



Review

Decision Making in Severe Equine Asthma—Diagnosis and Monitoring

Joana Simões 1,2,3,* and Paula Tilley 2,3

- Equine Health and Welfare Academic Division, Faculty of Veterinary Medicine, Lusófona University, Campo Grande 376, 1749-024 Lisbon, Portugal
- ² CIISA-Centre for Interdisciplinary Research in Animal Health, Faculty of Veterinary Medicine, University of Lisbon, 1300-477 Lisbon, Portugal; ptilley@fmv.ulisboa.pt
- ³ Associate Laboratory for Animal and Veterinary Sciences (AL4Animals), Faculty of Veterinary Medicine, University of Lisbon, 1300-477 Lisbon, Portugal
- * Correspondence: joana.simoes@ulusofona.pt

Simple Summary: Severe equine asthma (SEA) is an important respiratory disease affecting a large number of horses worldwide. This chronic disease induces cough and respiratory distress compromising the animals' athletic ability and welfare. A wide variety of diagnostic tests have been described for researching and diagnosing SEA, but not all are commonly available outside of large research or specialized diagnostic centers. Moreover, in routine ambulatory practice the process of decision making is not always easy, especially in more complex medical cases, but its importance is pivotal for individualized patient care. Thus, the aim of this paper is to develop a flow-chart to assist equine practitioners in the process of decision making associated with diagnosing and monitoring SEA.

Abstract: Decision making consists of gathering quality data in order to correctly assess a situation and determine the best course of action. This process is a fundamental part of medicine and is what enables practitioners to accurately diagnose diseases and select appropriate treatment protocols. Despite severe equine asthma (SEA) being a highly prevalent lower respiratory disease amongst equids, clinicians still struggle with the optimization of routine diagnostic procedures. The use of several ancillary diagnostic tests has been reported for disease identification and monitoring, but many are only suitable for research purposes or lack practicality for everyday use. The aim of this paper is to assist the equine veterinarian in the process of decision making associated with managing SEA-affected patients. This review will focus on disease diagnosis and monitoring, while also presenting a flow-chart which includes the basic data that the clinician must obtain in order to accurately identify severely asthmatic horses in their everyday routine practice. It is important to note that European and American board-certified specialists on equine internal medicine can provide assistance in the diagnosis and treatment plan of SEA-affected horses.

Keywords: equine; severe equine asthma; diagnosis; clinical history; clinical signs; diagnostic imaging; lung function; cytology; decision making



Citation: Simões, J.; Tilley, P. Decision Making in Severe Equine Asthma—Diagnosis and Monitoring. *Animals* 2023, 13, 3872. https://doi.org/10.3390/ani13243872

Academic Editors: Romain Paillot and Harold C. McKenzie III

Received: 19 October 2023 Revised: 22 November 2023 Accepted: 12 December 2023 Published: 16 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Severe equine asthma (SEA) is a chronic insidious respiratory disease which commonly affects mature adult horses [1,2]. Its estimated prevalence in the northern hemisphere is 20%, but the number of affected individuals continues to rise [3–5].

The precise immunological pathways of this multifactorial disease are complex and not yet fully understood [6–12], but it is known that when susceptible individuals are exposed to high concentrations of respirable particles they develop inflammation, bronchospasm and airway hyperreactivity [13–16]. Consequently these animals develop increased respiratory

Animals 2023, 13, 3872 2 of 17

effort at rest, cough and nasal discharge which, depending on inflammation severity, may impact athletic performance and the horse's well-being [17,18].

According to the type of inflammatory triggers associated with SEA exacerbation, two major disease phenotypes have been described [1]. The stable-associated SEA mainly involves exposure to organic respirable particles found indoors during the colder months, such as in bedding materials, hay and straw [19–21], whilst the pasture-associated SEA occurs in animals kept at pasture during the warmer season due to exposure to pollen [22–24].

However, inflammation can be triggered by a large number of molecules with a synergistic effect, such as LPS, pollen, mites, fungi spores, or even plastic particles, which can be found in the horses' habitat [19–21,25–28]. Because antigen avoidance can be extremely difficult to achieve, affected animals tend to present recurrent episodes of disease exacerbation [29].

Disease diagnosis is not always linear, and many practitioners struggle with decision making when confronted with a potential severely asthmatic horse. Empirical treatment, lack of compliance and erroneous posology are just some of the culprits for an unsuccessful treatment response [29], but misdiagnosis can further hinder this process [30]. Thus, thorough knowledge of the available diagnostic and therapeutical management options for SEA can help improve disease outcome. This paper provides a review of the currently available diagnostic tools, whilst offering a flow-chart for decision making in routine equine veterinary practice.

2. Diagnosis

In evidence-based medicine, reaching a correct diagnosis is paramount to produce individualized patient care and optimize clinical outcome [31]. Every patient is unique, as their medical conditions can vary in severity, underlying causes, and treatment response. Thus, by accurately identifying the patient's condition, veterinarians can tailor their treatment plans to address the specific needs of the animal and their owner, taking into account risks and potential benefits, ultimately maximizing the chances of therapeutic success and animal well-being [29,32]. Therefore, diagnosis must be sought using the most up-to-date research and clinical guidelines in the decision-making process. This approach ensures that patients are subjected to the most relevant clinical interventions and receive treatments that have been proven effective [30,31]. Also, regarding the practitioner, evidence-based medicine is also the best way to efficiently allocate resources and contribute to continuous learning and improvement in healthcare practice.

2.1. History, Clinical Signs and Clinical Scores

Severely asthmatic horses will have a history of recurrent respiratory disease with clinical signs being triggered by exposure to specific environmental factors [1]. Often these signs will be seasonal with coughing or labored breathing occurring during spring or autumn. Some horses will also develop clinical signs when performing physical exercise in dusty arenas or when moved to stables with poor air quality [33,34] which is frequently associated with dusty bedding materials, hay or straw and poor ventilation [35–37]. Thus, it is important to carefully examine the horses' environment but also to inquire about any recently made changes, since some owners may have decided to alter the animals' environment but have forgotten to report it during the initial consultation. Most commonly, owners will report that in response to being fed poor quality hay the horse will exhibit bouts of cough [38,39]. Two different phenotypes have been reported in SEA which means that asthmatic horses can develop clinical signs when exposed to stable environmental conditions (fungal spores, organic dust, mites, endotoxins, among others) or when exposed to a pasture environment (pollen and fungal spores) [4,19,40,41]. Identifying the horses' phenotype will determine the best treatment protocol for these animals, and intradermal testing can be useful for constructing an individualized allergen eviction protocol [42,43].

The clinical signs associated with SEA are quite distinctive [39,44], which can lead to the temptation of making a presumptive diagnosis based only on the patient's clinical

Animals 2023, 13, 3872 3 of 17

history and physical examination. Nonetheless, this should be discouraged since it can lead to an erroneous diagnosis especially in horses examined during disease remission or with low grade airway hyperreactivity.

Severely asthmatic horses have a history of chronic and persistent cough, which can be seasonal, associated with stabling and hay feeding or pasture [45]. Close inspection of the affected horses' living quarters and feed will normally reveal the triggering factor.

Airway inflammation, hyperreactivity and obstruction result in cough, exercise intolerance, increased respiratory effort at rest and nasal discharge [17,43,46], and the severity of the exhibited clinical signs tend to correlate with the degree and persistence of the inflammatory process [18,47,48].

Cough is usually the first clinical sign reported by owners and although the onset of exercise intolerance precedes it [49], the latter is not always reported early on. This could be related to the expected athletic level of the affected horse, since low grade inflammation may not impair the performance of less strenuous physical activities [50]. With disease progression the severity of cough increases and paroxysmal bouts of cough are typically observed [39,51].

The associated airway obstruction results in overt respiratory distress with consequent modification of the respiratory pattern, characterized by a short inspiration followed by prolonged exhalation and abdominal lift [52]. Affected animals present increased abdominal effort, with consequent hypertrophy of the external oblique muscles ('heave line') and nasal flaring as an attempt to reduce upper airway resistance [17,18,47,53].

Changes in mucus rheology along with its decreased clearance result in the accumulation of secretions in the tracheobronchial tree [54,55]. However, due to the regular swallowing of these secretions, horses only exhibit nasal discharge occasionally [56]. Nonetheless the presence of a tracheal rattle can be easily auscultated by placing the stethoscope over the tracheal area during physical examination, indicating the presence of mucus [57]. Since these animals are also at risk of contracting opportunistic respiratory infections, some horses may develop a purulent nasal discharge [58,59].

Thoracic auscultation of affected animals will reveal increased bronchovesicular sounds, end-expiratory wheezes due to airway narrowing, and inspiratory crackles caused by the opening of collapsed airways. An expanded pulmonary field can also be auscultated due to lung hyperinflation, secondary to air entrapment in the alveoli [1,2,60]. In some cases, lung auscultation will be unusually silent despite the horse exhibiting substantial respiratory effort due to a remarkable compromise of airflow associated with significant airway obstruction [60].

Loss of body mass and cachexia can also occur in horses with extreme respiratory distress due to a combination of reduced food intake and increased energy expenditure to overcome the expiratory obstruction [50,61,62].

Clinical scores can help assess and monitor disease severity and prognosis by evaluating common clinical signs observed during disease exacerbation, such as cough, nasal flaring, nasal discharge, abdominal lift, and exercise intolerance [17,18,46,63].

However, these scores alone can be insufficient, since mild or subclinical cases of severe equine asthma have little or no clinical signs despite maintaining some degree of airway inflammation and hyperreactivity [39,44,45,64]. Although scores based on nasal flaring and abdominal effort correlate with airway dysfunction, exercise intolerance does not, and cough may not occur during disease remission [47,53,65]. Examples of these scores are the ones published by Rush [65,66], Robinson [53,67] and Tesarowski [68,69].

The main limitation of clinical sign scores is therefore the correct diagnosis of severely asthmatic horses when in disease remission, making other ancillary diagnostic tests crucial for accurate disease characterization and exclusion of other alternative diagnoses [44,46,53]. Nonetheless, they can be useful in the initial triage and for the continuous assessment of treatment response, without resorting to more invasive diagnostic methods.

Furthermore, methods like the Horse Owner Assessed Respiratory Signs Index (HOARSI) [39,46], and the Visual Analog Scale (VAS) [70], can be particularly useful

Animals 2023, 13, 3872 4 of 17

for triage since they rely on information reported by horse owners to help identify the presence of respiratory disease. The HOARSI questionnaire scoring system is based on clinical history and on cough frequency, nasal discharge, breathing at rest and the horse's performance [15]. It has been primarily used to identify diseased asthmatic horses (with mild, moderate or severe equine asthma) and has shown good validation and repeatability [14,39,46]. However, it does not provide information on the degree of severity of each clinical entity and may misdiagnose horses in disease remission or in early inflammatory stages of SEA.

2.2. Diagnostic Imaging

Radiography and ultrasonography are routinely used ancillary diagnostic tests in equine ambulatory practice, since both are non-invasive and well accepted by horses [71,72].

In SEA, the radiographic findings have been found to correlate with the clinical signs and disease severity [17]. SEA-affected horses exhibit lung pattern changes, such as increased bronchovascular, interstitial and bronchial interstitial lung patterns, along with thickening of tracheal and bronchial walls [17,73]. In extreme cases, when disease progression results in lung remodeling, interstitial infiltration, increased lung radiopacity and bronchiectasis can also be observed [74,75]. Unfortunately thoracic x-rays are not able to detect discrete lung inflammation [18,76]; their interpretation can be challenging due to the superimposition of several anatomic structures and to fully observe the equine lung, x-rays of at least four different thoracic fields are required [77,78].

Ultrasound examination allows the clinician to attain more detailed information about the surface of the lung, including the assessment of effusion and its characteristics. However the most common ultrasonographic finding in severely asthmatic horses is the presence of comet tails which are non-specific artifacts associated with inflammation [79–81]. Alternatively, endobronchial ultrasound (ultrasonography via endoscopy) can be used to determine whether bronchial remodeling, a common feature of SEA, is present [82,83]. Using this ancillary test, one can distinguish SEA-affected horses in exacerbation from healthy individuals, but it fails to correctly identify severely asthmatic horses in remission [82].

Because the radiographic and ultrasonographic changes observed in severely asthmatic horses are not pathognomonic of the disease [76,80,81], it has been proposed that imaging is not required for the diagnosis of SEA in an ambulatory setting [2]. Nonetheless, the authors believe that these two techniques should be considered for disease staging and for excluding other potential differential diagnoses [1,17,71,72].

2.3. Endoscopy

Respiratory endoscopy alone or when combined with other sampling techniques can provide insight about the equine upper and lower airways [54,84,85]. Thus it can contribute to the characterization of SEA and to the exclusion of other differentials [1,17,55,85].

Endoscopic examination of severely asthmatic horses will reveal the presence of mucus in the trachea and bronchi and thickening of the carina (tracheal septum). The presence of bronchospasm and mucosal hyperemia associated with inflammation can also be assessed [17,55,86–90].

Tracheal secretions will usually form a small pool at the lowest point of the trachea. However, decreased mucus clearance and altered rheology increases the amount of mucus observed and mucus strings can be observed in the lateral and dorsal aspects of the trachea [55,91].

Most endoscopic scoring systems assess the quantity and quality of tracheal mucus [17,86–88,90], and mucus accumulation was found to correlate with clinical scores [92], cough frequency [51] and cytological indicators of airway inflammation [55]. The cytological findings associated with SEA are described in Section 2.5.

During disease remission, the amount of secretions found in the trachea of asthmatic horses significantly decrease and endoscopic mucus scores alone do not differentiate these animals from healthy ones [18,46]. Furthermore, mucus accumulation is not a sensitive

Animals 2023, 13, 3872 5 of 17

indicator of SEA since it is also present in other diseases, including mild and moderate equine asthma [1,84].

2.4. Lung Function Tests

Pulmonary function testing has become an important tool in the evaluation of the respiratory system since it provides information about ventilation and the dynamics of this system [71,93]. They are considered the gold standard for the diagnosis of asthma both in human and equine medicine [94]. In SEA they contribute to disease diagnosis and assessment of disease severity as well as treatment response. Nevertheless, no single test can be considered perfect, as they all have their strengths and weaknesses, and their selection will be determined by the clinician's needs, their practicality and availability. Moreover, despite technological evolution, some testing equipment are not suitable options for ambulatory settings, which is why lung function testing is mostly available at large research centers with few commercial options available in the market [2].

Arterial blood gas analysis assesses lung gas exchange and can be used in an ambulatory setting [95]. Severely asthmatic horses commonly present only hypoxemia, but lower values of pH and increased values of $PaCO_2$ have also been reported [18,96–99]. Its main disadvantage is that it lacks sensitivity for recognizing animals in remission [99].

The change in pleural pressure (ΔPpl), assessed by an oesophageal balloon catheter, is considered the gold standard for the diagnosis of SEA. The measured values can be interpreted on their own, with $\Delta Ppl > 15$ cm H_2O being considered the cut off value for disease diagnosis [1,46]. This method can easily be used in an ambulatory setting, but in the authors' experience some owners may be reticent to allow this test to be performed on their horses due to its invasive nature. When combined with a pneumotachograph, standard lung mechanics can be assessed, namely dynamic compliance (Cdyn), pulmonary resistance (RL), and work of breathing (W). Airway obstruction results in increased RL, W and ΔPpl , and decreased Cdyn [1,53,100]. Unfortunately ΔPpl alone will not help differentiate healthy animals from severely asthmatic ones in remission, nor will standard lung mechanics, as it has been reported to have a similar sensitivity to a clinical exam [44,53,63,101]. However, when combined with histamine bronchoprovocation, its sensitivity improves and, despite its limitations, it remains a valuable tool for assessing respiratory function in an ambulatory setting [102].

Flowmetrics is based in boxless plethysmography and it combines respiratory inductance plethysmography (RIP) with pneumotachography [103]. This system, which has been specifically developed for equines and was suited for field testing [104], is no longer commercially available. It had a sensitivity similar to that of pleural pressure but, when combined with histamine bronchoprovocation, allowed the detection of horses in disease remission [18,46,105,106].

Lung function testing can be associated with either a histamine or bronchodilator challenge [93]. Airway hyperreactivity, the reversible narrowing of airways in response to a bronchoconstrictor stimulus, such as histamine, occurs in all asthmatic horses, especially during disease exacerbation but is also present in cases of mild obstruction without apparent clinical signs [1,18,46,104,105,107–109]. After a base-line reading, increasing concentrations of a histamine solution (0, 2, 4, 8, 16 and 32 mg/mL) are progressively nebulized and following each administration, lung function is re-evaluated. If significant bronchoconstriction is observed or if clinical discomfort is noted, the test is immediately stopped. A healthy horse will tolerate the inhalation of the maximum histamine concentration, whilst asthmatic horses will tolerate significantly lower doses [18,46].

When asthmatic horses present with a severe compromise of baseline pulmonary function, it is recommended to perform a bronchodilator challenge instead [2,62]. A bronchodilator is administered (e.g., albuterol 450–900 μ g) after a base-line reading and 15 min later pulmonary function is re-assessed [1,93]. Within 10 min, a 50% improvement of airway resistance should occur in SEA-affected horses in exacerbation [93]. Furthermore,

Animals 2023, 13, 3872 6 of 17

both the maximum value and the magnitude of the observed bronchodilation can also predict, to some degree, the future therapeutic response.

The use of spirometry [44,110–112], electrical impedance tomography [113], and impulse oscillation system (IOS) for assessing the dynamics of equine lower airways have shown promising results, particularly IOS, which has been reported to differentiate severely asthmatic horses in remission from healthy controls [114–117]. At the present time these tests are more suited for research purposes and have yet to be perfected for everyday clinical use. Nonetheless, the authors believe that, similar to human medicine, lung function testing will become an indispensable diagnostic tool to monitor lower airway inflammation and assist in the diagnosis and staging of SEA.

2.5. Cytology

In equine ambulatory medicine, airway cytology remains a fundamental technique for diagnosing and monitoring SEA. It provides insight into the inflammatory status of airways and although it is not considered the gold standard for the diagnosis of this disease, its practicality in an ambulatory context has rendered this ancillary diagnostic test popular. Cytological samples can be collected using a wide variety of methods, including brush cytology, tracheal wash (TW), bronchoalveolar lavage fluid (BALF) or even bronchial biopsies [118–121].

Of these methods, BALF cytology is considered to be the one which most accurately reflects the cellular populations of the bronchi and alveoli and the consequent degree of inflammation found in the horses' lungs [120,122,123]. Sampling can easily be performed transendoscopically or 'blindly', via a balloon catheter, by instilling a volume of 250 to 500 mL of saline and a minimum of 400 cells should be counted [2,18,120,124,125]. When endoscopically aided, the BALF collection can be guided in order to sample either the right or left lung. This option can be useful if a localized disease or lesion is suspected (i.e., a lung abcess), but is unnecessary in SEA-affected horses where both lungs are equally affected [124].

BALF samples of healthy animals have <400 cells/ μ L and a superficial foam layer, indicating the presence of pulmonary surfactant. Alveolar macrophages (40–70%) and lymphocytes (30–60%) are the most commonly observed immune cells followed by neutrophils (<5%), mast cells (<2%) and eosinophils (<1%) [1]. The cytological profile of the severely asthmatic horse is usually characterized by neutrophilia (>20% neutrophils) and a reduction in macrophage and lymphocyte percentages [1,17,18,126–128]. An increased amount of mucus is also observed which can form Curschmann's spirals [57,129].

BALF differential cell counts correlate well with airway obstruction and hyperresponsiveness, and a higher percentage of neutrophils is associated with greater disease severity, coughing and worse mucus scores [17,47,51,129,130]. During clinical remission, affected horses maintain a slightly elevated neutrophil percentage [63], and despite corticosteroid treatment, BALF neutrophilia can persist when severely asthmatic horses continue to be exposed to the offending respirable particles [40,131–133].

Furthermore, it has been reported that the percentage of neutrophils in BALF can be used to classify the severity of SEA, as it correlates well with the clinical signs exhibited by asthmatic horses, changes observed in thoracic x-rays, mucus scores determined through endoscopy and airway remodeling [17,127]. It is important to mention that the bronchoalveolar lavage procedure should always be conducted after assessing lung function, as it has been shown to temporarily improve pulmonary resistance (RL), likely due to mucus clearance [134].

However, BALF cytology is not well accepted for the evaluation of high performance athletes [135]. Despite the reported poor correlation between neutrophil counts in the TW and in the BALF, which may indicate that the cellular population found in the tracheal lumen is not representative of lower airways [123,136,137], British race horse veterinarians prefer to perform TW instead of BALF collection [2,135]. Conversely, a study on a population of 154 horses showed a good correlation between the neutrophil percentage in TW

Animals 2023, 13, 3872 7 of 17

and BALF cytology with only 17.5% of the cases having been classified differently [138]. Nonetheless, until more studies are performed, BALF remains more suitable for the accurate diagnosis of SEA and when feasible, the combination of both TW and BALF can aid in disease diagnosis and characterization.

Diagnostic cytology should not be performed if the horse is receiving corticosteroids, since this will invariably modulate the inflammatory response associated with the disease and can result in an incorrect diagnosis. Furthermore, it should be noted that neutrophilia is not a prerequisite for the lung function deterioration observed in SEA and some horses may present an increased respiratory effort without BALF neutrophilia. Therefore, BALF cytology should not be used alone for diagnosing SEA [83,126,139].

The clinician should also be aware that signs of neutrophilic degeneration associated with the presence of bacteria or fungal spores/hyphae, either free or phagocyted, can be an indicator of a primary infectious condition or of an opportunistic infection. In such cases bacterial or mycological cultures should be considered to identify the causal agent and adjust the therapeutic protocol.

2.6. SEA Staging

One of the main challenges of managing a severely asthmatic patient is continuously monitoring disease severity and response to treatment. An initial detailed characterization of the disease may help select the best therapeutic protocol and optimize environmental management catering to the horses' and owners' individualized needs [94].

Staging methods were developed to help clinicians gather information about disease severity by combining history, physical examination and a variety of ancillary diagnostic tests.

A relatively complete SEA clinical staging method has been published, which encompasses clinical history reported by the owners and clinical signs observed during clinical examination, namely cough frequency, nasal flare and abdominal lift. It also uses ancillary diagnostic tests to quantify airway inflammation and remodeling, such as thoracic x-ray, endoscopy and BALF cytology [17]. This staging method has to be carried out in a hospital and only evaluates the present condition of the horse and does not take into account reported history. It also does not evaluate lung function which has been considered an important diagnostic indicator by the ECEIM 2016 consensus on inflammatory airway disease [1].

An alternative staging system for ambulatory practice that included lung function assessment was later developed [18]. This method included data of the horses' physical examination (clinical score), BALF cytology (neutrophil percentage), arterial blood oxygen pressure (PaO₂), pleural pressure (Δ Ppl) and histamine bronchoprovocation (maximum tolerated concentration). All the diagnostic procedures could easily be performed in the field, since all the equipment used was portable. However, this method has yet to be validated and the Open PlethTM used for lung function assessment during the histamine bronchoprovocation is no longer commercially available [2,18].

Staging methods are important tools for monitoring disease progression and treatment response [94]. In equine medicine, asthma staging still relies on invasive ancillary tests which may limit their routine application. For this reason, disease monitoring tends to rely on clinical scores or on owner reported information. We expect that with technological progress, less invasive and more sensitive ancillary tests will become available which will help the equine practitioner make informed decisions in their everyday practice.

2.7. SEA Characterization

Although the role of immunoglobulin E (IgE) in the pathophysiology of SEA remains controversial, differences in allergen-specific IgE concentrations in the sera and BALF between healthy and SEA-affected horses have been reported [26,140–143]. The measurement of allergen-specific IgE concentrations has helped ascertain the association between SEA and sensitization to fungi and mites [26,140,144]. Furthermore, using a microarray

Animals 2023, 13, 3872 8 of 17

platform, White and colleagues have reported that severely asthmatic horses can present different allergen sensitization profiles which usually involve exposure to fungi, mite, pollen proteins and, surprisingly, latex [20].

Intradermal tests (IDT) are used to assess the patient's reaction to an allergen. It requires the injection of specific allergens intradermally and if the horse is sensitized, a local allergic reaction occurs (papule). In SEA-affected horses the use of IDT allowed the identification of allergen sensitization [43,141]. Lo Feudo and colleagues reported that insects, the mite *Dermatophagoides* spp. and dog epithelium were the major allergen profiles associated with SEA [43].

In human medicine, skin prick tests (SPT) are commonly used in the diagnosis of allergic diseases since they are less invasive than IDT and also provide information about allergen-specific sensitization profiles. These ancillary tests can easily be performed in the horses in a clipped area of the neck by placing a drop of a specific allergen extract and penetrating the epidermis with a lancet. SPT successfully identified allergen sensitization in severely asthmatic horses [42], and has been proposed as a useful diagnostic tool for horses with insect bite hypersensitivity [145].

However, the IDT and SPT are not fully standardized in equine medicine. In IDT, a subjective method based on the wheals' size, discomfort, thickness, erythema and turgidity has been used to assess the grade of the reaction [43,146]. Nevertheless, in SPT, two very similar cut off values have been found for their use in horses, 1 cm [42] and 0.9 cm [145], which may already guide clinicians in the interpretation of these tests. SPT papule diameter values are calculated as the average of two orthogonal diameters.

Nonetheless, the measurement of allergen-specific IgE concentrations along with intradermal testing (IDT) and skin prick testing (SPT) allows the recognition of allergen-specific sensitization profiles which can be used to assist in the development of individualized allergen-avoidance protocols.

3. Diagnostic Flow-Chart

Because horses affected by severe equine asthma may not exhibit clinical signs during disease remission, disease diagnosis can be challenging, particularly to the less experienced practitioner. Thus, a flow-chart which aims to assist in the recognition of severely asthmatic horses by including the main steps required to correctly diagnose the disease, is proposed in Figure 1.

Although SEA is a common respiratory disease in equids, the clinician must be aware that other respiratory conditions, such as mild/moderate equine asthma, may result in lower airway inflammation which might mimic SEA in remission [1,147]. Furthermore, to a less trained eye, a bacterial/fungal pneumonia can be misdiagnosed as SEA, resulting in dire consequences should the affected horse be treated with corticosteroids [148]. A careful physical examination and owner interview, paired with environmental inspection are essential to prevent misdiagnosis. In SEA, the horses' respiratory signs will be recurrent, and in some cases irreversible, with owners reporting that they might worsen in specific seasons or when the animal is exposed to certain stimuli. Dry cough, respiratory effort with an abdominal pattern, exercise intolerance and nasal discharge are usually reported by owners [1,17,39,46,49]. Because these clinical signs are not pathognomonic of SEA, a thorough physical examination will help rule out other possible causes. The HOARSI questionnaire can be used to collect relevant information on the clinical history of the patient and help identify trigger factors [39]. Similarly, other clinical scores published by different research groups might be helpful in tracking the severity of clinical signs [53,65–69].

Animals 2023, 13, 3872 9 of 17

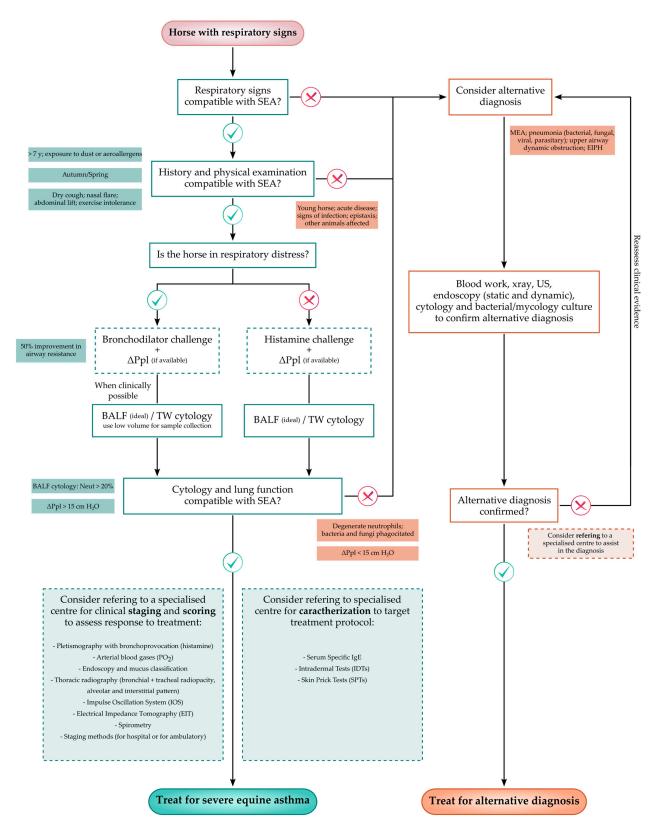


Figure 1. Flow-chart proposal for the diagnosis of severe equine asthma to be used in everyday veterinary practice [17,18,42,43,55,71,117]. MEA—Mild/Moderate Equine Asthma; EIPH—Exercise Induced Pulmonary Hemorrhage; BALF—Bronchoalveolar Lavage Fluid; TW—Tracheal Wash; ΔPpl—Change in pleural Pressure (indirect); SEA—Severe Equine Asthma; and Neut—Neutrophils.

Animals 2023, 13, 3872 10 of 17

Severely asthmatic horses will invariably exhibit airway inflammation, hyperreactivity and obstruction, and disease severity will determine the magnitude of these reversible findings [1]. When in remission, severely asthmatic horses will maintain low-grade inflammation and hyperreactivity but have little or no airway obstruction [18,46,104]. As previously mentioned, lung function tests are the best ancillary diagnostic tools for recognizing these cases. Lung function testing is not yet widely available in equine ambulatory practice and the most sensitive tests for detecting low-grade inflammation can only be found in research centers. However, clinicians can easily measure ΔPpl, using an esophageal balloon catheter and a pressure transducer, in order to assess the degree of airway obstruction [53,93]. The authors use VentiplotTM, a portable pleural pressure measurement device developed by Johannes Peter Schramel, Vetmeduni, Vienna. In order to improve the sensitivity of this test, histamine bronchoprovocation is used which allows the identification of horses in remission, since these animals will maintain some degree of airway hyperreactivity. This procedure should not be performed in horses with respiratory distress, since it will further impair the animal's ventilation. Alternatively, Δ Ppl can be coupled with a bronchodilator challenge and the clinicians should expect a 50% improvement of lung function readings [93].

If possible, cytology should always be performed on SEA-suspected horses. Lung neutrophilia is the hallmark of SEA, and BALF cytology will reveal a differential count of >20% neutrophils [1,149]. Cytology can also help rule out other causes of lower airway inflammation, such as mild/moderate equine asthma (>5% neutrophils and presence of mast cells) or infection (presence of degenerated neutrophils, bacteria and/or fungi) [1,150,151]. It is recommended to always perform cytology in combination with lung function testing since it has been reported that some severely asthmatic horses might have a paucigranulocytic cytological profile [83]. Additionally, when possible BALF cytology should be performed instead of TW to obtain a clearer image of the immune cells found in the horse's lungs [123,136]. Performing an endoscopically guided BALF collection allows the use of a small volume of fluid for sample collection, which can be useful in patients where instilling a large volume of saline is not recommended. The clinician should also be aware that when tracheal wash is used, the cut off of 20% neutrophils indicates only the presence of airway inflammation [128].

Thoracic radiography, ultrasonography and respiratory endoscopy can also contribute to disease diagnosis, not only by helping exclude alternative diagnostic possibilities but also by enabling SEA staging (endoscopic and radiographic examination) [17,18,46,81]. Whenever possible, these ancillary tests should be performed, although they are not an essential part of the basic diagnostic panel for this disease. Endoscopy can be used to evaluate tracheobronchial mucus accumulation, which in extreme cases will further compromise the animal's ventilation and might, therefore, constitute a potential therapeutic target for the improvement of clinical signs [54,55,91]. Similarly, performing a thoracic radiography will permit the assessment of the bronchial remodeling in response to bronchoconstriction and lung inflammation. Furthermore, thoracic radiography, ultrasonography and respiratory endoscopy can be performed in the field and this information can be included to help monitor treatment response [71]. Although the findings observed during the thoracic ultrasound of a SEA-affected horse are not specific to this disease, they may be used to assess the severity and chronicity of the inflammatory process. Ultrasound can also assist the clinician in ruling out potential infectious conditions, such as pneumonia/pleuropneumonia.

Although not mandatory, the authors recommend the use of a staging/scoring method to monitor disease progress. SEA-affected animals will require an individualized treatment protocol catered to both horse's and owner's needs [29]. Staging can also help the clinician identify potential lack of adhesion to the proposed treatment and can be used as a tool to confront non-adherent owners with the clinical evolution of their asthmatic horse.

Animals 2023, 13, 3872 11 of 17

4. Conclusions

The condition known today as severe equine asthma was first recognized several centuries ago. Since its first description, ancillary diagnostic tests have greatly evolved, but we have yet to develop a single diagnostic tool for disease identification and monitoring. A combination of clinical history, physical examination, lung function assessment and lower airway cytology are therefore fundamental to correctly diagnose affected animals.

Decision making, such as selecting the most suitable diagnostic approach, is a fundamental part of evidence-based medicine and can therefore make the difference between success and failure when managing a clinical case. We propose the use of a simple flow-chart diagram to help clinicians obtain the minimum database required for the diagnosis of SEA in their routine practice. Nonetheless, when possible, other ancillary tests should be performed to stage and characterize each clinical case, in order to improve diagnostic and treatment accuracy. These tests can also assist in monitoring the occurrence of pathological changes, such as mucus accumulation and bronchial smooth muscle hypertrophy secondary to the disease.

Author Contributions: J.S. and P.T. have contributed equally to the conceptualization, research, original draft preparation and review and editing of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Portuguese Foundation for Science and Technology (FCT) through grants UIDB/00276/2020 (CIISA) and LA/P/0059/2020 (AL4AnimalS).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: The authors would like to acknowledge the contribution of Hugo Rebelo for the flow-chart design.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Couëtil, L.L.; Cardwell, J.M.; Gerber, V.; Lavoie, J.P.; Léguillette, R.; Richard, E.A. Inflammatory Airway Disease of Horses-Revised Consensus Statement. *J. Vet. Intern. Med.* **2016**, *30*, 503–515. [CrossRef]
- 2. Couetil, L.; Cardwell, J.M.; Leguillette, R.; Mazan, M.; Richard, E.; Bienzle, D.; Bullone, M.; Gerber, V.; Ivester, K.; Lavoie, J.P.; et al. Equine Asthma: Current Understanding and Future Directions. *Front. Vet. Sci.* **2020**, *7*, 450. [CrossRef]
- 3. Hotchkiss, J.W.; Reid, S.W.J.; Christley, R.M. A survey of horse owners in Great Britain regarding horses in their care. Part 1: Horse demographic characteristics and management. *Equine Vet. J.* **2007**, *39*, 294–300. [CrossRef]
- 4. Hotchkiss, J.W.; Reid, S.W.J.; Christley, R.M. A survey of horse owners in Great Britain regarding horses in their care. Part 2: Risk factors for recurrent airway obstruction. *Equine Vet. J.* **2007**, *39*, 301–308. [CrossRef]
- 5. Ireland, J.L.; Christley, R.M.; McGowan, C.M.; Clegg, P.D.; Pinchbeck, G.L.; Martynski, P.D.; Payne, R.J.; Wylie, C.E.; Smith, S.; Marr, C.M.; et al. Prevalence of and Risk Factors for Recurrent Airway Obstruction in Geriatric Horses and Ponies. *Equine Vet. J.* **2015**, 47, 25. [CrossRef]
- 6. Giguère, S.; Viel, L.; Lee, E.; MacKay, R.J.; Hernandez, J.; Franchini, M. Cytokine induction in pulmonary airways of horses with heaves and effect of therapy with inhaled fluticasone propionate. *Vet. Immunol. Immunopathol.* **2002**, *85*, 147–158. [CrossRef]
- 7. Horohov, D.W.; Beadle, R.E.; Mouch, S.; Pourciau, S.S. Temporal regulation of cytokine mRNA expression in equine recurrent airway obstruction. *Vet. Immunol. Immunopathol.* **2005**, *108*, 237–245. [CrossRef]
- 8. Hulliger, M.F.; Pacholewska, A.; Vargas, A.; Lavoie, J.P.; Leeb, T.; Gerber, V.; Jagannathan, V. An integrative mirna-mrna expression analysis reveals striking transcriptomic similarities between severe equine asthma and specific asthma endotypes in humans. *Genes* 2020, *11*, 1143. [CrossRef]
- 9. Pacholewska, A.; Kraft, M.F.; Gerber, V.; Jagannathan, V. Differential expression of serum MicroRNAs supports CD4⁺ t cell differentiation into Th2/Th17 cells in severe equine asthma. *Genes* **2017**, *8*, 383. [CrossRef]
- 10. Padoan, E.; Ferraresso, S.; Pegolo, S.; Castagnaro, M.; Barnini, C.; Bargelloni, L. Real time RT-PCR analysis of inflammatory mediator expression in recurrent airway obstruction-affected horses. *Vet. Immunol. Immunopathol.* **2013**, *156*, 190–199. [CrossRef]
- 11. Ainsworth, D.M.; Wagner, B.; Franchini, M.; Grünig, G.; Erb, H.N.; Tan, J.Y. Time-dependent alterations in gene expression of interleukin-8 in the bronchial epithelium of horses with recurrent airway obstruction. *Am. J. Vet. Res.* **2006**, *67*, 669–677. [CrossRef]

Animals 2023, 13, 3872 12 of 17

12. Simões, J.; Batista, M.; Tilley, P. The Immune Mechanisms of Severe Equine Asthma—Current Understanding and What Is Missing. *Animals* **2022**, *12*, 744. [CrossRef]

- 13. Marti, E.; Gerber, H.; Essich, G.; Oulehla, J.; Lazary, S. The genetic basis of equine allergic diseases 1. Chronic hypersensitivity bronchitis. *Equine Vet. J.* **1991**, 23, 457–460. [CrossRef]
- 14. Gerber, V.; Baleri, D.; Klukowska-Rötzler, J.; Swinburne, J.E.; Dolf, G. Mixed inheritance of equine recurrent airway obstruction. *J. Vet. Intern. Med.* **2009**, 23, 626–630. [CrossRef]
- 15. Ramseyer, A.; Gaillard, C.; Burger, D.; Straub, R.; Jost, U.; Boog, C.; Marti, E.; Gerber, V. Effects of Genetic and Environmental Factors on Chronic Lower Airway Disease in Horses. *J. Vet. Intern. Med.* **2007**, *21*, 149–156. [CrossRef]
- 16. Schnider, D.; Rieder, S.; Leeb, T.; Gerber, V.; Neuditschko, M. A genome-wide association study for equine recurrent airway obstruction in European Warmblood horses reveals a suggestive new quantitative trait locus on chromosome 13. *Anim. Genet.* **2017**, *48*, 691–693. [CrossRef]
- 17. Tilley, P.; Sales Luis, J.P.; Branco Ferreira, M. Correlation and discriminant analysis between clinical, endoscopic, thoracic X-ray and bronchoalveolar lavage fluid cytology scores, for staging horses with recurrent airway obstruction (RAO). *Res. Vet. Sci.* 2012, 93, 1006–1014. [CrossRef]
- 18. Simões, J.; Sales Luís, J.; Tilley, P. Contribution of lung function tests to the staging of severe equine asthma syndrome in the field. *Res. Vet. Sci.* **2019**, *123*, 112–117. [CrossRef]
- 19. Moore-Colyer, M.J.S.; Taylor, J.L.E.; James, R. The Effect of Steaming and Soaking on the Respirable Particle, Bacteria, Mould, and Nutrient Content in Hay for Horses. *J. Equine Vet. Sci.* **2016**, *39*, *62*–68. [CrossRef]
- 20. White, S.J.; Moore-Colyer, M.; Marti, E.; Hannant, D.; Gerber, V.; Coüetil, L.; Richard, E.A.; Alcocer, M. Antigen array for serological diagnosis and novel allergen identification in severe equine asthma. *Sci. Rep.* **2019**, *9*, 15171. [CrossRef]
- 21. Pirie, R.S.; Collie, D.D.S.; Dixon, P.M.; McGorum, B.C. Inhaled endotoxin and organic dust particulates have synergistic proinflammatory effects in equine heaves (organic dust-induced asthma). *Clin. Exp. Allergy* **2003**, *33*, 676–683. [CrossRef]
- 22. Costa, L.R.R.; Johnson, J.R.; Baur, M.E.; Beadle, R.E. Temporal clinical exacerbation of summer pasture-associated recurrent airway obstruction and relationship with climate and aeroallergens in horses. *Am. J. Vet. Res.* **2006**, *67*, 1635–1642. [CrossRef]
- 23. Bullone, M.; Murcia, R.Y.; Lavoie, J.P. Environmental heat and airborne pollen concentration are associated with increased asthma severity in horses. *Equine Vet. J.* **2016**, *48*, 479–484. [CrossRef]
- 24. Ferrari, C.R.; Cooley, J.; Mujahid, N.; Costa, L.R.; Wills, R.W.; Johnson, M.E.; Swiderski, C.E. Horses With Pasture Asthma Have Airway Remodeling That Is Characteristic of Human Asthma. *Vet. Pathol.* **2018**, *55*, 144–158. [CrossRef]
- 25. McGorum, B.C.; Ellison, J.; Cullen, R.T. Total and respirable airborne dust endotoxin concentrations in three equine management systems. *Equine Vet. J.* **1998**, *30*, 430–434. [CrossRef]
- 26. Niedzwiedz, A.; Jaworski, Z.; Kubiak, K. Serum concentrations of allergen-specific IgE in horses with equine recurrent airway obstruction and healthy controls assessed by ELISA. *Vet. Clin. Pathol.* **2015**, 44, 391–396. [CrossRef]
- 27. Pacholewska, A.; Jagannathan, V.; Drögemüller, M.; Klukowska-Rötzler, J.; Lanz, S.; Hamza, E.; Dermitzakis, E.T.; Marti, E.; Leeb, T.; Gerber, V. Impaired cell cycle regulation in a natural equine model of asthma. *PLoS ONE* **2015**, *10*, e0136103. [CrossRef]
- 28. Klier, J.; Geis, S.; Steuer, J.; Geh, K.; Reese, S.; Fuchs, S.; Mueller, R.S.; Winter, G.; Gehlen, H. A comparison of nanoparticullate CpG immunotherapy with and without allergens in spontaneously equine asthma-affected horses, an animal model. *Immun. Inflamm. Dis.* **2018**, *6*, 81–96. [CrossRef]
- 29. Simões, J.; Sales Luís, J.P.; Tilley, P. Owner Compliance to an Environmental Management Protocol for Severe Equine Asthma Syndrome. *J. Equine Vet. Sci.* **2020**, *87*, 102937. [CrossRef]
- 30. Kavanagh, J.; Jackson, D.J.; Kent, B.D. Over-and under-diagnosis in asthma. Breathe 2019, 15, e20–e27. [CrossRef]
- 31. Masic, I.; Miokovic, M.; Muhamedagic, B. Evidence Based Medicine—New Approaches and Challenges. *Acta Inform. Medica* **2008**, 16, 219. [CrossRef]
- 32. Guilleminault, L.; Ouksel, H.; Belleguic, C.; Le Guen, Y.; Germaud, P.; Desfleurs, E.; Leroyer, C.; Magnan, A. Personalised medicine in asthma: From curative to preventive medicine. *Eur. Respir. Rev.* **2017**, *26*, 160010. [CrossRef]
- 33. Claußen, G.; Hessel, E.F. Particulate Matter in Equestrian Stables and Riding Arenas. J. Equine Vet. Sci. 2017, 55, 60–70. [CrossRef]
- 34. Lühe, T.; Volkmann, N.; Probst, J.; Dreyer-Rendelsmann, C.; Schulz, J.; Kemper, N. Bacterial Burden in the Air of Indoor Riding Arenas. *Agriculture* **2022**, *12*, 2111. [CrossRef]
- 35. Riihimäki, M.; Raine, A.; Elfman, L.; Pringle, J. Markers of respiratory inflammation in horses in relation to seasonal changes in air quality in a conventional racing stable. *Can. J. Vet. Res.* **2008**, 72, 432–439.
- 36. Mönki, J.; Saastamoinen, M.; Karikoski, N.; Norring, M.; Rajamäki, M.; Mykkänen, A. Effects of Bedding Material on Equine Lower Airway Inflammation: A Comparison of Two Peat Beddings, Wood Pellet, and Straw Pellet. *Front. Vet. Sci.* **2021**, *8*, 289. [CrossRef]
- Orard, M.; Hue, E.; Couroucé, A.; Bizon-Mercier, C.; Toquet, M.P.; Moore-Colyer, M.; Couëtil, L.; Pronost, S.; Paillot, R.; Demoor, M.; et al. The influence of hay steaming on clinical signs and airway immune response in severe asthmatic horses. *BMC Vet. Res.* **2018**, *14*, 345. [CrossRef]
- 38. Wasko, A.J.; Barkema, H.W.; Nicol, J.; Fernandez, N.; Logie, N.; Léguillette, R. Evaluation of a risk-screening questionnaire to detect equine lung inflammation: Results of a large field study. *Equine Vet. J.* **2011**, *43*, 145–152. [CrossRef]
- 39. Laumen, E.; Doherr, M.G.; Gerber, V. Relationship of horse owner assessed respiratory signs index to characteristics of recurrent airway obstruction in two Warmblood families. *Equine Vet. J.* **2010**, 42, 142–148. [CrossRef]

Animals 2023, 13, 3872 13 of 17

40. Holcombe, S.J.; Jackson, C.; Gerber, V.; Jefcoat, A.; Berney, C.; Eberhardt, S.; Robinson, N.E. Stabling is associated with airway inflammation in young Arabian horses. *Equine Vet. J.* **2001**, *33*, 244–249. [CrossRef]

- 41. Klier, J.; Lindner, D.; Reese, S.; Mueller, R.S.; Gehlen, H. Comparison of Four Different Allergy Tests in Equine Asthma Affected Horses and Allergen Inhalation Provocation Test. *J. Equine Vet. Sci.* **2021**, *102*, 103433. [CrossRef]
- 42. Tilley, P.; Sales Luis, J.P.; Branco Ferreira, M. Comparison of Skin Prick Tests with In Vitro Allergy Tests in the Characterization of Horses with Recurrent Airway Obstruction. *J. Equine Vet. Sci.* **2012**, *32*, 719–727. [CrossRef]
- 43. Lo Feudo, C.M.; Stucchi, L.; Alberti, E.; Conturba, B.; Zucca, E.; Ferrucci, F. Intradermal testing results in horses affected by mild-moderate and severe equine asthma. *Animals* **2021**, *11*, 2086. [CrossRef] [PubMed]
- 44. Couëtil, L.L.; Rosenthal, F.S.; DeNicola, D.B.; Chilcoat, C.D. Clinical signs, evaluation of bronchoalveolar lavage fluid, and assessment of pulmonary function in horses with inflammatory respiratory disease. *Am. J. Vet. Res.* **2001**, *62*, 538–546. [CrossRef] [PubMed]
- 45. Bosshard, S.; Gerber, V. Evaluation of coughing and nasal discharge as early indicators for an increased risk to develop equine recurrent airway obstruction (RAO). *J. Vet. Intern. Med.* **2014**, *28*, 618–623. [CrossRef] [PubMed]
- 46. Rettmer, H.; Hoffman, A.M.; Lanz, S.; Oertly, M.; Gerber, V. Owner-reported coughing and nasal discharge are associated with clinical findings, arterial oxygen tension, mucus score and bronchoprovocation in horses with recurrent airway obstruction in a field setting. *Equine Vet. J.* 2015, 47, 291–295. [CrossRef]
- 47. Bedenice, D.; Mazan, M.R.; Hoffman, A.M. Association between cough and cytology of bronchoalveolar lavage fluid and pulmonary function in horses diagnosed with inflammatory airway disease. *J. Vet. Intern. Med.* 2008, 22, 1022–1028. [CrossRef]
- 48. Janssen, P.; Tosi, I.; Hego, A.; Maréchal, P.; Marichal, T.; Radermecker, C. Neutrophil Extracellular Traps Are Found in Bronchoalveolar Lavage Fluids of Horses With Severe Asthma and Correlate With Asthma Severity. *Front. Immunol.* **2022**, *13*, 921077. [CrossRef]
- 49. Lo Feudo, C.M.; Stucchi, L.; Conturba, B.; Stancari, G.; Ferrucci, F. Impact of Lower Airway Inflammation on Fitness Parameters in Standardbred Racehorses. *Animals* **2022**, *12*, 3228. [CrossRef]
- 50. Robinson, N.E.; Derksen, F.J.; Jackson, C.A.; Peroni, D.; Gerber, V. Management of heaves. *Equine Vet. Educ.* **2001**, *13*, 247–259. [CrossRef]
- 51. Robinson, N.E.; Berney, C.; deFeijter-Rupp, H.L.; Jefcoat, A.M.; Cornelisse, C.J.; Gerber, V.M.; Derksen, F.J. Coughing, mucus accumulation, airway obstruction, and airway inflammation in control horses and horses affected with recurrent airway obstruction. *Am. J. Vet. Res.* 2003, 64, 550–557. [CrossRef] [PubMed]
- 52. Robinson, N.E.; Derksen, F.J.; Olszewski, M.A.; Buechner-Maxwell, V.A. The pathogenesis of chronic obstructive pulmonary disease of horses. *Br. Vet. J.* **1996**, *152*, 283–306. [CrossRef] [PubMed]
- 53. Robinson, N.E.; Olszewski, M.A.; Boehler, D.; Berney, C.; Hakala, J.; Matson, C.; Derksen, F.J. Relationship between clinical signs and lung function in horses with recurrent airway obstruction (heaves) during a bronchodilator trial. *Equine Vet. J.* **2000**, 32, 393–400. [CrossRef] [PubMed]
- 54. Gerber, V.; King, M.; Schneider, D.A.; Robinson, N.E. Tracheobronchial mucus viscoelasticity during environmental challenge in horses with recurrent airway obstruction. *Equine Vet. J.* **2000**, *32*, 411–417. [CrossRef] [PubMed]
- 55. Gerber, V.; Straub, R.; Marti, E.; Hauptman, J.; Herholz, C.; King, M.; Imhof, A.; Tahon, L.; Robinson, N.E. Endoscopic scoring of mucus quantity and quality: Observer and horse variance and relationship to inflammation, mucus viscoelasticity and volume. *Equine Vet. J.* **2004**, *36*, 576–582. [CrossRef]
- 56. Pirie, R.S. Recurrent airway obstruction: A review. Equine Vet. J. 2014, 46, 276–288. [CrossRef]
- 57. Couëtil, L.L.; Hawkins, J.F. Respiratory Diseases of the Horse: A Problem-Oriented Approach to Diagnosis & Management; CRC Press: Boca Raton, FL, USA, 2013; ISBN 9781840766479.
- 58. Theegarten, D.; Sachse, K.; Mentrup, B.; Fey, K.; Hotzel, H.; Anhenn, O. Chlamydophila spp. infection in horses with recurrent airway obstruction: Similarities to human chronic obstructive disease. *Respir. Res.* **2008**, *9*, 14. [CrossRef]
- 59. Juhn, Y.J. Risks for infection in patients with asthma (or other atopic conditions): Is asthma more than a chronic airway disease? *J. Allergy Clin. Immunol.* **2014**, 134, 247–257. [CrossRef]
- 60. Naylor, J.M.; Clark, E.G.; Clayton, H.M. Chronic obstructive pulmonary disease: Usefulness of clinical signs, bronchoalveolar lavage, and lung biopsy as diagnostic and prognostic aids. *Can. Vet. J.* **1992**, *33*, 591–598.
- 61. Couëtil, L.L.; Ward, M.P. Analysis of risk factors for recurrent airway obstruction in North American horses: 1444 Cases (1990–1999). J. Am. Vet. Med. Assoc. 2003, 223, 1645–1650. [CrossRef]
- 62. Mazan, M.R.; Deveney, E.F.; DeWitt, S.; Bedenice, D.; Hoffman, A. Energetic cost of breathing, body composition, and pulmonary function in horses with recurrent airway obstruction. *J. Appl. Physiol.* **2004**, *97*, 91–97. [CrossRef] [PubMed]
- 63. Miskovic, M.; Couëtil, L.L.L.; Thompson, C.A.A. Lung function and airway cytologic profiles in horses with recurrent airway obstruction maintained in low-dust environments. *J. Vet. Intern. Med.* **2007**, *21*, 1060–1066. [CrossRef] [PubMed]
- 64. Gerber, V.; Robinson, N.E.; Venta, P.J.; Rawson, J.; Jefcoat, A.M.; Hotchkiss, J.A. Mucin genes in horse airways: MUC5AC, but not MUC2, may play a role in recurrent airway obstruction. *Equine Vet. J.* **2003**, *35*, 252–257. [CrossRef] [PubMed]
- 65. Rush, B.R.; Raub, E.S.; Rhoads, W.S.; Flaminio, M.J.B.F.; Matson, C.J.; Hakala, J.E.; Gillespie, J.R. Pulmonary function in horses with recurrent airway obstruction after aerosol and parenteral administration of beclomethasone dipropionate and dexamethasone, respectively. *Am. J. Vet. Res.* **1998**, *59*, 1039–1043. [PubMed]

Animals 2023, 13, 3872 14 of 17

66. Rush, B.R.; Flaminio, M.J.B.F.; Matson, C.J.; Hakala, J.E.; Shuman, W. Cytologic evaluation of bronchoalveolar lavage fluid from horses with recurrent airway obstruction after aerosol and parenteral administration of beclomethasone dipropionate and dexamethasone, respectively. *Am. J. Vet. Res.* **1998**, *59*, 1033–1038. [PubMed]

- 67. Robinson, N.E.; Berney, C.; Behan, A.; Derksen, F.J. Fluticasone propionate aerosol is more effective for prevention than treatment of recurrent airway obstruction. *J. Vet. Intern. Med.* **2009**, 23, 1247–1253. [CrossRef] [PubMed]
- 68. Tesarowski, D.B.; Viel, L.; McDonell, W.N. Pulmonary function measurements during repeated environmental challenge of horses with recurrent airway obstruction (heaves). *Am. J. Vet. Res.* **1996**, *57*, 1214–1219.
- 69. Pirie, R.S.; Mueller, H.W.; Engel, O.; Albrecht, B.; von Salis-Soglio, M. Inhaled ciclesonide is efficacious and well tolerated in the treatment of severe equine asthma in a large prospective European clinical trial. *Equine Vet. J.* **2021**, 53, 1094–1104. [CrossRef]
- 70. Gerber, V.; Schott, H.C.; Robinson, N.E. Owner assessment in judging the efficacy of airway disease treatment. *Equine Vet. J.* **2011**, 43, 153–158. [CrossRef]
- 71. Kozłowska, N.; Wierzbicka, M.; Jasiński, T.; Domino, M. Advances in the Diagnosis of Equine Respiratory Diseases: A Review of Novel Imaging and Functional Techniques. *Animals* **2022**, *12*, 381. [CrossRef]
- 72. Ribonnet, C.; Palmers, K.; Saegerman, C.; Vanderperren, K.; van Galen, G. Perioperative lung ultrasonography in healthy horses undergoing general anesthesia for elective surgery. *J. Vet. Intern. Med.* **2022**, *36*, 1160–1172. [CrossRef] [PubMed]
- 73. Bakos, Z. Digital luminescence thoracic radiography in horses with recurrent airway obstruction. *Vet. Rec.* **2008**, *162*, 122–124. [CrossRef] [PubMed]
- 74. Lavoie, J.-P.; Dalle, S.; Breton, L.; Hélie, P. Bronchiectasis in Three Adult Horses with Heaves. *J. Vet. Intern. Med.* **2004**, *18*, 757–760. [CrossRef] [PubMed]
- 75. Allen, K.; Franklin, S. RAO and IAD: Respiratory disease in horses revisited. Practice 2007, 29, 76–82. [CrossRef]
- 76. Mazan, M.R.; Vin, R.; Hoffman, A.M. Radiographic scoring lacks predictive value in inflammatory airway disease. *Equine Vet. J.* **2005**, *37*, 541–545. [CrossRef] [PubMed]
- 77. Farrow, C.S. Radiography of the Equine Thorax: Anatomy and Technic. Vet. Radiol. 1981, 22, 62–68. [CrossRef]
- 78. Barton, A.K.; Schulze, T.; Doherr, M.G.; Gehlen, H. Influence of bronchoalveolar lavage on thoracic radiography in the horse. *J. Vet. Sci.* **2018**, *19*, 563–569. [CrossRef] [PubMed]
- 79. Bakos, Z.; Vörös, K.; Kellokoski, H.; Reiczigel, J. Comparison of the caudal lung borders determined by percussion and ultrasonography in horses with recurrent airway obstruction. *Acta Vet. Hung.* **2003**, *51*, 249–258. [CrossRef]
- 80. Siwinska, N.; Zak, A.; Slowikowska, M.; Krupinska, P.; Niedzwiedz, A. Prevalence and severity of ultrasonographic pulmonary findings in horses with asthma—A preliminary study. *Pol. J. Vet. Sci.* **2019**, 22, 653–659. [CrossRef]
- 81. Lo Feudo, C.M.; Stucchi, L.; Alberti, E.; Stancari, G.; Conturba, B.; Zucca, E.; Ferrucci, F. The role of thoracic ultrasonography and airway endoscopy in the diagnosis of equine asthma and exercise-induced pulmonary hemorrhage. *Vet. Sci.* **2021**, *8*, 276. [CrossRef]
- 82. Bullone, M.; Beauchamp, G.; Godbout, M.; Martin, J.G.; Lavoie, J.P. Endobronchial ultrasound reliably quantifies airway smooth muscle remodeling in an equine asthma model. *PLoS ONE* **2015**, *10*, e0136284. [CrossRef] [PubMed]
- 83. Bullone, M.; Lavoie, J.P. Science-in-brief: Equine asthma diagnosis: Beyond bronchoalveolar lavage cytology. *Equine Vet. J.* **2017**, 49, 263–265. [CrossRef] [PubMed]
- 84. Wysocka, B.; Kluciński, W. Cytological evaluation of tracheal aspirate and broncho-alveolar lavage fluid in comparison to endoscopic assessment of lower airways in horses with recurrent airways obstruction or inflammatory airway disease. *Pol. J. Vet. Sci.* 2015, 18, 587–597. [CrossRef] [PubMed]
- 85. Feudo, C.M.L.; Stancari, G.; Collavo, F.; Stucchi, L.; Conturba, B.; Zucca, E.; Ferrucci, F. Upper and Lower Airways Evaluation and Its Relationship with Dynamic Upper Airway Obstruction in Racehorses. *Animals* **2022**, *12*, 1563. [CrossRef] [PubMed]
- 86. Dixon, P.M.; Railton, D.I.; McGorum, B.C. Equine pulmonary disease: A case control study of 300 referred cases. Part 2: Details of animals and of historical and clinical findings. *Equine Vet. J.* 1995, 27, 422–427. [CrossRef] [PubMed]
- 87. Hare, J.E.; Viel, L. Pulmonary eosinophilia associated with increased airway responsiveness in young racing horses. *J. Vet. Intern. Med.* **1998**, 12, 163–170. [CrossRef]
- 88. Koch, C.; Straub, R.; Ramseyer, A.; Widmer, A.; Robinson, N.E.; Gerber, V. Endoscopic scoring of the tracheal septum in horses and its clinical relevance for the evaluation of lower airway health in horses. *Equine Vet. J.* **2007**, *39*, 107–112. [CrossRef]
- 89. Kutasi, O.; Balogh, N.; Lajos, Z.; Nagy, K.; Szenci, O. Diagnostic Approaches for the Assessment of Equine Chronic Pulmonary Disorders. *J. Equine Vet. Sci.* **2011**, *31*, 400–410. [CrossRef]
- 90. Koblinger, K.; Nicol, J.; Mcdonald, K.; Wasko, A.; Logie, N.; Weiss, M.; Léguillette, R. Endoscopic Assessment of Airway Inflammation in Horses. *J. Vet. Intern. Med.* **2011**, 25, 1118–1126. [CrossRef]
- 91. Gerber, V.; Lindberg, Å.; Berney, C.; Robinson, N.E. Airway Mucus in Recurrent Airway Obstruction-Short-Term Response to Environmental Challenge. *J. Vet. Intern. Med.* **2004**, *18*, 92–97. [CrossRef]
- 92. Rodrigues Costa, L.R.; Seahorn, T.L.; Moore, R.M.; Taylor, H.W.; Gaunt, S.D.; Beadle, R.E. Correlation of clinical score, intrapleural pressure, cytologic findings of bronchoalveolar fluid, and histopathologic lesions of pulmonary tissue in horses with summer pasture-associated obstructive pulmonary disease. *Am. J. Vet. Res.* 2000, *61*, 167–173. [CrossRef] [PubMed]
- 93. Mazan, M.R.; Hoffman, A.M. Clinical techniques for diagnosis of inflammatory airway disease in the horse. *Clin. Tech. Equine Pract.* **2003**, 2, 238–257. [CrossRef]
- 94. GINA. Global Strategy for Asthma Management and Prevention; GINA: Fontana, WI, USA, 2023.

Animals 2023, 13, 3872 15 of 17

95. Chevalier, H.; Divers, T.J. Pulmonary dysfunction in adult horses in the intensive care unit. *Clin. Tech. Equine Pract.* **2003**, 2, 165–177. [CrossRef]

- 96. Nuytten, J.; Deprez, P.; Picavet, T.; Van Den Hende, C.; Muylle, E. Comparison of different pulmonary function tests and their prognostic value in horses affected with COPD. *J. Equine Vet. Sci.* **1988**, *8*, 361–364. [CrossRef]
- 97. Couëtil, L.L.; Denicola, D.B. Blood gas, plasma lactate and bronchoalveolar lavage cytology analyses in racehorses with respiratory disease. *Equine Vet. J. Suppl.* **1999**, *30*, 77–82. [CrossRef] [PubMed]
- 98. Sánchez, A.; Couëtil, L.L.; Ward, M.P.; Clark, S.P. Effect of Airway Disease on Blood Gas Exchange in Racehorses. *J. Vet. Intern. Med.* 2005, 19, 87–92. [CrossRef]
- 99. Stopyra, A.; Sobiech, P.; Wacławska-Matyjasik, A. Acid-base indicators in the venous and arterial blood of horses affected by recurrent airway obstruction (RAO). *Pol. J. Vet. Sci.* **2012**, *15*, 463–467. [CrossRef]
- 100. Lavoie, J.P.; Leclere, M.; Rodrigues, N.; Lemos, K.R.; Bourzac, C.; Lefebvre-Lavoie, J.; Beauchamp, G.; Albrecht, B. Efficacy of inhaled budesonide for the treatment of severe equine asthma. *Equine Vet. J.* **2019**, *51*, 401–407. [CrossRef]
- 101. Derksen, F.J.; Slocombe, R.F.; Brown, C.M.; Rook, J.; Robinson, N.E. Chronic restrictive pulmonary disease in a horse. *J. Am. Vet. Med. Assoc.* **1982**, *180*, 887–889.
- 102. Doucet, M.Y.; Vrins, A.A.; Ford-Hutchinson, A.W. Histamine inhalation challenge in normal horses and in horses with small airway disease. *Can. J. Vet. Res.* **1991**, *55*, 285–293.
- 103. Hoffman, A.M.; Oura, T.J.; Riedelberger, K.J.; Mazan, M.R. Plethysmographic comparison of breathing pattern in heaves (recurrent airway obstruction) versus experimental bronchoconstriction or hyperpnea in horses. *J. Vet. Intern. Med.* **2007**, 21, 184–192. [CrossRef] [PubMed]
- 104. Nolen-Walston, R.D.; Kuehn, H.; Boston, R.C.; Mazan, M.R.; Wilkins, P.A.; Bruns, S.; Hoffman, A.M. Reproducibility of airway responsiveness in horses using flowmetric plethysmography and histamine bronchoprovocation. *J. Vet. Intern. Med.* **2009**, 23, 631–635. [CrossRef] [PubMed]
- 105. Wichtel, M.; Gomez, D.; Burton, S.; Wichtel, J.; Hoffman, A. Relationships between equine airway reactivity measured by flowmetric plethysmography and specific indicators of airway inflammation in horses with suspected inflammatory airway disease. *Equine Vet. J.* **2016**, *48*, 466–471. [CrossRef] [PubMed]
- 106. Dixon, C.E.; Bedenice, D.; Mazan, M.R. Comparison of Flowmetric Plethysmography and Forced Oscillatory Mechanics to Measure Airway Hyperresponsiveness in Horses. *Front. Vet. Sci.* **2021**, *7*, 511023. [CrossRef] [PubMed]
- 107. Derksen, F.J.; Robinson, N.E.; Armstrong, P.J.; Stick, J.A.; Slocombe, R.F. Airway reactivity in ponies with recurrent airway obstruction (heaves). *J. Appl. Physiol.* **1985**, *58*, 598–604. [CrossRef] [PubMed]
- 108. Vandenput, S.; Votion, D.; Duvivier, D.H.; Van Erck, E.; Anciaux, N.; Art, T.; Lekeux, P. Effect of a set stabled environmental control on pulmonary function and airway reactivity of COPD affected horses. *Vet. J.* 1998, 155, 189–195. [CrossRef] [PubMed]
- 109. Mazan, M.R.; Huffman, A.M.; Manjerovic, N. Comparison of forced oscillation with the conventional method for histamine bronchoprovocation testing in horses. *Am. J. Vet. Res.* **1999**, *60*, 174–180.
- 110. Burnheim, K.; Hughes, K.J.; Evans, D.L.; Raidal, S.L. Reliability of breath by breath spirometry and relative flow-time indices for pulmonary function testing in horses. *BMC Vet. Res.* **2016**, *12*, 268. [CrossRef]
- 111. Herholz, C.; Straub, R.; Braendlin, C.; Imhof, A.; Lüthi, S.; Busato, A. Measurement of tidal breathing flow-volume loop indices in horses used for different sporting purposes with and without recurrent airway obstruction. *Vet. Rec.* 2003, 152, 288–292. [CrossRef]
- 112. Raidal, S.L.; Burnheim, K.; Evans, D.; Hughes, K.J. Effects of sedation and salbutamol administration on hyperpnoea and tidal breathing spirometry in healthy horses. *Vet. J.* 2017, 222, 22–28. [CrossRef]
- 113. Secombe, C.; Adler, A.; Hosgood, G.; Raisis, A.; Mosing, M. Can bronchoconstriction and bronchodilatation in horses be detected using electrical impedance tomography? *J. Vet. Intern. Med.* **2021**, *35*, 2035–2044. [CrossRef] [PubMed]
- 114. Van Erck, E.; Votion, D.; Art, T.; Lekeux, P. Measurement of respiratory function by impulse oscillometry in horses. *Equine Vet. J.* **2004**, *36*, 21–28. [CrossRef] [PubMed]
- 115. Van Erck, E.; Votion, D.; Kirschvink, N.; Genicot, B.; Lindsey, J.; Art, T.; Lekeux, P. Influence of breathing pattern and lung inflation on impulse oscillometry measurements in horses. *Vet. J.* **2004**, *168*, 259–269. [CrossRef]
- 116. Richard, E.A.; Fortier, G.D.; Denoix, J.M.; Art, T.; Lekeux, P.M.; van Erck, E. Influence of subclinical inflammatory airway disease on equine respiratory function evaluated by impulse oscillometry. *Equine Vet. J.* **2009**, *41*, 384–389. [CrossRef] [PubMed]
- 117. Stucchi, L.; Ferrucci, F.; Bullone, M.; Dellacà, R.L.; Lavoie, J.P. Within-breath oscillatory mechanics in horses affected by severe equine asthma in exacerbation and in remission of the disease. *Animals* **2022**, *12*, 4. [CrossRef] [PubMed]
- 118. Lee, G.K.C.; Beeler-Marfisi, J.; Viel, L.; Piché, É.; Kang, H.; Sears, W.; Bienzle, D. Bronchial brush cytology, endobronchial biopsy, and SALSA immunohistochemistry in severe equine asthma. *Vet. Pathol.* **2022**, *59*, 100–111. [CrossRef] [PubMed]
- 119. Niedzwiedz, A.; Mordak, R.; Jaworski, Z.; Nicpon, J. Utility of the Histological Examination of the Bronchial Mucosa in the Diagnosis of Severe Equine Asthma Syndrome in Horses. *J. Equine Vet. Sci.* **2018**, *67*, 44–49. [CrossRef]
- 120. Hoffman, A.M. Bronchoalveolar lavage technique and cytological diagnosis of small airway inflammatory disease. *Equine Vet. Educ.* **1999**, *11*, 330–336. [CrossRef]
- 121. Dauvillier, J.; ter Woort, F.; van Erck-Westergren, E. Fungi in respiratory samples of horses with inflammatory airway disease. *J. Vet. Intern. Med.* **2019**, *33*, 968–975. [CrossRef]

Animals 2023, 13, 3872 16 of 17

122. Hoffman, A.M.; Mazan, M.R. Programme of lung function testing horses suspected with small airway disease. *Equine Vet. Educ.* **1999**, *11*, 322–328. [CrossRef]

- 123. Malikides, N.; Hughes, K.J.; Hodgson, D.R.; Hodgson, J.L. Comparison of tracheal aspirates and bronchoalveolar lavage in racehorses 2. Evaluation of the diagnostic significance of neutrophil percentage. *Aust. Vet. J.* **2003**, *81*, 685–687. [CrossRef] [PubMed]
- 124. Hermange, T.; Le Corre, S.; Bizon, C.; Richard, E.A.; Couroucé, A. Bronchoalveolar lavage fluid from both lungs in horses: Diagnostic reliability of cytology from pooled samples. *Vet. J.* **2019**, 244, 28–33. [CrossRef] [PubMed]
- 125. Fernandez, N.J.; Hecker, K.G.; Gilroy, C.V.; Warren, A.L.; Léguillette, R. Reliability of 400-cell and 5-field leukocyte differential counts for equine bronchoalveolar lavage fluid. *Vet. Clin. Pathol.* **2013**, 42, 92–98. [CrossRef] [PubMed]
- 126. Fairbairn, S.M.; Page, C.P.; Lees, P.; Cunningham, F.M. Early neutrophil but not eosinophil or platelet recruitment to the lungs of allergic horses following antigen exposure. *Clin. Exp. Allergy* 1993, 23, 821–828. [CrossRef] [PubMed]
- 127. Bullone, M.; Joubert, P.; Gagné, A.; Lavoie, J.P.; Hélie, P. Bronchoalveolar lavage fluid neutrophilia is associated with the severity of pulmonary lesions during equine asthma exacerbations. *Equine Vet. J.* **2018**, *50*, 609–615. [CrossRef] [PubMed]
- 128. Couetil, L.L.; Thompson, C.A. Airway Diagnostics: Bronchoalveolar Lavage, Tracheal Wash, and Pleural Fluid. *Vet. Clin. N. Am. Equine Pract.* **2020**, *36*, 87–103. [CrossRef]
- 129. Hoffman, A.M. Bronchoalveolar Lavage: Sampling Technique and Guidelines for Cytologic Preparation and Interpretation. *Vet. Clin. N. Am. Equine Pract.* **2008**, 24, 423–435. [CrossRef]
- 130. Moran, G.; Araya, O.; Ortloff, A.; Folch, H. Cytologic broncheoalveolar lavage findings and humoral immune response against Aspergillus fumigatus in Chilote horses with recurrent airway obstructions "heaves". *Arch. Med. Vet.* **2009**, *41*, 83–88.
- 131. Fairbairn, S.M.; Lees, P.; Page, C.P.; Cunningham, F.M. Duration of antigen-induced hyperresponsiveness in horses with allergic respiratory disease and possible links with early airway obstruction. *J. Vet. Pharmacol. Ther.* **1993**, *16*, 469–476. [CrossRef]
- 132. Couëtil, L.L.; Art, T.; De Moffarts, B.; Becker, M.; Mélotte, D.; Jaspar, F.; Bureau, F.; Lekeux, P. Effect of beclomethasone dipropionate and dexamethasone isonicotinate on lung function, bronchoalveolar lavage fluid cytology, and transcription factor expression in airways of horses with recurrent airway obstruction. *J. Vet. Intern. Med.* 2006, 20, 399–406. [CrossRef]
- 133. Leclere, M.; Lavoie-Lamoureux, A.; Joubert, P.; Relave, F.; Setlakwe, E.L.; Beauchamp, G.; Couture, C.; Martin, J.G.; Lavoie, J.P. Corticosteroids and antigen avoidance decrease airway smooth muscle mass in an equine asthma model. *Am. J. Respir. Cell Mol. Biol.* 2012, 47, 589–596. [CrossRef] [PubMed]
- 134. Léguillette, R.; Lavoie, J.P. Effects of the bronchoalveolar lavage procedure on lung function in horses with clinical exacervation of recurrent airway obstruction. *Am. J. Vet. Res.* **2006**, *67*, 1929–1933. [CrossRef] [PubMed]
- 135. Kinnison, T.; Cardwell, J.M. Conflict Between Direct Experience and Research-Based Evidence Is a Key Challenge to Evidence-Based Respiratory Medicine on British Racing Yards. *Front. Vet. Sci.* **2020**, *7*, 266. [CrossRef] [PubMed]
- 136. Derksen, F.J.; Brown, C.M.; Sonea, I.; Darien, B.J.; Robinson, N.E. Comparison of transtracheal aspirate and bronchoalveolar lavage cytology in 50 horses with chronic lung disease. *Equine Vet. J.* **1989**, 21, 23–26. [CrossRef] [PubMed]
- 137. Fraipont, A.; Van Erck, E.; Ramery, E.; Richard, E.; Denoix, J.M.; Lekeux, P.; Art, T. Subclinical diseases underlying poor performance in endurance horses: Diagnostic methods and predictive tests. *Vet. Rec.* **2011**, *169*, 154. [CrossRef] [PubMed]
- 138. Rossi, H.; Virtala, A.M.; Raekallio, M.; Rahkonen, E.; Rajamäki, M.M.; Mykkänen, A. Comparison of tracheal wash and bronchoalveolar lavage cytology in 154 horses with and without respiratory signs in a referral hospital over 2009–2015. *Front. Vet. Sci.* 2018, 5, 61. [CrossRef] [PubMed]
- 139. Couëtil, L.L.; Art, T.; de Moffarts, B.; Becker, M.; Mélotte, D.; Jaspar, F.; Bureau, F.; Lekeux, P. DNA binding activity of transcription factors in bronchial cells of horses with recurrent airway obstruction. *Vet. Immunol. Immunopathol.* 2006, 113, 11–20. [CrossRef]
- 140. Künzle, F.; Gerber, V.; Van Der Haegen, A.; Wampfler, B.; Straub, R.; Marti, E. IgE-bearing cells in bronchoalveolar lavage fluid and allergen-specific IgE levels in sera from RAO-affected horses. *J. Vet. Med. Ser. A Physiol. Pathol. Clin. Med.* **2007**, 54, 40–47. [CrossRef]
- 141. Tahon, L.; Baselgia, S.; Gerber, V.; Doherr, M.G.; Straub, R.; Robinson, N.E.; Marti, E. In vitro allergy tests compared to intradermal testing in horses with recurrent airway obstruction. *Vet. Immunol. Immunopathol.* **2009**, 127, 85–93. [CrossRef]
- 142. Hansen, S.; Otten, N.D.; Birch, K.; Skovgaard, K.; Hopster-Iversen, C.; Fjeldborg, J. Bronchoalveolar lavage fluid cytokine, cytology and IgE allergen in horses with equine asthma. *Vet. Immunol. Immunopathol.* **2020**, 220, 109976. [CrossRef]
- 143. Verdon, M.; Lanz, S.; Rhyner, C.; Gerber, V.; Marti, E. Allergen-specific immunoglobulin E in sera of horses affected with insect bite hypersensitivity, severe equine asthma or both conditions. *J. Vet. Intern. Med.* **2019**, *33*, 266–274. [CrossRef] [PubMed]
- 144. Eder, C.; Crameri, R.; Mayer, C.; Eicher, R.; Straub, R.; Gerber, H.; Lazary, S.; Marti, E. Allergen-specific IgE levels against crude mould and storage mite extracts and recombinant mould allergens in sera from horses affected with chronic bronchitis. *Vet. Immunojathol.* 2000, 73, 241–253. [CrossRef] [PubMed]
- 145. Pessoa, V.C.; Branco-Ferreira, M.; Jónsdóttir, S.; Marti, E.; Tilley, P. Comparison of Skin Prick Tests (SPT), Intradermal Tests (IDT) and In Vitro Tests in the Characterization of Insect Bite Hypersensitivity (IBH) in a Population of Lusitano Horses: Contribution for Future Implementation of SPT in IBH Diagnosis. *Animals* **2023**, *13*, 2733. [CrossRef] [PubMed]
- 146. Lorch, G.; Hillier, A.; Kwochka, K.W.; Saville, W.J.; Kohn, C.W.; Jose-Cunilleras, E. Results of intradermal tests in horses without atopy and horses with chronic obstructive pulmonary disease. *Am. J. Vet. Res.* **2001**, *62*, 389–397. [CrossRef] [PubMed]
- 147. Bond, S.; Léguillette, R.; Richard, E.A.; Couetil, L.; Lavoie, J.P.; Martin, J.G.; Pirie, R.S. Equine asthma: Integrative biologic relevance of a recently proposed nomenclature. *J. Vet. Intern. Med.* 2018, 32, 2088–2098. [CrossRef] [PubMed]

Animals **2023**, 13, 3872 17 of 17

148. Leclere, M. Corticosteroids and Immune Suppressive Therapies in Horses. *Vet. Clin. N. Am. Equine Pract.* **2017**, 33, 17–27. [CrossRef]

- 149. Uberti, B.; Morán, G. Role of neutrophils in equine asthma. Anim. Health Res. Rev. 2018, 19, 65–73. [CrossRef]
- 150. Jocelyn, N.A.; Wylie, C.E.; Lean, M.; Barrelet, A.; Foote, A.K. Association of neutrophil morphology with bacterial isolates in equine tracheal wash samples. *Equine Vet. J.* **2018**, *50*, 752–758. [CrossRef]
- 151. Davis, K.U.; Sheats, M.K. Bronchoalveolar lavage cytology characteristics and seasonal changes in a herd of pastured teaching horses. *Front. Vet. Sci.* **2019**, *6*, 74. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.