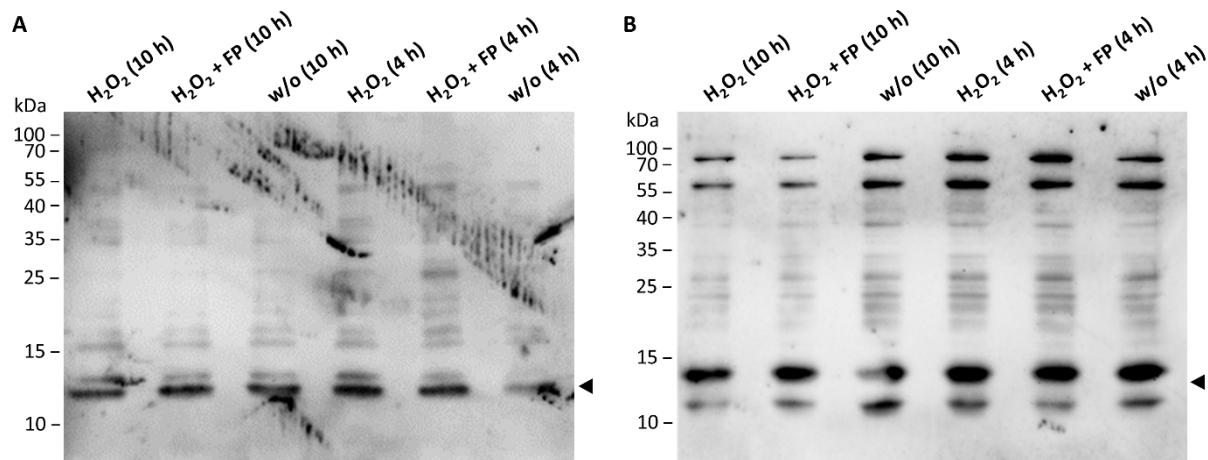


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2 **Figure S1.** *Ex vivo* cultivated human hpRPE cells show RPE-specific markers. (A-D) hpRPE were
3 pigmented and vital for 14 days on transwell filters. Shown are the cells of donor 2 at day (A) 2, (B) 4, (C)
4 10 and (D) 14 as an example. Scale bars 400 μm. Magnification 10x. (E-F) hpRPE were positive for (E) RPE65
5 mRNA and (F) RPE65 protein in immunostainings. (G-H) hpRPE were positive for (G) Bestrophin (BEST)
6 mRNA and (H) bestrophin protein in immunostainings. (F, H) Shown are representative examples after
7 10 days in culture. Scale bars 100 μm. Magnification 100x. (I, J) Marker for epithelial–mesenchymal
8 transition (I) mRNA SMA1 and (J) mRNA VIM were partly increased after properdin treatment of stressed
9 hpRPE cells.

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Figure S2. Western Blots of C3a and C5a in hpRPE cells. (A) C3a was detected in 5 h and 10 h stressed (H₂O₂, H₂O₂ + FP) and unstressed (w/o) hpRPE cells at approx. 12 kDa (arrow). Exemplarily shown for donor 6. (B) C5a was detected as double band in 5 h and 10 h stressed (H₂O₂, H₂O₂ + FP) and unstressed (w/o) hpRPE cells between 10 – 15 kDa (arrow). Exemplarily shown for donor 5. FP - properdin. Reduced conditions.

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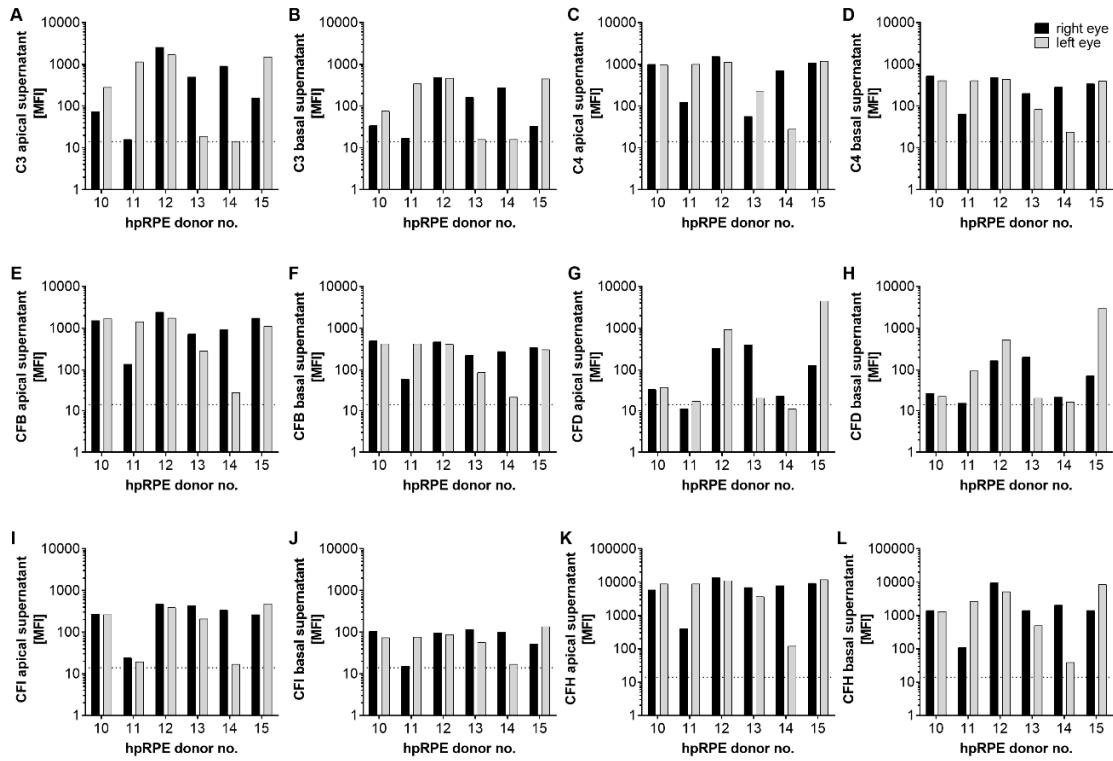
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Figure S3. Complement component secretion is different in hpRPE cells from left and right donor eyes. Complement secretion of (A, B) C3, (C, D) C4, (E, F) CFB, (G, H) CFD, (I, J) CFI and (K, L) CFH was increased at the (A, C, E, G, I, K) apical compared to the (B, D, F, H, J, L) basal side. hpRPE isolated from the right eye (black) showed different complement component secretion activity than hpRPE from the left eye (grey). Dotted line shows blank control.

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Table S1. In-house designed PCR primers.

transcript	gene accession number	name of sequence	application
ARMS_375	NG_011725	fw: CATTCTAAATCAAATCAAAACATTCA rv: CCTGGGGCTCTGTTGAATTG	
C2_116	NG_011730	fw: AGCACCATCTACACTTCGCC rv: CATGGCCCTTACCTCGACAT	
C2_144	NG_011730	fw: CAGGGGGAGATCAGAATCGTC rv: GTGGGGGCAGGGAAATTCTTT	
C3_147	NG_009557	fw: TTGCCTCTCTTAAGCCTGTG rv: GGCTAGGGTCTCACGAGG	
C3_223_2	NG_009557	fw: CTGGATGAAGAGGTACCCGC rv: CTTGGAACAGACCCCTGACA	
C9_623	NG_009894	fw: TCTGTGACAATGGCAGACAAC rv: TTTATTCCCCACTCTGTATCTT	
CFH_106	NG_007259	fw: CCTTGTTAGTAACCTTAGTCGTCTT rv: GGAGTAGGAGACCAGCCATT	hpRPE genotyping (2.2; Tab. 1)
CFH_109	NG_007259	fw: AGTCTACCTTGCTAACGGTTC rv: CCTCTAGATTGTAGTACCTGTTGCT	
CFH_121	NG_007259	fw: CAGATCCGTGTGAATATCCGA rv: TAAGAAGAGAGCCACCGGTCTCA	
CFH_570	NG_007259	fw: AGTGGTGAGGAAAAATGTAGAATGT rv: CATCTCCATCTTCTTAAATTCTGTGA	
CFH_618*	NG_007259	fw: GTGCATTCAAGGGCATATT rv: CCTGAAGACACAAATCTCACTGG	
CFI_100	NG_007569	fw: CCTGCAGTGTGGTCTGTGAG rv: CCAGTGCTACAAGGTGGAA	
CFI_141	NG_007569	fw: ATCCACTGATAACAAGCGCTCA rv: TTAAAATCGTCATGATGTTCAAAGC	
IL1B	NG_008851	fw: CTCGCCAGTGAAATGATGGCT rv: GTCGGAGATTCTGTAGCTGGAT	PCR (2.2/3; Fig. 5A) and RT- qPCR (2.3; Fig. 5B, C)
SMA1	NG_006672	fw: GCCTTGGTGTGACAATGG rv: AAAACAGCCCTGGGAGCAT	qPCR (2.3; Fig. S1)
VIM	NG_012413	fw: TGTCCAATCGATGTGGATTTTC rv: TTGTACCATTCTCTGCCTCCTG	

*transcript refers to SNP CFHR3/1_618

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Table S2 QuantiTect PrimerAssays (Qiagen)

mRNA transcript	gene accession number	PrimerAssay name	catalogue number
GAPDH	NG_007073	Hs_GAPDH_1_SG	QT00079247
C1Q	NG_007283	Hs_C1QB_1_SG	QT00003493
C3	NG_009557	Hs_C3_1_SG	QT00089698
C4A	NG_011638	Hs_C4A_1_SG	QT00237160
C4B	NG_011639	Hs_C4B_1_SG	QT00237167
C5	NG_007364	Hs_C5_1_SG	QT00088011
CFB	NG_008191	Hs_BF_1_SG	QT00012138
CFD	NG_007274	Hs_CFD_1_SG	QT00212191
CFI	NG_007569	Hs_CFI_1_SG	QT00213794
CFH	NG_007259	Hs_CFH_1_SG	QT00001624
CFP	NG_009893	Hs_CFP_1_SG	QT00010514
CD46	NG_009296	Hs_MCP_1_SG	QT00073689
CD59	NG_008057	Hs_CD59_1_SG	QT00035952
C3AR	NG_050736	Hs_C3AR1_1_SG	QT00090398
C5AR1	NM_001736.4	Hs_C5R1_1_SG	QT00997766
CD11B	NG_011719	Hs_ITGAM_1_SG	QT00031500
NLRP3	NG_007509	Hs_NLRP3_1_SG	QT00029771
RPE65	NG_008472	Hs_RPE65_1_SG	QT00001351
BEST	NG_009033	Hs_BEST1_1_SG	QT00023282

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