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Agroecological Zone-Specific Diet Optimization for Water Buffalo (*Bubalus bubalis*) through Nutritional and In Vitro Fermentation Studies

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Simple Summary: This study involves the formulation of distinct diets for water buffalo based on locally available feed resources to specific agroecological zones. The diets were categorized into three groups addressing the maintenance, growth, and lactation/production requirements of buffaloes. This study assessed the chemical composition and in vitro gas and methane emissions of each diet. The implication of this work suggests a promising future for buffalo feeding systems, as it focuses on need-based formulations using specific regional ingredients. This approach may enhance the efficiency and sustainability of buffalo farming in specific zones.

Abstract: The water buffalo faces challenges in optimizing nutrition due to varying local feed resources. In response to this challenge, the current study introduces originality by addressing the lack of region-specific feeding strategies for water buffaloes. This is achieved through the formulation of 30 different diets based on locally available resources, offering a tailored approach to enhance nutritional optimization in diverse agroecological contexts. These diets were segmented into three groups of ten, each catering to the maintenance (MD_1 to MD_{10}), growth (GD_1 to GD_{10}), and lactation/production (PD1 to PD10) needs of buffaloes. Utilizing local feed ingredients, each diet was assessed for its chemical composition, in vitro gas and methane emissions, and dry matter (DM) disappearance using buffalo rumen liquor. The production diets (127 and 32.2 g/kg DM) had more protein and fats than the maintenance diets (82.0 and 21.0 g/kg DM). There was less (p < 0.05) fiber in the production diets compared to the maintenance ones. Different protein components (P_{B1} , P_{B2}) were lower (p < 0.05) in the maintenance diets compared to the growth and production ones, but other protein fractions (P_{B3} , P_c) were higher (p < 0.05) in the maintenance diet. Furthermore, the growth diets had the highest amount of other protein components (P_A) , while the maintenance diets had the highest amount of soluble carbohydrates (586 g/kg DM), whereas the carbohydrate fraction (C_{B1}) was highest (p < 0.05) in the production diets (187 g/kg DM), followed by the growth (129 g/kg DM) and maintenance diets (96.1 g/kg DM). On the contrary, the carbohydrate C_A fraction was (p < 0.05) higher in the maintenance diets (107 g/kg DM) than in the growth (70.4 g/kg DM) and production diets (44.7 g/kg DM). The in vitro gas production over time (12, 24, and 48 h) was roughly the same for all the diets. Interestingly, certain components (ether extract, lignin, NDIN, ADIN, and P_{B3} and C_C) of the diets seemed to reduce methane production, while others (OM, NPN, SP, PA and PB1, tCHO and CB2) increased it. In simple words, this study reveals that different diets affect gas production during digestion, signifying a significant step towards a promising future for buffalo farming through tailored, region-specific formulations.



Citation: Singh, S.; Koli, P.; Kushwaha, B.P.; Anele, U.Y.; Bhattacharya, S.; Ren, Y. Agroecological Zone-Specific Diet Optimization for Water Buffalo (*Bubalus bubalis*) through Nutritional and In Vitro Fermentation Studies. *Animals* 2024, 14, 143. https:// doi.org/10.3390/ani14010143

Academic Editor: Daniel Mota-Rojas

Received: 23 November 2023 Revised: 27 December 2023 Accepted: 29 December 2023 Published: 31 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Keywords:** agroecological zone; buffalo; diet formulation; feeding system; fermentation; methane emission

1. Introduction

India's vast agroecological diversity offers a surplus of locally available feed resources. These diverse regions provide an opportunity to create diets specific to the needs of buffaloes, depending on their lifecycle stage, be it maintenance, growth, or lactation. The country's agricultural backbone stands not just on its crops but, significantly, also on its livestock, with buffaloes playing a pivotal role [1]. The buffalo, often deemed the 'Black Gold' of India and Pakistan, is central to the rural economy [2,3]. This is not surprising given that India boasts a multitude of buffalo breeds, each with its unique attributes, suiting the varied climatic and topographical conditions of the country. From the Murrah, known for its high milk yield, to the *Bhadawari*, appreciated for its adaptability, the diversity is truly expansive [4]. In India, buffalo and cattle farming face challenges with limited feed resources and insufficient farmer knowledge on animal nutrition, impacting dairy animal productivity, which varies across regions due to differences in feed availability, types, and adherence to scientific feeding practices [5]. Livestock is the primary contributor of 50% of the 14.17 Tg methane emission total that comes from the Indian agricultural sector [6]. Methane, an important GHG (greenhouse gas) about 22–25 times more potent than carbon dioxide [7], is produced by ruminants as an end product of microbial digestion [8]. During the fermentation process (digestion and metabolism) of diets in the gastrointestinal tract, between 2 and 12% of dietary gross energy is lost as methane [7]. Several factors, viz., animal species and size, animal physiological stage, feed intake, digestibility, diet composition, etc., influence enteric methane production [9,10]. Diet composition (chemical and physical qualities) and its intake level (quantity consumed) influence methane production due to their effect on the rate of digestion and the rate of passage [11]. Animal species and diet composition play an important role in methane production [7,12,13]. Animals have three main nutritional needs: staying healthy (maintenance), growing, and producing things like milk or offsprings (production). To meet these needs, we created three different diets for each region. These diets were made by combining different amounts of locally available food resources, like dry and green roughages, along with concentrated mixtures. Protein in vitro fermentation has been shown to be associated with lower CH₄ production than carbohydrates fermentation [14,15]. Dietary nitrogen (N) concentrations play an important role in influencing rumen methanogenesis [16], specifically where feed N is low [17], leading to the reduced microbial growth of methanogens, which face difficulty competing under low N conditions [18]. The type of carbohydrate being digested, such as cellulose, hemicelluloses, and soluble residue, holds a notable influence over methane production [19–21]. Moreover, a strong relationship is observed between CH_4 production and digestible neutral detergent fiber (NDF) for cows and calves [22].

The main goal of this study was to develop and evaluate 30 water buffalo diets tailored for various life stages and agroecological zones in India. The assessment involved scrutinizing their nutritional compositions and in vitro methane production. The ultimate aim was to redirect methane emissions into a valuable energy source, thereby improving livestock productivity and simultaneously addressing global environmental concerns.

2. Materials and Methods

2.1. Formulation of Concentrate Mixtures

Local feed ingredients and their use in feeding livestock according to their suitable agroecological regions were considered for the formulation of concentrate mixtures (CM) for different regions of the country. A total of nine CM were prepared using protein and energy sources in different proportion for use in different diets as described in Table 1.

Ingredients	CM ₁	CM_2	CM ₃	CM ₄	CM ₅	CM ₆	CM ₇	CM ₈	CM ₉
Mustard seed cake	35	40	-	-	-	-	40	45	-
Wheat bran	25	-	25	-	25	-	-	-	-
Maize grain	40	-	-	60	-	-	20	-	40
Cotton seed cake	-	-	35	40	-	-	-	-	45
Oat grain	-	-	40	-	-	60	-	-	-
Barley grain	-	60	-	-	40	-	-	-	-
Groundnut cake	-	-	-	-	35	40	-	-	-
Rice bran	-	-	-	-	-	-	40	55	15

Table 1. Proportion (%) and composition of ingredients in different concentrate mixtures.

CM: Concentrate mixture.

2.2. Preparation Diets/Rations

The nutritional requirements of livestock were classified into three categories based on animals' functional needs, viz., maintenance, growth, and production. For each category, ten diets/rations were prepared, and, hence, a total of thirty diets were formulated via the uniform mixing of dry and green fodder with concentrate mixtures in different proportions (Table 2).

Table 2. Composition of diets (ingredients and their proportions).

AER	Pagion	Maintenance			Growth				Production			
No.	Region	Diet	Composition	Proportions	Diet	Composition	Proportions	Diet	Composition	Proportions		
1	Western Himalayan region	MD_1	Grass: GOL	65:35	GD_1	SST:L:CM ₂	60:30:10	PD_1	WS:B:CM ₂	30:40:30		
2	Eastern Himalayan region	MD_2	Grass:LL	75:25	GD_2	RS:LL:CM1	50:35:15	PD_2	Grass: LL:CM ₁	35:40:25		
3	Eastern plateau and plains region	MD_3	RS:MG	20:80	GD_3	RS:NG:CM7	30:50: 20	PD_3	MST:NG:CM7	20:45:35		
4	Middle Gangetic plain	MD_4	WS:MG	50:50	GD_4	RS:B	40:60	PD_4	MST:CM ₆	60:40		
5	Trans and Upper Gangetic plain	MD_5	WS:B	70:30	GD_5	SST:B:CM ₂	60:25:15	PD_5	WS:B:CM ₃	30:40:30		
6	Central plateau and hills	MD_6	LS	100	GD_6	GS:CM ₂	80:20	PD_6	LS:CM5	60:40		
7	Western plateau and hills	MD_7	WS:SG	50:50	GD_7	SST/L/B	55:45	PD ₇	WS:B:CM ₄	35:35:30		
8	Southern plateau and hills region	MD_8	RS:L	65:35	GD_8	SST:ST:CM7	40:40:20	PD_8	SST:CM8	60:40		
9	Western dry zone	MD_9	PST:LL	75:25	GD9	PST: LL:CM ₂	55:30:15	PD_9	PST:CM ₂	60:40		
10	Coastal and island region	MD ₁₀	RS:LL	65:35	GD ₁₀	RS:LL:CM9	45:40:15	PD_{10}	RS:LL:CM9	30:35:35		

AER: Agroecological regions; CM: Concentrate mixture; MD: Maintenance diet; GD: Growth diet; PD: Production diet; GOL: *Grewia optiva* leaves; LL: *Leucaena leucocephala* leaves; MG: Maize green; RS: Rice straw; SST: Sorghum stover; L: Lucerne; WS: Wheat straw; B: Berseem; LS: Lentil straw; SG: sorghum green; PST: Pearl millet stover; NG: Napier grass; MST: Maize stover.

2.3. Determination of Chemical Composition

The dry matter (930.15), ash (932.05), N (976.05), and ether extract (EE, 920.39) contents of the diets' samples were estimated following the standard method of the Association of Official Analytical Chemists (AOAC) [23]. The nitrogen values were multiplied by 6.25 to convert them into crude protein (CP) values. Neutral detergent fiber (NDF), acid detergent fiber (ADF), cellulose, and lignin (sa) were estimated as per the sequential method [24] using a fiber analyzer (Fibra Plus FES 6, Pelican, Chennai, India). Both the NDF and the ADF were expressed inclusive of their residual ash. There was no complex plant matrix included in our diet compositions; therefore, heat-stable α -amylase and sodium sulfite were not used in NDF determination. The lignin (sa) was determined by solubilizing cellulose with 72% of sulfuric acid in the ADF residue [24]. The cellulose was calculated as the difference between the ADF and the lignin (sa) in the sequential analysis. The hemicellulose was calculated as the difference between the NDF and ADF contents.

2.4. Estimation of Carbohydrate Fractions

The carbohydrate (CHO) fractions of the different diets samples were estimated as per the Cornell Net Carbohydrate and Protein (CNCP) system [25]. This system classifies CHO fractions into four fractions, as follows: C_A indicates rapidly degradable sugars; C_{B1} classifies intermediately degradable starch and pectin; C_{B2} includes slowly degradable cell walls; and C_C comprises unavailable/lignin-bound cell walls based on their degradation rate. The diets' total CHO (tCHO; g/kg DM) was determined by subtracting the CP, ether extract (EE), and ash contents from 1000. The structural carbohydrates (SC) were calculated as the difference between the NDF and the neutral detergent-insoluble protein (NDIP), and the non-structural CHO were estimated as the difference between the tCHO and the SC [26]. For the starch estimation, samples were extracted with ethyl alcohol to solubilize free sugars, lipids, pigments, and waxes. The residue rich in starch was solubilized with perchloric acid, and the extract was treated with anthrone–sulfuric acid to determine glucose colorimetrically using a UV spectrophotometer (LABINDIA3000) at 630 nm [27].

2.5. Estimation of Protein Fractions

The CP fractions of the diets were partitioned into five fractions according to the CNCPS, [25] as modified previously [28]. These are the following: fraction P_A , indicating non-protein N, estimated as the difference between the total N and the true CP N precipitated with sodium tungstate (0.30 M) and 0.5 M of sulfuric acid; P_{B1} , the buffer-soluble protein calculated as the difference between the true protein and the buffer-insoluble protein, estimated with a boratephosphate buffer (pH 6.7–6.8) and a freshly prepared 0.10 M sodium azide solution; fraction P_{B2} , the neutral detergent-soluble protein, estimated as the difference between the ND-insoluble protein, estimated as the buffer-insoluble protein minus the ND-insoluble protein; fraction P_{B3} , the acid detergent-soluble CP, estimated as the difference between the ND-insoluble protein and the acid detergent-insoluble CP; and fraction P_C , assumed to be indigestible (protein associated with lignin, tannin–protein complexes, and Maillard products which are unavailable to animals).

2.6. In Vitro Incubation

The invitro gas production was determined using the pressure transducer technique [29]. Ruminal fluid was collected before feeding from two fistulated adult male Murrah water buffaloes (Bubalus bubalis) fed a wheat straw-concentrate diet (65:35 DM basis). The rumen fluid was filtered through a double layer of cheese cloth and bubbled with CO₂ before the commencement of incubation. The incubation medium was prepared by means of the sequential mixing of a buffer solution (NH₄HCO₃ and NaHCO₃), a macromineral solution, a micro-mineral solution, and resazurin solution [30]. Samples (1 g) of air-dry green forages were weighed into three serum bottles (150 mL capacity). Three serum bottles without substrate were used as blank cultures. The sample and control serum bottles were gassed briefly with CO_2 before adding 65 mL of medium. The bottles were continuously fluxed with CO_2 , and then 3 mL of reducing solution were added in each bottle. The gassing of bottles with CO₂ continued until the pink color turned colorless. Before inoculation, the gas pressure transducer was used to adjust the head-space gas pressure in each bottle (to adjust the zero reading on the LED display). The serum bottles were inoculated with 8 mL of ruminal fluid inoculum using a 10 mL syringe. The inoculated bottles were sealed and incubated at 39 °C. The samples were incubated in triplicates, and the gas production (mL) was measured at 12, 24, and 48 h of incubation. The whole process was repeated on a different day.

2.7. Methane Measurements

The methane (CH₄) in the total gas was measured from three bottles incubated for each of the thirty diets at the 12, 24, and 48 h timepoints using gas chromatography (Nucon 5765 microprocessor-controlled gas chromatograph (GC), Okhla, New Delhi, India) equipped with a stainless-steel column packed with Porapak-Q and a flame ionization detector. Gas (1 mL) was sampled from the gas produced using a Hamilton syringe and injected manually (pull and push method of sample injection) into a GC. The GC was calibrated using standard methane and CO₂. The methane level was additionally measured in blank samples at different fermentation stages, and these measurements were used to correct for methane produced by the inoculum. The methane measured was related to the total gas to estimate its concentration [31] and converted into energy and mass values using 39.54 kJ/L CH₄ and 0.716 mg/mL CH₄ factors, respectively [32].

2.8. In Vitro Dry Matter Digestibility (IVDMD) and Energy of Diets

For the determination of the IVDMD for the evaluated diets a standard method was followed [33], wherein a 0.5 g sample was incubated for 48 h and then digested with 0.1 g of pepsin (1:3000 Sisco Research Laboratories, Mumbai, India) and 2 mL of 6N HCl at 39 °C for 24 h. The samples were incubated in triplicate with ruminal inoculum from the two fistulated buffaloes described previously. A provision was also made for the blanks, as described for the in vitro gas production. The digestibility was estimated as the difference between the DM incubated and the residual DM at the end of the second stage of digestion. The gross energy (GE) of the forages was measured with a bomb calorimeter (Toshniwal Brothers CLOI/M2, Bangalore, India) using benzoic acid as the standard.

2.9. Statistical Analysis

The data were subjected to an analysis of variance using the GLM procedure of SAS (2002). The model was the following: Yij = [1] + Fi + Eij, where Yij represents the individual observation of the variable, and Fi is the fixed effect of the ith diet (i = 1–30). The overall mean is expressed as [1], and Eij is the random error associated with Yij, not accounted in the fixed effect. The means were separated using Fisher's LSD and all the statistical tests were at the p = 0.05 level of significance. The means of cereals, grasses, and legumes were compared using orthogonal contrasts (i.e., cereals vs. grasses, cereals vs. legumes, and grasses vs. legumes). The differences among forage means with p < 0.05 were accepted as statistically significant. A correlation analysis was used to establish relationships between chemical constituents, carbohydrate fractions, protein fractions, and CH₄ production. Pearson's correlation analysis was performed to establish the relationship of chemical composition with methane production, carbohydrate fractions, and protein fractions at level p < 0.05.

3. Results

3.1. Chemical Composition

The crude protein (CP) and ether extract (EE) values were significantly higher (p < 0.05) for the production diets (127 and 32.2) than the maintenance diets (82.0 and 21.0 g/kgDM), respectively. The CP values of all three diets including the MD, the GD, and the PD varied, measuring 69.8–96.1, 106–130, and 103–153 g/kg DM, respectively (Table 3), whereas, the concentrations of NDF, ADF, and cellulose were lower (p < 0.05) in the production diets (546, 333 and 245) than in the maintenance diets (618, 395 and 293 g/kg DM). Interestingly, no significant difference was observed in the lignin contents among all three diets.

Diet	СР	ОМ	EE	NDF	ADF	Cellulose	H cellulose	Lignin
MD ₁	76.0 ^{cd}	876 ^{cd}	32.1 ^a	646 ^c	453 ^a	298 ^c	193 ^{de}	93.4 ^b
MD_2	93.3 ^a	871 ^c	27.5 ^b	678 ^b	456 ^a	274 ^d	222 ^c	109 ^a
MD_3	84.3 ^b	923 ^a	17.8 ^d	668 ^{bc}	361 ^d	309 ^b	306 ^a	49.6 ^{de}
MD_4	69.8 ^{de}	903 ^b	14.4 ^e	651 ^c	379 ^c	318 ^b	271 ^b	45.7 ^{ef}
MD_5	96.1 ^a	884 ^c	18.1 ^d	573 ^e	386 ^c	301 ^c	187 ^e	49.7 ^{de}
MD_6	76.9 ^{cd}	914 ^a	13.4 ^e	537 ^f	386 ^c	283 ^d	151 ^f	93.9 ^b
MD_7	68.0 ^e	920 ^a	20.7 ^{cd}	713 ^a	406 ^b	345 ^a	307 ^a	42.6 ^f
MD_8	77.8 ^{bc}	857 ^e	26.4 ^b	591 ^d	391 ^{bc}	272 ^d	200 ^d	55.8 ^{cd}
MD ₉	81.8 ^{bc}	920 ^a	21.5 ^c	545 ^f	344 ^e	256 ^e	201 ^d	62.5 ^c
MD_{10}	95.5 ^a	852 ^e	17.9 ^d	575 ^e	392 ^{bc}	274 ^d	184 ^e	53.7 ^d
LSD	7.12	10.20	3.17	12.96	15.62	15.07	9.17	6.84
Diet	СР	ОМ	EE	NDF	ADF	Cellulose	H cellulose	Lignin
GD_1	121 ^{ab}	917 ^{ab}	19.7 ^{de}	610 ^b	411 ^b	311 ^b	199 ^{cde}	80.7 ^b
GD_2	116 ^{bcd}	874 ^d	32.4 ^a	527 ^e	341 ^e	231 ^c	186 ^e	55.4 ^{fg}
GD_3	88.7 ^e	856 ^e	24.7 ^{bc}	676 ^a	392 ^{cd}	334 ^a	284 ^a	55.9 ^{fg}
GD_4	111 ^{bcd}	864 ^{de}	18.6 ^e	619 ^b	411 ^{bc}	311 ^b	209 ^{cd}	52.0 ^g
GD_5	117 ^{bc}	922 ^a	26.7 ^b	610 ^b	391 ^d	303 ^b	219 ^c	64.2 ^{de}
GD_6	110 ^{cd}	909 ^{bc}	18.4 ^e	546 ^d	412 ^b	306 ^b	134 ^f	92.5 ^a
GD_7	106 ^d	899 ^c	17.7 ^e	584 ^c	389 ^d	306 ^b	195 ^{de}	71.6 ^{cd}
GD_8	111 ^{bcd}	916 ^{ab}	18.1 ^e	690 ^a	438 ^a	329 ^a	252 ^b	75.5 ^{bc}
GD_9	111 ^{bcd}	917 ^{ab}	23.1 ^e	493 ^f	300 ^f	213 ^d	193 ^{de}	62.5 ^{ef}
GD_{10}	130 ^a	872 ^d	35.5 ^a	512 ^e	328 ^e	220 ^d	183 ^e	52.0 ^g
LSD	10.87	10.29	3.35	18.25	18.32	11.30	22.70	7.90
Diet	СР	ОМ	EE	NDF	ADF	Cellulose	H cellulose	Lignin
PD_1	130 ^{bc}	901 ^c	18.9 ^{de}	491 ^e	345 ^b	262 ^c	$146^{\text{ f}}$	54.4 ^d
PD ₂	153 ^a	899 ^c	39.4 ^b	537 ^c	292 ^d	177 ^g	245 ^b	79.4 ^b
PD_3	103 ^e	882 ^d	33.6 ^{cd}	633 ^a	360 ^b	311 ^a	273 ^a	45.0 ^e
PD_4	116 ^d	912 ^{ab}	30.7 ^e	589 ^b	350 ^b	249 ^{de}	239 ^{bc}	68.1 ^c
PD_5	137 ^b	904 ^{bc}	26.3 ^f	529 ^{cd}	326 ^c	253 ^{cd}	203 ^d	49.7 ^{de}
PD_6	121 ^{cd}	918 ^a	32.8 ^{de}	453 ^f	288 ^d	203 ^f	165 ^e	74.3 ^b
PD_7	121 ^{cd}	917 ^a	33.4 ^{cd}	549 ^c	318 ^c	251 ^d	231 ^{bc}	51.1 ^d
PD_8	121 ^{cd}	902 ^c	35.7 ^c	634 ^a	455 ^a	301 ^b	179 ^e	98.0 ^a
PD_9	116 ^d	916 ^a	21.7 ^g	539 ^c	310 ^c	240 ^e	229 ^{bc}	49.3 ^{de}
PD_{10}	149 ^a	886 ^d	49.3 ^a	508 ^{de}	283 ^d	200 ^f	225 ^c	50.9 ^d
LSD	9.82	7.95	2.68	22.11	16.41	10.41	18.67	5.68

Table 3. Chemical composition of the maintenance diets (g/kg DM) *.

MD, maintenance diets; GD, growth diets; PD, production diets; CP, crude protein; OM, organic matter; EE, ether extract; NDF, neutral detergent fiber; ADF, acid detergent fiber; H cellulose, hemi cellulose; LSD, least significant difference at *p* value < 0.0001; different superscript letters within a column in the table signify statistical differences among the corresponding values; *, each value is a mean of four observations.

3.2. Nitrogen Fractions

The protein fraction values (P_{B1} and P_{B2} ; g/kg DM) followed a lower to higher (p < 0.05) order from the MD (150.2 and 357.3) to the GD (204.7 and 409.7), followed by the PD (217.4 and 412.3), while the mean concentration of the slowly degradable protein fraction (P_{B3}) and the lignin-bound protein fraction (Pc) were higher (p < 0.05) in the maintenance diets (205.6 and 181.3) than in the growth diets (113.9 and 114.8), followed by the production diets (151.5 and 104.0 g/kg DM) (Table 4). The average value of P_A (g/kg DM) was significantly higher (p < 0.05) for the growth diets (136.9) than in the production and maintenance diets, which had values of 114.8 and 105.6, respectively. In the maintenance diets, the affinity of protein binding to ADF was observed to be higher than in the growth and production diets, whereas the SP concentration was in the reverse order, meaning that the concentration in the maintenance diets was lower than in the two other diets.

Maintenance					Growth					Production							
Diet	PA	P _{B1}	P _{B2}	P _{B3}	P _C	Diet	P _A	P _{B1}	P _{B2}	P _{B3}	P _C	Diet	PA	P _{B1}	P _{B2}	P _{B3}	P _C
MD_1	35.6 ^{de}	101 ^f	358 ^{cd}	237 ^{cd}	268 ^a	GD_1	310 ^a	213 ^b	228 ^f	125 ^{cd}	123 ^{abc}	PD_1	124 ^d	242 ^{bcd}	423 ^c	95.2 ^{ef}	116 ^b
MD_2	18.1 ^e	95.3 ^f	287 ^{de}	301 ^b	299 ^a	GD_2	27.8 ^{fg}	85.4 ^d	547 ^a	194 ^a	146 ^a	PD_2	49.4 ^g	146 ^{fg}	364 ^d	291 ^b	148 ^a
MD_3	194 ^a	195 ^{ab}	233 ^e	157 e	220 ^b	GD_3	171 ^c	213 ^b	359 ^{de}	138 ^{bcd}	118 ^{abcd}	PD_3	249 ^a	279 ^{ab}	175 ^f	187 ^c	110 ^{bc}
MD_4	187 ^a	185 ^{bc}	327 ^d	88.8 ^f	211 ^{bc}	GD_4	172 ^c	202 ^b	385 ^{cde}	155 ^b	85.7 ^e	PD_4	158 ^b	296 ^a	304 ^e	133 ^d	108 ^{bc}
MD_5	173 ^a	213 ^a	429 ^{bc}	48.9 f	135 def	GD_5	167 ^c	295 ^a	336 ^e	114 ^{de}	87.2 ^{de}	PD_5	134 ^{cd}	259 ^{abc}	464 ^e	55.5 ^h	87.4 ^{cd}
MD_6	123 ^b	130 ^e	576 ^a	70.9 ^f	99.6 ^f	GD_6	141 ^d	292 ^a	429 ^{bc}	$28.4^{\text{ f}}$	110 ^{bcde}	PD_6	96.9 ^e	208 ed	508 ^{bc}	113 ^{de}	73.4 ^d
MD_7	188 ^a	218 ^a	24.3 ^f	425 a	145 ^{de}	GD_7	203 ^b	192 ^b	357 ^{de}	100 ^e	147 ^a	PD_7	71.3 ^f	171 ^{ef}	595 ^a	72.1 ^{fgh}	90.4 ^{cd}
MD_8	14.9 ^e	67.8 ^g	485 ^b	257 ^{bc}	175 ^{cd}	GD_8	109 ^e	218 ^b	416 ^{cd}	146 ^{bc}	110 ^{bcde}	PD_8	101 ^e	223 ^{cd}	492 ^b	65.6 ^{gh}	117 ^b
MD ₉	74.4 ^c	159 ^{cd}	412 ^{bc}	209 ^d	145 ^{de}	GD_9	50.8 ^f	188 ^b	490 ^{ab}	142 ^{bc}	129 ^{ab}	PD_9	143 ^c	228 ^{cd}	431 ^c	88.3 ^{efg}	108 ^{bc}
MD_{10}	47.7 ^d	138 ^{de}	440 ^b	261 ^{bc}	114 ^{ef}	GD_{10}	15.9 ^g	147 ^c	550 ^a	195 ^a	91.4 ^{cde}	PD_{10}	20.2 ^h	120 ^g	364 ^d	413 ^a	82.3 ^d
LSD	22.84	27.36	78.36	47.84	40.64	LSD	23.03	31.46	64.18	24.93	32.45	LSD	14.29	39.28	57.32	26.78	22.65

Table 4. Protein fractions of the diets (g/kg CP) *.

MD, maintenance diets; GD, growth diets; PD, production diets; P_A, non-protein nitrogen; P_{B1}, buffer-soluble protein; P_{B2}, neutral detergent-soluble protein; P_{B3}, acid detergent-soluble protein; P_C, indigestible protein; LSD, least significant difference at *p* value < 0.0001; different superscript letters within a column in the table signify statistical differences among the corresponding values; *, each value is a mean of four observations.

3.3. Carbohydrate Fractions

Among the carbohydrate fractions, no significant difference was observed among all three diets for the total carbohydrate levels (tCHO), while the SC contents were (p < 0.05) higher in the maintenance diets (586.2) than in the production diets (513.0 g/kg DM), respectively. The average value of the rapidly degradable carbohydrate fraction (C_{B1}) differed (p < 0.05) among the diets, being highest in the production diets (187.2), followed by the growth (129.5) and maintenance diets (96.1 g/kg DM; Table 5). The contrary carbohydrate C_A fraction was (p < 0.05) higher in the maintenance diets (107.1) than in the growth (70.4) and production diets (944.7 g/kg DM). The carbohydrate fractions C_{B2} and C_C were relatively lower in the production diets than in the maintenance and growth diets.

Diet	tCHO	NSC	SC	C _C	C _{B2}	C _{B1}	C _A
MD ₁	768 ^c	161 ^d	607 ^c	224 ^b	383 ^e	20.3 g	140 ^b
MD_2	751 ^d	128 ^e	622 ^{bc}	262 ^a	360 ^f	95.6 ^{cde}	32.9 ^d
MD_3	821 ^{ab}	185 ^c	636 ^b	119 ^{de}	517 ^b	74.5 ^{ef}	110 ^b
MD_4	818 ^{ab}	189 ^c	630 ^b	110 ^{ef}	520 ^b	87.6 ^{def}	101 ^c
MD_5	770 ^c	215 ^b	555 ^d	119 ^{de}	436 ^c	120 ^{bc}	94.8 ^c
MD_6	824 ^{ab}	300 ^a	523 ^f	225 ^b	298 ^g	178 ^a	122 ^b
MD ₇	832 ^a	157 ^d	674 ^a	102 ^f	572 ^a	114 ^{bcd}	43 ^d
MD_8	753 ^d	196 ^c	557 ^d	134 ^{cd}	423 ^{cd}	135 ^b	60.4 ^d
MD ₉	817 ^b	300 ^a	516 ^f	150 ^c	366 ^{ef}	58.4 ^f	242 ^a
MD_{10}	739 ^d	199 ^c	540 ^e	129 ^d	411 ^d	76.8 ^{ef}	123 ^b
LSD	14.23	15.14	14.70	16.43	18.56	30.53	31.46
Diet	tCHO	NSC	SC	C _C	C _{B2}	C _{B1}	CA
GD ₁	776 ^a	196 ^d	580 ^b	194 ^b	386 ^d	144 ^{cd}	51.9 ^{def}
GD_2	726 ^c	238 ^b	488 ^e	133 ^{fg}	355 ^e	175 ^{bc}	62.9 ^{cde}
GD_3	743 ^b	88.9 ^f	654 ^a	134 ^{fg}	520 ^a	59.2 g	29.8 ^{fg}
GD_4	735 ^{bc}	142 ^e	593 ^b	125 ^g	468 ^b	59.4 ^g	82.5 ^c
GD_5	779 ^a	192 ^d	586 ^b	154 ^{de}	433 ^c	155 ^{bc}	37.3 ^{efg}
GD_6	780 ^a	248 ^b	532 ^d	222 ^a	310 ^f	179 ^{ab}	68.7 ^{cd}
GD_7	776 ^a	218 ^c	558 ^c	172 ^{cd}	386 ^d	86.1 ^{fg}	132 ^b
GD_8	786 ^a	125 ^e	662 ^a	181 ^{bc}	480 ^b	108 ^{ef}	16.6 ^g
GD9	783 ^a	320 ^a	463 ^f	150 ^{ef}	313 ^f	121 ^{de}	199 ^a
GD ₁₀	707 ^d	232 ^{bc}	475 ^{ef}	125 ^g	350 ^e	208 ^a	23.5 ^{fg}
LSD	15.52	19.72	17.94	18.97	28.14	31.11	29.1
Diet	tCHO	NSC	SC	C _C	C _{B2}	C _{B1}	CA
PD ₁	752 ^{bc}	288 ^b	464 ^d	131 ^d	333 ^d	239 ^b	49.4 ^{bc}
PD ₂	707 ^d	237 ^d	470 ^d	190 ^b	280 ^f	194 ^d	43.1 ^c
PD_3	746 ^c	143 ^f	602 ^a	108 ^e	494 ^a	103 g	40.2 ^c
PD_4	765 ^{ab}	203 ^e	562 ^b	163 ^c	398 ^b	169 ^e	34.1 ^{cd}
PD_5	741 ^c	231 ^e	509 ^c	119 ^{de}	390 ^{bc}	133 ^f	98.0 ^a
PD_{6}	764 ^b	333 ^a	431 ^e	178 ^b	252 g	302 ^a	31.7 ^{cd}
PD ₇	762 ^b	233 ^d	529 ^c	122 ^d	407 ^b	217 ^{bcd}	16.1 ^d
PD_8	745 ^c	133 ^f	612 ^a	235 ^a	376 ^c	97.8 ^g	35.2 ^{cd}
PD_9	779 ^a	262 ^c	516 ^c	118 ^{de}	398 ^b	197 ^{cd}	65.4 ^b
PD_{10}	687 ^e	253 ^{cd}	434 ^e	122 ^d	312 ^e	219 ^{bc}	34.1 ^{cd}
LSD	13.55	22.04	22.31	13.64	19.94	24.31	19.49

Table 5. Carbohydrate fractions of maintenance diets (g/kg DM) *.

MD, maintenance diets; GD, growth diets; PD, production diets; tCHO, total carbohydrates; NSC, non-structural carbohydrates; SC, structural carbohydrates; C_C, unavailable/lignin-bound cell wall; C_{B2}, slowly degradable cell wall; C_{B1}, intermediately degradable starch and pectin; C_A, rapidly degradable CHO, including sugars; LSD, least significant difference at *p* value < 0.0001; different superscript letters within a column in the table signify statistical differences among the corresponding values; *, each value is a mean of four observations.

3.4. Gas and Methane Production Kinetics

The average values (mL/g DM) for the diets' in vitro gas production were found to have a consistent pattern at 0–12, 12–24, and 24–48 h. The observed values for the maintenance diets were 63.0, 52.0, and 48.15; for the growth diets, they were 63.8, 52.7, and 48.2, and, for the production diets, they were 63.5, 52.5, and 47.2. The cumulative gas production values (48 h) were close and similar 163, 165, and 163 mL/g DM for the maintenance, growth, and production diets, respectively (Table 6). The in vitro methane production mean values at 0–12, 12–24, and 24–48 h and the cumulative values of the maintenance diets tended (p > 0.05) to be lower than those of the growth and maintenance diets, whereas, the cumulative methane production was lower in the maintenance diets (28.4) than in the production diets (33.1 mL/g DM).

Table 6. Gas and methane production kinetics from the maintenance diets fermented in buffalo inoculums *.

		Gas	s (mL/g)			Methane (mL/g)				
Diets/Rations	0–12 h	12–24 h	24–48 h	Cumulative	0–12 h	12–24 h	24–48 h	Cumulative		
MD ₁	64.3 ^c	50.0 ^e	50.0 ^b	164 ^c	9.83 ^d	5.77 ^f	5.08 ^h	20.7 ^g		
MD_2	58.5 ^h	51.0 ^d	44.3 ^f	154 ^e	6.57 ^f	5.56 ^f	11.3 ^b	23.4 ^e		
MD_3	59.5 ^{gh}	55.8 ^b	47.6 ^d	163 ^c	11.2 ^c	12.3 ^a	16.3 ^a	39.8 ^a		
MD_4	63.6 ^{cd}	54.3 ^c	50.0 ^b	168 ^b	12.6 ^b	11.2 ^b	17.0 ^a	40.4 ^a		
MD_5	66.0 ^b	53.7 ^c	45.3 ^e	164 ^c	11.2 ^c	8.77 ^c	5.66 ^g	25.7 ^d		
MD_6	62.3 ^{de}	49.2 ^e	47.9 ^d	160 ^d	15.1 ^a	8.63 ^c	9.56 ^d	33.3 ^b		
MD_7	69.8 ^a	58.5 ^a	48.3 ^{cd}	178 ^a	12.1 ^b	7.89 ^d	8.32 ^e	28.3 ^c		
MD_8	63.8 ^{cd}	54.5 ^c	46.1 ^e	164 ^c	8.72 ^e	6.68 ^e	7.02 ^f	22.4 ^f		
MD ₉	61.8 ^{ef}	44.5 ^g	49.3 ^c	156 ^e	9.03 ^{de}	6.67 ^e	8.21 ^e	23.9 ^e		
MD_{10}	60.5 ^{fg}	47.8 ^t	52.6 ^a	160 d	7.12 ^t	7.98 ^d	10.8 °	25.9 d		
LSD	1.535	1.038	0.960	2.245	0.829	0.528	0.359	1.348		
Diets/Rations		Gas	s (mL/g)			Methan	e (mL/g)			
	0–12 h	12–24 h	24–48 h	Cumulative	0–12 h	12–24 h	24–48 h	Cumulative		
GD_1	65.0 ^b	50.3 ^f	48.8 ^{cd}	164 ^c	12.8 ^c	5.74 ^h	3.95 ⁱ	22.5 ^{gh}		
GD ₂	59.5 ^d	55.8 ^b	46.8 ^e	162 ^{de}	8.20 ^{ef}	10.1 ^b	15.3 ^c	33.6 ^c		
GD_3	62.4 ^c	55.2 ^{bc}	49.2 ^{bc}	167 ^b	10.5 ^d	10.6 ^a	15.8 ^b	36.9 ^b		
GD_4	64.8 b	52.6 ^e	48.8 ^{cd}	166 ^b	15.5 ^a	10.9 ^a	16.2 ^a	42.6 ^a		
GD_5	65.8 ^b	51.8 ^e	45.6 ^f	163 ^{cd}	13.2 °	7.39 ^f	6.40 g	26.96 ^t		
GD ₆	67.3 ^a	54.5 ^{cd}	44.3 g	166 ^b	10.9 d	7.96 ^e	4.73 ^h	23.6 g		
GD_7	67.8 ^a	57.0 ª	48.3 ^a	173 a	14.5 ^b	9.49 °	8.22 ^r	32.2 ^d		
GD ₈	62.8 °	54.0 ^d	49.8 ^b	167 ^b	9.09 °	6.07 s	6.40 ^g	21.6 ⁿ		
GD ₉	63.5 °	46.3 g	49.6 ^b	159 ¹	11.1 a	8.48 ^d	9.67 e	29.2 °		
	59.5 ^a 1 136	50.0 1	51.0 °° 0 729	161 er 1 836	7.871	8.50 ^a	10.27 ^u 0.357	26.6		
	1.150	0.044	(mL (a)	1.000	0.900	0.300	0.337	1.175		
Diets/Rations	0.12 h	10.04 h	24 48 h	Cumulativa	0.12 h	12.24 h	24 48 h	Cumulativa		
	0–12 n	12–24 n	24–48 n	Cumulative	0–12 h	12–24 n	24–48 n	Cumulative		
PD ₁	62.8 ^e	54.3 ^{cd}	44.0 ^g	161 ^d	13.9 ^b	12.7 ^a	14.5 ^b	41.1 ^a		
PD ₂	62.6 ^e	53.0 ^e	46.8 ^e	162 ^d	11.1 ^d	9.02 ^{de}	14.2 ^b	34.3 ^{cd}		
PD ₃	61.0 ^f	54.6 ^{bc}	48.8 ^{cd}	164 ^c	10.9 ^d	11.2 ^{bc}	16.0 ^a	38.1 ^b		
PD_4	64.3 ^d	54.0 ^d	50.0 ^{ab}	168 ^b	14.0 ^b	11.0 ^{bc}	15.9 ^a	40.9 ^a		
PD ₅	67.6 ^a	55.0 ^b	45.0 ^f	168 ^b	15.6 ^a	11.1 ^{bc}	8.03 ^d	34.7 ^c		
PD ₆	66.3 ^b	54.8 ^{bc}	44.1 g	165 ^c	15.0 ^a	11.4 ^b	6.94 ^e	33.4 ^{cd}		
PD ₇	66.0 ^b	58.6 ^a	45.3 ^f	170 ^a	13.0 ^c	10.5 ^c	6.94 ^e	30.4 ^e		
PD ₈	60.5 ^f	43.8 ^h	48.3 ^d	153 ^f	6.42 ^f	5.68 ^f	6.56 ^e	18.6 ^g		
PD ₉	65.0 ^c	47.5 ^g	49.4 ^{bc}	162 ^d	13.5 ^{bc}	9.41 ^d	9.75 ^c	32.7 ^d		
PD ₁₀	59.0 ^g	49.7 ^f	50.4 ^a	159 ^e	8.28 ^e	8.63 ^e	10.11 ^c	27.0 ^f		
LSD	0.663	0.557	0.796	1.278	0.883	0.741	0.786	1.970		

MD, maintenance diets; GD, growth diets; PD, production diets; LSD, least significant difference at *p* value < 0.0001; different superscript letters within a column in the table signify statistical differences among the corresponding values; *, each value is a mean of four observations.

3.5. Methane Production and Percentage Loss of Dietary Energy as Methane

The mean values of the in vitro methane production (mL/g DDM, g/kg DM and g/kg DDM) were similar in the diets formulated for maintenance (41.2, 13.3 and 29.5; Table 7), for growth (42.2, 14.3 and 30.2; Table 7), and for production (41.3, 15.9 and 29.6; Table 7). Furthermore, a similar trend was observed for the gross energy from each diet being lost as methane, with comparable values in the maintenance (1.57), growth (1.61), and production diets (1.58 kJ/g DDM), equivalent to 9.09, 9.37, and 9.14% of dietary energy lost as methane, respectively.

Diets/Rations	IVDMD g/kg DM	CH ₄ mL/g DDM 24h	CH ₄ g/kg DM	CH4 g/kg DDM	GE kJ/g	GE in CH ₄ g DDM	CH4 %GE DDM
MD_1	422 ^{bc}	37.2 ^f	11.2 ^c	26.7 ^{ef}	16.9 ^{cd}	1.42 ^{ef}	8.45 ^{ef}
MD_2	402 ^c	30.4 ^g	8.70 ^d	21.7 ^g	18.0 ^{ab}	1.16 ^g	6.46 ^g
MD_3	482 ^b	48.9 ^a	16.9 ^a	35.0 ^a	17.5 ^{bc}	1.87 ^a	10.7 ^{ab}
MD_4	468 ^b	49.4 ^a	17.1 ^a	35.4 ^a	16.3 ^d	1.89 ^a	11.6 ^a
MD_5	468 ^b	42.5 ^{bcd}	14.3 ^b	30.5 ^{bcd}	17.5 ^{bc}	1.63 ^{bcd}	9.31 ^{cde}
MD_6	584 ^a	43.2 ^{bc}	17.0 ^a	31.0 ^{bc}	18.7 ^a	1.66 ^{bc}	8.87 ^{de}
MD_7	417 ^{bc}	47.0 ^{ab}	14.4 ^b	33.7 ^{ab}	17.4 ^{bc}	1.80 ^{ab}	10.3 ^{bc}
MD_8	367 ^e	41.5 ^{cde}	11.0 ^c	29.8 ^{cde}	16.2 ^d	1.59 ^{cde}	9.8 ^{bcd}
MD ₉	474 ^{bc}	33.5 ^{fg}	11.3 ^c	24.1 ^{fg}	17.6 ^{bc}	1.28 ^{fg}	7.31 ^{fg}
MD_{10}	389 ^{de}	38.5 ^{de}	10.8 ^c	27.6 ^{de}	17.2 ^c	1.47 ^{de}	8.56 ^e
LSD	43.58	4.64	0.81	3.33	0.768	0.178	1.14
Diets/Rations	IVDMD g/kg DM	CH ₄ mL/g DDM 24 h	CH4 g/kg DM	CH4 g/kg DDM	GE kJ/g	GE in CH ₄ g DDM	CH4 %GE DDM
GD ₁	450 ^d	41.3 ^{bc}	13.27 ^e	29.6 ^{bc}	16.9 ^{cd}	1.57 ^{bc}	9.29 ^{bc}
GD_2	496 ^{bc}	37.0 ^{de}	13.14 ^e	26.5 ^{de}	17.5 ^{bc}	1.41 ^{de}	8.06 ^e
GD_3	530 ^b	39.9 ^{cd}	15.13 ^c	28.6 ^{cd}	16.9 ^{cd}	1.53 ^{cd}	9.07 ^{cd}
GD_4	628 ^a	42.1 ^{bc}	18.94 ^a	30.2 ^{bc}	17.0 ^{cd}	1.61 ^{bc}	9.49 ^{bc}
GD_5	466 ^{cd}	44.3 ^b	14.74 ^c	31.8 ^b	16.9 ^{cd}	1.7 ^b	10.0 ^b
GD_6	530 ^b	35.7 ^e	13.55 ^{de}	25.6 ^e	16.6 ^d	1.37 ^e	8.21 ^{de}
GD_7	409 ^e	58.7 ^a	17.21 ^b	42.1 ^a	17.9 ^{ab}	2.24 ^a	12.5 ^a
GD_8	372 ^f	40.9 ^{bc}	10.87 ^g	29.3 ^{bc}	16.7 ^d	1.57 ^{bc}	9.43 ^{bc}
GD_9	469 ^{cd}	41.9 ^{bc}	14.01 ^d	30.0 ^{bc}	18.3 ^a	1.61 ^{bc}	8.82 ^{cde}
GD ₁₀	408 ^e	40.1 ^{cd}	11.74 ^f	28.8 ^{cd}	17.4 ^{bc}	1.53 ^{cd}	8.83 ^{cde}
LSD	35.36	3.88	0.686	2.78	0.614	0.148	0.865
Diets/Rations	IVDMD g/kg DM	CH ₄ mL/g DDM 24h	CH ₄ g/kg DM	CH4 g/kg DDM	GE kJ/g	GE in CH ₄ g DDM	CH4 %GE DDM
PD_1	621 ^a	42.9 ^c	19.06 ^a	30.7 ^c	17.5 ^{bcd}	1.66 ^c	9.48 ^{cd}
PD ₂	605 ^a	33.2 ^{de}	14.39 ^d	23.8 ^{de}	17.7 ^{abc}	1.28 ^{de}	7.24 ^e
PD_3	600 ^{ab}	36.9 ^d	15.82 ^c	26.5 ^d	16.8 ^{ef}	1.41 ^d	8.40 ^d
PD_4	523 ^{cd}	47.9 ^{ab}	17.92 ^b	34.3 ^{ab}	16.9 ^{def}	1.82 ^{ab}	10.8 ^{ab}
PD_5	520 ^d	51.5 ^a	19.11 ^a	36.9 ^a	17.4 ^{bcd}	1.99 ^a	11.4 ^a
PD_6	563 ^{bc}	47.0 ^{bc}	18.96 ^a	33.7 ^{ab}	16.7 ^f	1.78 ^{bc}	10.7 ^{ab}
PD_7	496 ^d	47.4 ^{ab}	16.82 ^c	34.0 ^{ab}	18.1 ^a	1.82 ^{ab}	10.1 ^{bc}
PD_8	391 ^e	31.1 ^e	8.68 ^f	22.3 ^e	17.2 ^{cdef}	1.20 ^e	6.92 ^e
PD ₉	534 ^{cd}	43.1 ^c	16.4 ^c	30.9 ^c	17.3 ^{bcde}	1.66 ^c	9.59 ^c
PD_{10}	533 ^{cd}	31.8 ^e	12.1 ^e	22.8 e	17.8 ^{ab}	1.20 ^e	6.84 ^e
LSD	40.91	4.21	1.003	3.02	0.575	0.161	1.029

Table 7. Methane production and loss of dietary energy as methane from the maintenance diets *.

MD, maintenance diets; GD, growth diets; PD, production diets; IVDM, in vitro dry matter digestibility; GE, gross energy; LSD, least significant difference at p value < 0.0001; different superscript letters within a column in the table signify statistical differences among the corresponding values; *, each value is a mean of four observations.

3.6. Correlation between Chemical Constituents and Methane Production

Among the proximate constituents, the EE and lignin were significantly (p < 0.05) negatively associated (r = -0.422 ** and r = -0.365 **) with dietary methane production, while the OM contents of the diets were positively (p < 0.05 r = 0.266 *) correlated with methane production (Table 8). The protein fractions NDIP, ADIP, and P_{B3} of the diets were negatively associated with CH₄ production (r = -0.448 **, r = -0.272 **, and r = -0.341 **). On the other hand, the N fraction, the NPN, the SP, the P_A, and the P_{B1} fraction of the diets were positively associated (r = 0.450 **, 0.387 **, r = 0.412 **, and r = 0.284 **) with the in vitro CH₄ production. Among the diets, the tCHO and carbohydrate fraction C_{B2} contents were positively associated (r = 0.353 ** and 0.278 **) with the in vitro CH₄ production, while the carbohydrate fraction C_C DM was negatively associated (r = -0.365 **) with CH₄ production.

Table 8. Correlation between in vitro methane production and chemical constituents of the diets/rations.

Chemical Constituents	CH ₄ g/g DDM	Protein Fractions	CH ₄ g/g DDM	CHO Fractions	CH ₄ g/g DDM
СР	-0.134	NDIP	-0.448 (**)	tCHO	0.353 (**)
OM	0.266 (**)	ADIP	-0.272 (**)	NSC	0.115
EE	-0.422 (**)	SP	0.387 (**)	SC	0.083
NDF	-0.009	NPN	0.450 (**)	Starch % NSC	-0.104
ADF	-0.127	P_A	0.412 (**)	C _C DM	-0.365 (**)
Cellulose	-0.073	P _{B1}	0.284 (**)	C _{B2} DM	0.278 (**)
Hemi cellulose	0.130	P _{B2}	-0.053	C _{B1} DM	0.031
Lignin	-0.365 (**)	P _{B3}	-0.341 (**)	C _A DM	0.091
Energy	-0.032	P _C	-0.145		

CP, crude protein; OM, organic matter; EE, ether extract; NDF, neutral detergent fiber; ADF, acid detergent fiber; NDIP, neutral detergent-insoluble protein; ADIP, acid detergent-insoluble protein; SP, soluble protein; P_A, non-protein nitrogen; P_{B1}, buffer-soluble protein; P_{B2}, neutral detergent-soluble protein; P_{B3}, acid detergent-soluble protein; P_C, indigestible protein; tCHO, total carbohydrates; NSC, non-structural carbohydrates; SC, structural carbohydrates; C_C, unavailable/lignin-bound cell wall; C_{B2}, slowly degradable cell wall; C_{B1}, intermediately degradable starch and pectin; C_A, rapidly degradable CHO, including sugars; DM, dry matter; DDM, digestible dry matter; **, statistically significant.

4. Discussion

4.1. Chemical Composition

All three diet categories including maintenance, growth, and production showed crude protein (CP) levels equal to or exceeding the minimum required for microbial growth. A minimum of 7.0% CP is essential to optimize the growth and functionality of rumen microbes [34]. The reason for higher CP contents in the production diets (127) than in the growth (112) and maintenance diets (82.0 g/kg DM) may be due to the inclusion of a protein-rich concentrate mixture in the production diets. These values of CP were within the range (64.4 to 150.4 g/kg DM) of values reported for 45 rations [35]. Additionally, the higher values of NDF, ADF, and cellulose contents in the maintenance diets may be due to the sole roughage ingredients in its compositions. Further, the variability in the cell wall constituents of the maintenance, growth, and production diets may be attributed to the composition and level of diverse sources of dry, green, and concentrate mixtures. The average value of the CP and hemicelluloses obtained from the growth and production diets were in similar to the value reported in a combined mixed-diet ration containing low-protein and high-protein rations [36]. The OM contents of the maintenance, growth, and production diets evaluated in the present study were within the range of OM values observed in an experiment involving seven diets [37] and utilizing local-based feed resources and tropical grass pastures [38,39].

4.2. Carbohydrate and Protein Fractions

Carbohydrates and proteins are the two most important constituents of diets required for the different physiological functions of animals, viz., maintenance, growth, and production. The total carbohydrate (tCHO) and NSC content of the maintenance, growth, and production diets recorded in the present study were aligned within the range (773.3-859.4 and 95.1–335.6 g/kg DM) of values reported for 45 rations of different roughage and concentrate feed ratios formulated using different roughage and concentrate feed types [35]. A previous study conducted on top foliage [40] and concentrate feed [10] observed values within a similar range and following a comparable trend. The maintenance, growth, and production diets had the highest contents of carbohydrate fraction C_{B2} (429, 400, and 364 g/kg DM, respectively), following a similar pattern to that of the 45 rations reported by Dong and Zhao [35]; also, our diets' C_{B2} contents were within the range (344.8–588.2 g/kg DM) of values reported in the above-mentioned study. The variations in the concentration of the C_A , C_{B1} , C_{B2} , and C_C carbohydrate fractions of the maintenance, growth, and production diet are similar to those observed for the 45 rations reported in the earlier study mentioned previously [35]. The carbohydrate fractions C_{B2} and C_{B1} 's contents in most of our growth and production diets were within the range of values reported for six farm diets in a previous study [41]: the observed lower content of C_C fraction in the abovementioned study can likely be attributed to the elevated lignin levels in the diets analyzed in our study. The higher lignin content may hinder the release or accessibility of cellulose and hemicellulose, resulting in reduced C_C fraction values. This phenomenon suggests a potential influence of diet composition on the structural components of plant material, with a higher lignin content acting as a limiting factor for the measured C_C fraction. Further, the tCHO values of the growth and production diets were similar to the tCHO values of the diets reported in the above-mentioned study [41], while their NSC contents were relatively higher than our values. The difference in the protein/nitrogen fractions of the maintenance, growth, and production/lactation diets may be attributed to the differences in the proportion of different dietary ingredients and their chemical constituents.

4.3. Gas and Methane Production and Loss of Energy as Methane

The average values of the gas production kinetics at three time intervals (12, 24, and 48 h) and the cumulative gas production (mL/g DM) from the high-protein and low-protein diets showed values higher than the gas production values in the maintenance, growth, and production diets in our study [36]. The gas production from the total mixed rations collected from seven dairies ranged between 211 and 256 mL/g DM after 48 h, which was higher than our gas production values. This variation in gas production may be due to differences in the chemical constituents, mainly the cell wall fractions and the carbohydrate and protein fractions, and their degradability. The availability of nutrients to microbes influences gas production from any feed/diet [42,43]. The total gas production (mL/g DM) from 45 rations of various concentrate to roughage ratios ranged between 165 and 281 [35], which partially agrees with our gas production values. The relatively higher cumulative methane production (mL/g DM) at the 48 h timepoint in the lactation diets (33.1) compared to the maintenance diets (28.4) may be due to the higher digestibility of the production diets, as the degradability of a substrate influences both gas and methane production. In a previous study conducted on lactating cows' diets, the methane production (mL/200 g) reported was higher in the lactating ration (8.85) than in the dry ration (7.24), which substantiates our observations [44]. Further, in same study [44], they recorded higher gas production values in the lactation ration (54.4) than in the dry ration (43.0 mL/200 g). The methane production of 45 rations with varied roughage was the following: the concentrate ration ranged from 30 to 51 mL/g DM after 48 h of fermentation [35], which partially agreed with our results. A similar trend of methane production was observed in a study [45] where goats were fed three diets of different roughage to concentrate ratios (25:75, 50:50 and 73:27), and the values were 37.1, 36.4, and 34.5 g/kg DDM, respectively. Further, the CH_4 (%GE) for these three diets (8.6, 7.3, and 6.0%) differed significantly (p < 0.05), which agreed with our observations that the level of concentrate and the dietary ingredients' composition influences methane production. The percentages of CH₄ and GE were lower in the above-mentioned study than our average values in the maintenance, growth, and production diets.

4.4. Correlation between Methane Production and Chemical Constituents (Proximate Constituents, Carbohydrate Fractions, and Protein Fractions)

The correlations studies between chemical constituents and CH₄ production of forages and concentrate feeds are crucial for optimizing animal nutrition, reducing environmental impact, and improving overall feed efficiency in livestock production [34,46,47]. However, the information on the correlation between methane production and diets/rations' chemical constituents is limited. In this study, the EE and lignin from the proximate constituents and the NDIP and ADIP protein fractions were negatively associated with methane production. Similar to our observations, earlier studies [48–50] reported that EE, lignin, NDIP, and ADIP were negatively associated with methane production. Contrary, a positive correlation between EE and methane production was recorded by Ellis et al. [51]. Information on diets/rations' carbohydrate and/or protein fractions' relationship with in vitro methane production is scarce. In a study of 45 rations, a relationship between CNCPS carbohydrate fractions and methane production was reported [35]. They also reported that the carbohydrate fractions C_A , C_{B1} , and C_{B2} were positively related to methane production, and this agrees of our correlation results. In our study, $C_{\rm C}$ was negatively related to methane production, and this could be due to the unavailability of lignin-bound carbohydrates for digestion. The evaluated diets' soluble protein, NPN, P_A , and P_{B1} were positively associated with methane production. This is probably due to the ready availability of these more degradable protein fractions to microbial fermentation.

This study effectively created diets for water buffaloes, but it has limitations. It mainly looks at diet composition and gas emissions; therefore, future research should explore the diets' long-term effects on buffalo health, practical use on farms, and economic factors to get a fuller picture of their feasibility in real-world farming.

5. Conclusions

This study highlighted key findings on three categories (maintenance, growth, and production/lactation) of thirty different diet compositions for water buffaloes based on local resources, addressing the need for region-specific feeding strategies. The production diets exhibited higher crude protein contents, while the maintenance diets had more fiber. The soluble protein fractions (P_{B1} and P_{B2}) were more present in the production and growth diets and the indigestible fraction (P_C) in the maintenance diets. The higher levels of nonstructural carbohydrates in the production diets suggest dietary optimization possibilities. The loss of energy as CH_4 from the diets/feeding systems varied from 6.48% to 12.56 for the buffalos observed. Amongst the agroecological regions studied (AERs), the livestock from the AER-2 and AER-10 regions emitted the lowest CH₄. The diets in the AER-2 and AER-10 regions consisting of tree leaves as the green fodder source produced less CH₄, with lower losses of dietary energy as methane. The AER-10 diets supplemented with coconut cake as the protein source emitted less CH₄. These findings emphasize the importance of tailoring diets to meet the nutritional needs of buffaloes, marking a significant step forward in optimizing buffalo farming practices. Future implications involve refining agroecological regional feeding practices and considering correlations for a targeted and sustainable diet selection process, promoting both livestock health and environmental stewardship.

Author Contributions: Conceptualization, S.S., B.P.K., P.K., and Y.R.; methodology, S.S., B.P.K., P.K., U.Y.A., S.B., and P.K.; formal analysis, S.S., B.P.K., and P.K.; investigation, S.S., P.K., and B.P.K.; writing—original draft preparation, S.S., B.P.K., P.K., and Y.R.; writing—review and editing, S.S., P.K., U.Y.A., and Y.R.; visualization, S.S., P.K., and Y.R.; supervision, S.S., B.P.K., and Y.R.; project administration, S.S. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted according to the guidelines of animal welfare (animal ethics) being adopted by Indian Council of Agricultural Research (ICAR) institutes and approved by the Institutional Review Board of ICAR-Indian Grassland and Fodder Research Institute (2018).

Informed Consent Statement: Plant-Animal Relationship Division at ICAR-Indian Grassland and Fodder Research Institute, Jhansi, India, maintains fistulated/intact animals for in vitro and in sacco feed and fodder fermentation and degradability studies. The in-house and externally funded projects are presented in the Institute Research Committee (IRC), and the technical programs, including the requirements of animals and feeds/fodders, are approved in this house. The requirement/proposal is reviewed by the Head of the Division, who then approves the proposal to use fistulated buffaloes maintained at the experimental farm to collect rumen liquor for in vitro fermentation studies.

Data Availability Statement: Data are contained within the article.

Acknowledgments: The authors are thankful to Director IGFRI for providing financial support and to the Head Plant Animal Relationship Division for providing the laboratory and animal facilities required to carry out this research.

Conflicts of Interest: The authors have no conflicts of interest.

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