitative data were given, though). Since then, molecular biology technologies have found increasing applications in swine barn bioaerosol research. They constitute a key component of culture-independent bioaerosol measurement. Other applications of these technologies can be found in the next few sections. Both culture-independent and dependent methods have their advantages and limitations. A comparison of different counting technologies is given by Ref. [410].

#### 4.4.2. Viruses

It is important to recognize that the current measurement methods for airborne porcine viruses are largely exploratory and have yet to be well-established for large-scale field monitoring [411]. As an example, Ref. [247] developed a method capable of detecting airborne PRRSV-1 under experimental conditions; however, the field deployment of the method in a PRRSV-infected farm was unsuccessful. This demonstrates the complexity and technical challenges associated with field viral measurement and the necessity for further methodology development.

All the publications in Table 7 used qPCR or real-time RT-PCR for viral detection. The two methods are very similar in that they both offer quantitative results (in the copy

number of genes in a sample) by tracking the amplification efficiency of every PCR cycle. While qPCR works for DNA viruses, real-time RT-PCR is required for the detection of RNA viruses. Compared to qPCR, real-time RT-PCR includes an additional step, i.e., the creation of a complementary DNA (cDNA) sequence complementary to viral RNA through reverse transcription. The cDNA created is then quantified through regular qPCR.

For viral detection, qPCR and real-time RT-PCR are also used to determine if a sample is positive for a target virus based on a preset criterion or criteria of cycle threshold (Ct). The Ct is defined as the number of PCR cycles needed to reach a fluorescence signal threshold level (due to the use of fluorochromes in qPCR). A low Ct value suggests a high abundance of target viral RNA or DNA genes in the sample. Several different Ct criteria were selected in the literature. A bioaerosol sample was considered positive when its Ct value was below 35 [240,244,246,248] or 36 [249]; or suggestively positive when its Ct value was below 38.5 [249] or 40 [240,244,246]. Bioaerosols generally contained a lower amount of genomic material than pig nasal swab or oral fluid samples [247]. Accordingly, different Ct criteria may be applied for positive sample determination.

The measurement procedure for airborne porcine viruses consists of three steps: bioaerosol sampling, RNA or DNA extraction, and PCR (qPCR or real-time RT-PCR).

- Wet cyclones were the most prevalent bioaerosol samplers in the literature [101, 136,240–242,244,247,249], followed by filter samplers [216,245,247,249,250] and impingers [217,243]. The selected filters include gelatin filters [216,250] and PTFE filters [245,249]. Non-viable cascade impactors were used to study the size distribution of viral particles, with samples collected on aluminum plates [246]. One study collected deposited dust samples for viral measurement [248]. Despite the prevalence of wet cyclones, no studies have yet demonstrated or affirmed their superiority to other samplers. A lower IAV detection rate was derived from Coriolis<sup>®</sup>  $\mu$  (a wet cyclone) than filter samplers [249].
- Limited information is available about RNA or DNA extraction from collected samples. For liquid samples acquired from wet cyclones and impingers, they can be directly submitted for DNA or RNA extraction. For solid samples derived from filter samplers or sedimentation plates, they can be extracted into a liquid solution and optionally further pelleted before extraction [216,245,250]. No extraction kit information is available for DNA viruses. In principle, many commercial DNA extraction kits should work. For RNA viruses, the selected extraction kits include QIAamp Viral RNA Mini Kit [136,245] and MagMAX 96 Viral RNA Isolation Kit [240].
- Real-time RT-PCR and qPCR were often conducted with commercial kits, e.g., Takara one-step qRT-PCR Kit [243] and Superscript III Platinum one-step qRT-PCR Kit [249]. Information about the selected PCR primers can be found in corresponding publications. The establishment of a calibration curve often involves several steps [249]. First, the genes of a target virus were inserted into plasmid vectors. The vectors were then cloned in host bacterial cells (e.g., *E. coli*). For RNA viruses, the next step was to transcribe the inserted genes. Finally, the transcripts or cell extracts were serially diluted and analyzed to build the calibration curve.

Special attention should be paid to the selection of bioaerosol samplers. Most commercial bioaerosol or PM samplers are not designed for the collection of airborne viruses [9], especially when the viruses occur in the air as individual particles. AGI-30 impingers have a 50% cut size of 0.3  $\mu\text{m}$  [412] and Coriolis<sup>®</sup>  $\mu$  samplers are stated to effectively collect PM larger than 0.5  $\mu\text{m}$ . Both are larger than the size of viral particles (0.02–0.5  $\mu\text{m}$ ) and that of the minor viral PSD peak (0.4  $\mu\text{m}$ ) reported by Ref. [246]. PM filters generally perform better than wet cyclones and impingers in collecting submicron particles. However, little is known about their performance for particles <0.1  $\mu\text{m}$  or the viability of viruses on the filter surface. To make measurement results comparable, a standard sampling protocol needs to be developed.

#### 4.4.3. Microbial Composition

This section provides a brief overview of relevant methods that have been adopted for swine barn PM research. Readers should bear in mind that numerous methods are available for microbial composition analysis and that new methods continue to emerge. A detailed discussion of the methods is beyond the scope of the review and can be found in Refs. [405–407].

##### Culture-Dependent Methods

In culture-dependent methods, after airborne microbes are collected on or transferred to an agar plate, they are cultivated to form visible colonies. A colony can be identified for its microbial species or group based on its appearance [158,159] including the size, shape, color, surface texture, elevation, and margin (i.e., edge) of the colony. The identification work usually involves a comparison of formed colonies with a reference library or colonies prepared from pure cultures. A microscope may be used to assist the visual observation and/or comparison of colony characteristics [59,158].

A formed colony can be isolated and cultivated in a selective or differential medium or a set of such media for species identification or categorization [413]. For example, Ref. [159] grew colony isolates (from R2A agar plates) with five selective/differential agars. Seven bacterial genera were, thus, identified. Identification can also be conducted through biochemical testing. For example, Gram staining can distinguish Gram-negative bacteria from Gram-positive bacteria [43,167]. Hydrogen peroxide can be used to detect the occurrence of catalase, thereby distinguishing *Streptococcus* from *Staphylococcus* [414]. Many of these biochemical tests have been incorporated as part of selective or differential media [182].

A recent development is to identify colonies or affirm identification results (from the above tests) through genomic or proteomic analysis. To do it, colony isolates were subjected to PCR [101,183,193] or MALDI-TOF MS analysis [185,192,193,210]. Both PCR and MALDI-TOF MS are molecular biology methods: PCR for genomes and MALDI-TOF MS for proteins. As they were used for the analysis of individual colonies, they should be considered as part of culture-dependent methods. However, they can also be part of culture-independent methods, as reviewed in the following section.

##### Culture-Independent Methods

Culture-independent methods require no cultivation steps. Instead, biological molecules characteristic of a species or a group of species are extracted from bioaerosol samples and subjected to molecular biology analysis. A measurement procedure typically consists of three steps: sampling, extraction (including purification and/or amplification), and molecular biology analysis.

Various devices and media were selected for swine barn PM sampling. The selected samplers include IOM or SKC personal PM samplers [169,262,271], isokinetic TSP sampler [209], filter cassettes [87,218–220,263,267,274], BTPM-H1 ambient air PM samplers [270], TH-150F PM samplers [272], Andersen multistage non-viable cascade impactors [192], AGI or SKC impingers [217,261,268], Coriolis<sup>®</sup>  $\mu$  samplers [221], ESP samplers [210,265,269], and Andersen multistage viable cascade impactors and alike [192,273]. Many of them (devices listed before impingers) are filter samplers. The selected filters include gelatin filters [169,262], glass fiber filters [192,209,267,272,274], cellulose nitrate filters [218–220], and Teflon filters [270]. To prevent microbial contamination, these filters were often autoclaved or baked before use [209,218,219,270,272]. An ESP collects bioaerosols on a collecting electrode or electrodes and the collected bioaerosols can later be dislodged from the collecting electrode. An Andersen viable cascade impactor collects bioaerosols to agar plates. An impinger and a Coriolis<sup>®</sup>  $\mu$  sampler collect bioaerosols into a liquid medium. The selection of sampling devices and media has a large influence on bioaerosol extraction.

The culture-independent methods can be classified into two categories: proteomics and genomics. Their sample extraction and analysis are presented separately here. For proteomic analysis, MALDI-TOF MS is the only analytical tool that has been applied to swine barn bioaerosols. Two studies [192,210] used MALDI-TOF MS to identify microbial species in colony isolates, i.e., colonies formed on agar plates. Thus, their measurement would fall into the category of culture-dependent methods. The only study using MALDI-TOF MS for culture-independent microbial detection was Ref. [261] in which bioaerosols were collected into a PBS solution using an impinger. The liquid was then filtered sequentially with two cell strainers of different pore sizes (70  $\mu\text{m}$  and 40  $\mu\text{m}$ ). The sequential (cascade) filtration, coupled with ultra-high-speed centrifuging, led to the separation of microbes into two size fractions: 20  $\mu\text{m}$  and 5  $\mu\text{m}$ . The bioaerosol samples were pelleted (upon centrifuging) and re-suspended in a matrix solution before being submitted for MALDI-TOF MS analysis. The analysis generated high-resolution mass spectra of bioaerosols. By comparing the generated mass spectra with reference spectra derived from pure cultures, bacteria were identified at the genus or species level. Collections of microbial reference mass spectra have been available, e.g., BDAL standard library and Filamentous fungi library. They, along with special software (MALDI Biotype, Bruker Corp., Billerica, MA, USA), were used to aid the identification of airborne microbes in swine barns [192,210]. Detailed information about MALDI-TOF MS and its application to microbial analysis can be found in Ref. [415].

Several genomic methods (also known as nucleic acid-based methods) were selected to study microbial composition in swine barn bioaerosols: DGGE, FISH, clone library, NGS, and shotgun metagenomic sequencing (Table 9). A comprehensive review of these methods is available in Refs. [405,407]. Among these methods, FISH is the only method that requires no PCR products and, thus, it is discussed separately. Others are discussed after a review of PCR steps in the literature.

As an in-situ technology, FISH can be directly applied to environmental bioaerosol samples without extraction or purification. It uses a DNA or RNA strand conjugated with fluorescent molecules (i.e., fluorochromes) as a probe to selectively hybridize and bond with a target gene in the microbes of interest, thereby enabling the detection of specific microbes. FISH can also be used to determine the presence of specific functional genes. The detection/identification is usually conducted with a fluorescence microscope (that excites the fluorochromes and visualizes the cells or cell organelles carrying the target genes). Only one study used FISH to study the composition of swine barn bioaerosols [87], with one general bacterial fluorescent probe and ten specific probes selected for microbial identification. The hybridization was conducted on gelatin-coated glass slides and the visualization was performed with an Axioscope II epifluorescence microscope (Carl Zeiss, Oberkochen, Germany).

PCR is fundamental to many nucleic acid-based methods. It selectively amplifies a target DNA or RNA gene or gene segment, which occurs at trace amounts in a sample, to such an extent that molecular analysis of the gene becomes feasible. Section 4.4.1 discusses a special type of PCR known as qPCR that can quantify the copies of a target gene in a bioaerosol sample. Similar to qPCR, PCR uses a pair of primers to define and delimit the region of genes to be amplified. The selection of primers depends on the microbes of interest and the subsequent analysis. Relevant information is given when discussing specific analytical methods. Similar to qPCR, PCR is usually conducted through thermal cycling—DNA strands are synthesized at low temperatures (from free nucleic acids) and denatured at high temperatures (deconjugated as free single strands) repeatedly over multiple cycles.

Before PCR, the genetic material (usually DNA) of microbes must be extracted and purified. Many studies used a soil DNA extraction kit (e.g., Power Soil Kit) for this purpose [87,192,210,218–220,263,266,267,270,273,274]. Other extraction kits selected include QIAamp DNA Mini Kit [169,262], Fast DNA SPIN Kit [87,272], GenElute Plant Genomic DNA Miniprep Kit [261], Qiagen DNA Minikit [221], NucliSens Magnetic Extract Kit [268], and Nucleospin 8 Plant II Kit [269]. Two studies used a customized extraction proto-

col [209,217]. For filter samples, a PM-laden filter can be sliced and then subjected to DNA extraction [209,218–220,263,270–272], or PM can be washed off the filter and pelleted (through centrifuging) for DNA extraction [169,262,267,274]. For bioaerosols sampled with ESP, a mixed solution of NaCl and Tween 80 was used to extract bioaerosols from ESP [210]. A similar procedure was adopted by Ref. [265]. After DNA extraction and PCR, the following analyses were carried out:

- DGGE. Three early studies adopted this technology [169,209,262]. DGGE was extensively used in microbial community studies but has been largely replaced by sequencing in recent years. DGGE separates a mixture of amplified DNA sequences (PCR products) into multiple bands through electrophoresis, based on their differences in melting in a denaturing agent. The number, location, and intensity of the bands can be used to characterize and compare the richness and diversity of microbial communities. The bands can be excised for sequencing to further identify associated microbial species. In the literature, the V3 [169,262] and V4–V5 regions [209] of 16S rRNA gene sequences were PCR'd before the DGGE analysis. The 16S rRNA gene refers to the DNA sequence that codes 16S ribosomal RNA in a prokaryotic cell; and it is the most prevalent fingerprint for bacterial/archaeal identification and phylogenetic analysis [407].
- Clone library. Five studies adopted this technology. Among them, two excised DGGE bands [169,262] and three collected purified PCR products (amplicons) [87,217,261] for clone library construction. The construction is usually conducted using a TOPO TA cloning kit. Specifically, the purified PCR products are ligated into plasmid vectors in host bacterial cells (e.g., *E. coli*) and cloned by cultivating the cells. The recombinant plasmids from clone colonies are then sequenced for microbial identification. An advantage of the clone library method is that it enables long reads (typically >1000 base pairs (bps)) from sequencing and, thus, allows for microbial identification (taxonomic/phylogenetic assignment) at high resolutions, e.g., at species or strain levels. The clone library method is often coupled with Sanger sequencing, a traditional low-throughput DNA sequencing technology but capable of generating long reads. Due to the long reads, the PCR primers selected for clone library construction are usually different than those for other sequencing technologies. The primer pairs selected include 63f/1387r [169], 8f/1492r [87], 515f/1391r [217], and 27f/1492r [261] for bacteria, Ar3f/Ar1492r for archaea [262], ARC8f/ARC1492r for archaea and fungi [87], and nu-SSU0817f/nu-SSU-1536r for fungal phyla Ascomycota and Basidiomycota [87].
- NGS. Compared with clone library sequencing, NGS is a high-throughput sequencing technology and can offer millions of reads in a single run. However, the reads are relatively short. Thus, NGS generally cannot provide the same taxonomic resolution as clone library sequencing. NGS is the most prevalent sequencing method in the literature for its massive data output and affordability. Two NGS techniques were used: (1) barcoded 454 pyrosequencing (454 Life Sciences, Branford, CT, USA) and (2) Illumina sequencing (Illumina Inc., San Diego, CA, USA). The former technique, now discontinued, was used by Refs. [209,263]; while the latter was used in all other NGS studies (Table 9). Illumina sequencing provides more reads in a single run but the reads are generally shorter than 454 pyrosequencing (150 bps vs. 500 bps). A technical review of Illumina sequencing and 454 pyrosequencing is beyond our expertise. Additional information can be found from the companies' websites or Ref. [416]. The PCR primers selected for NGS include 519f/926r [209], 338f/533r [218], 338f/519r [219], 27f/338r [263], 341f/805r [265,271], 515f/806r [221,274], 515f/805r [210], 515f/813r [268], and 338f/806r [270,272,273] for bacteria; and ITS1FI2f/ITS2r [220], ITS1f/ITS1r [192,210], ITS3f/ITS4r [268], and ITS1f/ITS2r [270] for fungi.
- Shotgun metagenomic sequencing. Only two studies adopted this technology [267,269]. Instead of using PCR primers to target fingerprint DNAs, shotgun metagenomic sequencing, in principle, sequences all genomic DNAs in a bioaerosol sample. To do it, DNA molecules extracted from the sample are broken down into short fragments

(e.g., 350 bps [267]), (optionally) barcoded, and submitted for NGS sequencing. The acquired sequence reads are processed and assembled (based on their overlaps) into long sequences for taxonomic assignment and classification. Compared with PCR-based sequencing, shotgun metagenomic sequencing requires no PCR, provides superior taxonomic resolution, identifies prokaryotes and eukaryotes concurrently, and is capable of profiling functional genes (e.g., ARGs). However, it has limited coverage of bacterial species, requires a greater amount of DNA input, and is prone to interference from host (e.g., swine) DNA. In Ref. [267], ~5 µg of DNA per sample was gathered for shotgun metagenomic analysis on an Illumina HiSeq 2000 platform. Functional annotations and antibiotic resistance genes were also reported. To overcome the issue of limited DNA material in bioaerosols, a Kapa Hyper Prep Kit was used by Ref. [269] to amplify certain genes and prepare a library for metagenomic sequencing.

For sequencing-based microbial identification, bioinformatic analysis and reference databases are critical. Numerous bioinformatics tools (for purposes such as alignment, trimming, error correction, and taxonomic annotation) were used in the literature, and a review of these tools is beyond the scope of this review. Regarding reference databases, the databases for bacterial identification include the National Center for Biotechnology Information (NCBI) BLAST [169,217,262], Ribosomal Database Project (RDP) [209,261], Silva 16S rRNA [87,210,219,265–268,273,274], EzTaxon-e [218], Greengenes [263], and Genbank [261]; and the ones for fungi include Silva 18S rRNA [87], NCBI BLAST [220], UNITE [192,210], and GenBank [268].

#### 4.4.4. ARB and ARGs

##### ARB Measurement

ARB are often detected with culture-dependent methods and the detection involves the use of selective or differential cultivation media. A selective medium is used to isolate a target group of bacteria by inhibiting the growth of other microbes. A differential medium is used to distinguish a target group or species from closely resembled groups. Various selective/differential media are commercially available (Table 22). These media must be incubated under appropriate conditions (temperature, time duration, and atmosphere) to ensure the reliability of measurement results.

In addition to commercial media, self-prepared media can be used for ARB detection. These media are often prepared by amending a general-purpose bacterial cultivation medium (e.g., TSA or R2A) with antibiotics or antimicrobials. For example, Ref. [186] added tetracycline and erythromycin each to an LB nutrient agar and used the self-prepared selective media to detect airborne tetracycline- and erythromycin-resistant bacteria in swine barns. Ref. [263] used kanamycine-amended Luria–Bertani agar plates for the detection of kanamycin-resistant bacteria.

Selective/differential media can be loaded in an agar plate-based impactor for direct ARB measurement [178,179,183,185] or used as a screening tool to identify ARB in samples from other types of bioaerosol samplers. For PM-laden filters, bioaerosols are first extracted into liquids. Liquid samples from PM extraction, wet cyclones, or impingers are diluted and then plated on selective/differential agar plates [101,175,176]. The liquids can also be filtered and the filters are then pressed on the agar plates for ARB cultivation and identification [175,290].

Antimicrobial susceptibility testing (AMT) is a prevalent approach to identifying ARB colonies formed on a cultivation medium, either general-purpose or selective. Refs. [160,163] adopted a three-step procedure for airborne ARB detection. In the first step, bioaerosols were collected and grown on TSA agar plates. In the second step, selective agars were used to identify the bacteria of interest (e.g., *S. aureus*) with the replica plate method. In the final step, the colony isolates of known bacteria were subjected to AMT by challenging these isolates (on Mueller–Hinton agar and TSA plates) with antibiotics. The final step followed the Kirby–Bauer diffusion disk method. The same method was adopted by Ref. [286]; however, the study used a selective medium for *E. coli* collection and growth and, thus,

had no separate identification step. Ref. [285] employed a three-step procedure similar to Refs. [160,163] but with two differences: (1) an impinger was selected for bioaerosol collection and the acquired liquid was diluted and cultivated on a selective medium for *Enterococcus* identification, and (2) in the final step, the MIC agar dilution method was selected for AMT analysis.

The ARB isolates can be further analyzed for ARGs [101,175,176,183], resistance to other antibiotics [183,289], and microbial composition [179,186,263]. For MRSA, *mec(A)* genes are often analyzed through PCR to confirm the isolates' methicillin resistance, and Pantone–Valentine leucocidin or *nuc* genes can be PCR'ed to confirm the presence of *S. aureus*. Other post-analysis of MRSA isolates include *spa* typing (PCR detection of *spa* gene subtypes) and coagulase reactions (positive for *S. aureus*).

Regarding sampling methods, filter samplers, agar plates-based impactors, impingers, and wet cyclones have all been reported. Among them, viable Andersen cascade impactors were most commonly selected. These impactors are prone to overloading when measuring airborne culturable bacterial counts in swine barns. Accordingly, the sampling typically took only tens of seconds or a few minutes. This issue could be slightly relaxed for ARB sampling when selective media are used for ARB collection and the target ARB accounts for only a small portion of total culturable bacteria.

#### ARG Measurement

Two types of nucleic acid-based methods were used for ARG detection: PCR (including qPCR) and metagenomic sequencing (Table 11). Both PCR and qPCR use the same ARG primers. While regular PCR can identify the presence of target ARGs, qPCR can quantify the copies of target ARGs in a DNA extract. For each ARG subtype (e.g., *mec(A)* and *tet(H)*), the sequence information of primers can be found in corresponding publications.

Metagenomic sequencing can be classified into (1) amplicon and (2) shotgun metagenomic sequencing. Amplicons refer to PCR products containing selectively amplified DNA sequences or genes. Several types of amplicons are frequently used for the analysis of microbial communities in bioaerosol samples: 16S rRNA amplicons for bacteria and archaea, ITS amplicons for fungi, and 18S rRNA amplicons for fungi and other eukaryotes. None of them can detect ARGs. Comparatively, shotgun metagenomic sequencing requires no amplicons, i.e., PCR products. A brief description of this technology is given in Section 4.4.3. As it is whole-genome sequencing, the DNA sequences generated from shotgun sequencing can be compared against ARG sequence databases to identify the presence of ARGs. The ARG databases selected include the Antibiotic Resistance Database (ARDB) [267] and the Comprehensive Antibiotic Resistance Database (CARD) [274].

Various sampling devices have been used for ARG measurement, including Andersen viable impactors [286,291], impingers [172], wet cyclone (e.g., Coriolis<sup>®</sup> $\mu$ ) samplers [103], ESP samplers [269], and filter samplers [209,218,219,267,272,274]. For filter samplers, glass fiber filters were most commonly used, followed by cellulose nitrate filters. Various kits have been used for DNA extraction from collected samples, including QIAamp DNA Mini Kit, Power Soil Kit, Fast DNA Spin Kit, and Nucleospin 8 Plant II Kit. Additional information regarding DNA extraction can be found in Section 4.4.3.

#### 4.4.5. Bioaerosol Markers

##### Endotoxin

Endotoxins can be analyzed with GC-MS or LAL assays. The GC-MS method quantifies 3-hydroxylated fatty acids (3-OH FAs) in lipid A, a major component of endotoxin molecules [417]. The typical chain lengths of 3-OH FAs are C<sub>10</sub>, C<sub>12</sub>, C<sub>14</sub>, C<sub>16</sub>, and C<sub>18</sub> [418]. The analysis consists of three steps. First, endotoxins are extracted from a filter sample in a hot methanolic HCl solution. Secondly, the extracts are subjected to trimethylsilyl derivatization. Finally, the trimethylsilyl derivatives are analyzed using GC-MS or GC-MS-MS [419]. The measured concentrations are presented in the unit of ng 3-OH FAs m<sup>-3</sup>. As the GC-MS method measures specific chemicals with a well-refined wet chemistry

procedure, the analysis results are highly replicable—an advantage over the LAL assay method [420]. However, 3-OH FAs concentrations are not a direct measure of toxicity. Only one endotoxin study in swine barns used the GC-MS method [349]. PM sampling lasted for only 3 h in the study and, thus, the measurement results can hardly be compared with those from other studies.

The LAL assay is the de facto standard method for endotoxin analysis. LAL is an aqueous extract from the blood of *Limulus polyphemus*, a horseshoe crab. The animal is highly sensitive to infection by Gram-negative bacteria, and once infected, blood clotting quickly occurs [421]. Five types of LAL assays are commercially available: gel clot, endpoint fluorescent, endpoint chromogenic, kinetic chromogenic, and kinetic turbidimetric. Different types of LAL assays could produce different quantification results [326,422]. Most swine barn studies used the kinetic chromogenic LAL assay (Table 13) for its great sensitivity and wide detection range.

A chromogenic LAL assay is made up of a colorless substrate and a proenzyme extracted from amoebocyte cells in the blood of *Limulus polyphemus*. The proenzyme becomes an active enzyme in the presence of endotoxins. The enzyme then dissociates the colorless substrate into a short peptide segment and a p-nitroaniline (pNA; a yellow organic compound). The pNA can be photometrically quantified at 405 nm. In the kinetic assay, the pNA-associated color development is semi-continuously monitored at 37 °C with an incubating microplate reader. The onset time of a test sample (i.e., the reaction time needed to reach a pre-set absorbance level at 405 nm) is compared with the onset time of endotoxin standards to calculate the endotoxin concentration in the test sample.

The LAL assay analysis results are highly dependent on experimental procedures. For airborne endotoxin measurement, endotoxins are first extracted from collected PM samples and the extracts are then diluted to ensure that the diluted concentration stays within the detection range of a selected assay. Many factors can affect the potency of endotoxins and accordingly the analysis results, including PM sampling media, solvents for extraction and dilution, containers, and extraction methods [423–425]. Thus, caution must be taken when comparing the measurement results from different studies. Efforts have been made to develop a SOP [423–426]. However, no such protocol has been established. According to Ref. [420], LAL assays would be suitable for comparing endotoxin contamination in similar environments; and when environmental conditions are substantially different, the GC-MS method could be a better choice.

As the LAL assay method is based on enzyme reactions, factors that promote or inhibit enzyme activation and activity can interfere with the analysis results [421]. To identify possible inhibition or enhancement, positive product control (PPC) samples are often analyzed, along with test samples and endotoxin standards. When significant inhibition or enhancement is observed, the test samples should be re-extracted or diluted to another ratio. To address the interference issue, a special kinetic chromogenic LAL assay named the kinetic *Limulus* assay with resistant-parallel-line estimation (KLARE) was developed by Ref. [211]. Two previous studies used the KLARE assay for airborne endotoxin measurement in swine barns [55,326]. However, according to Ref. [326], the KLARE assay could underestimate endotoxin concentrations when glass fiber filters were used for PM sample collection.

Due to the ubiquitous presence of Gram-negative bacteria, caution must be taken to prevent possible contamination of PM samplers and filters before, during, and after field sampling. Prevention measures may include sterilizing samplers and filters (through UV radiation, ozonation, or baking (for glass or quartz fiber filters only)), keeping filters in sterile containers, storing filters in an ultralow temperature freezer, and sterilizing filter handling tools using lab micro-incinerators [20,302].

#### (1→3)-β-D-glucan

Two methods are available for (1→3)-β-D-glucan analysis: inhibition enzyme immunoassay (EIA) and LAL assay [337]. They both have been used for PM studies in swine barns. However, no SOP has been available for airborne (1→3)-β-D-glucan measurement.

The EIA method was developed by Ref. [45]. It is a competitive enzyme-linked immunosorbent assay (ELISA) with (1→3)-β-D-glucans as the antigen. In this method, a glucan conjugate solution is injected into animals, e.g., rabbits, to create anti-β (1→3)-glucan antibodies in the blood, and the antibodies are then extracted and purified for detection purposes. As an ELISA method, the EIA requires a primary and a secondary antibody, prepared with different animals. A downside of the method is its limited sensitivity. To ensure that the (1→3)-β-D-glucan concentration in a PM extract is greater than the detection limit, preparing a 1-mL extract requires 25 mg PM [338]. A revised EIA named monoclonal antibody-based two-site enzyme immunoassay (mAb-EIA) was developed with improved selectivity and it was used for bioaerosol measurement in swine barns [344].

The LAL assays for (1→3)-β-D-glucan analysis are similar to those for endotoxins. The only difference lies in activating factors. While activating factor C is used in endotoxin LAL assays, activating factor G is used in (1→3)-β-D-glucan assays. Only (1→3)-β-D-glucans can activate factor G, thereby initiating an enzyme-catalyzed color- or turbidity-yielding process [427]. Among various assays, a kinetic chromogenic LAL assay was often selected (Table 14) due to its large detection range and superior sensitivity. The experimental procedure is similar to that for airborne endotoxin analysis. However, an alkaline solution is advised to use for PM extraction to release (1→3)-β-D-glucans from fungal cell walls while destroying endotoxins. Compared to the EIA, the kinetic chromogenic LAL assay is more sensitive, accurate, and specific; and it requires less lab equipment [338]. The kinetic chromogenic LAL assay with activating factor G has been extensively used for airborne (1→3)-β-D-glucan assessment in various environments, including swine barns. The assay requires a nearly neutral pH condition. A pH adjustment must be conducted to the alkaline extract before submitting it for analysis [20].

#### 4.5. Chemical Analysis

##### 4.5.1. Elemental Composition

###### Individual Particles

SEM-EDX is the most prevalent method for elemental composition analysis of individual particles, including swine barn PM. To do it, PM must be first collected on filters [80,104,148] or microscopy grids [125]. Both polycarbonate filters [80,148] and quartz fiber filters [104] were used. However, a quartz fiber filter is not recommended due to its rough surface (providing a poor background). The sampling time should be carefully regulated to avoid the overlapping of particles on the filter surface.

In an SEM-EDX system, SEM is used to identify and magnify a target particle, thus enabling the subsequent EDX analysis. As an accessory of SEM, EDX energizes a particle with an X-ray or electron beam and detects the X-rays emitted from the particle. X-rays are emitted when electrons in the particle's atoms restore from excited (energized) to ground states. As the energy of emitted X-ray photons is element-specific, EDX can identify chemical elements in the particle and further enable quantitative analysis based on photon counts. However, the quantitative analysis using SEM-EDX is known to carry large uncertainties. For example, the lack of calibration standards with X-ray generation and propagation manners similar to actual PM samples can compromise the accuracy of quantitative analysis [428]. Thus, SEM-EDX may better serve as a particle classifier—as in [80,148]—than a quantitative tool.

###### Bulk PM

No method information is available in Ref. [124]. Ref. [85] used X-ray fluorescence (XRF) and ICP-AES to determine the elemental composition of PM<sub>2.5</sub> and PM<sub>10</sub> samples from swine barns. Both XRF and ICP-AES are listed by the U.S. Environmental Protection Agency (EPA) as standard methods (Method IO-3.3 [429] and Method IO-3.4 [430]) for ambient PM analysis and have been extensively used worldwide. Examples of their applications include the CSN and IMPROVE networks in the U.S. Another EPA standard

method is Method IO-3.5 that uses inductively coupled plasma-mass spectrometry (ICP-MS) [431]. However, its application to swine barn PM has not been reported.

The detection principle of ICP-AES can be found in Ref. [430]. In brief, the instrument nebulizes a test solution or suspension and injects formed droplets into a test chamber where an argon plasma (also known as argon torch) bombards off the water and breaks down molecules into their respective atoms. In the plasma, the atoms are ionized (lose electrons) and quenched (regain electrons) repeatedly, emitting light with wavelengths characteristic of the elements contained. For ICP-AES analysis, PM samples are first collected on a filter. Teflon filters and acid-washed quartz fiber filters are normally used due to their low impurity contents. Next, PM is extracted with hot acids or microwave extraction. A hot acid extraction method (a mixture of hydrochloric acid and nitric acid, heated at 160 °C) was used by [85]. The extract can then be submitted for ICP-AES analysis. A similar sampling and extraction protocol applies to ICP-MS analysis. ICP-MS is selected when exceptional sensitivity is required.

The detection principle of XRF is similar to that of EDX [429]. For XRF analysis, PM is usually sampled on a ringed Teflon filter (e.g., Teflo™ from Pall Corp., Port Washington, NY, USA). The filter is exceptionally thin and light and contains a very low impurity level. These features are critical for XRF analysis. However, XRF may be unsuitable for swine barn PM analysis, especially the quantification of light elements [85]. Due to high PM concentrations in swine barns, the PM deposited on a filter surface can easily exceed 0.1 mg cm<sup>-2</sup> filter area, causing significant attenuation of low-energy fluorescence X-rays emitted from light elements. The XRF analysis of PM samples also involves several other assumptions (e.g., PM size and composition) and restrictions. A detailed discussion about the issues can be found in [85]. Compared to ICP-AES, XRF is a non-destructive method. Thus, the samples after XRF analysis can be submitted for other physical and chemical tests [429].

#### 4.5.2. PM-Borne Odors

The measurement of PM-borne odors typically consists of three steps: (1) sampling, (2) sample extraction or preparation, and (3) odor or odorant analysis. As many PM-borne odorants in swine barns are volatile organic compounds (VOCs), the measurement protocols developed for indoor PM or ambient PM-borne VOCs were usually followed. However, a unique environment condition in swine barns (e.g., exceptionally high VOC concentrations in the gas phase) poses a challenge to odorant measurement. A discussion of the challenge and possible solutions is attempted in this section.

#### Settled Dust Sampling

Few technical details are available in the literature regarding the collection of settled dust. Ref. [357] placed aluminum foil-covered sedimentation plates 2 m above the floor to collect settled dust. Ref. [363] collected settled dust from exhaust fans and kept it in 10-mL glass vials. Ref. [364] used a vacuum sampler to collect dust into sample bags. Ref. [146] scraped dust from exhaust fans, pen dividers, and pipelines using a stainless-steel knife and kept the samples in Petri dishes at 4 °C. The collected settled dust samples were directly submitted for sample extraction/preparation [146,357,363], resuspended in the lab for PM sampling [354], or size separation [364] before moving to the next experimental step.

#### PM Sampling

PM sampling setup has a large influence on odorants measurement. Relevant information, however, is limited in the literature. Regarding samplers, an ESP was used by [355] for TSP collection, in which particles were attracted onto a collecting electrode. All other studies used filter samplers; but the specific setup differed among publications.

- Filters. Several studies used a single glass fiber filter for PM sampling [277,350,365]. Glass fiber filters are inexpensive and easy to clean (via baking). However, they are known to absorb significant amounts of VOCs from the air, thereby leading to an

overestimate of PM-borne odors [367]. The same sampling artifact also occurs to quartz fiber filters [368]. To address the issue, [277,368] dislodged particles from filters for odorant analysis. However, it only works when PM loadings are high on filters. Another solution is to use dual glass or quartz fiber filters: a front filter and a backup filter. With a long enough sampling time, both filters would have been saturated by VOCs from the air. By subtracting an odorant's concentration on the backup filter from that on the front filter, the odorant concentration in collected PM can be calculated. Dual filters were attempted by Ref. [355] and later adopted by Refs. [367,369] in the studies of swine barn PM-borne odors. According to Ref. [367], a backup filter should also be installed when sampling PM with a Teflon-coated filter. Teflon-coated filters absorb lesser but still significant amounts of odorants from the air. Ref. [366] collected PM on Teflon filters with even less gas adsorption.

- Denuders. A denuder is a non-obstructive scrubber that can remove certain VOCs from the air (depending on coatings). Only one study included denuders [367]. However, the purpose was to capture VOCs in the air for odorant analysis. In a classic setup for PM-borne VOCs sampling, a denuder is installed before filter holders to pre-remove gaseous VOCs in the air. Dual filters may also be used; but in this case, the concentration of a PM-borne odorant would be equal to the sum of its concentration on the front filter and that on the backup filter [23]. The backup filter can be replaced by a sorption column to capture all the VOCs bypassing or released from the front filter.
- Filter samplers. Various filter samplers were used, including isokinetic TSP samplers [277], Harvard impactors [277], open cassettes [365,369], high volume PM<sub>10</sub> sampler [368], and TEOM [366]. In theory, VOC sorption and desorption on a PM-laden filter or a denuder can be affected by linear velocity (which is equal to a sampling airflow rate divided by an effective filter area) [432]. Thus, the specifications of a PM sampler can also affect odorant sampling. However, no relevant information is available in the literature. Teflon filters in a TEOM are usually kept at 50 °C to prevent condensation. This may cause the volatilization loss of volatile odorants [366].

Upon the analysis of the sampling protocols, we recommend the use of dual filters for high or middle-volume PM sampling (e.g., 16.7 LPM for 24 h) and the use of a denuder plus a filter plus a sorption column for low-volume PM sampling (e.g., 5 LPM for 2 h). In the latter setup, glass or quartz fiber filters can be used due to their large VOCs adsorption capacity. An ESP sampler may also be selected for its exemption from filter-induced sampling artifacts. The PM samples collected should be immediately extracted [365] or stored in an ultra-low temperature freezer (−80 °C) to minimize volatilization loss [277].

#### Sampling Extraction/Preparation

Various sampling extraction or preparation methods were used, including solvent extraction, headspace SPME, P&T, and thermal desorption. Among them, solvent extraction was the most prevalent method. The solvents selected include a water–ether mixture [354], diethyl ether [277,355], ethanol [357], methanol [363], dichloromethane [365,369], and acetone [364]. These solvents differ in their properties (e.g., polarity) and, thus, extract an odorant at different efficiencies. An extraction efficiency (EE) can further be affected by factors such as filter material, extraction temperature and time, and agitation strength. EE is especially important for quantitative analysis. To assess and address EE, dust samples or PM filters can be spiked with internal standards with known concentrations before extraction [277]. The acquired extract was usually concentrated in a pure N<sub>2</sub> stream [277,355,364,365,369] before odorant analysis. Solvent extraction is a classic method for the measurement of PM-borne VOCs. It offers accurate quantitative results when properly operated. However, solvent extraction is time-consuming and labor-intensive compared to methods such as headspace SPME and purge and trap (P&T).

PM extracts can be further processed through adsorption, derivatization, extraction, evaporation, etc. These post-solvent extraction steps were common in early studies but have not been seen in the literature since 1990. Ref. [354] used a cold trap to condense PM

extract vapors and then concentrated acquired liquids on a water bath. Ref. [355] conducted a series of wet chemistry experiments to further separate and purify acids, phenols, and carbonyls from PM extracts. Ref. [357] alkalized a PM extract and then acidified it to separate phenolic/indolic compounds from fatty acids.

Headspace SPME is a technology developed by Dr. Pawliszyn and his colleagues at the University of Waterloo, Canada, in the early 1990s [433]. It features the use of coated silica fibers to sorb (absorb and/or adsorb) and enrich VOCs from the air in a headspace. The technology was initially designed for the measurement of VOCs in water samples but quickly found applications for other environmental samples, including PM [434]. For PM-borne VOCs measurement, a PM sample is placed in an enclosed vial and the VOCs released from the sample are sorbed onto a pre-conditioned SPME fiber or fibers. Headspace SPME was employed by three studies [365,366,368,369] to extract odorants from swine barn PM; however, no quantitative analysis was conducted. The SPME filters tested include polydimethylsiloxane (PDMS), Carbowax/divinylbenzene, and Carboxen/PDMS. Headspace SPME can be used for quantitative VOC analysis; but in general, it is less prevalent than solvent extraction. When using SPME, the GC must be equipped with an SPME injector or injector liner.

P&T is a popular technology to analyze VOCs in water and soil samples. It uses an inert gas stream to purge a liquid or solid sample and a cold or sorbent trap to capture (and enrich) the VOCs purged off the sample. At elevated temperatures, the captured VOCs are released from the trap and transferred to GC for analysis. Only one study used P&T for swine barn PM samples [365,369] and it was limited to qualitative analysis.

Thermal deposition (TD) desorbs PM-borne odors (odorants) at increased temperatures. The method was adopted by two previous studies but in different fashions. Ref. [130] built a temperature-controllable convective chamber, inside which a settled dust sample was purged with pure air and heated to 105 °C. The air effluent from the chamber was collected for NH<sub>3</sub> and odor analysis. Ref. [367] placed a PM filter in an empty steel tube and heated the tube to 100 °C, 200 °C, and 290 °C with a TD module. The module was part of a TD-GC-MS system. The gas desorbed from the PM filter was immediately analyzed by GC-MS. Quantitative results were available from both studies.

#### Odor/Odorant Analysis

Most of the previous studies used GC to analyze odorants, especially organic odorants in swine barn PM samples. In GC, a packed or capillary column separates multiple gas analytes based on their different affinities to the column packings or coatings and accordingly different mobilities in the column. Capillary columns were usually selected for their superior separation performance. Packed columns were only used in early studies [354,355,357]. The capillary columns selected in the literature included weakly polar columns (DB-5 and alike) [364,365,368,369] and polar columns (DB-wax and alike) [277,363,366,367]. During capillary GC analysis, a column oven was programmed to raise its temperature from 40–80 °C to 150–260 °C at a rate of 4–10 °C min<sup>-1</sup>. The temperature of GC injectors was typically maintained at 250 °C. A variable injector temperature program (−50 to 250 °C at 12 °C min<sup>-1</sup>) was adopted by Ref. [277]. For injection of PM extracts, both splitless [277] and split modes [363] were reported.

Two types of GC detectors were commonly selected for odorant analysis: FID [354,357,362,363] and mass selective detector (MSD or MS) [277,354,364–368]. An FID uses a hydrogen flame to ionize organic molecules and detects induced currents. An MSD breaks down a molecule into ionized fragments and measures the mass-to-charge ratios of produced fragments—known as a mass spectrum. As a mass spectrum is molecule specific; it enables the quick identification of an odorant. Comparatively, an FID relies on a comparison of retention time with standards for odorant identification. The temperatures of FID and MSD (Note: MSD transfer lines) were typically set at 280 °C to avoid the condensation of semi-volatiles. A higher MSD transfer line temperature (320 °C) was adopted by Ref. [368]. For both GC-FID and GC-MS, the quantification of odorants requires calibration standards. They

can be external or internal standards. A unique calibration method known as stable isotope dilution was employed by Ref. [277] which featured the spiking of PM filter samples with stable isotope internal standards. However, this method only works for GC-MS. Other GC systems include a GC with a sulfur chemiluminescence detector for H<sub>2</sub>S detection and quantification [364].

SIFT-MS was used for odorant analysis [368]. It is a real-time chemical ionization MS technology and can measure multiple gases simultaneously. Additional information about this technology can be found in Ref. [435]. It is noteworthy that Ref. [368] used SIFT-MS to study the sorption of four odorants on swine barn PM but still used GC-MS to detect odorants in actual PM samples.

A few studies analyzed NH<sub>3</sub> and odor emission potency in swine barn PM [36,146,356]. However, only one study entailed analytical methods [146]. For odor, the gas sample from thermal desorption was transferred into a gas sampling bag and then shipped to an olfactometry lab for odor concentration measurement. In the lab, an olfactometer serially diluted the sample until odor panelists were unable to detect the odor in the diluted sample. The dilution ratio was taken as the odor threshold concentration. For NH<sub>3</sub>, the gas sample from thermal desorption was immediately transferred into a chemiluminescence NH<sub>3</sub> analyzer. NH<sub>3</sub>/NH<sub>4</sub><sup>+</sup> in swine barn PM can also be quantified by ion chromatography or colorimetry after being extracted in water [85].

In summary, various methods have been adopted to detect and quantify PM-borne odors or odorants in swine barns. For qualitative analysis, a three-step procedure is recommended: (1) ESP sampling, (2) headspace SPME, and (3) GC-MS analysis. A dual-filter setup is not recommended, because a correction of sampling artifacts requires quantitative information. For quantitative analysis, the three steps are (1) ESP or dual-filter sampling, (2) solvent extraction, and (3) GC-MS analysis. We suggest spiking PM samples with internal standards before solvent extraction so that the recovery efficiency of an odorant during sample extraction and analysis can be accurately assessed and considered.

## 5. Mitigation Technologies

PM could exert adverse health effects on pigs and farm workers and compromise equipment and materials inside swine barns. Upon being emitted into the atmosphere, PM could spread odors, pathogens, and other hazardous substances to nearby livestock farms and communities. PM mitigation, thus, benefits both pig production and environmental stewardship.

According to their installation locations, PM mitigation technologies can be generalized into two categories: in-barn and end-of-pipe. In-barn mitigation occurs inside a swine barn, with the primary goal of reducing indoor PM concentrations. End-of-pipe mitigation occurs at the exhaust of a barn to reduce PM emissions into the atmosphere. Only in-barn mitigation technologies are reviewed (Table 23) as this report focuses on PM in swine barns. A review of end-of-pipe PM mitigation technologies, such as cyclones and wet scrubbers, can be found in Refs. [115,436].

**Table 23.** In-barn PM mitigation technologies tested in swine barns since 1990.

Reference	Technology	Reduction Effectiveness <sup>a</sup>	Barn and Ventilation Type	Location	Main Findings
	Oil/water sprinkling				
[123]	Sprinkling a mineral oil (~15 mL oil m <sup>-2</sup> day <sup>-1</sup> ) to barn floors	75% for respirable 76% for inhalable	Grower–finisher; MV	Saskatchewan, Canada	Right after oil sprinkling, PM concentrations were close to those in a nearby office.
[44]	Sprinkling a rapeseed oil-water mixture (5–10 mL oil pig <sup>-1</sup> day <sup>-1</sup> )	76% for respirable 54% for respirable 52% for respirable	Nursery; n/a Grower; n/a Finisher; n/a	Demark	No improved pig performance was observed.
[389]	Spraying 24–60 mL m <sup>-2</sup> to barn floors	Up to 73% for respirable	Farrowing; MV	Alberta, Canada	–

Table 23. Cont.

Reference	Technology	Reduction Effectiveness <sup>a</sup>	Barn and Ventilation Type	Location	Main Findings
[390]	Sprinkling canola oil at various rates	37–89% for TSP	Grower–finisher; MV	Saskatchewan, Canada	At the same total volume of oil applied, PM reduction increased with oil sprinkling frequency. Other benefits included a 94% reduction in personal PM exposure, a 92% reduction in airborne endotoxins, and an 89% reduction in personal endotoxin exposure. A specially designed oil sprinkling system was developed.
[49]	Sprinkling canola oil (on average 6.7 mL oil m <sup>-2</sup> day <sup>-1</sup> )	~84% for TSP	Grower–finisher; MV	Saskatchewan, Canada	
[437]	Sprinkling canola oil (on average 4.3 mL oil m <sup>-2</sup> day <sup>-1</sup> ) Spraying a rapeseed oil-water mixture	87% for TSP 90% for inhalable 86% for respirable	Grower–finisher; MV	Saskatchewan, Canada	
[438]	(3.1–3.9 g oil m <sup>-2</sup> day <sup>-1</sup> ), coupled with adding 4% of fat to the feed	~80% (35–95%) for TSP	Finisher; MV (chimney)	Denmark	Animal activity controlled spraying reduced oil-water usage.
[439]	Sprinkling canola oil	85% at 10 mL g oil m <sup>-2</sup> day <sup>-1</sup> 92% at 20 mL g oil m <sup>-2</sup> day <sup>-1</sup> 95% at 30 mL g oil m <sup>-2</sup> day <sup>-1</sup>	Finisher; lab chambers	Quebec, Canada	–
[440]	Sprinkling vegetable oil (on average 6.7 mL oil m <sup>-2</sup> day <sup>-1</sup> )	59% for TSP 65% for respirable 79% for inhalable	Finisher; side-curtain	Minnesota	The operating cost was USD 0.58 per pig.
[61]	Sprinkling soybean oil (on average 6.7 mL oil m <sup>-2</sup> day <sup>-1</sup> )	70% for all sizes of particles	Finisher; MV	Illinois	–
[62]	Spraying an oil-water mixture (7 g soybean oil pig <sup>-1</sup> day <sup>-1</sup> or 8 g canola oil pig <sup>-1</sup> day <sup>-1</sup> )	~52% for respirable	Finisher; n/a	Iowa	No difference between soybean and canola oils.
[65]	Atomizing an acid-oil-alcohol mixture (45 mL oil m <sup>-2</sup> day <sup>-1</sup> )	75–90% for TSP, PM <sub>10</sub> , and PM <sub>2.5</sub>	Finisher; MV (tunnel)	Mid-Atlantic region, USA	>90% reduction in the levels of total viable bacteria, Gram-negative bacilli, and <i>Enterococcus</i> spp. was also achieved.
[67]	Sprinkling soybean oil	67% for TSP	Finisher; MV	US Midwest	–
[165]	Spraying seven different liquids	~70% for TSP when spraying soybean oil	Grower; MV	Korea	Except for soybean oil, dust reduction by other liquids diminished after 3 h.
[441]	Sprinkling canola, soybean, or sunflower oil (10 mL oil m <sup>-2</sup> day <sup>-1</sup> )	68–72% for TSP	Finisher; lab chambers	Quebec, Canada	–
[166]	Sprinkling a canola oil-water mixture (3 g oil pig <sup>-1</sup> day <sup>-1</sup> )	28% for respirable 43% for inhalable 42% for total culturable bacteria	Nursery; n/a	Australia	–
[442]	Sprinkling canola oil (on average 6.7 mL oil m <sup>-2</sup> day <sup>-1</sup> )	17% for respirable 46% for inhalable	Grower; n/a		Total viable bacterial levels increased after treatment.
[83]	Sprinkling canola oil (on average 6.7 mL oil m <sup>-2</sup> day <sup>-1</sup> )	~57% for respirable	Grower–finisher; MV	Saskatchewan, Canada	~32% reduction in personal exposure to respirable PM.
		86% for TSP 82.5% for endotoxins 32% for total culturable bacteria 52% for total culturable fungi	Grower–finisher; MV	Saskatchewan, Canada	The reduction was greater for larger than smaller particles.

Table 23. Cont.

Reference	Technology	Reduction Effectiveness <sup>a</sup>	Barn and Ventilation Type	Location	Main Findings
[443]	Sprinkling a canola oil-water mixture (6.3 g oil m <sup>-2</sup> day <sup>-1</sup> )	35% for respirable 51% for inhalable 55% for total culturable bacteria 13% for respirable 53% for inhalable	Nursery; MV  Grower; NV	Australia	–
[100]	Sprinkling an oil-water mixture (3–6 mL oil pig <sup>-1</sup> day <sup>-1</sup> )	29–83% for TSP 20–80% for PM <sub>10</sub>	Finisher; MV	Germany	Total viable bacterial levels increased after treatment. The sprinkling system with smaller nozzles delivered better reduction than that with larger nozzles.
Ionization					
[138]	Commercial negative ionization systems (–9300 and –16,500 V)	Up to 46% at low ventilation rates; as low as 3% at high ventilation rates	Grower–finisher; MV	Saskatchewan, Canada	Reduction efficiency decreased with PM buildup on electrode surface.
[444]	Self-customized electrostatic precipitators made from charging wires and collection pipes (–24 kV)	36% for respirable 58% for TSP	Nursery; MV	Iowa	Better removal performance was observed for larger particles.
[380,445]	Electrostatic space discharge systems (ESDS)	50% for respirable 45% for TSP 40–46% for PM <sub>2.5</sub> 32–58% for PM <sub>10</sub> 45–83% for TSP	Farrowing; MV  Finisher; MV	n/a, likely U.S. Midwest	–
[446]	Air cleaners (plasma-based ionization)	78% for PM <sub>1</sub> 79% for PM <sub>2.5</sub> 22% for total culturable 25% for total culturable fungi	n/a; MV (cross-flow)	Korea	No reduction in PM <sub>10</sub> or TSP was seen.
[447]	Commercial electrostatic particle ionization (EPI) systems (–30 kV)	47% for PM <sub>2.5</sub> 58% for PM <sub>10</sub>	Nursery; n/a	n/a, likely U.S. Midwest	–
[448]	Commercial negative corona ionization (NCI) systems (–30 kV)	36% for PM <sub>10</sub>	Finisher; MV	The Netherlands	–
[133]	Commercial electrostatic filtration unit (EFU) systems	28% for PM <sub>10</sub>	Commercial EPI systems (–30 kV)	Kansas	–
[388]	Commercial EPI systems (–30 kV)	By particle counts: 46% for PM <sub>10</sub> 44% for PM <sub>2.5</sub>	Nursery; n/a	Minnesota	Reduction in viable PRRSV and IAV was observed.
[254]	Commercial EPI systems (–30 kV)	76–82% for PM ≥ 1 μm 52–56% for PM < 1 μm	Nursery; environment-controlled chambers	Manitoba, Canada	PM reduction performance decreased with raised ventilation rates.
[122]	Commercial EPI systems (–30 kV)	>94% for PM ≥ 0.6 μm 100% for PM ≥ 6 μm 68–96% for PPRSV	No pigs; test chambers	Manitoba, Canada	PM reduction performance decreased with raised ventilation rates.
Alternation of feed and feeders					
[449]	Adding 2% canola oil to feed	32% for inhalable	Grower–finisher; MV	Saskatchewan, Canada	Increases in respirable particle count and bacterial count were observed.
[47]	Pelletizing feed into 3-mm pellets	40% for respirable	n/a; lab chamber	United Kingdom	–
[450]	Coating feed pellets with 2% lignin	33% for respirable			
[450]	Coating feed pellets with 2% fat	25% for respirable			
[47]	Adding 4% animal fat to feed	37% for TSP 57% for respirable 42% for TSP 39% for respirable	Nursery; MV (chimney) Finisher; MV (chimney)	Denmark	A 47% reduction in TSP exposure was observed. A 67% reduction in TSP exposure was observed.
[450]	Adding 3% corn oil to feed	40% for TSP	Grower–finisher; MV	South Dakota	–
[442]	Reducing crude proteins in feed diet (19.5% to 15% during growing; 18.2% to 12% during finishing)	~23% for respirable	Grower–finisher; MV	Saskatchewan, Canada	–

Table 23. Cont.

Reference	Technology	Reduction Effectiveness <sup>a</sup>	Barn and Ventilation Type	Location	Main Findings
		Recirculating air filtration			
[451]	Recirculating air filter	50–60% for TSP and total culturable bacteria	Nursery; MV	United Kingdom	Sedimentation was even more impactful on dust removal.
[157]	Recirculating air with three-stage fabric filters	18–64% for inhalable 10–50% for total culturable bacteria	Grower; MV	British Columbia, Canada	Electrostatic filters had higher dust removal efficiency only in spring and winter.
	Recirculating air with electrostatic precipitators	20–66% for inhalable 20–52% for total culturable bacteria	Finisher; MV		
[93]	Recirculating air with Shaker dust collectors	33% for inhalable 41% for respirable	Farrowing; MV	Iowa	The system had no significant effects on NH <sub>3</sub> or CO <sub>2</sub> .
[100]	Recirculating air with wet scrubbers (using water or acid) for dust and gas removal	52–72% for TSP 52–67% for PM <sub>10</sub>	Finisher; MV	Germany	Reduction efficiency was greater for larger particles than smaller particles.
[101]	Recirculating air filter (MERV 5–6 pocket filter) <sup>b</sup>	8% for TSP compared to a barn w/o filter	Finisher; MV	Germany	Airborne culturable bacteria were also reduced.
[187]	Recirculating air filter (ISO 50% coarse) <sup>c</sup> with UVC light	78% for TSP 37% for total culturable bacteria	Nursery; MV	Germany	PM reduction was determined against a barn with no filter.

<sup>a</sup> The reduction effectiveness in bioaerosol is also summarized. It is noteworthy that bioaerosol reduction results from (1) physical removal of dust and (2) inactivation of biological agents. Although a mitigation technology could act on both, these two factors were not assessed separately in most previous studies. <sup>b</sup> MERV—Minimum efficiency reporting value. <sup>c</sup> ISO—International Organization for Standardization.

Ventilation is not covered in this review. Ventilation can be considered as an in-barn PM mitigation technology [436]. A higher ventilation rate brings in more fresh air, resulting in a usually lower indoor PM concentration. Ventilation system design and operation, thus, can impact PM concentrations and emissions. However, the primary function of a swine barn's ventilation system is to ensure an appropriate thermal environment for pigs. In most barns, ventilation rates are regulated based on indoor temperatures only. Furthermore, the effect of ventilation rates on indoor PM concentrations has yet to be fully understood. Although an increased ventilation rate promotes dilution, the elevated air velocity it creates could enhance the suspension and re-suspension of dust particles [96]. Ventilation also affects indoor PM concentrations via its regulation of humidity. High air humidity levels could suppress PM suspension inside animal barns [452]. The effect of ventilation systems and rates on PM characteristics can be found in previous sections. For example, a discussion about their effects on PM concentrations is available in Section 3.1.1.

It is also noteworthy that the PM reduction effectiveness derived from stationary samplers (i.e., samplers installed at a fixed location) was usually different than that of personal (exposure) samplers (Table 23). As aforementioned, personal samplers are carried by caretakers or other staff to assess their PM exposure during a work shift or shifts. Due to large spatial and temporal variability in PM generation inside swine barns, personal and stationary samplers usually read different concentrations.

### 5.1. Oil/Water Sprinkling

To our knowledge, the first attempt at sprinkling oil/water for swine barn PM control was reported by Dr. Takai at Aarhus University in Denmark [453]. It quickly attracted great interest from academia, especially in the 1990s and 2000s. To date, oil/water sprinkling has been the most studied in-barn PM mitigation method for swine and poultry barns. Oil/water sprinkling for dust control is not a new idea and it has been extensively used to reduce the emissions of fugitive dust such as road dust and mine tailings dust [454]. The working principle of the method is straightforward. Liquid droplets sprayed in the room air can wash out airborne particles—a process similar to the wet deposition of atmospheric

PM. When the liquid reaches the floors or other room surfaces, it bonds small particles together, thereby reducing the suspension and resuspension of particles [455].

Pure water was usually not used [165,438]. No specific explanation was given in the literature, but it is likely for several reasons. First, water has a higher vapor pressure so it evaporates out faster than oils. Thus, the dust suppression effect of water sprinkling does not persist long. Secondly, the viscosity of water is around two-fold lower than those of plant oils [456], meaning that water applied on the floor or other room surfaces is more prone to aerosolization (i.e., becoming particles) than oils. Thirdly, a very high moisture environment in swine barns, resulting from frequent water applications, could facilitate the growth and reproduction of microorganisms, causing animal health and odor concerns [457]. Fourthly, some organic particles are hydrophobic, with poor wettability in pure water. Wettability is a key parameter affecting the bonding (coagulation) of particles and accordingly particle suspension [458]. Due to the aforementioned reasons, oils or oil–water mixtures were commonly used.

All but one study selected plant oils (e.g., canola and soybean oils), possibly due to the oils' biological benignness and availability to pork producers. When blended with water, the percentage of oils varied greatly among studies, ranging from 5% [62] to 40% [166,443]. Surfactants (e.g., Tween 80 and Span 80) were often added to the oil–water mixture to promote emulsions and accordingly the formation of a stable, uniform mixture of water and oils [62,166,440,443,453]. However, the high material cost of surfactants could impede their use in commercial barns.

The sprinkling rate was typically measured as the mass or volume of oils applied per pig (or per m<sup>2</sup> of floor area) per day and it ranged from 3.1 to 45 mL oil pig<sup>-1</sup> day<sup>-1</sup>. Oil/water can be sprinkled at a fixed rate or variable rates. Ref. [390] compared six oil sprinkling treatments in grower–finisher barns and recommended a variable-rate treatment upon considering a trade-off between effectiveness and economics: 40 mL oil m<sup>-2</sup> day<sup>-1</sup> in the first two days, 20 mL oil m<sup>-2</sup> day<sup>-1</sup> in the next two days, and 5 mL oil m<sup>-2</sup> day<sup>-1</sup> in the remain days (an average sprinkling rate of 6.7 mL oil m<sup>-2</sup> day<sup>-1</sup>). This treatment plan was adopted by many later studies [49,83,440,442,443]. A larger sprinkling rate and a higher application frequency were found to improve dust control efficiency. However, a daily oil application rate (dosage) approaching or exceeding 20 mL m<sup>-2</sup> would result in slippery walkways [390]. A feasible way to reduce the oil application dosage while keeping dust control effective is sprinkling oil/water when pigs are active. This could be achieved by wiring a sprinkling system controller to animal activity sensors [438].

Oil/water can be sprinkled with low-pressure sprayers [123,389,390] or high-pressure pipe systems [44,62,166,438,442]. The former method is affordable, installation-free, and easy to implement. However, the application height was typically <1 m to the floor, limiting the floor area radius that each spray could cover. Due to their low operating pressures, the low-pressure sprayers were less capable of generating small, uniformly-sized droplets than high-pressure pipe systems. Several customized, high-pressure pipe systems were tested in previous oil/water sprinkling studies [44,62,437,440]. These systems each included an oil reservoir, a high-pressure pump, pipes, and nozzles. The nozzles were typically installed near the ceiling of a swine barn (at a height of 2.0–3.5 m above the floor) to maximize the coverage area of each nozzle. Only a few studies provided specific design information. Among the limited reports, the operating pressure of these high-pressure systems ranged from 241 kPa [62] to 5 MPa [438], and the median/mean diameter of droplets was 110–600 µm [44,438]. However, with no design and operation details available, it is impossible to conduct a systematic comparison of those sprinkling systems.

Oil/water sprinkling was also tested without pigs in lab chambers for suppression of PM generation from feeds, a primary source of PM in swine barns [459–461]. These studies, however, are excluded in this review as it is difficult to directly translate their findings to real-world swine barn PM control efforts.

## 5.2. Ionization

Ionization aims to charge airborne particles inside a swine barn. Due to electrostatic forces, charged particles are attracted to the barn's room surfaces or specialized collectors. Charged particles also tend to clump together, thereby precipitating out faster than uncharged particles. The use of ionization in animal barns can date back to the 1960s [462,463]. These early studies focused on the possible influence of air ions on animal production performance. To our knowledge, the first attempt at using ionization for PM control in swine barns was reported by Dr. Bundy at Iowa State University [29]. Since then, a variety of ionization techniques have been tested inside swine barns to reduce indoor PM concentrations (Table 23) or at the barns' air exhaust to mitigate PM emissions (which is beyond the scope of this review).

Technically, ionization-based PM control is based on the same principle as electrostatic precipitation (ESP). The latter has been used to control PM emissions from industrial sources (e.g., coal-fired power plants) for over a century. In reality, some researchers used the terms ionization and ESP interchangeably [464]. However, it is noteworthy that although a few studies used industrial ESP designs [465,466], most ionization systems tested in swine barns employed a simpler design than commercial ESP systems.

In an ionization system, a high DC voltage is applied across two electrodes: a charging electrode and a collection electrode. The charging electrode can be a wire, a barbed wire, a metal bar with teeth (or other types of tapered sections), etc., sharing a common feature of a small surface area and a large curvature. The purpose of this feature is to create an electric field near the electrode surface which is strong enough to ionize air molecules and/or particles. The collection electrode can be a plate, a tube, a cup, etc., with an extended surface area for particle collection. The charging and collection electrodes are often placed in parallel with small spacing in between. Particle removal by an ionization system is a four-step process: air ionization, charging, migration, and collection.

- **Air ionization.** The extremely strong electric field adjacent to the charging electrode surface dissociates air molecules into electrons and positive ions. Based on the polarity of the charging electrode, there are two types of ionization systems: positive and negative. In a positive system (Note: it is also called a positive corona system; a corona is formed due to the recombination of positive ions and electrons), the positive charging electrode attracts and scavenges electrons while repelling positive ions to space. The positive ions gain additional energy when migrating in the electric field. When the gained energy exceeds a certain level, the collision of the positive ions with other air molecules would create more positive ions. In a negative system (also known as a negative corona system), a similar process occurs but the cascade ionization process is primarily caused by electrons in the space. Most previous ionization systems in swine barns were negative systems (Table 23), likely due to their better voltage/current characteristics than positive systems [116]. However, positive systems produce less ozone than negative systems and, thus, could be a better option from the animal health perspective.
- **Charging.** Particles gain electric charges when collided by randomly moving electrons or positive ions in the space—a mechanism termed diffusion charging or diffusional charging. Particles can also become charged via field charging in which the collision is driven by the electromigration of electrons or ions in the electric field. The charging process can occur repeatedly until the charges on a particle become saturated. Particles would eventually carry positive charges in a positive ionization system and negative charges in a negative system.
- **Migration.** The electric field drives charged particles to migrate towards an electrode with the opposite polarity. In both positive and negative ionization systems, particles migrate towards the collection electrode. The final migration velocity of a charged particle, which is often referred to as a drift velocity, is determined by a balance between the electrostatic force and the air friction exerted on the particle. The drift

velocity (similar to the terminal settling velocity for gravitational settling) is a key design parameter for ionization systems [116].

- Collection. Once a particle reaches a collection electrode, it transfers part of the charges it carries to the electrode. The remaining charges make the particle attach to the electrode surface due to an electrostatic force. Particles accumulated on the surface of collection electrodes can be mechanically removed after a certain period.

Most previous swine barn PM studies used commercial ionization systems, with no design specifications available. Thus, it is difficult to make a comprehensive comparison of different systems. In general, these ionization systems use only a single section of paired electrodes (Note: commercial ESP systems have multiple sections in the direction of airflow to achieve an exceptionally high PM removal efficiency), with larger spacing between electrodes than that in commercial ESP systems. As a result, those systems have a generally lower PM mitigation efficiency than ESP systems; and particle deposition occurs on the surfaces of not only collection electrodes but also other objects inside a swine barn.

An ionization system consumes no water, oil, or chemicals during operation. Another possible advantage is its low power consumption—due to the high resistivity of air, the current in an ionization system is expected to be very low (milliamperes). Only a few previous studies reported current consumption data [444,448]. However, no further normalization of the data with information such as the number/weight of pigs or barn areas was reported, making it difficult to compare energy consumption between different systems.

As aforementioned, drift velocity is a key parameter for ionization system design. It is related to not only electric field strength, but also particle characteristics such as particle size and dielectric constant. Ref. [467] measured the properties of organic and respirable particles in swine barns, including particle size and resistivity. The acquired data, however, has yet to be effectively utilized.

### 5.3. Alternation of Feed and Feeders

Feed is a primary source of PM in swine barns. Thus, any measures that suppress the suspension of feed particles can help reduce indoor PM concentrations. These measures include pelletization of feed, the use of wet feeders, and the modification of feed diet. An early review can be found in Ref. [452].

Pelleted feed was reported to result in lower PM concentrations than meal (mash or ground) feed [30,449,468], for two possible reasons. First, due to its large granular size and physical integrity, pelleted feed is less likely to disaggregate and suspend as airborne particles. Secondly, the pelleted feed was found to increase feed efficiency and cause less feed spillage in feeders [469], which in turn could reduce the suspension of feed particles. Although early studies all indicated the benefit of pelleted feed, a recent study in nursery barns found that pelleted feed resulted in higher indoor PM concentrations than coarsely ground meal [470]. The authors ascribed this to the formation of small particles during feed pelleting.

Wet feeders amend dry feed (either in the form of pellets or powders) with water. The high moisture content of feed in theory would suppress the suspension of feed particles [29]. The benefit of wet feeders for swine barn PM management was first identified by several independent studies in Europe [452] where wet feeders remain prevalent as of today. A similar benefit was observed by Ref. [96] from a multi-farm monitoring project in the U.S. Midwest; however, according to the authors, swine barn types could confound the observed difference between dry and wet feeders. The effectiveness of wet feeders for PM mitigation was challenged by Ref. [468] who found that the barns with wet feeders were among the dustiest of surveyed confined swine farms. In two independent studies, no significant difference in PM concentrations was noted between dry and wet feeders [438,471]. Such inconsistency in the literature could be related to changes in dry feeder designs [472].

Modification of feed diet aims to increase the physical integrity of feed particles while maintaining (if not improving) pig performance. The most studied diet modification is adding fat or oil to swine feed. Although adding fat as an energy source to swine feed

has been practiced for >60 years, the first study of its mitigation of swine barn PM was reported by Ref. [376]. Dietary fat was thought to serve as a binder agent to bind small particles together during feed storage, delivery, and feeding, thereby reducing the potency of dust suspension. By adding 2.5%, 5.0%, and 7.5% of tallow (an animal fat) to the diet of grower–finisher pigs, the average indoor PM concentrations were reduced by 21%, 50%, and 56%, respectively [202,376]. Vegetable oils were found to function similarly to animal fats. Ref. [473] added 5% soybean oil to nursery swine diets and found a reduction in settled dust of 45–47% and a reduction in total viable bacteria of 27%. Similar observations were made by [474,475]. Since 1990, a few follow-up studies have been conducted, with a consistent reduction in TSP or inhalable particles observed (Table 23). For respirable particles, not every effort was successful. Ref. [122] reported a 46% increase in respirable particle counts after oil treatment. They suggested that the interception and scavenging of fine particles (e.g., respirable) by coarse particles (e.g., inhalable) could play a role in the settling removal of fine particles and, thus, a decreased inhalable particle concentration upon oil treatment could discourage respirable particle removal. Other than oil and fat, the feed additives tested include lecithin [474] and lignin [449], both resulting in a significant PM reduction. Molasses was proposed by Ref. [452] as a feed additive for in-barn PM mitigation; however, it has yet to be tested.

The purpose of feed diet modification is multifaceted. For air quality management, most previous studies focused on odor mitigation. PM mitigation was often investigated as a side-benefit. A comprehensive review of the effect of feed diets on odor is given by Ref. [476]. In principle, any diet modification can be tested for PM concentrations. In addition to feed additives, reducing the dietary crude protein content was reported to reduce PM concentrations inside grower–finisher barns [442]. The reason is unknown. Reduced crude proteins were believed to decrease total nitrogen excretion from pigs, thereby reducing the generation and emissions of ammonia and odors [477].

In summary, although most previous studies offered positive results, no agreement has been reached regarding the effectiveness of pelleted feed or wet feeders for PM mitigation. Adding fat/oil to feed was effective, particularly for large particles. Compared to oil/water sprinkling and ionization, feed and feeder alternation receives less research interest, especially in recent years. This is likely because any alternation of feed and/or feeders could significantly impact pig performance. Swine nutrition and physiology carry greater weight than air quality when it comes to feeds and feeders.

#### 5.4. Recirculating Air Filtration

Recirculating air filtration, also known as internal air filtration or indoor air purification or air cleaning, features a recirculated air loop that feeds PM-laden air inside a built environment to an air cleaner and releases clean air back to the environment. The concept of recirculating air filtration can date back to the early 20th century [478] when multiple studies examined its effectiveness in temperature and odor control. To our knowledge, the idea of using recirculating air filtration for animal environmental control was first proposed by Ref. [479] and the first study of its effectiveness in swine barn PM reduction was reported by Ref. [201]. Since 1990, six relevant studies have been reported. Five of them used fabric filters, one used ESP systems, and one used wet scrubbers for in-barn PM removal (Table 23).

PM removal by fabric filters is governed by three mechanisms: impaction, interception, and diffusion [116]. Impaction occurs when a particle strikes a filter fiber in a face-to-face “direct hit” fashion. In comparison, interception can be considered as a “side slam”—the moving trajectory of a particle stays off a fiber but due to the definite size of the particle, it still strikes the fiber. Diffusion occurs when a small particle would miss the fiber but strikes it due to Brownian motion. It is noteworthy that the high efficiency of fabric filters is largely ascribed to dust layers built up on the fabric which fill interstitial holes. The three mechanisms apply to the “dirty” fabric filters as well.

PM removal by wet scrubbers is primarily contributed by the impaction of particles onto liquid droplets. In a wet scrubber, a liquid (usually water) is pressurized to form fine liquid droplets using nozzles, venturi, or venturi jets so as to increase the contact of particles with the liquid. Various wet scrubbers designs are available, including spray-chamber, cyclone spray-chamber, wet-impingement, and venturi scrubbers. The one adopted by Ref. [100] is a typical horizontal spray-chamber scrubber, with a entrainment eliminator installed downstream to remove liquid droplets; a sulfuric acid solution was selected as the liquid for simultaneous PM and NH<sub>3</sub> removal.

Detailed information about fabric filters, ESP systems (Refer to Section 5.2), and wet scrubbers can be found from Ref. [116]. In general, fabric filters offer a greater PM removal efficiency but create a higher pressure drop than ESP systems and wet scrubbers. A higher pressure drop means that additional energy is needed to recirculate and clean a given volume of barn air.

In reality, several fabric filters tested in swine barns held only a moderate filtration efficiency. No reason was given regarding filter selection; it was likely due to energy consideration. For example, an ISO 50% coarse filter [187] is <50% effective in PM<sub>10</sub> removal; and an MERV 5–6 pocket filter [101] has a removal efficiency of 20–49% for particles with diameters of 3–10 µm. High-efficiency fabric filters or filter sets were also tested. Ref. [157] used a three-stage filter set, with two coarse filters for pre-filtration and a high-efficiency particulate air (HEPA) filter as the final stage. A commercial shaker dust collector (Model: SDC-140) was tested by Ref. [93]. The collector consisted of high-efficiency fabric bags, with >99% efficiency for general industrial dust.

It is important to note that filtration or scrubbing efficiency is different from overall in-barn PM reduction. The latter is related to not only the efficiency of recirculating air filters but also barn volume, recirculating airflow, in-barn PM generation, PM sedimentation, ventilation, infiltration, exfiltration, etc. A classic indoor air quality box model considering recirculated air can be found from Ref. [480]. The model is based on material balance and can be used to determine the recirculating airflow required for target PM reduction. However, many of the model parameters (e.g., ventilation rates) were unavailable in the previous swine barn studies, making it difficult to conduct a thorough analysis of system performance and a comparison among different filtration systems.

## 6. Knowledge Gaps and Future Research Needs

Decades of research has profoundly advanced our knowledge of PM in swine barns, including sources, characteristics, measurement, and mitigation. However, PM continues to be an environmental challenge facing pork producers. Some fundamental questions remain unanswered, calling for future research input. Detailed information can be found in the previous sections. This section lists several major knowledge gaps identified from the review effort.

- Little is known about the concentration and size distribution of submicron particles (i.e., PM with diameters < 1 µm) in swine barns. These particles are of growing concern, as exemplified by the PM<sub>1</sub> standard being rigorously discussed by the U.S. EPA and the European Union. Due to their ultra-small size, submicron particles can enter alveoli and be directly absorbed into pulmonary cells. No PM sources in swine barns have been known to emit a significant amount of submicron particles. However, this may be ascribed to the lack of monitoring data. Furthermore, submicron particles can be associated with airborne porcine viruses. As mentioned in Section 3.2.2, some porcine viruses could occur as individual airborne particles, with diameters <1 µm. This size fraction of viruses is hard to remove with regular PM mitigation technologies. Thus, the measurement of submicron PM in swine barns can improve our understanding of airborne viral transmission in and around infected barns and aid the development of mitigation technologies.
- An updated model to simulate in-barn PM concentrations is lacking. Previous studies have investigated the effect of various environmental and operational factors such as

barn type, ventilation, outdoor temperature, and stock density. Based on the investigation results, semi-mechanistic or statistical models were developed to predict in-barn PM concentrations from these easily measurable factors [11,20,325,481]. However, no further modeling effort was seen in the past decade. In the meantime, the increasing use of farm information systems and real-time PM monitors have generated massive data. A thorough analysis of the big data is needed to further assess the effect of individual factors. A more robust, accurate, and comprehensive model is anticipated, with the data and advanced data analytics tools (e.g., deep learning). Knowledge of PM concentrations is critical as many air hazards (e.g., odor levels and pathogen concentrations) in swine barns can correlate with PM concentrations.

- Additional research is needed to further investigate the spatial distribution of PM in swine barns to better assess PM exposure risks for human and pigs. Previous investigations predominantly focused on PM concentrations. Little is known about the distribution of other PM characteristics such as PSD, bacterial/fungal counts, viruses, and microbial composition. Microbial composition may be a good subject to start with for its strong implications for human and animal health. The investigation can share the same filters as PM concentration monitoring and relevant analysis (e.g., PCR and DNA sequencing) is becoming increasingly affordable. In addition to grid sampling (adopted by most of the previous investigations), future research may consider sampling PM in different function or risk zones inside swine barns.
- Limited information is available about the size distribution of some key bioaerosols. These include ARGs, viruses, and size-segregated microbial composition. Special attention should be paid to zoonotic pathogens, including their concentrations and viability in different size fractions. Size is a critical factor affecting the transport and fate of bioaerosols. Pathogens and allergens can bind to large particles or occur as individual particles in the air. Knowledge of their size distribution is essential for the development of cost-effective mitigation or prevention strategies. Another factor related to size is the mixing state of bioaerosols, i.e., the way of microbes mixed with abiotic components in a particle; and it remains largely unexplored. The mixing state is expected to have a substantial influence on the viability and infectivity of bioaerosols. Bioaerosol size and mixing state can be measured using microscopy coupled with microbial identification technologies and/or size segregated sampling.
- For the U.S. pork industry, updated information is needed regarding bioaerosols in swine barns, especially culturable bacterial/fungal counts, total bacterial/fungal counts, microbial composition, and ARGs. Most field research projects were conducted over ten years ago. Their results may not represent the current status due to the continual consolidation of pork production and changes in swine nutrition and veterinary practices. The past ten years also witnessed the rapid development and deployment of molecular biology technologies. These technologies are becoming increasingly powerful, accessible, and affordable. A revisit to swine barn bioaerosols with the new technologies can profound our understandings of bioaerosols in various aspects: concentration, composition, viability, health impacts, transport, mitigation, etc. Furthermore, advances in bioinformatics have made it possible to unravel the interrelations between different bioaerosols (e.g., ARGs and bacterial composition) and between bioaerosols and abiotic PM components (e.g., PM-borne odorants). All these advancements together provide an unprecedented opportunity to address pork production-associated bioaerosol issues. In addition to their direct health implication, bioaerosols (as well as several other PM components (e.g., EC)) were found to create reactive oxygen species upon inhalation to the human and animal lung [482,483]. The oxidative potential—as a collective measure of PM toxicity—of swine barn PM should be assessed in future studies.
- Mathematical modeling is needed to assess and understand the role of PM in causing community odor nuisances. The previous studies have identified and qualified numerous odorants in PM. The data, however, have yet to be effectively utilized for

odor assessment. A conceptual exposure model is needed to describe the pathways for PM-borne odors to transport from swine barns to a neighboring community. For each pathway, environmental transport modeling (e.g., air dispersion modeling) needs to be conducted to predict the occurrence and concentration of odors at the neighboring community. Odor exposure routes (via inhalation) should also be modeled, thereby completing the modeling framework. It is noteworthy that the previous modeling work on PM-borne odors focused on in-barn transport and exposure, and requires significant revisions for community odor nuisance simulation.

- Additional research is needed for source apportionment of PM and bioaerosols. Most PM source apportionment research projects were conducted over ten years ago. In the U.S., no relevant studies have been reported since 1990. PM source apportionment is critical, as it enables cost-effective control of in-barn PM by addressing major sources. As mentioned in Section 3.4, PM source apportionment can be conducted through microscopic analysis of individual particles or receptor modeling. The latter requires the measurement of PM chemical composition (which has been conducted by several studies). The source information of bioaerosols is equally important but remains largely unexplored. Similarly, both microscopic analysis and receptor modeling can be useful for bioaerosol source apportionment. The latter may be conducted with the microbial composition information derived from high-throughput DNA sequencing.
- The performance of low-cost PM sensors has yet to be assessed in swine barns. Numerous low-cost PM sensors have become available in the past few years. They constitute a central component of affordable handheld or wearable air quality meters. Although their extensive use in swine barns has yet to come, these sensors may create numerous challenges and opportunities. Regarding challenges, the meters may be used by non-farm groups to survey air quality around swine farms. Regarding opportunities, the meters may offer a less expensive and more convenient approach for in-barn air quality monitoring and personal exposure assessment. The low-cost sensors may also help to achieve the precision management of in-barn PM—mitigation (e.g., oil sprinkling) is more cost-efficient when PM concentrations are available as a control input. Efforts should be made to test whether the sensors perform properly for swine barn PM and to identify the best-performing sensors if possible.
- Innovative in-barn PM mitigation solutions are needed. Nearly all existing mitigation solutions fall into four technological categories, as summarized in Section 5. Their history all dates back to the 1970s or 1980s. Decades of research has generated massive data. However, none of them have been widely used in commercial barns. Further development of these conventional technologies is necessary. Meanwhile, efforts should be encouraged to develop innovative mitigation technologies or solutions. PM mitigation is anticipated to be part of an integrated precision swine farming system, thereby enabling the “smart” control of PM generation, concentrations, and emissions in and from swine barns. The effectiveness of conventional and innovative technologies in bioaerosol (e.g., PRRSV and *S. aureus*) reduction should receive particular attention due to the health implications of bioaerosols.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/atmos13030490/s1>, Table S1: List of odorants identified in swine barn PM (1979-Present). Refs. are [36,146,277,354,355,357,363–368] cite in the supplementary materials.

**Author Contributions:** Conceptualization, X.Y.; methodology, X.Y. and D.A.; software, X.Y. and A.O.; validation, X.Y., N.H. and D.A.; formal analysis, X.Y., N.H., A.O. and Z.C.; investigation, X.Y., N.H., A.O. and D.A.; resources, X.Y.; data curation, X.Y., N.H., A.O., Z.C. and K.L.A.; writing—X.Y., N.H. and A.O.; writing—review and editing, X.Y., N.H., A.O., K.L.A. and D.A.; visualization, X.Y.; supervision, X.Y.; project administration, X.Y.; funding acquisition, X.Y. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was sponsored by the National Pork Board (Grant No. 20-140). The authors also wish to thank the support of the USDA NIFA Hatch (SD00H709-20) and Multistate Hatch Projects (SD00R690-20) through the South Dakota Agricultural Experimental Station.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest. The discussion and statements made in the review only represent the authors' opinions and should not be taken as the viewpoints of the National Pork Board or South Dakota State University.

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