



Supplementary Table S1. Old and New World *Leishmania* species considered a potential source of animal and human infection without LEVs research previously considered or tested (***) open field for future research) [8,20,24,26,27,121].

Species	Geographic Distribution	Clinical Disease in Humans	Hosts	Experimental Activatory Stimuli	LEVs Isolation Methods	LEVs type and Sub-Cellular Origin	Size (Mean/Range)	Major LEVs Content	Method of LEVs Analysis
<i>L. (M.) enriettii</i>	Present in Brazil		Life cycle remains poorly understood						
		Human infections undetected. Emerging as new human pathogens, causing VL and CL in HIV ⁺ and HIV ⁻ patients	The identity of vectors responsible for transmission has not been confirmed for any species yet	***	***	***	***	***	***
		Prevalence of undiagnosed cases and asymptomatic infections?	Infect domestic Guinea pigs. Detected in wildlife kept in captivity.						
<i>L. (M.) macropodum</i>	Present in Australia	Human infections undetected. Prevalence of undiagnosed cases and asymptomatic infections?	Life cycle remains poorly understood	***	***	***	***	***	***
			Vector: (possibly, Phlebotomine sand fly species and biting midges (<i>Forcipomyia</i> sp.))						
<i>L. (M.) martiniquensis</i>	Present in Martinique island, Florida,	CL, DCL, VL	Detected in wildlife kept in captivity (Kangaroo species)						
		Prevalence of undiagnosed cases and	Life cycle remains	***	***	***	***	***	***

	Switzerland, asymptomatic Germany and infections? south-east Asia	poorly understood							
		VL in BALB/c mice	Vectors (?) Detected in human and domestic animals						
		CL, DCL, VL	Life cycle remains poorly understood						
<i>L. (M.) orientalis</i>	Present in south-east Asia	Prevalence of undiagnosed cases and asymptomatic infections?	The identity of vectors responsible for transmission has not been confirmed for any species yet.	***	***	***	***	***	***
			Detected in humans.						
		CL	Life cycle remains poorly understood						
<i>L. (M.) sp.</i> Ghana that is still formally undescribed	Present in Africa	Prevalence of undiagnosed cases and asymptomatic infections?	The identity of vectors responsible for transmission has not been confirmed for any species yet.	***	***	***	***	***	***
			Detected in human.						
			Vector: Phlebotomine sand fly species (<i>Phlebotomus</i>)						
<i>L. (L.) aethiopica</i>	Highlands of Ethiopia and Kenya, highlands of south-west Ethiopia	CL, DCL, MCL, Chronic oriental sore Zoonotic	Infect wild rodents.	***	***	***	***	***	***

			Rock hyraxes, <i>Procavia</i> <i>capensis</i> , <i>Heterohyrax</i> <i>brucei</i> (reservoir hosts)						
			Vector: Phlebotomine sand fly species (<i>Phlebotomus</i>)						
<i>L. (L.) gerbilli</i>	Eastern Russia, Mongolia	Human infection unknown	Great gerbil <i>Rhombomys</i> <i>opimus</i> (reservoir host)	***	***	***	***	***	***
			Vector: Phlebotomine sand fly species (<i>Phlebotomus</i>)						
<i>L. (L.) turanica</i>	Central Asia	Human infection unknown	Great gerbil <i>Rhombomys</i> <i>opimus</i> (reservoir host)	***	***	***	***	***	***
			Vector: Phlebotomine sand fly species (<i>Phlebotomus</i>)						
<i>L. (L.) arabica</i>	Saudi Arabia	Human infection unknown	Fat sand-rat <i>Psammomys</i> <i>obesus</i> (reservoir host)	***	***	***	***	***	***
			Vector: Phlebotomine sand fly species (<i>Phlebotomus</i>)						
<i>L. (L.) tropica</i>	Central to south-west Asia, North, equatorial and southern Africa, Middle east, Iran, Afghanistan, Sub-Saharan Africa, Kenya, Ethiopia and Namibia. Mediterranean	CL, VL Oriental sore (dry form), leishmaniasis recidivans	Infect wild rodents and wild canids? cats? dogs (CL), and humans. Humans are reservoir hosts (peridomestic).	***	***	***	***	***	***

			Probably rock hyraxes and <i>Procapra capensis</i> .						
		CL, DCL	Vector: Unknown (<i>Lutzomyia</i> ?)						
<i>L. (L.) venezuelensis</i>	Northern Venezuela, in the States of Lara and Yaracuy, Dominican Republic		Infect domestic cats? humans? wild and synanthropic rodents? Presumably zoonotic and sylvatic, but reservoir host unknown.	***	***	***	***	***	***
			Vector: Phlebotomine sand fly species (<i>Lutzomyia</i>)						
<i>L. (V.) colombiensis</i>	Colombia, Panama, Colombia, Venezuela, forests of Brazil and Peruvian lowlands, other Latin American countries	CL, VL	Infect sloths <i>Choloepus hoffmanni</i> (reservoir host) and humans (sylvatic)	***	***	***	***	***	***
			Vector: Phlebotomine sand fly species (<i>Lutzomyia</i>)						
<i>L. (V.) lainsoni</i>	Forested areas of Northern Pará Brazil, Guianas, Peru and Bolivia	CL	<i>Cuniculus paca</i> (reservoir host)	***	***	***	***	***	***
			Presumably zoonotic and sylvatic						
			Vector: Phlebotomine sand fly species (<i>Lutzomyia</i>)						
<i>L. (V.) lindenbergi</i>	Degraded forest in Belém, Pará, Brazil		Infect humans. It is suspected that the wild animal	***	***	***	***	***	***
		CL							

			reservoirs are probably terrestrial						
			Vector: Phlebotomine sand fly species (<i>Lutzomyia</i>)						
<i>L. (V.) naiffi</i>	States of Pará and Amazonas (Brazil), French Guyana, Panama	CL	Nine- banded armadillo <i>Dasypus novemcinctus</i> (reservoir host)	***	***	***	***	***	***
			Presumably zoonotic and sylvatic.						
			Vector: Phlebotomine sand fly species (<i>Lutzomyia</i>)						
<i>L. (V.) panamensis</i>	South and Central America, West of Andes, northern Venezuela, Panama	CL, MCL	Infect Synanthropic and Neotropical rodents, edentates, marsupials, procyonids, erethizontides, monkeys, hunting dogs and humans (primary forest, zoonotic).	***	***	***	***	***	***
			Sloth <i>Choloepus hoffmanni</i> and probably other animals (reservoir host)						
<i>L. (V.) peruviana</i>	South America, Arid valleys of the western Peruvian Andes	CL, MCL	Vector: Phlebotomine sand fly species (<i>Lutzomyia</i>)	***	***	***	***	***	***

Infect dogs
(CL) and
humans.

Reservoir
hosts: Rodents
(*Phyllotis
andinum* ?),
marsupials
(*Didelphis
marsupialis* ?)
and dogs
(zoonotic).

Transmission
is thought
mainly to be
dependent on
humans
(peridomestic
and sylvatic)

1. Centrifugation 2. Filtration, 3. Concentration by ultrafiltration/high molecular weight cut-off filter, 4. Sequential/serial centrifugation 5. Ultracentrifugation, 6. Buoyant density on Optiprep gradient fractionation, 7. Buoyant density on sucrose gradient fractionation, 8. Precipitation by exo-prep kit, 9. Gel exclusion chromatography, 10. Size exclusion chromatography, 11. Dissection/Suspension in PBS FP, flagellar pocket, PM, plasma membrane [8]