

Supplemental materials

Phenotypic adaptation of *Pseudomonas aeruginosa* in the presence of siderophore-antibiotic conjugates during epithelial cell infection

Table S1. Strains and plasmids used in this study.

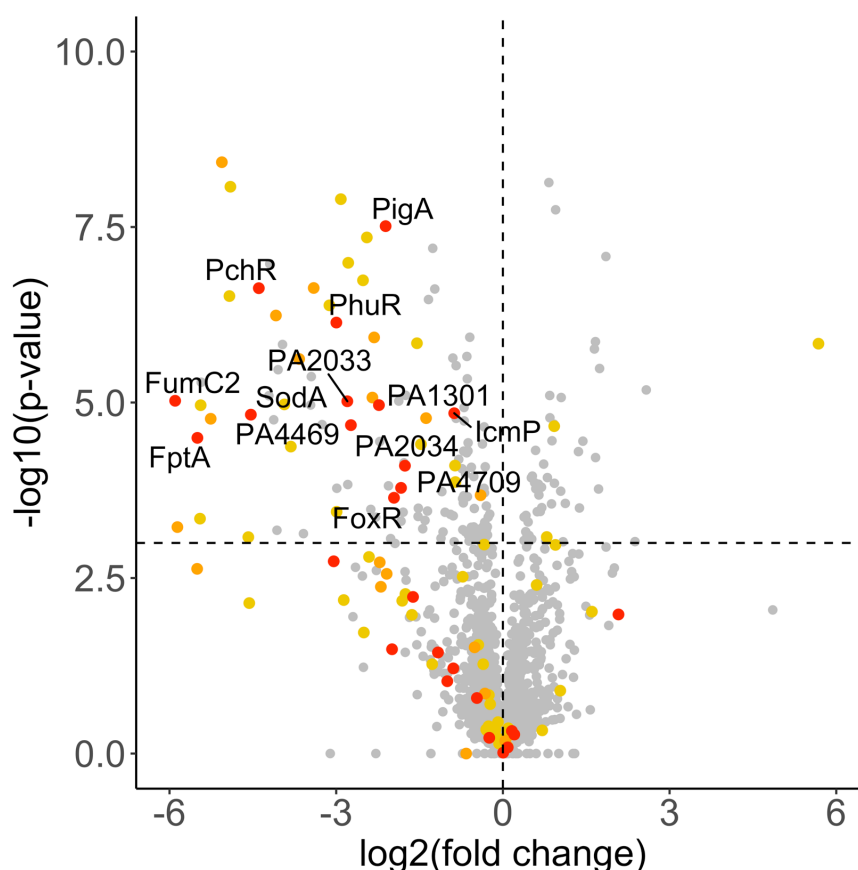
Strain	Collection ID	Relevant characteristics	Reference
<i>Pseudomonas aeruginosa</i>			
PAO1	PAO1	Wild-type strain	
$\Delta pvdF\Delta pchA$	PAS283	PAO1; <i>pvdF</i> and <i>pchA</i> chromosomally deleted	[1]
$\Delta pvdF\Delta pchA\Delta pfeA$	PAS294	PAO1; <i>pvdF</i> , <i>pchA</i> and <i>pfeA</i> chromosomally deleted	[2]
$\Delta pvdF\Delta pchA\Delta pirA$	PAS348	PAO1; <i>pvdF</i> , <i>pchA</i> and <i>pirA</i> chromosomally deleted	[1]
$\Delta pvdF\Delta pchA\Delta pfeA\Delta pirA$	PAS351	PAO1; <i>pvdF</i> , <i>pchA</i> , <i>pfeA</i> and <i>pirA</i> chromosomally deleted	[1]
$\Delta pvdF\Delta pchA\Delta fiuA$	PAS534	PAO1; <i>pvdF</i> , <i>pchA</i> and <i>fiuA</i> chromosomally deleted	[3]
<i>pfeEmcherry</i>	PAS355	PAO1; <i>pfeEmcherry</i> chromosomally integrated	[4]

Table S2. Primers used for RT-qPCR analysis

Primer ID	Target	Sequence
<i>uvrD</i> F	<i>uvrD</i>	CTACGGTAGCGAGACCTACAACAA
<i>uvrD</i> R	<i>uvrD</i>	GCGGCTGACGGTATTGGA
<i>GAPDH</i> F	<i>GAPDH</i>	TGCACCACCAACTGCTTAGC
<i>GAPDH</i> R	<i>GAPDH</i>	GGCATGGACTGTGGTCATGAG
<i>aprA</i> F	<i>aprA</i>	AACCAGAAGATCAACCTCAACGA
<i>aprA</i> R	<i>aprA</i>	TCGACACATTGCCCTTCAAC
<i>exoY</i> F	<i>exoY</i>	AATGGATGGCGGAGCCTATA
<i>exoY</i> R	<i>exoY</i>	CAAGGCGTTGCCGAGAGAT
<i>lasB</i> F	<i>lasB</i>	CGCCTGGGCGAGAACA

<i>lasB</i> R	<i>lasB</i>	GGGAATCAGGTAGGAGACGTTGT
<i>phzA2</i> F	<i>phzA2</i>	GGCACAACGTGCGGATCT
<i>phzA2</i> R	<i>phzA2</i>	CGCACTCGACCCAGAAGTG
<i>piv</i> F	<i>piv</i>	GCGTGGGCCTGAAAACG
<i>piv</i> R	<i>piv</i>	CGATCCATTCGAGGGTTGTC
<i>toxA</i> F	<i>toxA</i>	CCCGGCGAAGCATGAC
<i>toxA</i> R	<i>toxA</i>	GGGAAATGCAGGCGATGA
<i>amrZ</i> F	<i>amrZ</i>	TCGCTCGCAGCCATCAC
<i>amrZ</i> R	<i>amrZ</i>	TCGAGTCGGGCGATGATC
<i>fecA</i> F	<i>fecA</i>	GATCGACGACCTGATCCTCAA
<i>fecA</i> R	<i>fecA</i>	GGTCATCGCCGAAAACGT
<i>pirA</i> F	<i>pirA</i>	GCCTGAACGCTTCCCAAA
<i>pirA</i> R	<i>pirA</i>	TGAAGGCCCGTGCGATA
<i>pfeA</i> F	<i>pfeA</i>	GCCGAGACCAGCGTGAAC
<i>pfeA</i> F	<i>pfeA</i>	GGCCGGATTGATCTTGTT
<i>fpvA</i> F	<i>fpvA</i>	AGCCGCCTACCAGGATAAGC
<i>fpvA</i> R	<i>fpvA</i>	TGCCGTAATAGACGCTGGTTT
<i>fptA</i> F	<i>fptA</i>	GCGCCTGGGCTACAAGATC
<i>fptA</i> R	<i>fptA</i>	CCGTAGCGTTGTTCCAGTT
<i>foxA</i> F	<i>foxA</i>	AAGGGCTCGGATACCCAGTT
<i>foxA</i> R	<i>foxA</i>	CGTTGGGATCGTGTTGCA
<i>fiuA</i> F	<i>fiuA</i>	GCCGCGACAAGAAGTTCAG
<i>fiuA</i> R	<i>fiuA</i>	ACGACTCCGCATAGGAGATATAGG
<i>feoA</i> F	<i>feoA</i>	CCTACCGCATCACCGTTAT
<i>feoA</i> R	<i>feoA</i>	ACAGGCGTTGGCGATAGC
<i>hasR</i> F	<i>hasR</i>	AGCGCCTGCAGTTCAGCTA
<i>hasR</i> R	<i>hasR</i>	GTTCTCGGTGTTGAGCATGTTG
<i>phuR</i> F	<i>phuR</i>	GGTCGAACTGCCCAACGA
<i>phuR</i> R	<i>phuR</i>	TACGATGTCCGGATCGACGTA

11



12

13 **Figure S1. Differential production of iron-regulated proteins in the presence of ALBO based on**
 14 **the Palma, Worgall, and Quadri (2003) and Ochsner et al. (2002) studies [5,6].** Both studies were
 15 transcriptome analyses of the *P. aeruginosa* response to iron. The analysis presented here was performed
 16 on *P. aeruginosa* PAO1 cells after a 3-h infection of A549 epithelial cells in RPMI medium, in the
 17 presence or absence of 10 μ M ALBO. The yellow points correspond to proteins identified in one of the
 18 two analyses, the orange points to proteins identified in the two analyses, and the red points to proteins
 19 belonging to the direct Fur regulon, as described in the Collectf database.

20

21

22 **Synthesis of TCVL6.**

23 **General informations**

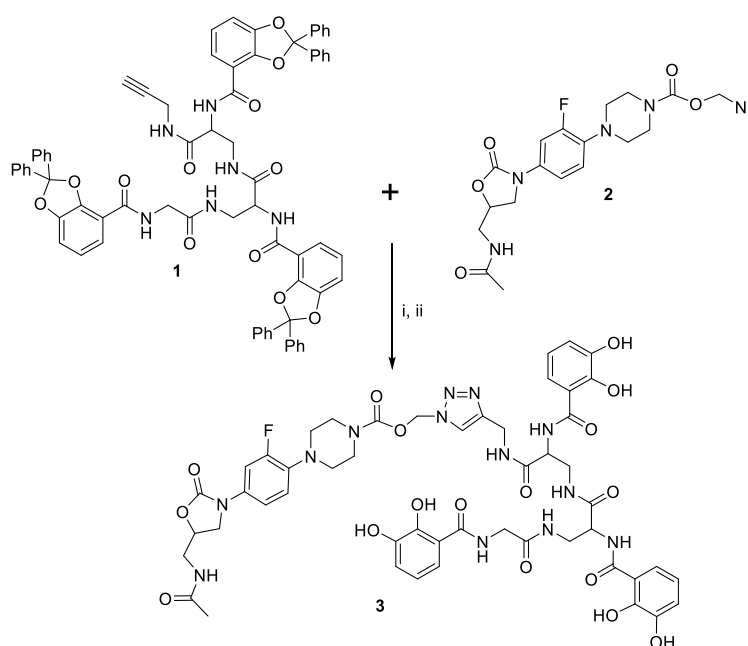
24 TCVL6 was synthesized as described in 1S Scheme. Linezolid was purchased from Sigma-
 25 Aldrich. Protected TCV **1** and linezolid-azide derivative **2** were synthesized according
 26 previously described protocols [7,8]. All reactions were carried out under argon (technical

quality, Air products). Solvents used were of analytical grade purity (>99.9%). When necessary, solvents and bases were purchased extra-dry. All other chemicals were obtained from commercial suppliers and were used as received, unless otherwise stated. All reactions were monitored by thin-layer chromatography (TLC) using *Merck* precoated silica gel 60F²⁵⁴ (0.25 mm). TLC are visualized using UV (254 nm/365 nm, *Vilber Lourmat*, VL-4LC) and/or using classical revelation mixtures (sulfuric vaniline, potassium permanganate, ninhydrin reagent). Final conjugate **3** was also detected on TLC using a 2% hydromethanolic solution of FeCl₃ (2%). Before chromatographic purification reaction mixtures were adsorbed on silica gel (60-200 µm, VWR Chemicals). Chromatographic purifications were performed on a *Reveleris* (*Grace Davison Discovery Sciences*) purification device using PuriFlash® pre-packed silica gel columns (*Interchim*, Montluçon, France). NMR spectra were recorded on Bruker Avance 400 (¹H: 400 MHz, ¹³C: 100 MHz, ¹⁹F : 376 MHz) or Avance 500 (¹H: 500 MHz, ¹³C: 125 MHz, ¹⁹F : 470 MHz), using the residual non-deuterated solvent as reference. The chemical shifts (δ) and coupling constants (*J*) are expressed in ppm and hertz respectively. Multiplicity is indicated as follows: s for singlet, d for doublet, t for triplet, q for quadruplet, quint for quintuplet and m for a multiplet. The letter "b" before the multiplicity indicates a broadened signal. Mass spectra were recorded in the *Service Commun d'Analyses (SCA) de la Faculté de Pharmacie de l'Université de Strasbourg* and were measured in ES-TOF experiments on a *Bruker Daltonic* MicroTOF mass spectrometer. LC/HRMS were measured after calibration on an *Agilent* QToF.

Protocol

Linezolid azide **2** (1 eq.) and alkyne-vector **1** (1,2 eq.) were solubilized in THF (final concentration 0.1 M). An aqueous solution of CuSO₄ (200 mg/mL, 1 eq.) and sodium ascorbate (5 eq.) were successively added [9]. The mixture was sonicated and further stirred at 20°C

under argon till the total consumption of limiting reagent (linezolid azide). The mixture was then filtered through a celite pad and the filtrate was adsorbed on silical gel. The crude mixture was then purified on silica gel using a gradient of EtOH in CH₂Cl₂. The resulting white solid was dissolved in the mixture CH₂Cl₂/TFA/Triisopropylsilane/EtOH (70/20/5/5, 5 mL for 0.1 mmol of starting linezolid azide **2**). The solution was stirred at 20°C till the total consumption of the starting material. Solvents were then evaporated from the mixture under reduced pressure. The oily residue was dissolved in a minimum volume of THF then cyclohexane was added dropwise till the precipitation of the expected conjugate as a thin powder. The pure conjugate was filtered off on a Hirsch funnel.

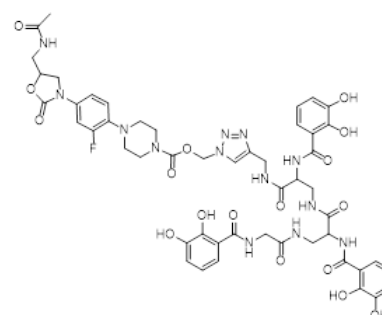


Scheme S1: Synthesis of TCVL6 (3). i. CuSO₄, sodium ascorbate, THF/H₂O, ((, 20°C. ii. CH₂Cl₂/TFA/Triisopropylsilane/EtOH (70/20/5/5, 20°C.

Spectroscopic analysis of TCVL6.

¹H NMR (500 MHz, DMSO-*d*₆): δ 11.98 (s, 1H), 11.73 (m, 2H), 9.05 (br s, 3H), 8.79 (s, 1H), 8.56 (d, *J* = 6.9 Hz, 1H), 8.53 (d, *J* = 6.9 Hz, 1H), 8.45 (t, *J* = 5.5 Hz, 1H), 8.18 (t, *J* = 5.5 Hz, 1H), 8.02 (t, *J* = 5.5 Hz, 1H), 7.93 (t, *J* = 5.5 Hz, 1H), 7.81 (s, 1H), 7.46 (t, *J* = 7.2 Hz, 1H), 7.28-7.22 (m, 1H), 7.08-6.99 (m, 3H), 6.95-6.89 (m, 1H), 6.84-6.78 (m, 1H), 6.72-6.66 (m, 3H), 6.49-6.40 (m, 3H), 6.03 (s, 2H), 4.35-4.28 (m, 1H), 4.19-4.08 (m, 2H), 4.02-3.91 (m, 2H), 3.89-3.85 (m, 1H), 3.71-3.65 (m, 1H), 3.55-2.91 (m, 14H), 1.44 (s, 3H). **¹⁹F NMR** (125 MHz, DMSO-*d*₆): δ -73.72, -121.19. **¹³C NMR** (125 MHz, DMSO-*d*₆): δ 169.7, 169.6, 169.5, 169.4, 169.3, 169.2, 169.1, 162.5, 149.2, 149.0, 146.1, 146.0, 145.4, 137.1, 132.8, 129.7, 128.7, 124.1, 119.9, 118.9, 118.8, 118.2, 118.1, 117.8, 117.7, 115.5, 114.1, 71.6, 68.4, 53.9, 53.8, 53.7, 50.1, 47.3, 47.2, 43.7, 42.4, 41.5, 10.5, 40.3, 40.2, 40.1, 35.9, 30.9, 27.5, 23.4, 23.3, 22.5, 21.8, 21.7. **HRMS:** C₅₀H₅₄FN₁₃O₁₇: calcd: 1127.37447, found: 1127.37501.

LC-HRMS Spectra of compound 3 (TCVL6)



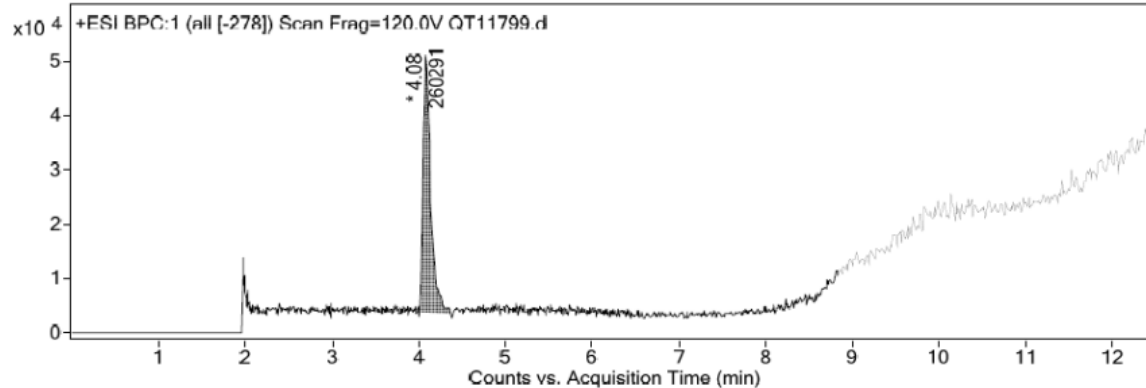
Chemical Formula: C₅₀H₅₄FN₁₃O₁₇
 Exact Mass: 1127.37447
 Molecular Weight: 1128.05440

Data Filename: QT11799.d Sample Name: NZ108
 Inj. Vol: 0.1 Position: P1-B3
 Instrument Name: SCA Illkirch QToF User Name: PW
 Acq Method: C18-2,1x5x1,8DMSO.m Acquired Time: 2/4/2016 12:07:19 PM
 IRM Calibration Status: Success DA Method: C18-2,1x5x1,8.m
 Comment:

Sample Group: Info.

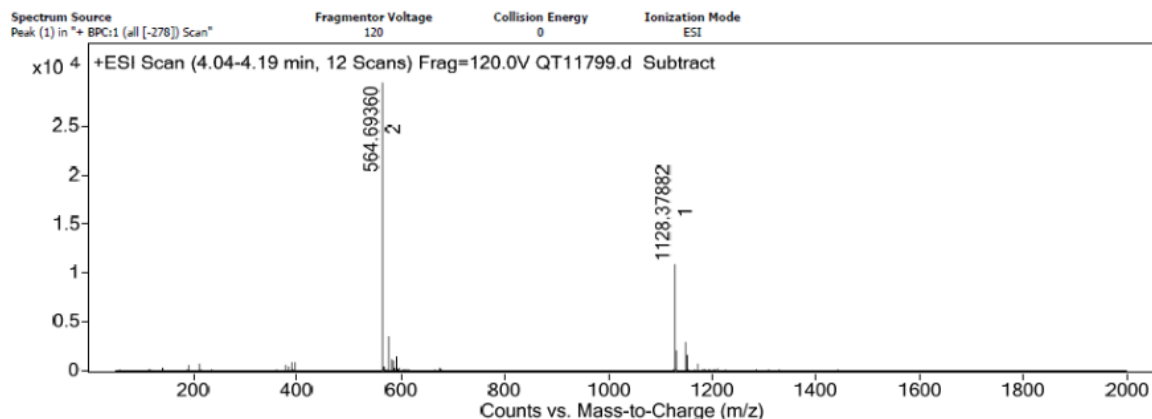
User Chromatograms

Fragmentor Voltage: 120 Collision Energy: 0 Ionization Mode: ESI



Integration Peak List

Start	RT	End	Height	Area	Area %	AreaSum%
3.97	4.08	4.37	47512	260291	100	100



Peak List

m/z	z	Abund	Formula	Ion
564.6936	2	29418.6	C50 H56 F N13 O17	(M+2H)+2
565.19508	2	17423.8	C50 H56 F N13 O17	(M+2H)+2
565.69599	2	6106.5	C50 H56 F N13 O17	(M+2H)+2
575.68361	2	3582.9		
576.18663	2	2021		
1128.37882	1	10820.6	C50 H55 F N13 O17	(M+H)+
1129.38218	1	6280.4	C50 H55 F N13 O17	(M+H)+
1130.38535	1	2133	C50 H55 F N13 O17	(M+H)+
1150.36048	1	2936.7	C50 H54 F N13 Na O17	(M+Na)+
1151.36413	1	1682.2	C50 H54 F N13 Na O17	(M+Na)+

Formula Calculator Results

Formula	Best	Mass	Tgt Mass	Diff (ppm)	Mz	Ion Species	Score
C46 H50 F N19 O15		1127.37167	1127.37178	0.1	1128.37882	C46 H51 F N19 O15	99.57
C49 H49 N19 O14		1127.37166	1127.37064	-0.91	1128.37882	C49 H50 N19 O14	98.92
C45 H54 F N15 O19		1127.37165	1127.37044	-1.07	1128.37882	C45 H55 F N15 O19	98.77
C62 H50 F N11 O10		1127.3716	1127.37262	0.9	1128.37882	C62 H51 F N11 O10	98.03
C53 H53 N13 O16		1127.37162	1127.37332	1.51	1128.37882	C53 H54 N13 O16	97.9
C48 H53 N15 O18		1127.37164	1127.3693	-2.08	1128.37882	C48 H54 N15 O18	96.54
C50 H54 F N13 O17	TRUE	1127.37163	1127.37447	2.52	1128.37882	C50 H55 F N13 O17	95.18
C54 H49 N17 O12		1127.37164	1127.37466	2.68	1128.37882	C54 H50 N17 O12	94.19
C57 H50 F N13 O12		1127.37162	1127.36859	-2.68	1128.37882	C57 H51 F N13 O12	94.08
C51 H50 F N17 O13		1127.37165	1127.3758	3.69	1128.37882	C51 H51 F N17 O13	90.37
C46 H50 F N19 O15		1127.37139	1127.37178	0.34	1150.36048	C46 H50 F N19 Na O15	98.44
C49 H49 N19 O14		1127.37139	1127.37064	-0.66	1150.36048	C49 H49 N19 Na O14	98.12
C45 H54 F N15 O19		1127.37137	1127.37044	-0.82	1150.36048	C45 H54 F N15 Na O19	98.11
C48 H53 N15 O18		1127.37136	1127.3693	-1.83	1150.36048	C48 H53 N15 Na O18	96.2
C62 H50 F N11 O10		1127.37132	1127.37262	1.15	1150.36048	C62 H50 F N11 Na O10	96.17
C53 H53 N13 O16		1127.37134	1127.37332	1.76	1150.36048	C53 H53 N13 Na O16	96.08
C57 H50 F N13 O12		1127.37134	1127.36859	-2.43	1150.36048	C57 H50 F N13 Na O12	93.62
C50 H54 F N13 O17	TRUE	1127.37135	1127.37447	2.77	1150.36048	C50 H54 F N13 Na O17	93.18
C54 H49 N17 O12		1127.37136	1127.37466	2.92	1150.36048	C54 H49 N17 Na O12	92.17
C60 H49 N13 O11		1127.37133	1127.36745	-3.45	1150.36048	C60 H49 N13 Na O11	89.46
C53 H53 N13 O16		1127.37264	1127.37332	0.6	564.6936	C53 H55 N13 O16	99.64
C46 H50 F N19 O15		1127.3727	1127.37178	-0.81	564.6936	C46 H52 F N19 O15	99.61
C62 H50 F N11 O10		1127.37262	1127.37262	-0.01	564.6936	C62 H52 F N11 O10	98.85
C50 H54 F N13 O17	TRUE	1127.37265	1127.37447	1.61	564.6936	C50 H56 F N13 O17	98.67
C49 H49 N19 O14		1127.37269	1127.37064	-1.82	564.6936	C49 H51 N19 O14	98.32
C54 H49 N17 O12		1127.37267	1127.37466	1.77	564.6936	C54 H51 N17 O12	98.14
C45 H54 F N15 O19		1127.37267	1127.37044	-1.98	564.6936	C45 H56 F N15 O19	97.96
C51 H50 F N17 O13		1127.37267	1127.3758	2.78	564.6936	C51 H52 F N17 O13	96.21
C48 H53 N15 O18		1127.37267	1127.3693	-2.99	564.6936	C48 H55 N15 O18	95.76
C42 H53 N19 O19		1127.37271	1127.37651	3.37	564.6936	C42 H55 N19 O19	94.43

91

92

93 References

- 94 1. Gasser, V.; Baco, E.; Cunrath, O.; August, P.S.; Perraud, Q.; Zill, N.; Schleberger, C.; Schmidt,
95 A.; Paulen, A.; Bumann, D.; et al. Catechol siderophores repress the pyochelin pathway
96 and activate the enterobactin pathway in *Pseudomonas aeruginosa*: an opportunity for
97 siderophore-antibiotic conjugates development. *Environ. Microbiol.* **2016**, *18*, 819–832,
98 doi:10.1111/1462-2920.13199.
- 99 2. Paulen, A.; Gasser, V.; Hoegy, F.; Perraud, Q.; Pesset, B.; Schalk, I.J.; Mislin, G.L.A.
100 Synthesis and antibiotic activity of oxazolidinone-catechol conjugates against
101 *Pseudomonas aeruginosa*. *Org. Biomol. Chem.* **2015**, *13*, 11567–11579,
102 doi:10.1039/c5ob01859e.
- 103 3. Normant, V.P.; Josts, I.; Kuhn, L.; Perraud, Q.; Fritsch, S.; Hammann, P.; Mislin, G.L.A.;
104 Tidow, H.; Schalk, I.J. Nocardamine-dependent iron uptake in *Pseudomonas aeruginosa*:
105 exclusive involvement of the FoxA outer membrane transporter. *ACS Chem. Biol.* **2020**,
106 *15*, 2741–2751. doi: 10.1021/acscchembio.0c00535. .
- 107 4. Perraud, Q.; Moynié, L.; Gasser, V.; Munier, M.; Godet, J.; Hoegy, F.; Mély, Y.; Mislin,
108 G.L.A.; Naismith, J.H.; Schalk, I.J. A Key Role for the Periplasmic PfeE Esterase in Iron
109 Acquisition via the Siderophore Enterobactin in *Pseudomonas aeruginosa*. *ACS Chem.*
110 *Biol.* **2018**, *13*, 2603–2614, doi:10.1021/acscchembio.8b00543.
- 111 5. Palma, M.; Worgall, S.; Quadri, L.E.N. Transcriptome analysis of the *Pseudomonas*
112 *aeruginosa* response to iron. *Arch. Microbiol.* **2003**, *180*, 374–379, doi:10.1007/s00203-
113 003-0602-z.

- 114 6. Ochsner, U.A.; Wilderman, P.J.; Vasil, A.I.; Vasil, M.L. GeneChip expression analysis of the
115 iron starvation response in *Pseudomonas aeruginosa*: identification of novel pyoverdine
116 biosynthesis genes. *Mol. Microbiol.* **2002**, *45*, 1277–87.
- 117 7. Baco, E.; Hoegy, F.; Schalk, I.J.; Mislin, G.L. Diphenyl-benzo[1,3]dioxole-4-carboxylic acid
118 pentafluorophenyl ester: a convenient catechol precursor in the synthesis of siderophore
119 vectors suitable for antibiotic Trojan horse strategies. *Org. Biomol. Chem.* **2014**, *12*, 749–
120 57, doi:10.1039/c3ob41990h.
- 121 8. Paulen, A.; Hoegy, F.; Roche, B.; Schalk, I.J.; Mislin, G.L.A. Synthesis of conjugates between
122 oxazolidinone antibiotics and a pyochelin analogue. *Bioorg. Med. Chem. Lett.* **2017**, *27*,
123 4867–4870, doi:10.1016/j.bmcl.2017.09.039.
- 124 9. Rostovtsev, V.V.; Green, L.G.; Fokin, V.V.; Sharpless, K.B. A Stepwise Huisgen Cycloaddition
125 Process: Copper(I)-Catalyzed Regioselective “Ligation” of Azides and Terminal Alkynes.
126 *Ang. Chem. Int. Ed.* **2002**, *41*, 2596–2599, doi:10.1002/1521-
127 3773(20020715)41:14<2596::AID-ANIE2596>3.0.CO;2-4.
- 128

129