

Review

# Biomarkers Associated with the Outcome of Traumatic Brain Injury Patients

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Received: 29 August 2017; Accepted: 20 October 2017; Published: 27 October 2017

**Abstract:** This review focuses on biomarkers associated with the outcome of traumatic brain injury (TBI) patients, such as caspase-3; total antioxidant capacity; melatonin; S100B protein; glial fibrillary acidic protein (GFAP); glutamate; lactate; brain-derived neurotrophic factor (BDNF); substance P; neuron-specific enolase (NSE); ubiquitin carboxy-terminal hydrolase L-1 (UCH-L1); tau; decanoic acid; and octanoic acid.

**Keywords:** biomarkers; prognosis; mortality; traumatic brain injury; caspase-3; total antioxidant capacity; melatonin; malondialdehyde; substance P

## 1. Introduction

Traumatic brain injury (TBI) results in many deaths, disabilities, and the consumption of healthcare resources each year [1]. All these facts have motivated interest in research on prognostic biomarkers in TBI patients [2–10]. This review focuses on biomarkers associated with the outcome of TBI patients, such as caspase-3, total antioxidant capacity, melatonin, S100B protein, glial fibrillary acidic protein (GFAP), glutamate, lactate, brain-derived neurotrophic factor (BDNF), substance P, neuron-specific enolase (NSE), ubiquitin carboxy-terminal hydrolase L-1 (UCH-L1), tau, decanoic acid, and octanoic acid. The interest in these biomarkers lies in that they could be used as prognostic biomarkers and that the modulation of some of them could open new research lines in the treatment of TBI patients to reduce the risk of mortality.

## 2. Biomarkers

### 2.1. Caspase-3

Cell death by apoptosis occurs mainly through the activation of the extrinsic pathway (death receptor pathway in type I cells) or intrinsic pathway (mitochondrial pathway in type II cells) [11–15]. Apoptosis begins in type I cells when the union between the surface death receptor of tumor necrosis factor receptor superfamily (TNFRSF) and the ligand of tumor necrosis factor superfamily (TNFSF) occurs, which produces the appearance of a death signal and causes pro-caspase-8 to be cleaved in active caspase-8, resulting in the activation of caspase-3. Apoptosis begins in type II cells by the action of pro-inflammatory cytokines such as interleukin (IL)-1 and IL-6, and of oxygen free radicals; afterwards, cytochrome-c is released from mitochondria to cytosol and caspase-3 is activated. Therefore, in both pathways of apoptosis (intrinsic and extrinsic pathways), caspase-3 is activated, and death cell occurs.

In brain tissue samples from animals [16–18] and humans [19,20], apoptotic changes have been found after a TBI; also, in brain tissues from animal models of TBI [21–29], an increase of caspase-3 has been found [21–29]. Higher caspase-3 levels in TBI patients compared to controls in cerebrospinal fluid [30–32] and in brain tissue [19,33] have been found. In addition, higher caspase-3 levels in

the brain tissue (periischemic zone of traumatic cerebral contusions) of non-surviving compared to surviving TBI patients have been found [33]. In a study by our team, we found that there is an association between serum levels of caspase-3 and TBI mortality [34]. I think that these higher serum caspase-3 levels in non-survivor patients could lead to a higher degree of apoptosis.

## 2.2. Total Antioxidant Capacity (TAC)

An increase in reactive oxygen species (ROS) production occurs in TBI and is involved in the development of secondary brain injury [35–38], by means of cellular dysfunction, microvascular regulation loss, vasogenic edema formation, and post-traumatic ischemia. Antioxidants attempt to compensate this increased ROS production to avoid oxidation. TAC determination may give relevant information about the antioxidant status [39,40].

In two studies, significant differences were not found in circulating TAC levels between non-survivor and survivor patients [41,42], and in three studies higher circulating TAC levels were found in non-survivor compared to survivor patients [43–45]. In a study published by our team, an association between serum TAC and mortality in TBI patients was found, as well as an association between serum levels of TAC and TBI severity assessed by the Acute Physiology and Chronic Health Evaluation-II score and the Glasgow Coma Scale, and a positive association between serum levels of TAC and malondialdehyde (MDA) levels [44]. In another study, higher serum TAC levels at admission, 24 h, and 48 h were found in patients with a poor functional outcome at six months post-TBI, in comparison to patients with a good functional outcome [45].

The increase of ROS leads to lipid peroxidation, and MDA is an end-product of this lipid peroxidation due to cellular membrane phospholipids degradation [46,47]. MDA is released into extracellular space and afterwards appears into the blood. The determination of circulating MDA levels has been used in other clinical circumstances, such as sepsis [48,49], brain infarction [50], or hepatocellular carcinoma [51], as an effective biomarker of lipid oxidation. Higher levels of MDA have been found in patients with TBI compared to healthy controls [9,52–56]. In addition, higher levels of MDA were found in non-surviving compared to surviving TBI patients in erythrocytes [55,56] or serum [9,57]. Moreover, in a previous study by our team [9], an association was found between serum MDA levels and mortality in TBI patients.

I believe that these increased serum TAC levels in non-survivor patients compared to survivors may be due to an attempt to compensate the higher free radicals production and higher peroxidation (according to the higher serum MDA concentrations); however, this increase in TAC is not enough to compensate for this unfavorable clinical state.

## 2.3. Melatonin

Melatonin is synthesized with a circadian rhythm (with higher production during the night than during the day) in the pineal gland, and also without circadian rhythm in other organs such as bone marrow, retina, gastrointestinal tract, and thymus [58]. Melatonin, in addition to participating in sleep regulation [59], could have antioxidant, anti-inflammatory, and antiapoptotic effects [60–69].

In some studies, lower concentrations of melatonin were found in TBI patients compared to healthy controls in saliva [70] or serum [71–73]; however, in one study, higher levels of melatonin were found in TBI patients compared to healthy controls in cerebrospinal fluid [74]. In a study recently published by our team, an association between serum levels of melatonin and mortality in TBI patients, as well as a positive association of serum levels of melatonin with serum TAC levels and with serum MDA levels were found [75].

I think that those increased serum melatonin levels in non-survivor TBI patients compared to survivor patients could be due to an attempt to compensate for the unfavorable clinical situation with higher ROS production and higher oxidant states (assessed by increased serum levels of MDA); however, this compensation is not achieved, and finally the death of the patient occurs.

#### 2.4. S100B Protein

This protein has been found in astrocytes and Schwann cells, and also has been detected in other tissues such as melanocytes, bone marrow cells, lymphocytes, chondrocytes, and adipocytes. S100B protein plays a role in the regulation of intracellular levels of calcium and is eliminated by kidneys [76,77]. In some studies, higher circulating S100B levels were found in non-survivor compared to survivor TBI patients [78–84]. It could be possible that those high levels in non-survivor patients indicate the expression of astrocyte damage.

#### 2.5. Glial Fibrillary Acidic Protein (GFAP)

This is the main protein of cytoskeletal filaments in astrocytes, although it is also present in Schwann cells, spleen, and bone marrow. Higher circulating GFAP levels in non-survivor than in survivor TBI patients have been found [85–87]. Those GFAP levels in non-survivor patients could represent higher astrocyte damage.

#### 2.6. Glutamate

Glutamate is the main excitatory amino acid in humans, and high glutamine levels in cerebral microdialysis have been associated with a poor prognosis in TBI patients [88–90]. Recently, one study found higher glutamate levels in cerebrospinal fluid in non-survivor compared to survivor TBI patients [91]. Glutamate at high concentrations could be toxic to neurons involving an excessive influx of  $\text{Na}^+$  and  $\text{Ca}^{2+}$ , mitochondrial dysfunction, and apoptosis [90].

#### 2.7. Lactate

During brain hypoxia, an increase of lactate levels and lactate/pyruvate ratio appears to maintain energy production [92,93]; thus, high lactate levels could represent hypoxia in TBI patients. High levels of lactate have been found in TBI patients with unfavorable outcome in microdialysis [94], as well as in cerebrospinal fluid [91,95,96].

#### 2.8. Brain-Derived Neurotrophic Factor (BDNF)

BDNF is a neurotrophin that protect neurons against glutamate excitotoxicity [97]. Studies on BDNF levels in TBI patients have been scarce, and their results have been controversial; thus, the role in TBI remains unclear. On the one hand, an association between TBI patient outcome and BDNF levels in serum [98] or cerebrospinal fluid [91] have not been found. On the other hand, an association between poor TBI patient outcome and low BDNF levels in serum [99] as well as high BDNF levels in cerebrospinal fluid [100] have been found.

#### 2.9. Substance P

Substance P is a neuropeptide included in the tachykinin family that is mainly synthesized in the central and peripheral nervous systems [101]. Substance P exhibits pro-inflammatory effects, and has been associated with vascular permeability, development of brain edema, and functional deficits after TBI in an animal model [102].

Increased Substance P (SP) in brain tissue samples from TBI patients with neuropathological abnormalities compared with patients without abnormalities have been found [103]. In a study by our team, we found that serum SP levels were associated with TBI severity as well as with early mortality [8].

#### 2.10. Neuron-Specific Enolase (NSE)

NSE is an enzyme present in the cytoplasm of neurons that participates in the glycolysis pathway. An association between high circulating levels of NSE and unfavorable outcome in TBI patients has

been found [104–108]. Those high NSE levels in unfavorable outcome patients could represent greater damage of neurons.

### 2.11. Ubiquitin Carboxy-Terminal Hydrolase L-1 (UCH-L1)

UCH-L1 is an enzyme present in the soma of neurons. Higher circulating levels of UCH-L1 has been found in non-survivor compared to survivor TBI patients [109,110], and may be the expression of neuron damage.

### 2.12. Tau

Tau is a protein of the microtubule-associated protein (MAP) family that stabilizes microtubular assembly in the axons of neurons. High Tau levels in cerebrospinal fluid has been associated with a poor outcome in TBI patients [111]. Those tau levels in non-survivor patients could represent higher astrocyte damage.

### 2.13. Metabolomics

An alteration in brain metabolism appears in TBI, and metabolomics (that consist of a large-scale study of metabolites in a biosample) could help to better our knowledge of TBI pathophysiology [112]. In a recently published metabolomics study, 465 metabolites were determined in serum samples from 211 TBI patients [113]. Out of 465 metabolites, 49 were significantly different between patients with unfavorable and favorable outcomes, most of them upregulated in poor outcome patients. For example, high levels of decanoic acid and octanoic acid (both medium-chain fatty acids) were associated with unfavorable outcome; also, both medium-chain fatty acids have been associated with mitochondrial dysfunction [114].

## 3. Future

The interest in these biomarkers lies in that they could be used as prognostic biomarkers. I think that the evidence that we have at present about these biomarkers will not change the daily clinical practice, and that the sole use of these biomarkers should be taken with caution; however, their use could help in the mortality prediction of TBI patients estimated by other prognostic factors (such as APACHE-II, Glasgow Coma Scale (GCS), age, and computer tomography findings).

Another point of interest concerning these biomarkers is the fact that is possible that the modulation of some of these biomarkers by the administration of different agents could incite research on new lines in the treatment of patients with TBI. In respect to caspase-3, the administration of caspase-3 inhibitors has reduced caspase-3 activity and apoptosis in brain tissues in rat models [24–29]. Regarding the total antioxidant capacity, a reduction of MDA have been found with the administration of different antioxidant agents (memantine, amantadine sulphate, melatonin); for example, a reduction of MDA levels in brain tissues has been achieved in animal models with the administration of memantine [115], as well as a reduction of circulating MDA levels with the administration of amantadine sulphate in TBI patients [116], and a reduction of MDA levels in brain tissues in animal models with the administration of melatonin [117,118]. With respect to melatonin, the administration of melatonin in animal models has led to antioxidant effects [117–123], anti-apoptotic effects [123], a reduction in brain edema [121,122,124], and anti-inflammatory effects [120,125]. With respect to substance P, the administration of an antagonist of neurokinin-1 receptor (which is the receptor of substance P) has reduced substance P activity and brain edema formation, and improved functional outcome [102].

## 4. Conclusions

Circulating levels of caspase-3, total antioxidant capacity, melatonin, S100B protein, glial fibrillary acidic protein, brain-derived neurotrophic factor (BDNF), substance P, neuron-specific enolase (NSE),

ubiquitin carboxy-terminal hydrolase L-1 (UCH-L1), decanoic acid, and octanoic acid, as well as levels in microdialysis and cerebrospinal fluid of glutamate, lactate, and cerebrospinal fluid tau levels have been found to be associated with the outcome of TBI patients. These biomarkers could help in the prognostic classification of TBI patients and could open new research lines in the treatment of patients with TBI.

**Acknowledgments:** This study was supported by a grant from Instituto de Salud Carlos III (INT16/00165) (Madrid, Spain) and co-financed by Fondo Europeo de Desarrollo Regional (FEDER).

**Conflicts of Interest:** The author declares no conflict of interest.

## References

1. Brain Trauma Foundation; American Association of Neurological Surgeons; Congress of Neurological Surgeons. Guidelines for the management of severe traumatic brain injury. *J. Neurotrauma* **2007**, *24*, S1–106.
2. Thelin, E.P.; Zeiler, F.A.; Ercole, A.; Mondello, S.; Büki, A.; Bellander, B.M.; Helmy, A.; Menon, D.K.; Nelson, D.W. Serial Sampling of Serum Protein Biomarkers for Monitoring Human Traumatic Brain Injury Dynamics: A Systematic Review. *Front. Neurol.* **2017**, *8*, 300. [[CrossRef](#)] [[PubMed](#)]
3. Sandmark, D.K. Clinical Outcomes after Traumatic Brain Injury. *Curr. Neurol. Neurosci. Rep.* **2016**, *16*, 52. [[CrossRef](#)] [[PubMed](#)]
4. Toman, E.; Harrisson, S.; Belli, T. Biomarkers in traumatic brain injury: A review. *J. R. Army Med. Corps* **2016**, *162*, 103–108. [[CrossRef](#)] [[PubMed](#)]
5. Lorente, L. New Prognostic Biomarkers in Patients with Traumatic Brain Injury. *Arch. Trauma Res.* **2015**, *4*, e30165. [[CrossRef](#)] [[PubMed](#)]
6. Lorente, L.; Martín, M.M.; López, P.; Ramos, L.; Blanquer, J.; Cáceres, J.J.; Solé-Violán, J.; Solera, J.; Cabrera, J.; Argueso, M.; et al. Association between serum tissue inhibitor of matrix metalloproteinase-1 levels and mortality in patients with severe brain trauma injury. *PLoS ONE* **2014**, *9*, e94370. [[CrossRef](#)] [[PubMed](#)]
7. Lorente, L.; Martín, M.M.; González-Rivero, A.F.; Ramos, L.; Argueso, M.; Cáceres, J.J.; Solé-Violán, J.; Serrano, N.; Rodríguez, S.T.; Jiménez, A.; et al. Serum soluble CD40 Ligand levels are associated with severity and mortality of brain trauma injury patients. *Thromb. Res.* **2014**, *134*, 832–836. [[CrossRef](#)] [[PubMed](#)]
8. Lorente, L.; Martín, M.M.; Almeida, T.; Hernández, M.; Ramos, L.; Argueso, M.; Cáceres, J.J.; Solé-Violán, J.; Jiménez, A. Serum substance P levels are associated with severity and mortality in patients with severe traumatic brain injury. *Crit. Care* **2015**, *19*, 192. [[CrossRef](#)] [[PubMed](#)]
9. Lorente, L.; Martín, M.M.; Abreu-González, P.; Ramos, L.; Argueso, M.; Cáceres, J.J.; Solé-Violán, J.; Lorenzo, J.M.; Molina, I.; Jiménez, A. Association between serum malondialdehyde levels and mortality in patients with severe brain trauma injury. *J. Neurotrauma* **2015**, *32*, 1–6. [[CrossRef](#)] [[PubMed](#)]
10. Lorente, L.; Martín, M.M.; González-Rivero, A.F.; Argueso, M.; Ramos, L.; Solé-Violán, J.; Cáceres, J.J.; Jiménez, A.; Borreguero-León, J.M. Serum levels of caspase-cleaved cytokeratin-18 in patients with severe traumatic brain injury are associated with mortality: A pilot study. *PLoS ONE* **2015**, *10*, e0121739. [[CrossRef](#)] [[PubMed](#)]
11. Cavallucci, V.; D'Amelio, M. Matter of life and death: The pharmacological approaches targeting apoptosis in brain diseases. *Curr. Pharm. Des.* **2011**, *17*, 215–229. [[CrossRef](#)] [[PubMed](#)]
12. Wang, K.; Liu, B.; Ma, J. Research progress in traumatic brain penumbra. *Chin. Med. J.* **2014**, *127*, 1964–1968. [[PubMed](#)]
13. Rovegno, M.; Soto, P.A.; Sáez, J.C.; von Bernhardi, R. Biological mechanisms involved in the spread of traumatic brain damage. *Med. Intensiv.* **2012**, *36*, 37–44. [[CrossRef](#)] [[PubMed](#)]
14. Kunz, A.; Dirnagl, U.; Mergenthaler, P. Acute pathophysiological processes after ischaemic and traumatic brain injury. *Best Pract. Res. Clin. Anaesthesiol.* **2010**, *24*, 495–509. [[CrossRef](#)] [[PubMed](#)]
15. Liu, X.; Zou, H.; Slaughter, C.; Wang, X. DFF, a Heterodimeric Protein That Functions Downstream of Caspase-3 to Trigger DNA Fragmentation during Apoptosis. *Cell* **1997**, *89*, 175–184. [[CrossRef](#)]
16. Raghupathi, R.; Conti, A.C.; Graham, D.I.; Krajewski, S.; Reed, J.C.; Grady, M.S.; Trojanowski, J.Q.; McIntosh, T.K. Mild traumatic brain injury induces apoptotic cell death in the cortex that is preceded by decreases in cellular Bcl-2 immunoreactivity. *Neuroscience* **2002**, *110*, 605–616. [[CrossRef](#)]

17. Villapol, S.; Byrnes, K.R.; Symes, A.J. Temporal dynamics of cerebral blood flow, cortical damage, apoptosis, astrocyte-vasculature interaction and astrogliosis in the pericontusional region after traumatic brain injury. *Front. Neurol.* **2014**, *5*, 82. [[CrossRef](#)] [[PubMed](#)]
18. Chen, R.; Wang, J.; Jiang, B.; Wan, X.; Liu, H.; Liu, H.; Yang, X.; Wu, X.; Zou, Q.; Yang, W. Study of cell apoptosis in the hippocampus and thalamencephalon in a ventricular fluid impact model. *Exp. Ther. Med.* **2013**, *6*, 1463–1468. [[CrossRef](#)] [[PubMed](#)]
19. Clark, R.S.; Kochanek, P.M.; Chen, M.; Watkins, S.C.; Marion, D.W.; Chen, J.; Hamilton, R.L.; Loeffert, J.E.; Graham, S.H. Increases in Bcl-2 and cleavage of caspase-1 and caspase-3 in human brain after head injury. *FASEB J.* **1999**, *13*, 813–821. [[PubMed](#)]
20. Miñambres, E.; Ballesteros, M.A.; Mayorga, M.; Marin, M.J.; Muñoz, P.; Figols, J.; López-Hoyos, M. Cerebral apoptosis in severe traumatic brain injury patients: An in vitro, in vivo, and postmortem study. *J. Neurotrauma* **2008**, *25*, 581–591. [[CrossRef](#)] [[PubMed](#)]
21. Jia, F.; Mao, Q.; Liang, Y.M.; Jiang, J.Y. Effect of post-traumatic mild hypothermia on hippocampal cell death after traumatic brain injury in rats. *J. Neurotrauma* **2009**, *26*, 243–252. [[CrossRef](#)] [[PubMed](#)]
22. Eberspächer, E.; Heimann, K.; Hollweck, R.; Werner, C.; Schneider, G.; Engelhard, K. The effect of electroencephalogram-targeted high- and low-dose propofol infusion on histopathological damage after traumatic brain injury in the rat. *Anesth. Analg.* **2006**, *103*, 1527–1533. [[CrossRef](#)] [[PubMed](#)]
23. Shah, S.A.; Prough, D.S.; Garcia, J.M.; DeWitt, D.S.; Hellmich, H.L. Molecular correlates of age-specific responses to traumatic brain injury in mice. *Exp. Gerontol.* **2006**, *41*, 1201–1205. [[CrossRef](#)] [[PubMed](#)]
24. Saykally, J.N.; Rachmany, L.; Shaer, A.; Rubovitch, V.; Pick, C.G.; Citron, B.A. The nuclear factor erythroid 2-like Hatic, H2 activator, tert-butylhydroquinone, improves cognitive performance in mice after mild traumatic brain injury. *Neuroscience* **2012**, *223*, 305–314. [[CrossRef](#)] [[PubMed](#)]
25. Abrahamson, E.E.; Ikonomovic, M.D.; Ciallella, J.R.; Hope, C.E.; Paljug, W.R.; Isanski, B.A.; Flood, D.G.; Clark, R.S.; DeKosky, S.T. Caspase inhibition therapy abolishes brain trauma-induced increases in Abeta peptide: Implications for clinical outcome. *Exp. Neurol.* **2006**, *197*, 437–450. [[CrossRef](#)] [[PubMed](#)]
26. Soustiel, J.F.; Palzur, E.; Nevo, O.; Thaler, I.; Vlodavsky, E. Neuroprotective anti-apoptosis effect of estrogens in traumatic brain injury. *J. Neurotrauma* **2005**, *22*, 345–352. [[CrossRef](#)] [[PubMed](#)]
27. Clausen, F.; Lundqvist, H.; Ekmark, S.; Lewén, A.; Ebendal, T.; Hillered, L. Oxygen free radical-dependent activation of extracellular signal-regulated kinase mediates apoptosis-like cell death after traumatic brain injury. *J. Neurotrauma* **2004**, *21*, 1168–1182. [[CrossRef](#)] [[PubMed](#)]
28. Clark, R.S.; Kochanek, P.M.; Watkins, S.C.; Chen, M.; Dixon, C.E.; Seidberg, N.A.; Melick, J.; Loeffert, J.E.; Nathaniel, P.D.; Jin, K.L.; et al. Caspase-3 mediated neuronal death after traumatic brain injury in rats. *J. Neurochem.* **2000**, *74*, 740–753. [[CrossRef](#)] [[PubMed](#)]
29. Sanchez Mejia, R.O.; Ona, V.O.; Li, M.; Friedlander, R.M. Minocycline reduces traumatic brain injury-mediated caspase-1 activation, tissue damage, and neurological dysfunction. *Neurosurgery* **2001**, *48*, 1393–1399. [[PubMed](#)]
30. Uzan, M.; Erman, H.; Tanrıverdi, T.; Sanus, G.Z.; Kafadar, A.; Uzun, H. Evaluation of apoptosis in cerebrospinal fluid of patients with severe head injury. *Acta Neurochir.* **2006**, *148*, 1157–1164. [[CrossRef](#)] [[PubMed](#)]
31. Härter, L.; Keel, M.; Hentze, H.; Leist, M.; Ertel, W. Caspase-3 activity is present in cerebrospinal fluid from patients with traumatic brain injury. *J. Neuroimmunol.* **2001**, *121*, 76–78. [[CrossRef](#)]
32. Hentze, H.; Schwoebel, F.; Lund, S.; Keel, M.; Ertel, W.; Wendel, A.; Jäättelä, M.; Leist, M. In vivo and in vitro evidence for extracellular caspase activity released from apoptotic cells. *Biochem. Biophys. Res. Commun.* **2001**, *283*, 1111–1117. [[CrossRef](#)] [[PubMed](#)]
33. Nathoo, N.; Narotam, P.K.; Agrawal, D.K.; Connolly, C.A.; van Dellen, J.R.; Barnett, G.H.; Chetty, R. Influence of apoptosis on neurological outcome following traumatic cerebral contusion. *J. Neurosurg.* **2004**, *101*, 233–240. [[CrossRef](#)] [[PubMed](#)]
34. Lorente, L.; Martín, M.M.; Argueso, M.; Ramos, L.; Solé-Violán, J.; Riaño-Ruiz, M.; Jiménez, A.; Borreguero-León, J.M. Serum caspase-3 levels and mortality are associated in patients with severe traumatic brain injury. *BMC Neurol.* **2015**, *15*, 228. [[CrossRef](#)] [[PubMed](#)]
35. Ikeda, Y.; Long, D.M. The molecular basis of brain injury and brain edema: The role of oxygen free radicals. *Neurosurgery* **1990**, *27*, 1–11. [[CrossRef](#)] [[PubMed](#)]

36. McCall, J.M.; Braughler, J.M.; Hall, E.D. Lipid peroxidation and the role of oxygen radicals in CNS injury. *Acta Anaesthesiol. Belg.* **1990**, *38*, 373–379.
37. Warner, D.S.; Sheng, H.; Batinić-Haberle, I. Oxidants, antioxidants and the ischemic brain. *J. Exp. Biol.* **2004**, *207*, 3221–3231. [CrossRef] [PubMed]
38. Hall, E.D. Lipid antioxidants in acute central nervous system injury. *Ann. Emerg. Med.* **1993**, *22*, 1022–1027. [CrossRef]
39. Young, I.S.; Woodside, J.V. Antioxidants in health and disease. *J. Clin. Pathol.* **2001**, *54*, 176–186. [CrossRef] [PubMed]
40. Ghiselli, A.; Serafini, M.; Natella, F.; Scaccini, C. Total antioxidant capacity as a tool to assess redox status: Critical view and experimental data. *Free Radic. Biol. Med.* **2000**, *29*, 1106–1114. [CrossRef]
41. Sögüt, O.; Kaya, H.; Gökdemir, M.T.; Solduk, L.; Dokuzoglu, M.A.; Sayhan, M.B.; Kaya, A.; Koçyigit, A. Early oxidative status in adult patients with isolated traumatic brain injury. *Turk. J. Med. Sci.* **2012**, *42*, 1010–1019.
42. Kaya, H.; Sögüt, O.; Gökdemir, M.T.; Albayrak, A.T. The role of oxidative status in initial evaluation of paediatric patients with graded traumatic brain injury. *Hong Kong J. Emerg. Med.* **2013**, *23*, 225–233.
43. Kavaklı, H.S.; Erel, O.; Karakayali, O.; Neselioglu, S.; Tanrıverdi, F.; Coskun, F.; Kahraman, A.F. Oxidative stress in isolated blunt traumatic brain injury. *Sci. Res. Essays* **2010**, *5*, 2832–2836.
44. Lorente, L.; Martín, M.M.; Almeida, T.; Abreu-González, P.; Ramos, L.; Argueso, M.; Riaño-Ruiz, M.; Solé-Violán, J.; Jiménez, A. Total antioxidant capacity is associated with mortality of patients with severe traumatic brain injury. *BMC Neurol.* **2015**, *15*, 115. [CrossRef] [PubMed]
45. Rodríguez-Rodríguez, A.; Egéa-Guerrero, J.J.; Vilches-Arenas, Á.; Gordillo-Escobar, E.; Ruiz de Azúa-López, Z.; Murillo-Cabezas, F. Prognostic value of total antioxidant capacity to predict functional outcome in traumatic brain injury patients. *Clin. Chem. Lab. Med.* **2017**, *55*, e265–e267. [CrossRef] [PubMed]
46. Draper, H.H.; Hadley, M. Malondialdehyde determination as index of lipid peroxidation. *Methods Enzymol.* **1990**, *186*, 421–431. [PubMed]
47. Dalle-Donne, I.; Rossi, R.; Colombo, R.; Giustarini, D.; Milzani, A. Biomarkers of oxidative damage in human disease. *Clin. Chem.* **2006**, *52*, 601–623. [CrossRef] [PubMed]
48. Lorente, L.; Martín, M.M.; Abreu-González, P.; Domínguez-Rodríguez, A.; Labarta, L.; Díaz, C.; Solé-Violán, J.; Ferreres, J.; Borreguero-León, J.M.; Jiménez, A.; et al. Prognostic value of malondialdehyde serum levels in severe sepsis: A multicenter study. *PLoS ONE* **2013**, *8*, e53741. [CrossRef] [PubMed]
49. Lorente, L.; Martín, M.M.; Abreu-González, P.; Domínguez-Rodríguez, A.; Labarta, L.; Díaz, C.; Solé-Violán, J.; Ferreres, J.; Cabrera, J.; Igéño, J.C.; et al. Sustained high serum malondialdehyde levels are associated with severity and mortality in septic patients. *Crit. Care* **2013**, *17*, R290. [CrossRef] [PubMed]
50. Lorente, L.; Martín, M.M.; Abreu-González, P.; Ramos, L.; Argueso, M.; Solé-Violán, J.; Riaño-Ruiz, M.; Jiménez, A. Serum malondialdehyde levels in patients with malignant middle cerebral artery infarction are associated with mortality. *PLoS ONE* **2015**, *10*, e0125893. [CrossRef] [PubMed]
51. Lorente, L.; Rodriguez, S.T.; Sanz, P.; Abreu-González, P.; Díaz, D.; Moreno, A.M.; Borja, E.; Martín, M.M.; Jiménez, A.; Barrera, M.A. Association between Pre-Transplant Serum Malondialdehyde Levels and Survival One Year after Liver Transplantation for Hepatocellular Carcinoma. *Int. J. Mol. Sci.* **2016**, *17*, 500. [CrossRef] [PubMed]
52. Hu, S.; Zheng, L.; Chen, B.; Xie, J.; Yang, C. The role of the leukocytes in pathogenesis of secondary brain injury. *Hunan Yi Ke Da Xue Xue Bao* **1999**, *24*, 56–58. [PubMed]
53. Hohl, A.; Gullo-Jda, S.; Silva, C.C.; Bertotti, M.M.; Felisberto, F.; Nunes, J.C.; de Souza, B.; Petronilho, F.; Soares, F.M.; Prediger, R.D.; et al. Plasma levels of oxidative stress biomarkers and hospital mortality in severe head injury: A multivariate analysis. *J. Crit. Care* **2012**, *27*, 523.e11–523.e19. [CrossRef] [PubMed]
54. Cristofori, L.; Tavazzi, B.; Gambin, R.; Vagnozzi, R.; Vivenza, C.; Amorini, A.M.; Di Pierro, D.; Fazzina, G.; Lazzarino, G. Early onset of lipid peroxidation after human traumatic brain injury: A fatal limitation for the free radical scavenger pharmacological therapy? *J. Investigig. Med.* **2001**, *49*, 450–458. [CrossRef] [PubMed]
55. Nayak, C.; Nayak, D.; Bhat, S.; Raja, A.; Rao, A. Relationship between neurological outcome and early oxidative changes in erythrocytes in head injury patients. *Clin. Chem. Lab. Med.* **2007**, *45*, 629–633. [CrossRef] [PubMed]
56. Kasprzak, H.A.; Woźniak, A.; Drewna, G.; Woźniak, B. Enhanced lipid peroxidation processes in patients after brain contusion. *J. Neurotrauma* **2001**, *18*, 793–797. [CrossRef] [PubMed]

57. Paolin, A.; Nardin, L.; Gaetani, P.; Rodriguez, Y.; Baena, R.; Pansarasa, O.; Marzatico, F. Oxidative damage after severe head injury and its relationship to neurological outcome. *Neurosurgery* **2002**, *51*, 949–954. [PubMed]
58. Cagnacci, A. Melatonin in relation to physiology in adult humans. *J. Pineal Res.* **1996**, *21*, 200–202. [CrossRef] [PubMed]
59. Dawson, D.; Encel, N. Melatonin and sleep in humans. *J. Pineal Res.* **1993**, *15*, 1–12. [CrossRef] [PubMed]
60. Galano, A.; Tan, D.X.; Reiter, R.J. Melatonin as a natural ally against oxidative stress: A physicochemical examination. *J. Pineal Res.* **2011**, *51*, 1–16. [CrossRef] [PubMed]
61. Mauriz, J.L.; Collado, P.S.; Veneroso, C.; Reiter, R.J.; Gonzalez-Gallego, J. A review of the molecular aspects of melatonin’s anti-inflammatory actions: Recent insights and news perspectives. *J. Pineal Res.* **2013**, *54*, 1–14. [CrossRef] [PubMed]
62. Kurdi, M.S.; Patel, T. The role of melatonin in anaesthesia and critical care. *Indian J. Anaesth.* **2013**, *57*, 137–144. [CrossRef] [PubMed]
63. Bourne, R.S.; Mills, G.H. Melatonin: Possible implications for the postoperative and critically ill patient. *Intensive Care Med.* **2006**, *32*, 371–379. [CrossRef] [PubMed]
64. Reiter, R.J.; Paredes, S.D.; Manchester, L.C.; Tan, D.X. Reducing oxidative/nitrosative stress: A newly-discovered genre for melatonin. *Crit. Rev. Biochem. Mol. Biol.* **2009**, *44*, 175–200. [CrossRef] [PubMed]
65. Esposito, E.; Cuzzocrea, S. Antiinflammatory activity of melatonin in central nervous system. *Curr. Neuropharmacol.* **2010**, *8*, 228–242. [CrossRef] [PubMed]
66. Samantaray, S.; Das, A.; Thakore, N.P.; Matzelle, D.D.; Reiter, R.J.; Ray, S.K.; Banik, N.L. Therapeutic potential of melatonin in traumatic central nervous system injury. *J. Pineal Res.* **2009**, *47*, 134–142. [CrossRef] [PubMed]
67. Maldonado, M.D.; Murillo-Cabezas, F.; Terron, M.P.; Flores, L.J.; Tan, D.X.; Manchester, L.C.; Reiter, R.J. The potential of melatonin in reducing morbidity-mortality after craniocerebral trauma. *J. Pineal Res.* **2007**, *42*, 1–11. [CrossRef] [PubMed]
68. Naseem, M.; Parvez, S. Role of melatonin in traumatic brain injury and spinal cord injury. *Sci. World J.* **2014**, *2014*, 586270. [CrossRef] [PubMed]
69. Fernández-Gajar, R.; Matamala, J.M.; Carrasco, R.; Gutiérrez, R.; Melo, R.; Rodrigo, R. Novel therapeutic strategies for traumatic brain injury: Acute antioxidant reinforcement. *CNS Drugs* **2014**, *28*, 229–248. [CrossRef] [PubMed]
70. Shekleton, J.A.; Parcell, D.L.; Redman, J.R.; Phipps-Nelson, J.; Ponsford, J.L.; Rajaratnam, S.M. Sleep disturbance and melatonin levels following traumatic brain injury. *Neurology* **2010**, *74*, 1732–1738. [CrossRef] [PubMed]
71. Seifman, M.A.; Gomes, K.; Nguyen, P.N.; Bailey, M.; Rosenfeld, J.V.; Cooper, D.J.; Morganti-Kossmann, M.C. Measurement of serum melatonin in intensive care unit patients: Changes in traumatic brain injury, trauma, and medical conditions. *Front. Neurol.* **2014**, *5*, 237. [CrossRef] [PubMed]
72. Paul, T.; Lemmer, B. Disturbance of circadian rhythms in analgosedated intensive care unit patients with and without craniocerebral injury. *Chronobiol. Int.* **2007**, *24*, 45–61. [CrossRef] [PubMed]
73. Paparrigopoulos, T.; Melissaki, A.; Tsekou, H.; Efthymiou, A.; Kribeni, G.; Baziotis, N.; Geronikola, X. Melatonin secretion after head injury: A pilot study. *Brain Inj.* **2006**, *20*, 873–878. [CrossRef] [PubMed]
74. Seifman, M.A.; Adamides, A.A.; Nguyen, P.N.; Vallance, S.A.; Cooper, D.J.; Kossmann, T.; Rosenfeld, J.V.; Morganti-Kossmann, M.C. Endogenous melatonin increases in cerebrospinal fluid of patients after severe traumatic brain injury and correlates with oxidative stress and metabolic disarray. *J. Cereb. Blood Flow Metab.* **2008**, *28*, 684–696. [CrossRef] [PubMed]
75. Lorente, L.; Martín, M.M.; Abreu-González, P.; Pérez-Cejas, A.; Ramos, L.; Argueso, M.; Solé-Violán, J.; Cáceres, J.J.; Jiménez, A.; García-Marín, V. Serum melatonin levels in survivor and non-survivor patients with traumatic brain injury. *BMC Neurol.* **2017**, *17*, 138. [CrossRef] [PubMed]
76. Donato, R.; Sorci, G.; Riuzzi, F.; Arcuri, C.; Bianchi, R.; Brozzi, F.; Tubaro, C.; Giambanco, I. S100B’s double life: Intracellular regulator and extracellular signal. *Biochim. Biophys. Acta* **2009**, *793*, 1008–1022. [CrossRef] [PubMed]
77. Rezaei, O.; Pakdaman, H.; Gharehgozli, K.; Simani, L.; Vahedian-Azimi, A.; Asaadi, S.; Sahraei, Z.; Hajiesmaeli, M. S100 B: A new concept in neurocritical care. *Iran J. Neurol.* **2017**, *16*, 83–89. [PubMed]

78. Pelinka, L.E.; Kroepfl, A.; Leixnering, M.; Buchinger, W.; Raabe, A.; Redl, H. GFAP versus S100B in serum after traumatic brain injury: Relationship to brain damage and outcome. *J. Neurotrauma* **2004**, *21*, 1553–1561. [CrossRef] [PubMed]
79. Petzold, A.; Green, A.J.; Keir, G.; Fairley, S.; Kitchen, N.; Smith, M.; Thompson, E.J. Role of serum S100B as an early predictor of high intracranial pressure and mortality in brain injury: A pilot study. *Crit. Care Med.* **2002**, *30*, 2705–2710. [CrossRef] [PubMed]
80. Dimopoulou, I.; Korfiatis, S.; Dafni, U.; Anthi, A.; Psachoulia, C.; Jullien, G.; Sakas, D.E.; Roussos, C. Protein S-100b serum levels in trauma-induced brain death. *Neurology* **2003**, *60*, 947–951. [CrossRef] [PubMed]
81. Thelin, E.P.; Johannesson, L.; Nelson, D.; Bellander, B.M. S100B is an important outcome predictor in traumatic brain injury. *J. Neurotrauma* **2013**, *30*, 519–928. [CrossRef] [PubMed]
82. Korfiatis, S.; Stranjalis, G.; Boviatidis, E.; Psachoulia, C.; Jullien, G.; Gregson, B.; Mendelow, A.D.; Sakas, D.E. Serum S-100B protein monitoring in patients with severe traumatic brain injury. *Intensive Care Med.* **2007**, *33*, 255–260. [CrossRef] [PubMed]
83. Rothoerl, R.D.; Woertgen, C.; Holzschuh, M.; Metz, C.; Brawanski, A. Rapid evaluation of S-100 serum levels. Case report and comparison to previous results. *Brain Inj.* **1999**, *13*, 387–391. [PubMed]
84. Ucar, T.; Baykal, A.; Akyuz, M.; Dosemeci, L.; Toptas, B. Comparison of serum and cerebrospinal fluid protein S-100b levels after severe head injury and their prognostic importance. *J. Trauma* **2004**, *57*, 95–98. [CrossRef] [PubMed]
85. Pelinka, L.E.; Kroepfl, A.; Schmidhammer, R.; Krenn, M.; Buchinger, W.; Redl, H.; Raabe, A. Glial fibrillary acidic protein in serum after traumatic brain injury and multiple trauma. *J. Trauma* **2004**, *57*, 1006–1212. [CrossRef] [PubMed]
86. Raheja, A.; Sinha, S.; Samson, N.; Bhoi, S.; Subramanian, A.; Sharma, P.; Sharma, B.S. Serum biomarkers as predictors of long-term outcome in severe traumatic brain injury: Analysis from a randomized placebo-controlled Phase II clinical trial. *J. Neurosurg.* **2016**, *125*, 631–641. [CrossRef] [PubMed]
87. Mondello, S.; Papa, L.; Buki, A.; Bullock, M.R.; Czeiter, E.; Tortella, F.C.; Wang, K.K.; Hayes, R.L. Neuronal and glial markers are differently associated with computed tomography findings and outcome in patients with severe traumatic brain injury: A case control study. *Crit. Care* **2011**, *15*, R156. [CrossRef] [PubMed]
88. Bullock, R.; Zauner, A.; Woodward, J.J.; Myseros, J.; Choi, S.C.; Ward, J.D.; Marmarou, A.; Young, H.F. Factors affecting excitatory amino acid release following severe human head injury. *J. Neurosurg.* **1998**, *89*, 507–518. [CrossRef] [PubMed]
89. Koura, S.S.; Doppenberg, E.M.; Marmarou, A.; Choi, S.; Young, H.F.; Bullock, R. Relationship between excitatory amino acid release and outcome after severe human head injury. *Acta Neurochir. Suppl.* **1998**, *71*, 244–246. [PubMed]
90. Chamoun, R.; Suki, D.; Gopinath, S.P.; Goodman, J.C.; Robertson, C. Role of extracellular glutamate measured by cerebral microdialysis in severe traumatic brain injury. *J. Neurosurg.* **2010**, *113*, 564–570. [CrossRef] [PubMed]
91. Stefani, M.A.; Modkowska, R.; Hansel, G.; Zimmer, E.R.; Kopczynski, A.; Muller, A.P.; Strogulski, N.R.; Rodolphi, M.S.; Carteri, R.K.; Schmidt, A.P.; et al. Elevated glutamate and lactate predict brain death after severe head trauma. *Ann. Clin. Transl. Neurol.* **2017**, *4*, 392–402. [CrossRef] [PubMed]
92. Carpenter, K.L.; Jalloh, I.; Hutchinson, P.J. Glycolysis and the significance of lactate in traumatic brain injury. *Front. Neurosci.* **2015**, *9*, 112. [CrossRef] [PubMed]
93. Lazaridis, C.; Andrews, C.M. Brain tissue oxygenation, lactate-pyruvate ratio, and cerebrovascular pressure reactivity monitoring in severe traumatic brain injury: Systematic review and viewpoint. *Neurocrit. Care* **2014**, *21*, 345–355. [CrossRef] [PubMed]
94. Timofeev, I.; Carpenter, K.L.; Nortje, J.; Al-Rawi, P.G.; O’Connell, M.T.; Czosnyka, M.; Smielewski, P.; Pickard, J.D.; Menon, D.K.; Kirkpatrick, P.J.; et al. Cerebral extracellular chemistry and outcome following traumatic brain injury: A microdialysis study of 223 patients. *Brain* **2011**, *134*, 484–494. [CrossRef] [PubMed]
95. Thelin, E.P.; Nelson, D.W.; Ghatal, P.H.; Bellander, B.M. Microdialysis Monitoring of CSF Parameters in Severe Traumatic Brain Injury Patients: A Novel Approach. *Front. Neurol.* **2014**, *5*, 159. [CrossRef] [PubMed]
96. DeSalles, A.A.; Kontos, H.A.; Becker, D.P.; Yang, M.S.; Ward, J.D.; Moulton, R.; Gruemer, H.D.; Lutz, H.; Maset, A.L.; Jenkins, L. Prognostic significance of ventricular CSF lactic acidosis in severe head injury. *J. Neurosurg.* **1986**, *65*, 615–624. [CrossRef] [PubMed]

97. Almeida, R.D.; Manadas, B.J.; Melo, C.V.; Gomes, J.R.; Mendes, C.S.; Grãos, M.M.; Carvalho, R.F.; Carvalho, A.P.; Duarte, C.B. Neuroprotection by BDNF against glutamate-induced apoptotic cell death is mediated by ERK and PI3-kinase pathways. *Cell Death Differ.* **2005**, *12*, 1329–1343. [CrossRef] [PubMed]
98. Simon, D.; Nascimento, R.I.; Filho, E.M.; Bencke, J.; Regner, A. Plasma brain-derived neurotrophic factor levels after severe traumatic brain injury. *Brain Inj.* **2016**, *30*, 23–28. [CrossRef] [PubMed]
99. Korley, F.K.; Diaz-Arrastia, R.; Wu, A.H.; Yue, J.K.; Manley, G.T.; Sair, H.I.; Van Eyk, J.; Everett, A.D.; TRACK-TBI investigators; Okonkwo, D.O.; et al. Circulating Brain-Derived Neurotrophic Factor Has Diagnostic and Prognostic Value in Traumatic Brain Injury. *J. Neurotrauma* **2016**, *33*, 215–225. [CrossRef] [PubMed]
100. Failla, M.D.; Conley, Y.P.; Wagner, A.K. Brain-Derived Neurotrophic Factor (BDNF) in Traumatic Brain Injury-Related Mortality: Interrelationships between Genetics and Acute Systemic and Central Nervous System BDNF Profiles. *Neurorehabilit. Neural Repair.* **2016**, *30*, 83–93. [CrossRef] [PubMed]
101. Vink, R.; Gabrielian, L.; Thornton, E. The Role of Substance P in Secondary Pathophysiology after Traumatic Brain Injury. *Front. Neurol.* **2017**, *8*, 304. [CrossRef] [PubMed]
102. Donkin, J.J.; Nimmo, A.J.; Cernak, I.; Blumbergs, P.C.; Vink, R. Substance P is associated with the development of brain edema and functional deficits after traumatic brain injury. *J. Cereb. Blood Flow Metab.* **2009**, *29*, 1388–1398. [CrossRef] [PubMed]
103. Zacest, A.C.; Vink, R.; Manavis, J.; Sarvestani, G.T.; Blumbergs, P.C. Substance P immunoreactivity increases following human traumatic brain injury. *Acta Neurochir. Suppl.* **2010**, *106*, 211–216. [PubMed]
104. Beers, S.R.; Berger, R.P.; Adelson, P.D. Neurocognitive outcome and serum biomarkers in inflicted versus non-inflicted traumatic brain injury in young children. *J. Neurotrauma* **2007**, *24*, 97–105. [CrossRef] [PubMed]
105. Olivecrona, Z.; Bobinski, L.; Koskinen, L.O. Association of ICP, CPP, CT findings and S-100B and NSE in severe traumatic head injury. Prognostic value of the biomarkers. *Brain Inj.* **2015**, *29*, 446–454. [CrossRef] [PubMed]
106. Rodríguez-Rodríguez, A.; Egea-Guerrero, J.J.; Gordillo-Escobar, E.; Enamorado-Enamorado, J.; Hernández-García, C.; Ruiz de Azúa-López, Z.; Vilches-Arenas, Á.; Guerrero, J.M.; Murillo-Cabezas, F. S100B and Neuron-Specific Enolase as mortality predictors in patients with severe traumatic brain injury. *Neurol. Res.* **2016**, *38*, 130–137. [CrossRef] [PubMed]
107. Woertgen, C.; Rothoerl, R.D.; Holzschuh, M.; Metz, C.; Brawanski, A. Comparison of serial S-100 and NSE serum measurements after severe head injury. *Acta Neurochir.* **1997**, *139*, 1161–1164. [CrossRef] [PubMed]
108. Žurek, J.; Fedora, M. The usefulness of S100B, NSE, GFAP, NF-H, secretagogin and Hsp70 as a predictive biomarker of outcome in children with traumatic brain injury. *Acta Neurochir.* **2012**, *154*, 93–103. [CrossRef] [PubMed]
109. Brophy, G.M.; Mondello, S.; Papa, L.; Robicsek, S.A.; Gabrielli, A.; Tepas, J., 3rd; Buki, A.; Robertson, C.; Tortella, F.C.; Hayes, R.L.; et al. Biokinetic analysis of ubiquitin C-terminal hydrolase-L1 (UCH-L1) in severe traumatic brain injury patient biofluids. *J. Neurotrauma* **2011**, *28*, 861–870. [CrossRef] [PubMed]
110. Mondello, S.; Linnet, A.; Buki, A.; Robicsek, S.; Gabrielli, A.; Tepas, J.; Papa, L.; Brophy, G.M.; Tortella, F.; Hayes, R.L.; et al. Clinical utility of serum levels of ubiquitin C-terminal hydrolase as a biomarker for severe traumatic brain injury. *Neurosurgery* **2012**, *70*, 666–675. [PubMed]
111. Ost, M.; Nylén, K.; Csajbok, L.; Ohrfelt, A.O.; Tullberg, M.; Wikkelsö, C.; Nellgård, P.; Rosengren, L.; Blennow, K.; Nellgård, B. Initial CSF total tau correlates with 1-year outcome in patients with traumatic brain injury. *Neurology* **2006**, *67*, 1600–1604. [CrossRef] [PubMed]
112. Posti, J.P.; Dickens, A.M.; Orešić, M.; Hyötyläinen, T.; Tenovuo, O. Metabolomics Profiling As a Diagnostic Tool in Severe Traumatic Brain Injury. *Front. Neurol.* **2017**, *8*, 398. [CrossRef] [PubMed]
113. Orešić, M.; Posti, J.P.; Kamstrup-Nielsen, M.H.; Takala, R.S.; Lingsma, H.F.; Mattila, I.; Jäntti, S.; Katila, A.J.; Carpenter, K.L.; Ala-Seppälä, H.; et al. Human Serum Metabolites Associate With Severity and Patient Outcomes in Traumatic Brain Injury. *EbioMedicine* **2016**, *12*, 118–126. [CrossRef] [PubMed]
114. Schuck, P.F.; Ferreira Gde, C.; Tonin, A.M.; Viegas, C.M.; Busanello, E.N.; Moura, A.P.; Zanatta, A.; Klamt, F.; Wajner, M. Evidence that the major metabolites accumulating in medium-chain acyl-CoA dehydrogenase deficiency disturb mitochondrial energy homeostasis in rat brain. *Brain Res.* **2009**, *1296*, 117–126. [CrossRef] [PubMed]
115. Ozsüer, H.; Görgülü, A.; Kiriş, T.; Cobanoğlu, S. The effects of memantine on lipid peroxidation following closed-head trauma in rats. *Neurosurg. Rev.* **2005**, *28*, 143–147. [CrossRef] [PubMed]

116. Sanova, B.; Drobny, M.; Lehotsky, J.; Sulaj, M.; Schudichova, J. Biochemical and clinical improvement of cytotoxic state by amantadine sulphate. *Cell. Mol. Neurobiol.* **2006**, *26*, 1475–1482. [[CrossRef](#)] [[PubMed](#)]
117. Kerman, M.; Cirak, B.; Ozguner, M.F.; Dagtekin, A.; Sutcu, R.; Altuntas, I.; Delibas, N. Does melatonin protect or treat brain damage from traumatic oxidative stress? *Exp. Brain Res.* **2005**, *163*, 406–410. [[CrossRef](#)] [[PubMed](#)]
118. Horakova, L.; Onrejickova, O.; Barchrrata, K.; Vajdova, M. Preventive effect of several antioxidants after oxidative stress on rat brain homogenates. *Gen. Physiol. Biophys.* **2000**, *19*, 195–205. [[PubMed](#)]
119. Messenge, C.; Margail, I.; Verrecchia, C.; Allix, M. Protective effect of melatonin in a model of traumatic brain injury in mice. *J. Pineal Res.* **1998**, *25*, 41–46. [[CrossRef](#)]
120. Tsai, M.C.; Chen, W.J.; Tsai, M.S.; Ching, C.H.; Chuang, J.I. Melatonin attenuates brain contusion-induced oxidative insult, inactivation of signal transducers and activators of transcription 1, and upregulation of suppressor of cytokine signaling-3 in rats. *J. Pineal Res.* **2011**, *51*, 233–245. [[CrossRef](#)] [[PubMed](#)]
121. Dehghan, F.; Khaksari Hadad, M.; Asadikram, G.; Najafipour, H.; Shahrokhi, N. Effect of melatonin on intracranial pressure and brain edema following traumatic brain injury: Role of oxidative stresses. *Arch. Med. Res.* **2013**, *44*, 251–258. [[CrossRef](#)] [[PubMed](#)]
122. Ding, K.; Wang, H.; Xu, J.; Li, T.; Zhang, L.; Ding, Y.; Zhu, L.; He, J.; Zhou, M. Melatonin stimulates antioxidant enzymes and reduces oxidative stress in experimental traumatic brain injury: The Nrf2-ARE signaling pathway as a potential mechanism. *Free Radic. Biol. Med.* **2014**, *73*, 1–11. [[CrossRef](#)] [[PubMed](#)]
123. Yürüker, V.; Naziroğlu, M.; Şenol, N. Reduction in traumatic brain injury-induced oxidative stress, apoptosis, and calcium entry in rat hippocampus by melatonin: Possible involvement of TRPM2 channels. *Metab. Brain Dis.* **2015**, *30*, 223–231. [[CrossRef](#)] [[PubMed](#)]
124. Kabadi, S.V.; Maher, T.J. Posttreatment with uridine and melatonin following traumatic brain injury reduces edema in various brain regions in rats. *Ann. N. Y. Acad. Sci.* **2010**, *1199*, 105–113. [[CrossRef](#)] [[PubMed](#)]
125. Ding, K.; Wang, H.; Xu, J.; Lu, X.; Zhang, L.; Zhu, L. Melatonin reduced microglial activation and alleviated neuroinflammation induced neuron degeneration in experimental traumatic brain injury: Possible involvement of mTOR pathway. *Neurochem. Int.* **2014**, *76*, 23–31. [[CrossRef](#)] [[PubMed](#)]



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