

High performance liquid chromatography (HPLC) analysis of phenolic and terpenophenolic compounds

The HPLC apparatus consisted of a two PU-2080 PLUS chromatographic pump, a DG-2080-54 line degasser, a mix-2080-32 mixer, diode array (DAD), an AS-2057 PLUS autosampler and a CO-2060 PLUS column thermostat (all from Jasco, Tokyo, Japan). Integration was performed by ChromNAV2 Chromatography software. Before injecting in the HPLC apparatus, hemp extracts were centrifuged at 3500× g for 15 min, and supernatant diluted at 1 mg/mL for the identification and quantitative analysis of phenolic and terpenophenolic compounds through comparison with respective standards.

Regarding the phenol quantitative determination, the extracts were analyzed using a reversed-phase HPLC-DAD in gradient elution mode (Table S1). The separation was conducted within 60 min of the chromatographic run, starting from the following separation conditions: 97 % water with 0.1 % formic acid, 3 % methanol with 0.1 % formic acid. The separation was performed on an Infinity lab Poroshell 120-SB reverse phase column (C18, 150 × 4.6 mm i.d., 2.7 µm; Agilent, Santa Clara, CA, USA). Column temperature was set at 30 °C. Quantitative determination of phenolic compounds was performed via a DAD detector. Quantification was done through 7-point calibration curves, with linearity coefficients (R^2) > 0.999, in the concentration range 2–140 µg/mL. All standards were purchased from Sigma Aldrich (Milan, Italy), and have a purity ≥ 95 %. The limits of detection were lower than 1 µg/mL for all assayed analytes (Table S2). The area under the curve from HPLC chromatograms was used to quantify the analyte concentrations in the extract. The wavelength of quantification with the Photo Diode Array Detector was set at 254 nm.

The quantitative determination of terpenophenols was conducted using a reversed-phase HPLC–UV-MS in gradient elution mode. The separation was conducted within 30 min, starting from the following conditions: 0.007% formic acid, 7% water, 93% acetonitrile. The details about gradient are listed in Table S3. The separation was performed on an Infinity lab Poroshell 120 reverse phase column (C18, 150 mm × 4.6 mm i.d., 2.7 µm) (Agilent Santa Clara, CA, USA). Column temperature was set at 30 °C.

Quantitative determination of terpenophenolic compounds (Table S4) was performed via DAD detector at 230 nm wavelength. Quantification was performed through 7-point calibration curves, with linearity coefficients (R^2) > 0.99, in the concentration range 2–160 µg/mL. The area under the curve from HPLC chromatograms was used to quantify the terpenophenol concentration in the extract. The limits of detection were lower than 0.9 µg/mL for all assayed analytes.

Table S1. Gradient Elution Conditions

| TIME (Min.) | COMPOSITION A% (Water+Formic acid 0.1%) | COMPOSITION B% (Methanol+Formic acid 0.1%) | FLOW (mL/min) |
|-------------|---|---|---------------|
| 1.00 | 97.0 | 3.0 | 0.600 |
| 5.00 | 77.0 | 23.0 | 0.600 |
| 12.00 | 73.0 | 27.0 | 0.600 |
| 18.00 | 57.0 | 43.0 | 0.600 |
| 25.00 | 52.0 | 48.0 | 0.600 |
| 32.00 | 50.0 | 50.0 | 0.600 |
| 34.00 | 50.0 | 50.0 | 0.600 |
| 37.00 | 35.0 | 65.0 | 0.600 |
| 40.00 | 5.0 | 95.0 | 0.600 |
| 47.00 | 5.0 | 95.0 | 0.600 |
| 48.00 | 97.0 | 3.0 | 0.600 |
| 60.00 | 97.0 | 3.0 | 0.600 |

Table S2. Retention times and wavelength of quantification of the standards used to identify and quantify phenolics in the extracts.

| | Standard | Wavelength (nm) | Retention Time (min) |
|----|--------------------------|-----------------|----------------------|
| 1 | Gallic acid | 254 | 8.770 |
| 2 | 3-Hydroxytyrosol | 254 | 11.483 |
| 3 | Caftaric acid | 254 | 12.693 |
| 4 | Catechin | 254 | 15.007 |
| 5 | 4-Hydroxybenzoic acid | 254 | 16.117 |
| 6 | Loganic acid | 254 | 16.503 |
| 7 | Chlorogenic acid | 254 | 16.750 |
| 8 | Vanillic acid | 254 | 18.567 |
| 9 | Caffeic acid | 254 | 18.887 |
| 10 | Epicatechin | 254 | 19.273 |
| 11 | Syringaldehyde | 254 | 21.833 |
| 12 | Chicoric acid | 254 | 22.283 |
| 13 | Coumaric acid | 254 | 23.043 |
| 14 | Ferulic acid | 254 | 24.133 |
| 15 | Benzoic acid | 254 | 26.407 |
| 16 | Hyperoside | 254 | 26.970 |
| 17 | Rutin | 254 | 27.263 |
| 18 | Resveratrol | 254 | 28.787 |
| 19 | Isoquercetin | 254 | 27.347 |
| 20 | Resveratrol | 254 | 27.980 |
| 21 | Rosmarinic acid | 254 | 28.633 |
| 22 | t-Cinnamic acid | 254 | 34.453 |
| 23 | Quercetin | 254 | 35.747 |
| 24 | Naringenin | 254 | 36.647 |
| 25 | 2,3-Dimethylbenzoic acid | 254 | 37.743 |
| 26 | Luteolin | 254 | 38.907 |
| 27 | Hesperitin | 254 | 39.420 |
| 28 | Kaempferol | 254 | 41.827 |
| 29 | Carvacrol | 254 | 44.717 |
| 30 | Thymol | 254 | 44.837 |
| 31 | Flavone | 254 | 45.607 |
| 32 | 3-Hydroxyflavone | 254 | 45.940 |
| 33 | Emodin | 254 | 47.683 |

| Table S3 Gradient elution of HPLC-DAD | | | |
|---------------------------------------|---------------|------|------|
| Time (min.) | Flow (ml/min) | %A | %B |
| 0 | 0.750 | 32.5 | 67.5 |
| 0.5 | 0.750 | 32.5 | 67.5 |
| 14 | 0.750 | 7 | 93 |
| 22 | 0.750 | 7 | 93 |
| 22.1 | 1.05 | 32.5 | 67.5 |
| 28 | 1.05 | 32.5 | 67.5 |
| 28.1 | 0.750 | 32.5 | 67.5 |
| 30 | 0.750 | 32.5 | 67.5 |

Table S4. Retention times and wavelength of quantification of the standards used to identify and quantify terpenophenolics in the extracts.

| | Standard | Wavelength (nm) | Retention time (min) |
|---|----------|-----------------|----------------------|
| 1 | CBD A | 230 | 9.433 |
| 2 | CBGA | 230 | 9.967 |
| 3 | CBD | 230 | 10.600 |
| 4 | THC-d3 | 230 | 15.433 |
| 5 | CBC | 230 | 16.967 |
| 6 | THCA | 230 | 17.633 |