



Article Acaricidal Activity of Tea Tree and Lemon Oil Nanoemulsions against Rhipicephalus annulatus

Samar M. Ibrahium ^{1,*}, Ahmed A. Wahba ², Ahmed A. Farghali ³, Abdel-Azeem S. Abdel-Baki ⁴, Shaimaa A. A. Mohamed ⁵, Saleh Al-Quraishy ⁶, Ahmed O. Hassan ⁷ and Shawky M. Aboelhadid ^{8,*}

- ¹ Department of Parasitology, Animal Health Research Institute, Fayum Branch 16101, Egypt
- ² Department of Parasitology, Animal Health Research Institute, Dokki Branch 12611, Egypt
- ³ Materials Science and Nanotechnology Department, Faculty of Postgraduate Studies for Advanced Sciences, Beni-Suef University, Beni-Suef 62511, Egypt
- ⁴ Zoology Department, Faculty of Science, Beni-Suef University, Beni-Suef 62511, Egypt
- ⁵ Health Administration, Egyptian Ministry of Health, Beni-Suef 62511, Egypt
- ⁶ Zoology Department, College of Science, King Saud University, Riyadh 12372, Saudi Arabia
- ⁷ Department of Medicine, Washington University School of Medicine, St. Louis, MO 63110, USA
- ⁸ Parasitology Department, Faculty of Veterinary Medicine, Beni-Suef University, Beni-Suef 62511, Egypt
- * Correspondence: drsamarmahmoud333@yahoo.com (S.M.I.); shawky.abohadid@vet.bsu.edu.eg (S.M.A.)

Abstract: Tick infestation is a serious problem in many countries since it has an impact on the health of animals used for food production and pets, and frequently affects humans. Therefore, the present study aimed to investigate the acaricidal effects of nanoemulsions of essential oils of Melaleuca alternifolia (tea tree, TT) and Citrus limon (lemon oil, CL) against the different stages (adult, eggs, and larvae) of deltamethrin-resistant Rhipicephalus annulatus ticks. Three forms of these oils were tested: pure oils, nanoemulsions, and a binary combination. Tea tree and lemon oil nanoemulsions were prepared, and their properties were assessed using a zeta droplet size measurement and a UV-Vis spectrophotometer. The results showed that TT and CL exhibited higher adulticidal effects in their pure forms than in their nanoemulsion forms, as demonstrated by the lower concentrations required to achieve LC_{50} (2.05 and 1.26%, vs. 12.8 and 11.4%, respectively) and LC_{90} (4.01% and 2.62%, vs. 20.8 and 19.9%, respectively). Significant larvicidal activity was induced by the TTCL combination, and LC₅₀ was reached at a lower concentration (0.79%) than that required for the pure and nanoemulsion forms. The use of pure CL oil was found to have the most effective ovicidal effects. In conclusion, pure TT and CL have potent acaricidal effects against phenotypically resistant R. annulatus isolates. It is interesting that the activity levels of TT and CL EOs' binary and nanoemulsion forms were lower than those of their individual pure forms.

Keywords: Melaleuca alternifolia; Citrus limon; essential oil; tick; resistant

1. Introduction

Rhipicephalus annulatus is an obligate blood-feeding ectoparasite of cattle that is widely distributed in tropical and subtropical regions [1]. Tick infestations of cattle can harm hides and reduce production. In addition, they have the ability to transmit pathogens, including *Babesia* species and *Anaplasma marginale* [2].

Chemical acaricides are frequently used to control ticks. Pyrethroid acaricides are commonly used to control tick populations. There are serious environmental and public health issues associated with its extensive use. Deltamethrin, the most common pyrethroid, can bind to sodium channels and cause hyperexcitability in neurons and tick death [3,4]. Deltamethrin has an adverse effect on humans, animals, and the environment [5]. Due to its overuse, the *R. annulatus* tick has evolved phenotypic and genotypic resistance to deltamethrin [1,6]. It is, therefore, necessary to search for alternative products or strategies for tick control. Essential oils represent prospective substitutes to the acaricides now in use,



Citation: Ibrahium, S.M.; Wahba, A.A.; Farghali, A.A.; Abdel-Baki, A.-A.S.; Mohamed, S.A.A.; Al-Quraishy, S.; Hassan, A.O.; Aboelhadid, S.M. Acaricidal Activity of Tea Tree and Lemon Oil Nanoemulsions against *Rhipicephalus annulatus*. *Pathogens* **2022**, *11*, 1506. https://doi.org/10.3390/ pathogens11121506

Academic Editors: Craig A. Miller and Mason V. Reichard

Received: 30 September 2022 Accepted: 6 December 2022 Published: 9 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). since various studies have shown that several plants have toxicological effects on ticks [7]. The current use of essential oils is safe for humans, animals, and the environment, and the development of resistance is uncommon [8,9]. The mechanisms of action of EOs include many target locations, such as cell wall disintegration, membrane protein obliteration, and increased cell wall permeability [10].

Melaleuca alterniolia (tea tree oil) (TT) has an acaricidal effect on nymphs of *Ixodes ricinus* [11]. TT and its nanostructured form cause a mortality rate of 70%, and a 100% reduction in the egg laying and hatchability phases of female *Rhipicephalus microplus* ticks [12]. The application of *Citrus limon* (Lemon oil) (CL) was proven to induce a mortality rate of 98.68% in the mealy bug *Dysmicoccus brevipes*, with the LC₅₀ and LC₉₅ attained at concentrations of 0.72% and 2.91%, respectively [13]. Additionally, CL was used in rabbits to treat mange, the most severe ectoparasitic disease caused by infestation by the mite *Sarcoptes scabiei* with a 100% mortality rate in mites and complete eradication of the disease's clinical manifestations [14]. Additionally, CL showed significant acaricidal activity against engorged *Hyalomma dromedarii* females [15].

The low solubility in the aqueous phase, high volatility, and low long-term stability are limiting factors for the use of EOs as natural acaricides. A promising solution is the development of essential oil nanoemulsions, which are prepared by the mixing of two immiscible liquids (oil and water) to form a single phase by means of an emulsifying agent, also known as a surfactant. The resultant nanoemulsion typically has droplet sizes between 20 and 200 nm [16]. Recently, it was found that nanocapsules and nanoemulsions containing cinnamon oil have significant activity against engorged *R*. (*B.*) *microplus* females [17,18]. Additionally, Boito et al. [19] reported that *tea tree* nanoemulsions interfere with the reproduction of *R*. (*B.*) *microplus*. Ibrahium et al. [20] found that geranium nanoemulsions were much more effective against *R. annulatus* than its pure form.

In this study, we aimed to assess the effectiveness of TT and CL nanoemulsions compared with their pure forms and binary combination against the deltamethrin phenotypicresistant tick *Rhipicephalus annulatus*.

2. Materials and Methods

2.1. Ethical Approval

These experiments were conducted in accordance with the ethics of the Faculty of Veterinary Medicine, Beni-Suef University, Egypt, under approval number (021-172).

2.2. Source of Compounds of the Study

M. alternifolia (*tea tree* oil) (TT) and *C. limon* (CL) were purchased from a commercial source (Trust Scientific for Natural Products, Cairo, Egypt). The oils were dissolved in 70% ethanol at rates of 10, 5, 2.5, 1.25, and 0.625% (volume/volume). Binary mixtures from these oils were added at a rate of (1:1) for all five concentrations. This step was performed to investigate the synergistic/antagonistic activity between essential oil binary mixtures, and the synergistic factor (SF) was calculated according to Suwannayod et al. [21].

$$SF = \frac{LC50 \text{ for the oil alone}}{LC50 \text{ for the combination}}$$

SF > 1, indicates synergism; SF < 1, indicates antagonism.

2.3. GC-MS of TT and CL Oil

The GC-MS analysis was carried out using a gas chromatography–mass spectrometry instrument (TRACE GC Ultra Gas Chromatographs (THERMO Scientific Corp., Santa Clara, CA, USA). The chemical constituents of the essential oils were identified using AMDIS software (www.amdis.net) as well as by assessing their retention indices (relative to n-alkanes C8-C22), matching their mass spectra to authentic standards and consulting the Wiley Spectral Library collection and the NSIT Library database (Nawah Scientific Educational Research Center, Cairo, Egypt).

2.4. Nanoemulsion Preparation of TT and CL

TT and CL nanoemulsions were formulated using three components in accordance with Nirmala et al. [22]: TT or CL oil, Tween 80, and distilled water. The nanoemulsion formulation process was a two-step process, initiated with preparations of oil/water macroemulsions by blending TT or CL oil, surfactant (Tween 80), and water, at a specific concentration of one oil to two T80 using a magnetic stirrer at a speed of 500 rpm for 10 min. The TT or CL oil concentration was 10%. Then, the emulsions were prepared by forming the respective nanoemulsions using an ultrasonicator with a 750 W input power processor (Branson Probe sonicator-Advanced model, 20 kHz, Swedesboro, NJ, USA). Each concentration was subjected to different sonication time periods of 5, 10, and 15 min, respectively. The turbulence produced by the high-energy shockwaves disrupted the droplets. The mild heat generated was regulated by positioning the sample in a container filled with ice.

2.5. Characterization of Nanoemulsion

The absorbance of the emulsions was measured at 270–345 nm with the help of a UV-visible spectrophotometer (UV-2600, Shimadz, Japan). The zeta potential, droplet size distribution (d, nm) (analysis by volume) and polydispersity index (PDI) of the nanoemulsions were measured by a zeta sizer apparatus (dynamic light scattering technique) (Nano-ZS90, Malvern, UK). Prior to the experiment, all samples were diluted to 10% with deionized water in order to reduce the effects caused by multiple scattering effects.

2.6. Acaricide Activity of the Prepared Materials

2.6.1. Preparation of Ticks

Female *R. annulatus* ticks engorged by blood were collected from naturally infested cattle from El-Fayoum Governorate, Egypt (85 km southwest of Cairo). The ticks were collected from cattle with a history of recurrent tick infestation that had not received any treatment for four weeks. The collected ticks were transported to the Parasitology Laboratory, Faculty of Veterinary Medicine, Beni-Suef University. They were identified in accordance with Estrada-Peña [23]. The ticks were divided into Petri dishes containing 10 ticks each, for the adult immersion bioassay. A second group of collected ticks was incubated in a BOD incubator and allowed to oviposit. Eggs were collected, mixed, weighed, and then allocated to test tubes in 50 mg lots for the ovicidal activity test. Some of the allotted eggs were then incubated until hatching. The hatched larvae were used for the larval packet test.

2.6.2. Adult Immersion Test (AIT)

The prepared concentrations of EOs (TT and CL), nanoemulsions (TTN and CLN), and their combinations (TTCL) were tested at 5 different concentrations against adult engorged female ticks. The ticks were immersed in petri dishes containing 10 mL samples of the tested solutions for 2 min and dried with filter paper. Then, they were incubated in a BOD incubator (26–28 °C and 80% relative humidity) [24]. For each concentration, 5 replicates (10 ticks for each replicate) were used. The negative control was ticks treated with 70% ethyl alcohol. The positive controls were: one treated with 2 mL/L deltamethrin (Butox®, CAS: 52918-63-5) (pyrethroid insecticide as sodium channel modulator), and the other one treated with 0.5 mL/L Phoxim 50% (Sebacil®, CAS: 14816-18-3) (organophosphate insecticide as an acetylcholinesterase inhibitor). The number of dead adult ticks were counted after 14 days, and the egg production index (EPI) was reported for live ticks [25]. The egg production index (EPI) was calculated as follows: EPI = weight of egg mass/initial weight of engorged females \times 100.

2.6.3. Ovicidal Activity

Fifty micrograms of tick eggs were placed on filter paper that was 4×7 cm in size. Then, 1 mL of the tested concentration was applied to the eggs in plastic mesh for 2 min.

Then, the eggs were dried on filter paper and incubated in glass tubes closed with cotton [24]. The control eggs were treated with 70% ethanol, 50% phoxim, and 5% deltamethrin. The eggs from the treated groups were incubated in a BOD incubator until hatching. For each concentration, three replicates were used.

2.6.4. Larvicidal Activity

A modified larval packet technique (LPT) described by [26,27] was used. Using a paint brush, approximately one hundred larvae were placed on the center of 7×7 cm filter papers. Then, 100 µL samples of the test solutions were added. The papers were then enclosed to form packets. Control group samples were treated with ethanol (70%). For each concentration, 5 replicates were used. The treated packets were investigated after 24 h to record mortality rates.

2.6.5. Statistical Analysis

The results of the different treatments were statistically analyzed using IBM SPSS for Windows, v.22 (IBM, Armonk, NY, USA). The difference between treatments was compared using ANOVA, and subsequent Duncan's tests were used to estimate the differences between means ($\alpha = 0.05$). The lethal concentrations and 50%, and 90% mortalities were calculated with SPSS v.22.

3. Results

3.1. GC-Mas Analysis of Tea Tree and Citrus Limon Oils

Terpinene (52.24%), dihydro- α -terpineol (5.97), and diterpene (2.87%) are the main constituents of TT (Table 1). Meanwhile, (L)-alpha-terpineol (18.32%), alpha-terpinol (13.43%), trans-4-thujanol (9.64%), α - terpinolene (5.81%), citral propylene glycol acetal (5.73%), geranial propylene glycol acetal (4.00%), and α -terpineol acetate (3.60%) are the major components of CL (Table 1).

RT	Compounds in TT	Area%	RT	Compounds in CL	Area%
3.30	Terpinene	52.24	4.54	trans-4-Thujanol	9.64
5.10	Dihydro-α-terpineol	5.97	4.87	alpha-terpineol	13.43
5.60	Diterpene	2.87	5.37	beta-Fenchyl alcohol	0.68
16.57	4-methyl-3-cyclohexen-1-yl	5.42	5.95	α- Terpinolene	5.81
24.02	Pentatriacontane	2.91	6.14	Dipropylene glycol	0.27
24.45	1-Octatriacontanol, pentafluoropropionate	4.20	6.61	Camphene hydrate	0.58
25.46	N-pentatriacontane	3.8	6.99	1-Terpinenol	0.56
25.86	17-Pentatriacontene	3.93	7.24	Menthol terpine hydrate	1.70
26.35	Octatriacontyl 2,2,3,3,3-pentafluoropropanoate	2.55	8.34	(L)-alpha-Terpineol	18.32
26.46	Tetrapentacontane, 1,54-dibromo-	3.31	9.16	cis-Citral	1.57
26.84	1,54-dibromotetrapentacontane	4.58	9.36	Linalool acetate	0.63
27.21	Octatriacontyl pentafluoropropionate	4.88	9.79	β-Geranial	2.21
28.16	Nonyl tetracosyl ether	3.35	11.4	α-Terpineol acetate	3.6
			12.74	Diphenyl ether	24.56
		-	13.53	Widdrol hydroxyether	3.96
	T ()	-	13.65	methyl ester	2.76
	lotal	100	14.18	Citral propylene glycol acetal	5.73
		-	14.32	Geranial propylene glycol acetal	4.00
		-		Total	100

Table 1. GC-Mas of tea tree (TT) and Citrus limon (CL) oils.

3.2. Characterization of Oil Nanoemulsions

The UV-Vis spectrophotometer detected the absorbance peaks of CLN and TTN at 345 nm and 270 nm, respectively (Figure 1). Zeta potential measurements revealed negative charges on the outer surface of TT N (-2.02 mV) and CLN (-2.30 mV) and the hydrodynamic particle sizes of TTN and CLN with mean droplet sizes of approximately 188.4 d.nm and 50.66 d.nm and PDI values of 0.527 and 0.171, respectively (Figures 2 and 3). The low PDI values reflect the homogeneous size distribution of the EO droplets (Table 2).



Figure 1. UV-vis spectrophotometer absorbance of tea tree (TT) and limon (CL) oils.



Figure 2. Size distribution of *tea tree* (TT) and lemon oils (CL) by intensity using Zeta apparatus.



Figure 3. Zeta potential distribution of tea tree (TT) and lemon (CL) oils by Zeta apparatus.

 Table 2. Essential oil nanoemulsion characteristics.

Eos	TT	CL				
The absorbance peak by UV-vis (nm)	270	345				
Droplet size (d.nm)	188	50.66				
PDI (d.nm)	0.527	0.171				
Potential charges	-2.02	-2.30				
DI – polydispersity index						

PDI = polydispersity index.

3.3. Adulticidal Activity of Oils against R. Annulatus Tick

The adulticidal activities of TT and CL against *R* annulatus ticks were significant (p < 0.05), especially at concentrations of 5 and 10%, where they induced mortality rates of 96.60 and 100%, respectively (Table 3). No deaths were reported in the deltamethrin (2 μ L/mL) treated ticks. Meanwhile, 100% mortality was observed in the Phoxim-treated ticks (Table 3).

Concentrations %	10	5	2.50	1.25	0.625
TT (DF %)	100.0 ± 00.00 *	96.66 \pm 1.527 *	56.33 ± 8.020 *	41.00 ± 5.291 *	23.66 ± 3.785 *
TTN (DF %)	33.66 ± 2.081 *	17.00 ± 1.000	$14.66 \pm 0.1.527$	5.666 ± 2.516	2.666 ± 1.527
CL (DF %)	100.0 ± 0.000 *	100.0 ± 0.000 *	80.66 ± 2.081 *	63.00 ± 4.358 *	36.33 ± 5.131 *
CLN (DF %)	37.33 ± 3.055	22.00 ± 3.000	13.33 ± 1.527	9.666 ± 0.577	3.666 ± 1.154
TTCL (DF %)	76.33 \pm 0.577 *	53.00 ± 1.000 *	43.33 ± 2.081 *	35.66 ± 2.516 *	24.00 ± 2.645 *
Control (Deltamethrin 2 μL/mL)			0.000 ± 0.000		
Control (Phoxim 0.50 μL /mL)			100.0 ± 0.000		
Control (Ethyl alcohol 70%)			0.000 ± 0.000		

Table 3. Adulticidal activity of different forms of *tea tree* (TT) and *citrus lemon* (CL) oils on *R. annulatus* (the ratio between oils binary mixture was 1:1) at 14 days post application.

(*) significant compared with the control (Ethyl alcohol) (p < 0.05), DF % (female mortality %), TT (*tea tree* oil), TTN (*tea tree* nanoemulsion), CL (*citrus lemon* oil), CLN (*citrus lemon* nanoemulsion), TTCL (*tea tree* oil + Citrus limon oil).

The lowest dosages of TT and CL exhibited lower egg production indices (EPI) than control ticks (treated with ethyl alcohol 70%), whereas TTN and CLN were associated with similar EPI values to the control ticks (Table 4). The LC_{50} values for the pure TT and CL oils were 2.05 and 1.26%, respectively, whereas they were 12.3 and 11.4%, respectively, for the nanoemulsion forms (Figure 4). Furthermore, the corresponding LC_{90} values for TT and CL were 4.00 and 2.62%, but they were 20.8% and 19.9% for TTN and CLN.

Table 4. Egg production index (EPI) of different forms of TT and CL against *R.* (*B.*) *annulatus* at 14 days post application.

Concentrations %	10	5	2.50	1.25	0.625
TT (EPI %)	0.000 ± 0.000 *	3.163 ± 0.233 *	23.50 ± 1.600	25.10 ± 1.110	30.40 ± 0.088
TTN (EPI %)	23.16 ± 1.813	36.06 ± 2.674	43.72 ± 1.041	55.01 ± 2.602	65.33 ± 0.770
CL (EPI %)	0.000 ± 0.000 *	0.000 ± 0.000 *	8.263 ± 3.209 *	21.93 ± 0.474 *	28.09 ± 2.404
CLN (EPI %)	25.09 ± 1.127	30.39 ± 0.078	33.62 ± 3.298	47.48 ± 0.893	53.99 ± 1.923
TTCL (EPI %)	21.59 ± 0.923 *	24.49 ± 0.823	28.08 ± 2.055	30.25 ± 2.463	34.34 ± 1.006
Control (Deltamethrin 2 µL/1mL)			59.34 ± 7.987		
Control (Phoxim 0.50 µL/1mL)			0.000 ± 0.000		
Control (Ethyl alcohol 70%)			64.43 ± 7.623		

(*) significant compared with the control (Ethyl alcohol) (p < 0.05), DF % (female mortality %), TT (*tea tree* oil), TTN (*tea tree* nanoemulsion), CL (*citrus lemon* oil), CLN (*citrus lemon* nanoemulsion), TTCL (*tea tree* oil + Citrus limon oil).



Figure 4. LC₅₀ and LC ₉₀ of *tea tree* and *citrus lemon* oils against (**A**) adult of *R. annulatus*, and (**B**) larvae of *R. annulatus*.

3.4. Ovicidal Activity of EO Formulations

CL and CLN significantly inhibited egg hatching ($p \le 0.05$), achieving inhibition rates of 90.66 and 69.00%, respectively, at the highest concentration (10%). At the same time, neither TT nor TTN had significant effects on the egg hatching percentage at any of the tested concentrations, and the treated eggs hatched normally (Table 5). The eggs treated with Phoxim 50% showed significant inhibition of egg hatching, achieving an inhibition percentage of 82.66%, whereas deltamethrin-treated eggs were found to have no effect on the hatching rate, achieving the same hatchability percentage as the untreated control eggs (Table 5).

Table 5. Ovicidal effect of different TT and CL EO forms and nanoemulsion forms.

Concentrations %	10	5	2.50	1.25	0.625
TT (Percentage of egg hatching inhibition mean \pm SD)	13.66 ± 2.081	11.33 ± 2.516	8.566 ± 1.504	5.000 ± 1.000	2.333 ± 1.154
TTN (Percentage of egg hatching inhibition mean \pm SD)	13.00 ± 2.000	10.66 ± 2.081	8.666 ± 3.055	5.666 ± 1.527	2.313 ± 1.527
CL (Percentage of egg hatching inhibition mean \pm SD)	90.66 ± 3.055 *	69.00 ± 4.000 *	47.66 ± 3.055 *	5.666 ± 1.527	4.000 ± 1.000
CLN (Percentage of egg hatching inhibition mean \pm SD)	69.00 ± 3.605 *	26.33 ± 3.511 *	15.00 ± 2.000	7.666 ± 1.527	4.666 ± 1.527
Control (Deltamethrin 2 µL/mL)			8.666 ± 1.527		
Control (Phoxim 0.50 µL/1mL)			82.66 ± 1.527		
Control (Ethyl alcohol 70%)			5.666 ± 2.082		

(*) significant compared with the control (Ethyl alcohol) (*p* < 0.05), TT (*tea tree* oil), CL (*Citrus limon* oil), TTN (*tea tree* nanoemulsion), CLN (*citrus lemon* nanoemulsion).

3.5. Larvicidal Activity of EO Formulations

TT and TTN only exhibited significant larval mortality at high concentrations, as the mortality rate reached 100% and 92.3% at a concentration of 10% and 100%, and 83.3% at a concentration of 5% for TT and TTN, respectively, but CL and CLN were not associated with significant larval mortality, even at high concentrations (Table 6). The LC₅₀ values for TT and TTN were 3.11% and 3.55%, respectively, whereas they were much higher for CL (30.1%) and CLN (32.27%) (Table 7) (Figure 4).

Table 6. Larvicidal effect of different forms of TT and CL (the ratio between oils in the 1:1 binary mixture) (mortality rate %).

Concentrations %	10	5	2.50	1.25	0.625
TT	100.0 ± 0.000 *	92.33 ± 2.516 *	22.00 ± 3.000	16.66 ± 3.785	11.66 ± 2.886
TTN	100.0 ± 0.000 *	83.33 ± 3.785 *	18.00 ± 4.582	11.33 ± 4.041	7.333 ± 2.516
CL	8.333 ± 3.511	7.000 ± 2.645	5.333 ± 1.527	4.333 ± 1.527	4.000 ± 1.000
CLN	8.666 ± 3.214	6.333 ± 2.516	4.666 ± 1.527	3.666 ± 1.527	3.333 ± 0.577
TTCL	100.0 ± 0.000 *	100.0 ± 0.000 *	100.0 ± 0.000 *	$78.00 \pm 2.645 *$	47.66 \pm 2.516 *
Control Deltamethrin (2 µL/1mL)			20.33 ± 0.577		
Control (Phoxim 0.50 μL/1mL)			100.0 ± 0.000		
Control (Ethyl alcohol 70%)			10.23 ± 0.432		

(*) significant compared with the control (Ethyl alcohol) (*p* < 0.05), TT (*tea tree* oil), TTN (*tea tree* nanoemulsion), CL (*citrus lemon* oil), CLN (*citrus lemon* nanoemulsion), TTCL (*tea tree* oil and *citrus lemon* oil).

Table 7. LC₅₀, LC₉₀, and synergistic factor for the adulticidal and larvicidal activity of TT and CL against *R*. (*B*.) *annulatus* (the ratio between oils in the 1:1 binary mixture).

0:1	Adulticidal Activity			Larvicidal Activity		
Oli	LC ₅₀ %	LC ₉₀ %	Synergistic Factor	LC ₅₀ %	LC ₉₀ %	Synergistic Factor
TT	2.05	4.00		3.112	5.155	
TTN	12.3	20.8	0.167 (Antagonism)	3.554	5.705	0.875 (Antagonism)
CL	1.26	2.62		30.13	49.87	
CLN	11.4	19.9	0.110 (Antagonism)	32.27	53.67	0.933 (Antagonism)
TTCL	4.93	12.1	0.416 (Antagonism)	0.790	1.386	3.939 (Synergism)

TT (tea tree oil), TTN (tea tree nanoemulsion), CL (citrus lemon oil), CLN (citrus lemon nanoemulsion), TTCL (tea tree oil and citrus lemon oil).

3.6. Synergistic/Antagonistic Effects of Binary Mixtures of the Used Oils

In the adult immersion test, the TTCL combination resulted in a lower mortality rate than using TT or CL alone (Table 3). The synergistic ratio of 0.416 indicated that this combination had an antagonistic effect (Table 7, Figure 4). TTCL showed significant larval mortality even at low concentrations, with an LC₅₀ value of 0.79%, indicating a synergistic effect (Table 7, Figure 4).

4. Discussion

The principal strategy used in tick control is the application of synthetic chemical acaricides. However, widespread and improper use of acaricides for tick control has resulted in significant resistance issues. Recently, there have been numerous reports on resistance to pyrethroid acaricides in ticks, for example, resistance to deltamethrin by *Rhipicephalus annulatus* [6]. Therefore, there is an urgent need to develop safe alternative

strategies to effectively control tick populations, and EOs have been proposed as one such strategy [28].

In the current investigation, the application of TT and CL at all tested concentrations resulted in significant adult mortality (p < 0.05). However, CL only produced a significant ovicidal impact at high concentrations (10 and 5%), while TT had a non-significant ovicidal effect. The nanoemulsion forms of TTN and CLN showed non-significant mortality rates against adult ticks. At concentrations of 5% and 10%, TT and TTN showed considerable rates of larval death; meanwhile, CL and CLN did not appear to induce any larval deaths at any of the tested concentrations. Similar results were obtained by Pazinato et al. [12] who demonstrated 100% reproductive suppression and recorded a 70% acaricidal impact of TT and TT nanoparticles against *Rhipicephalus microplus* females. Additionally, TT oil showed effectiveness against *Ixodes ricinus* nymphs with the mortality rate reaching 80% at a concentration of 10 μ L/mL [11]. Terpinene-4-ol, the primary constituent of TT oil, was shown to be the compound responsible for acaricidal activity [29].

Habeeb et al. [15] found significant toxic impacts of *Citrus limon* and *Citrus sinensis* against *Hyalomma dromedarii* ticks. Additionally, lemon oil was shown to be efficient for eradicating *Sarcoptes scabiei* mites in rabbits. Furthermore, treatment with citrus lemons significantly reduced the mosquito species' larvae population [30,31] and produced significant insecticidal action against *Spodoptera frugiperda* [32] and *Aedes alopictus* larvae [33].

It is noteworthy that the current EO nanoemulsions displayed lower activity levels than the pure oils against adult ticks. Galli et al. [34] reported similar levels of effectiveness for pure, nanoemulsion, and nanocapsule forms of Eucalyptus globulus EO against Rhipicephalus *microplus* ticks. This result can be explained by the fact that a variety of factors, such as the temperature, storage time, pH, and ionic strength, have negative impacts on nanoemulsion stability. Furthermore, when an emulsion is kept at a certain temperature during its synthesis, the thermal energy of the droplets increases, raising the possibility of droplet collisions. Additionally, this lessens the viscosity of the interface, facilitating faster droplet coalescence and film drainage [35]. Although the EOs did not cause significant mortality rates in their nanoemulsion forms compared with their ordinary forms, they significantly reduced the reproductive efficacy of the treated ticks. In contrast, Dos Santos et al. [17] discovered that the nanostructured form of cinnamon essential oils had a high level of acaricidal effectiveness against R. (B.) annulatus ticks. In addition, Ibrahium et al. [20] found that a geranium oil nanoemulsion was more effective than the pure form against *R. annulatus.* The increased effectiveness of nanoemulsions over pure oils is a result of the smaller droplet size of EOs, which enhances the number of active molecule interactions with the biological target membranes and promotes their transfer through them [36]. Additionally, Gadelhaq et al. [37] showed that the D-limonene nanoemulsion maintains its efficacy for 2 months and has better efficacy than D-limonene in its unemulsified form. Furthermore, by lowering the LC₅₀ to its half value, EO nanoemulsions showed enhanced effectiveness against *Aedes aegypti* larvae [38].

The insecticidal activity of an essential oil depends not only on its primary molecules but also on how all of its constituent parts interact with the physiology and behavior of an insect [39,40]. In addition to the essential oils' toxicity, it is important to note that larvicidal effectiveness can be improved by using straightforward binary combinations [8]. The adulticidal activity level of the EO binary mix (TTCL), when a ratio of 1:1 was used, was less than those of the pure TT and CL oils alone. As a result, the combination had non-significant LC_{50} and LC_{90} values (4.93 and 12.1%, respectively) against adult ticks and an antagonistic effect with a synergistic factor of less than 1, although the TTCL was associated with high proportions of larval death at all concentrations. Additionally, the mixture had considerable LC_{50} and LC_{90} values (0.790 and 1.386%, respectively), demonstrating a synergistic impact with a synergistic factor greater than 1. Similarly, Benelli et al. [8] investigated binary combinations of various essential oils and discovered that *Pinus nigra* and *Aloysia citriodora* had a strong synergistic effect on *Culex quinquefasciatus* larvae. Meanwhile, they reported a low effectiveness for *Hyssopus officinalis* and *Pelargonium graveolens*.

5. Conclusions

The pure TT and CL EOs had lower LC_{50} and LC_{90} values than the other two types (nanoemulsion and combination) against adult tick stages. The high CL concentrations (10 and 5%) showed the highest effective ovicidal activity levels. In contrast to their pure and nanoemulsion forms, the TT and CL binary mixture displayed potent larvicidal activity. Given that CL essential oil is affordable, further research into the use of CL on animals harmed by ticks needs to be undertaken.

Author Contributions: Conceptualization, S.M.A.; methodology, S.M.I. and S.M.A.; software, S.A.A.M.; validation, S.M.I.; formal analysis, S.M.I. and S.M.A.; investigation, S.M.I. and S.M.A.; resources, A.O.H.; data curation, S.M.I.; writing—original draft, S.M.I.; writing—review & editing, A.-A.S.A.-B. and S.M.A.; visualization, S.M.I. and S.M.A.; supervision, A.A.W., A.A.F. and S.M.A.; funding acquisition, A.-A.S.A.-B. and S.A.-Q. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the Researcher Supporting Project (RSP-2021/3), King Saud University.

Institutional Review Board Statement: The study was approved by the Faculty of Veterinary Medicine, Beni-Suef University, Egypt, under approval number (021-172).

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors appreciated the helping of veterinarians in collecting samples of this work.

Conflicts of Interest: The authors declare that they have no conflict of interest.

List of Abbreviations

TT (tea tree); TTN (*tea tree* nanoemulsion); CL (citrus lemon); CLN (citrus lemon nanoemulsion); *R. annulatus* (*Rhipicephalus annulatus*); EOs (essential oils); GC-MS (gas chromatography–mass spectrometry); PDI (polydispersity index); ALT (adult immersion test); LPT (larval packet test); EPI (egg production index); LC₅₀ (lethal concentration of 50% of the population); LC₉₀ (lethal concentration of 90% of the population).

References

- Aboelhadid, S.M.; Arafa, W.M.; Mahrous, L.N.; Fahmy, M.M.; Kamel, A.A. Molecular detection of *Rhipicephalus (Boophilus)* annulatus resistance against deltamethrin in middle Egypt. Vet. Parasitol. Reg. Stud. Rep. 2018, 13, 198–204. [CrossRef] [PubMed]
- Pérez de León, A.A.; Mitchell, R.D., 3rd; Watson, D.W. Ectoparasites of cattle. Vet. Clin. N. Am. Food Anim. Pract. 2020, 36, 173–185. [CrossRef] [PubMed]
- 3. Davies, T.G.E.; Field, L.; Usherwood, P.N.R.; Williamson, M.S. DDT, pyrethrins, pyrethroids and insect sodium channels. *IUBMB Life* 2007, *59*, 151–162. [CrossRef] [PubMed]
- Field, L.M.; Emyr Davies, T.G.; O'Reilly, A.O.; Williamson, M.S.; Wallace, B.A. Volt-age-gated sodium channels as targets for pyrethroid insecticides. *Eur. Biophys. J.* 2017, 46, 675–679. [CrossRef] [PubMed]
- Pawa, A.; Mali, G.; Deshmukh, H. Biodegradation of Deltamethrin by using Indigenous Bacteria Isolated from Contaminated Soil. Int. J. Curr. Microbiol. Appl. Sci. 2016, 5, 258–265. [CrossRef]
- Arafa, W.M.; Klafke, G.M.; Tidwell, J.P.; Perez de leon, A.A.; Esteve-Gassent, M. Detection of single nucleotide polymorphism in the para-sodium channel gene of *Rhipicephalus annulatus* populations from Egypt resistant to deltamethrin. *Tick Tick-Borne Dis.* 2020, 11, 101488. [CrossRef]
- 7. Bagavan, A.; Kamaraj, C.; Elango, G.; Abduz Zahir, A.; Abdul Rahuman, A. Adulticidal and larvicidal efficacy of some medicinal plant extracts against tick, fluke and mosquitoes. *Vet. Parasitol.* **2009**, *166*, 286–292. [CrossRef]
- Benelli, G.; Pavela, R.; Canale, A.; Cianfaglione, K.; Ciaschetti, G.; Conti, F.; Nicoletti, M.; Senthil-Nathan, S.; Mehlhorn, H.; Maggi, F. Acute larvicidal toxicity of five essential oils (*Pinus nigra, Hyssopus officinalis, Satureja montana, Aloysia citrodora* and *Pelargonium* graveolens) against the filariasis vector *Culex quinquefasciatus*: Synergistic and antagonistic effects. *Parasitol. Int.* 2017, 66, 166–171. [CrossRef]
- Aboelhadid, S.M.; Arafa, W.M.; Abdel-Baki, A.-A.S.; Sokmen, A.; Al-Quraishy, S.; Hassan, A.O.; Kamel, A.A. Acaricidal activity of *Foeniculum vulgare* against *Rhipicephalus annulatus* is mainly dependent on its constituent from trans-anethone. *PLoS ONE* 2021, 16, e0260172. [CrossRef]

- 10. Nazzaro, F.; Fratianni, F.; De Martino, L.; Coppola, R.; De Feo, V. Effect of Essential Oils on Pathogenic Bacteria. *Pharmaceuticals* **2013**, *6*, 1451–1474. [CrossRef]
- 11. Iori, A.; Grazioli, D.; Gentile, E.; Marano, G.; Salvatore, G. Acaricidal properties of the essential oil of *Melaleuca alternifolia* Cheel (*tea tree* oil) against nymphs of *Ixodes ricinus. Vet. Parasitol.* **2005**, 129, 173–176. [CrossRef] [PubMed]
- Pazinato, R.; Klauck, V.; Volpato, A.; Tonin, A.; Santos, R.C.; De Souza, M.E.; Vaucher, R.A.; Raffin, R.; Gomes, P.; Felippi, C.C.; et al. Influence of *tea tree* oil (*Melaleuca alternifolia*) on the cattle tick *Rhipicephalus microplus*. *Exp. Appl. Acarol.* 2013, 63, 77–83. [CrossRef] [PubMed]
- Martins, G.D.S.O.; Zago, H.B.; Costa, A.V.; Junior, L.M.D.A.; De Carvalho, J.R. Chemical Composition and Toxicity of Citrus Essential Oils on *Dysmicoccus brevipes* (Hemiptera: Pseudococcidae). *Rev. Caatinga* 2017, 30, 811–817. [CrossRef]
- 14. Aboelhadid, S.M.; Mahrous, L.N.; Hashem, S.A.; Abdel-Kafy, E.M.; Miller, R.J. In vitro and in vivo effect of *Citrus limon* essential oil against sarcoptic mange in rabbits. *Parasitol. Res.* **2016**, *115*, 3013–3020. [CrossRef]
- 15. Habeeb, S.M.; El Namaky, A.H.; Kamel, R.O. In vivo evaluation of toxic effects of avermectin, *Citrus sinensis* var. balady and *C. limon* on female *Hyalomma dromedarii* (acari: Ixodidae). *Acarologia* **2008**, *49*, 13–22.
- 16. Wang, L.; Li, X.; Zhang, G.; Dong, J.; Eastoe, J. Oil-in-water nanoemulsions for pesticide formulations. *J. Colloid Interface Sci.* 2007, 314, 230–235. [CrossRef]
- Dos Santos, D.S.; Boito, J.P.; Santos, R.C.V.; Quatrin, P.M.; Ourique, A.; Dos Reis, J.H.; Gebert, R.R.; Glombowsky, P.; Klauck, V.; Boligon, A.A.; et al. Nanostructured cinnamon oil has the potential to control *Rhipicephalus microplus* ticks on cattle. *Exp. Appl. Acarol.* 2017, 73, 129–138. [CrossRef]
- Lazcano, D.E.; Padilla, C.E.; Castillo, H.G.A.; Estarrón, E.M.; Espinosa, A.H.; Paniagua, B.N.A.; Gutiérrez, O.A.; Martínez, V.M. Development of essential oil-based phyto-formulations to control the cattle tick *Rhipicephalus microplus* using a mixture design approach. *Exp. Parasitol.* 2019, 201, 26–33. [CrossRef]
- 19. Boito, J.P.; Santos, R.C.V.; Vaucher, R.A.; Raffin, R.; Machado, G.; Tonin, A.A.; Da Silva, A.S. Evaluation of *tea tree* oil for controlling *Rhipicephalus microplus* in dairy cows. *Vet. Parasitol.* **2016**, 225, 70–72. [CrossRef]
- Ibrahium, S.M.; Aboelhadid, S.M.; Wahba, A.A.; Farghali, A.A.; Miller, R.J.; Abdel-Baki, A.-A.S.; Al-Quraishy, S. Preparation of geranium oil formulations effective for control of phenotypic resistant cattle tick *Rhipicephalus annulatus*. Sci. Rep. 2022, 12, 11693. [CrossRef]
- Suwannayod, S.; Sukontason, K.L.; Pitasawat, B.; Junkum, A.; Limsopatham, K.; Jones, M.K.; Somboon, P.; Leksomboon, R.; Chareonviriyaphap, T.; Tawatsin, A.; et al. Synergistic Toxicity of Plant Essential Oils Combined with Pyrethroid Insecticides against Blow Flies and the House Fly. *Insects* 2019, 10, 178. [CrossRef] [PubMed]
- 22. Nirmala, M.J.; Durai, L.; Rao, K.A.; Nagarajan, R. Ultrasonic Nanoemulsification of *Cuminum cyminum* Essential Oil and Its Applications in Medicine. *Int. J. Nanomed.* **2020**, *15*, 795–807. [CrossRef] [PubMed]
- 23. Estrada-Peña, A. Ticks of Domestic Animals in the Mediterranean Region: A Guide to Identification of Species; University of Zaragoza: Zaragoza, Spain, 2004.
- Drummond, R.O.; Ernst, S.E.; Trevino, J.L.; Gladney, W.J.; Graham, O.H. Boophilus annulatus and B. microplus: Laboratory Tests of Insecticides. J. Econ. Entomol. 1973, 66, 130–133. [CrossRef] [PubMed]
- 25. Klafke, G.M.; Thomas, D.B.; Miller, R.J.; de León, A.A.P. Efficacy of a water-based botanical acaricide formulation applied in portable spray box against the southern cattle tick, *Rhipicephalus (Boophilus) microplus (Acari: Ixodidae), infesting cattle. Ticks Tick-Borne Dis.* **2021**, *12*, 101721. [CrossRef]
- Matos, R.S.; Daemon, E.; Monteiro, C.M.D.O.; Sampieri, B.R.; Marchesini, P.B.C.; Delmonte, C.; Camargo-Mathias, M.I. Thymol action on cells and tissues of the synganglia and salivary glands of *Rhipicephalus sanguineus* sensu lato females (Acari: Ixodidae). *Ticks Tick-Borne Dis.* 2018, 10, 314–320. [CrossRef] [PubMed]
- 27. Stone, B.F.; Haydock, K.P. A method for measuring the acaricide-susceptibility of the cattle tick *Boophilus microplus* (Can.). *Bull. Entomol. Res.* **1962**, *53*, 563–578. [CrossRef]
- Coles, T.B.; Dryden, M.W. Insecticide/acaricide resistance in fleas and ticks infesting dogs and cats. *Parasites Vectors* 2014, 7, 8. [CrossRef]
- 29. Walton, S.F.; McKinnon, M.; Pizzutto, S.; Dougall, A.; Williams, E.; Currie, B.J. Acaricidal activity of *Melaleuca alternifolia* (Tea tree) oil. *Arch. Dermatol.* **2015**, 140, 563–566. [CrossRef]
- Melliou, E.; Michaelakis, A.; Koliopoulos, G.; Skaltsounis, A.L.; Magiatis, P. High quality bergamot oil from Greece: Chemical analysis using enantiomeric GC-MS and larvicidal activity against the West Nile virus vector. *Molecules* 2009, 14, 839–849.
- Michaelakis, A.; Papachristos, D.; Kimbaris, A.; Koliopoulos, G.; Giatropoulos, A.; Polissiou, M.G. Citrus essential oils and four enantiomeric pinenes against *C. pipiens* (Diptera: Culicidae). *Parasitol. Res.* 2009, 105, 769–773. [CrossRef]
- 32. Villafañe, E.; Tolosa, D.; Bardón, A.; Neske, A. Toxic effects of *Citrus aurantium* and *C. lemon* essential oils on *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Nat. Prod. Commun.* **2011**, *6*, 1389–1392.
- Giatropoulos, A.; Papachristos, D.P.; Kimbaris, A.; Koliopoulos, G.; Polissiou, M.G.; Emmanouel, N.; Michaelakis, A. Evaluation
 of bioefficacy of three *Citrus* essential oils against the dengue vector *Aedes albopictus* (Diptera: Culicidae) in correlation to their
 components enantiomeric distribution. *Parasitol. Res.* 2012, 111, 2253–2263. [CrossRef] [PubMed]
- Galli, G.M.; Volpato, V.; Santos, R.C.V.; Gebert, R.R.; Quatrin, P.; Ourique, A.F.; Klein, B.; Wagner, R.; Tonin, A.A.; Baldissera, M.D.; et al. Effects of essential oil of *Eucalyptus globulus* loaded in nanoemulsions and in nanocapsules on reproduction of cattle tick (*Rhipicephalus microplus*). Arch. Zootec. 2018, 67, 494–498. [CrossRef]

- 35. Sagalowicz, L.; Leser, M.E. Delivery systems for liquid food products. Curr. Opin. Colloid Interface Sci. 2010, 15, 61–72. [CrossRef]
- 36. Sugumar, S.; Clarke, S.; Nirmala, M.; Tyagi, B.; Mukherjee, A.; Chandrasekaran, N. Nanoemulsion of eucalyptus oil and its larvicidal activity against *Culex quinquefasciatus*. *Bull. Entomol. Res.* **2014**, *104*, 393–402. [CrossRef]
- Gadelhaq, S.M.; Aboelhadid, S.M.; Abdel-Baki, A.S.; Hassan, K.M.; Arafa, W.M.; Ibrahium, S.M.; Al-Quraishy, S.; Hassan, A.O.; Abd El-Kareem, S.G. D-limonene nanoemulsion: Lousicidal activity, stability, and effect on the cuticle of *Columbicola columbae*. *Med. Vet. Entomol.* 2022. [CrossRef] [PubMed]
- Botas, G.d.S.; Cruz, R.A.S.; de Almeida, F.B.; Duarte, J.L.; Araújo, R.S.; Souto, R.N.P.; Ferreira, R.; Carvalho, J.C.T.; Santos, M.G.; Rocha, L.; et al. *Baccharis reticularia* DC. and limonene nanoemulsions: Promising larvicidal agents for *Aedes aegypti* (Diptera: Culicidae) control. *Molecules* 2017, 22, 1990. [CrossRef] [PubMed]
- 39. Pavela, R. Essential oils for the development of eco-friendly mosquito larvicides: A review. *Ind. Crops Prod.* 2015, 76, 174–187. [CrossRef]
- 40. Ebadollahi, A. Plant essential oils from Apiaceae Family as alternatives to conventional insecticides. Ecol. Balk. 2013, 5, 149–172.