

Method. *Nuclear factor-erythroid-2-related factor 2 (Nrf-2) myonuclear localization*

A detailed subcellular localization analysis of Nrf2 in myonuclei indirectly is an index of the transcription factor activation ratio.

Human skeletal muscle biopsy cryosections (8 μ m thickness), from VL (n=7) pre-, post- and rec-BR groups, were double immunolabelled with anti-Dystrophin (mouse monoclonal 107416, Novocastra, Leica Biosystems.com) and anti-Nrf-2 (rabbit polyclonal, sc-13032, Santa Cruz, Santa Cruz Biotechnology, Inc.) antibodies. In all Nrf-2 immunostaining protocols goat anti-rabbit Alexa-488 for Nrf-2 and goat anti-mouse Alexa-555 for Dystrophin were used. Nuclei were routinely counterstained with blue DAPI.

Nrf-2 double immunostained muscle cryosections were inspected by confocal microscopy (Leica TCS SP8, leica-microsystems.com) for high resolution immunosignal detection of Nrf2-accumulation in myonuclei. At least 70 myonuclei were scored for each pre-, post-BR, and R+10 muscle biopsy/subject.

Figure S 2. Anti-oxidative master gene Nrf-2 regulation in VL of bed rest subjects. **a.** Representative image Nrf-2 immunopositive myonuclei (green) in VL before (Pre) and after (Post) bedrest in CTR and Cocktail group. Dystrophin (red) used as reference marker to label the subsarcolemmal plasma membrane border; nuclei were counterstained with DAPI (blue). Inset, magnification Nrf-2 positive myonuclei for comparison with Nrf-2 negative myonuclei and nuclei outside the myofiber. **b.** Quantitative analysis of Nrf-2 positive myonuclei in VL. The total number of Nrf-2 positive myonuclei was decreased in both VL CTR (Pre, $18.95 \pm 3.56\%$, Post, $2.38 \pm 0.94\%$, $p \leq 0.001$) and Cocktail (Pre, $23.4 \pm 4.43\%$, Post, $8.27 \pm 1.94\%$, $p \leq 0.05$) group. (* = significant difference, ANOVA and *t*-test, n= 10, *p*-value ≤ 0.05). The graph represents the mean \pm SEM.

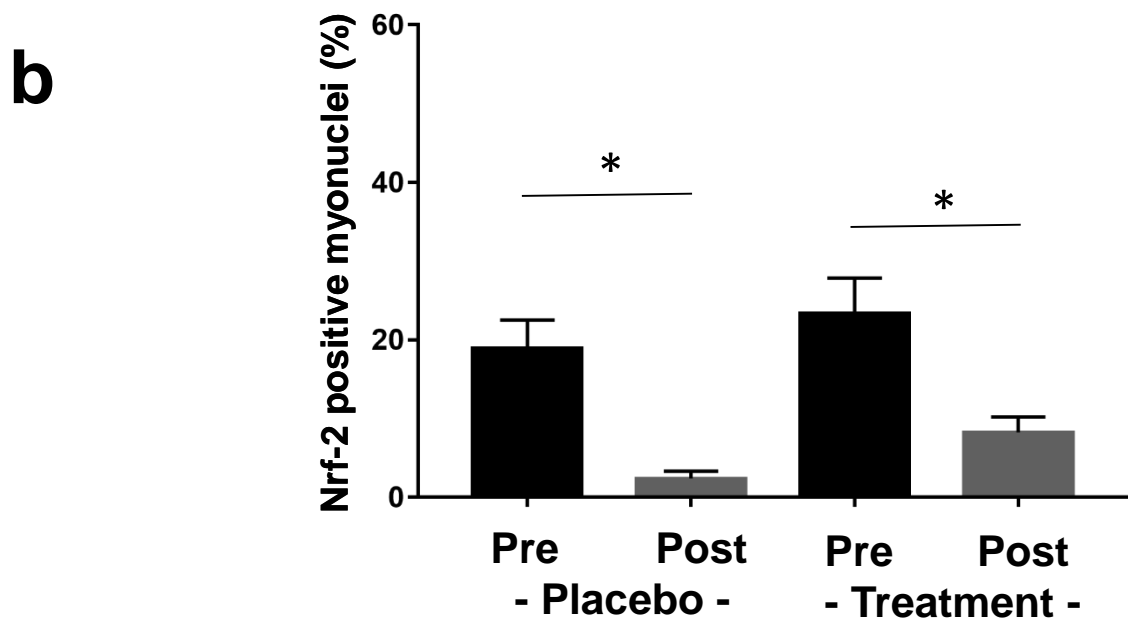
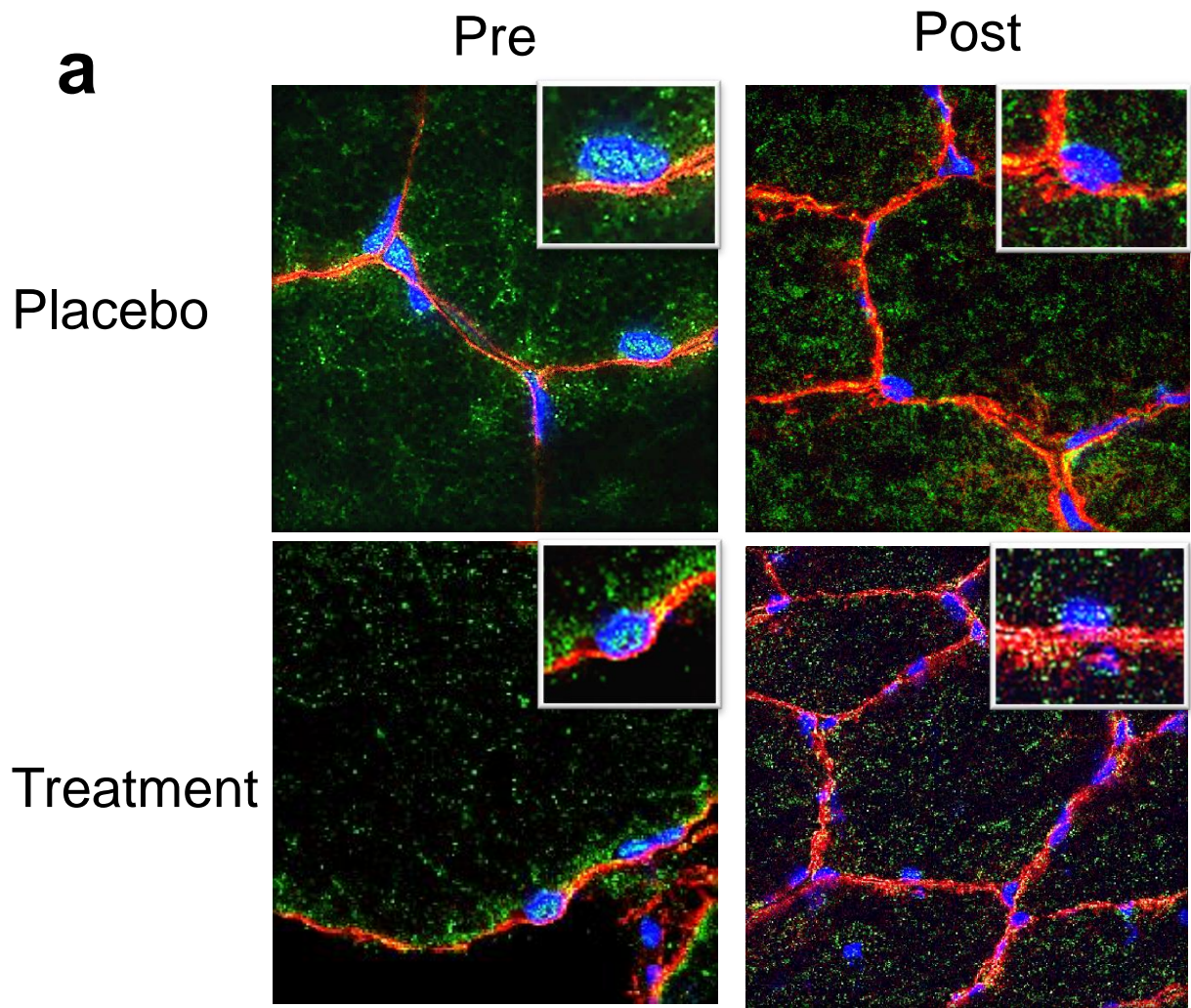


Figure S4