



Review

Role of Iron Oxide (Fe₂O₃) Nanocomposites in Advanced Biomedical Applications: A State-of-the-Art Review

Mehrab Pourmadadi ¹, Erfan Rahmani ¹, Amin Shamsabadipour ¹, Shima Mahtabian ²,
Mohammadjavad Ahmadi ¹, Abbas Rahdar ^{3,*} and Ana M. Díez-Pascual ^{4,*}

¹ School of Chemical Engineering, College of Engineering, University of Tehran, Tehran 14174, Iran

² Department of Materials Engineering, Shahreza Branch, Islamic Azad University, Shahreza, Isfahan 61349-37333, Iran

³ Department of Physics, Faculty of Sciences, University of Zabol, Zabol 538-98615, Iran

⁴ Universidad de Alcalá, Facultad de Ciencias, Departamento de Química Analítica, Química Física e Ingeniería Química, Ctra. Madrid-Barcelona, Km. 33.6, 28805 Alcalá de Henares, Madrid, Spain

* Correspondence: a.rahdar@uoz.ir (A.R.); am.diez@uah.es (A.M.D.-P.)

Abstract: Nanomaterials have demonstrated a wide range of applications and recently, novel biomedical studies are devoted to improving the functionality and effectivity of traditional and unmodified systems, either drug carriers and common scaffolds for tissue engineering or advanced hydrogels for wound healing purposes. In this regard, metal oxide nanoparticles show great potential as versatile tools in biomedical science. In particular, iron oxide nanoparticles with different shape and sizes hold outstanding physiochemical characteristics, such as high specific area and porous structure that make them idoneous nanomaterials to be used in diverse aspects of medicine and biological systems. Moreover, due to the high thermal stability and mechanical strength of Fe₂O₃, they have been combined with several polymers and employed for various nano-treatments for specific human diseases. This review is focused on summarizing the applications of Fe₂O₃-based nanocomposites in the biomedical field, including nanocarriers for drug delivery, tissue engineering, and wound healing. Additionally, their structure, magnetic properties, biocompatibility, and toxicity will be discussed.

Keywords: iron oxide nanoparticles; nanomaterials; nanotreatment; nanocarrier; drug delivery; tissue engineering; wound dressing



Citation: Pourmadadi, M.; Rahmani, E.; Shamsabadipour, A.; Mahtabian, S.; Ahmadi, M.; Rahdar, A.; Díez-Pascual, A.M. Role of Iron Oxide (Fe₂O₃) Nanocomposites in Advanced Biomedical Applications: A State-of-the-Art Review. *Nanomaterials* **2022**, *12*, 3873. <https://doi.org/10.3390/nano12213873>

Academic Editors: Nelson Marmiroli, Jason C. White and Luca Pagano

Received: 8 October 2022

Accepted: 27 October 2022

Published: 2 November 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Transferring from the macro-sized world to nano-sized materials seems to change the world immensely, and the fast-growing advances in nanotechnology have led to excellent clinical achievements [1]. Nanotechnology offers tremendous applications in biomedical engineering for loading therapeutic cures, specifically in cancer treatments. Thanks to their unique surface features, nanoparticles (NPs) can offer a promising platform for loading therapeutic agents in drug delivery systems [2–8]. The interaction of NPs with biological systems has offered the simultaneous possibility of targeted drug delivery, monitoring, and therapy. Anti-cancer loaded Nanocarriers could escape from the recognition of the reticuloendothelial system (RES), increasing circulation time makes them more preferable than the traditional cancer therapies, such as chemotherapy, by enhancing drug accumulation at tumor sites to reduce systemic toxicity. Free anti-cancer drugs damage health cells which cause harmful side effects [9,10]. Additionally, nanomaterial-based carriers improve the oral bioavailability and solubility of the chemotherapeutic agents and lessen the required administrated doses of drugs [11–14]. Another merit of nano delivery systems is their ability to decrease drug resistance, which could be employed to treat resistant cancers by utilizing co-delivery systems [15]. Hence, nanocarriers could be used in drug delivery systems to load the therapeutic agent, protect it from physicochemical degradation, target

ligands because of their functional groups, enter the cells, and release the anti-cancer agent with high therapeutic efficiency.

In contrast with other inorganic NPs (e.g., gold and carbon-based NPs), magnetic NPs can be decomposed to Fe, especially in acidic cell compartments (such as lysosomes), declining the potential long-term toxicity of NPs [16]. Additionally, iron oxide nanoparticles (IONPs) can be coated in a core-shell structure to prevent their agglomeration as a cluster. In this way, they could be gradually eliminated from the bloodstream [17]. The surface coating also improves the vascular permeability of the drug and facilitates its transport across the endothelial barrier [18]. It can also attach to particular aptamers or antibodies to target specific cells. The modified surfaces could offer diverse targeted moieties such as peptides, antibodies, and small molecules that can be conveniently coated on magnetic nanoparticles' surfaces thanks to the intrinsic magnetic features [19,20].

Iron oxide has unique properties that make it an attractive nanoparticle in biomedical applications [21–23]. Especially, Fe₂O₃ nanoparticles can enhance the stability and permeability of therapeutic agents through tissues, bringing about a high circulation time [24]. Consequently, applying Fe₂O₃-based nanocarriers presents an efficient treatment with less drug dosage requirement. Additionally, according to their capacity for loading drugs, they would provide controlled release of drugs, which minimizes toxicity and eliminates overdose by increasing the accumulation of the medicine in non-healthy sites [25]. Hence, these advantages, alongside the biocompatibility of Fe₂O₃ NPs, have attracted a significant amount of attention among the researchers during past few decades. Characteristics of these NPs depend on their morphology and crystal structure, size, and preparation route [26].

As a transition metal oxide, iron oxide possesses a variety of stoichiometric and crystalline structures, such as magnetite (Fe₃O₄), maghemite (γ -Fe₂O₃), Hematite (α -Fe₂O₃), Barite, (β -Fe₂O₃) maghemite, and (ϵ -Fe₂O₃) magnetite wüstite (FeO) [27]. Hematite (α -Fe₂O₃) has shown the highest stability at ambient conditions due to its small bandgap (2.0–2.2 eV). A hematite crystal consists of iron atoms surrounded by six oxygen atoms (hexagonal). Two-thirds of the hexagonal interstices in α -Fe₂O₃ are occupied by Fe³⁺ cations in the (001) basal planes, leaving the tetrahedral sites unoccupied, while oxide ions (O²⁻) reside along the (001) plane of a hexagonal closed-packed lattice. This cationic arrangement would lead to the formation of pairs of FeO₆ octahedrons, whose edges are shared by three neighboring octahedrons of the same plane and one face with an octahedron in an adjacent plane along the (001) direction [28]. With C_{3v} symmetry, Hematite exhibits two Fe-O bond lengths. Tauc plot reveals the indirect bandgap for the α -Fe₂O₃ involving d-d transitions and a direct transition from O (2p) to Fe (3d) occurring for E_g > 3.2 eV [29]. Hematite is an environmentally friendly n-type semiconductor (E_g = 2.1 eV) with diverse applications in lithium-ion batteries, water treatment, gas sensors, photocatalysis, and water splitting for H₂ generation.

The fundamental of targeted delivery of anti-cancer agents is enhancing permeability and retention (EPR), and one of its techniques is taking advantage of the physiological differences between normal and tumor cells. For example, cancerous sites have lower pH (5.4) in comparison with healthy ones' (pH 7.4). Facilitating pH-responsive systems is an amazing way to enhance the accumulation of drugs in tumors. Surface modifications of Iron oxide nanoparticles, mainly because of their high specific area and porosity, make them a great candidate for being used as pH-sensitive vehicles in drug delivery systems (DDSs) [30]. Furthermore, the magnetic properties of iron oxide NPs can be applied as a novel route for delivering drugs and optimizing their dispensation through an exterior magnetic field to guide iron oxide-based nanocarriers to deliver drugs into a specific location in the body. Magnetic guidance can also boost the drug release at the tumor site [31].

In addition, magnetic nanoparticles have electrocatalytic features that give rise to rapid readout in redox-responsive delivery systems [32]. Controlled disassembly of redox-responsive drug delivery system results in rapid release of encapsulated medicines.

Nanocarriers can be functionalized by enzyme-labile connections to achieve an on-demand enzyme-sensitive drug release system to decrement the adverse effects of therapeutic agents. An efficient enzyme-sensitive nanosystem can penetrate tumor tissues with odd features such as leaky vasculature and various enzyme expression profiles. Relevant enzymes can be employed to prepare Fe₂O₃ NPs with promising features. Due to their endogenous nature, enzymes exhibit superior selectivity and catalytic features in various biological processes. Therefore, they have been widely explored as target agents in diverse endogenous and exogenous stimuli-sensitive systems [33]. Moreover, enzyme sensitivity increases both the rate of drug release into the tumor and the amount of drug that accumulates within it. Thus, redox responsive dual-stimuli responsive DDSs can enhance anti-cancer therapy by raising drug accumulation, promoting tumor targeting, and accelerating drug release. Further stimuli can be incorporated into redox reactive delivery systems to control release and minimize the side effects effectively [34].

In addition, to enhance mechanical and biological performances, nanoparticles, because of their small size that helps them to simulate extracellular matrix components of tissues, have been used recently in tissue engineering as substitutes for damaged and diseased tissues [35].

Specifically, different structures of iron oxide nanocomposites have a high surface-to-volume ratio, which has increased their application in this field of biomedical engineering. This feature can increase drug loading and entrapment efficiency and can cause a gradual drug release pattern [36,37]. It should be noticed that the extended surface area is produced in iron oxide nanocomposites due to the addition of iron oxide nanoparticles to the prepared hydrogels, in other words, by adding iron oxide nanoparticles to the fabricated hydrogel, these nanoparticles introduce porosity to the system and the final iron oxide nanocomposites would be a porous platform with a considerable ratio of surface area to volume. The BET assessment that has been carried out by Nurul Hidayah et al. [38], revealed a significant increase in the specific surface area of reduced graphene oxide from 25 m²/g to 51 m²/g after incorporation with iron oxide nanoparticles.

For example, the physical and chemical properties of Fe₂O₃ nanoparticles can apply growth factors are applied to scaffolds to induce bone-forming cell regeneration that would overcome the difficulties of producing complicated organs such as bones [39,40]. Furthermore, for tissue engineering applications, scaffolds needed to be porous for better mass transportation of O₂, CO₂, and other nutrients with extended surface area [41,42]. These requirements can be perfectly fulfilled by employing iron oxide nanoparticles in the structural fabrication of the scaffolds.

In addition, iron oxide nanoparticles benefit shapes and non-toxic nature that make them applicable for wound healing, too. They could be used in wound dressing materials to stimulate the movements through the different healing phases in acute and chronic wounds [43]. Moreover, some characteristics of IONPs including spacious surface, high specific area, and porous structure give the Fe₂O₃-based nanocomposites and hydrogel higher absorption capacity, water, and oxygen permeability [44–46]. These special features of IONPs can introduce an applicable wound dressing for biomedical applications.

Potential of Fe₂O₃ for being functionalized and coated by suitable polymers for making these NPs non-toxic, super magnetic, biocompatible, and biodegradable, introducing them as the best materials for biomedical applications including drug delivery (magnetic-sensitive), tissue engineering (bone tissue regeneration), and wound dressing. In this study, different structures and specific characteristics of IONPs were examined, and previous investigations on their advanced biomedical applications were reviewed.

2. Structure of Iron oxide Nano Particles

Fe₂O₃ and Fe₂O₃-based materials with diverse sizes and shapes (NPs, nanotubes, nanorods, spindles, hollow, and porous nanostructures) were prepared by hydrothermal, chemical precipitation, sol-gel, anodization, and thermal decomposition approaches. The hydrothermal method is desirable as it offers efficient control of size, morphology, and

agglomeration while lower impurities enter the hydrolyzed product. It also benefits from relatively low reaction temperature, cost-effectiveness, and environmental compatibility. Moreover, Fe_2O_3 possesses two interchangeable crystalline forms ($\alpha\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$). The structure of $\alpha\text{-Fe}_2\text{O}_3$ is represented in Figure 1. At higher temperatures, $\gamma\text{-Fe}_2\text{O}_3$ (maghemite) can be transformed to $\alpha\text{-Fe}_2\text{O}_3$ (Hematite) as $\alpha\text{-Fe}_2\text{O}_3$ is the most thermodynamically stable phase. The transition from $\alpha\text{-Fe}_2\text{O}_3$ to $\gamma\text{-Fe}_2\text{O}_3$ requires a nitrogen atmosphere with adequate temperature control or the application of a reducing agent. The morphology generally remains unchanged by the transition between $\alpha\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$, which could be an applicable approach to provide the controllable preparation of Fe_2O_3 with a modulated crystalline structure. There are a variety of magnetic phenomena in the $\alpha\text{-Fe}_2\text{O}_3$, which is due to the exchange bias and memory effects. Furthermore, in small Fe_2O_3 nanoparticles (smaller than 10 nm), superparamagnetic behavior and spin-glass properties are observed [47]. These hexagonal-structured nanoparticles are chemically active, avoid clearance by immune cells, and have a noticeable retention time, electrical conductivity, and biocompatible nature, all of which make them good candidates for biomedical applications [48,49].

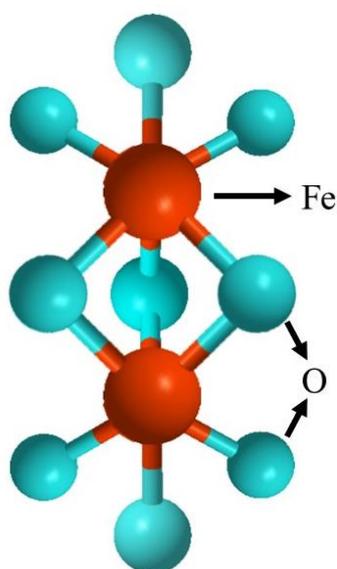


Figure 1. Structure of $\alpha\text{-Fe}_2\text{O}_3$ (Hematite).

3. Drug Delivery Application of Fe_2O_3 -Based Nanocomposites

In biomedical settings, pH-responsive systems may benefit from inducing physicochemical alterations due to the wide range of pH values seen in the human body [27,50,51]. Gastric passage elevates pH in oral medication delivery systems, including insulin administration, causing acrylic-based polymers to expand and discharge the medicine [52]. A negatively charged surface would be formed in pH 7.4 settings, causing the nanoparticle to discharge the loaded medicine. This is another advantage of functionalized nanoparticles with positively charged surfaces that trigger anionic drug loading at lower pH [53]. By covering Eudragit-S100 polymer using ibuprofen-loaded Fe_2O_3 magnetic mesoporous silica nanocomposites tablets, Xing et al. [54] create a dual-stimulus-responsive platform. Nanoparticles with tailored releasing capability were created by the magnetic characteristics of Fe_2O_3 when used in conjunction with an externally applied magnetic field. More medication was delivered into modeled proximal intestinal fluid when the pH-sensitive polymer was used, as opposed to modeled stomach fluid.

pH-responsive mechanisms may also be used at the cellular level. DNA and other therapeutic compounds are damaged when the pH of early endosomes, sorting endosomes, and multivesicular bodies drops rapidly following endocytosis [55]. Injury caused by intracellular distribution may be avoided using polymers that safeguard endosomal compartments.

A group led by Alexander et al. [56] using γ -Fe₂O₃@polymerized 2-(dimethyl amino) ethyl methacrylate developed dual-responsive core-shell nanoparticles for plasmid DNA delivery throughout CHO-K1 cells. Researchers found that, compared to polyethyleneimine, magnetic core-shell nanoparticles might experience bidirectional pH-dependent temperature-induced aggregation and enhanced efficacy of gene transport with no extra cytotoxic effects. Chemotherapy, which is the typical cancer treatment strategy, can assault both diseased and normal cells due to the application of nonspecific targeting medicines [57]. By increasing the specificity of drug-loaded NPs and ameliorating permeability and retention (EPR) events, intelligent pH-responsive platforms can help reduce complications [58,59].

Tumors develop rapidly, accumulating lactic acid and lowering the pH of these areas (pH 4–6) in comparison to physiological status (pH 7.4) [50,60]. The highly expanded surface area, appropriate porosity, and capacity to be coated by polymers as core-shell nanoparticles make Fe₂O₃ NPs such an ideal carrier for pH-responsive drug delivery platforms, moreover, there is a potential for the reduction of Fe₂O₃ based nanocarriers for loading biomolecules (Figure 2) [61–66]. These characteristics provide sufficient room for loading antitumor medications. Regarding the delivery of doxorubicin, Sheng et al. [67] created polyethylene glycol-functionalized γ -Fe₂O₃ nanoparticles, then tested for pH influences on drug distribution and alternating magnetic field treatment. They observed that total discharges were 32 and 63% at pH levels of 7.2 and 5.5, respectively. Daunorubicin hydrochloride was loaded into a γ -Fe₂O₃/ZnO nanocomposite by Maiti et al. [63], and then drug delivery was measured at pH 5.5 and 7.5. When used as a model for tumor locations, the acidic setting aided in the drug delivery. To make things even better for medication distribution, mesoporous ZnO can load large amounts of medicines. With curcumin anticancer medication, Patil and colleagues created functionalized chitosan-coated γ -Fe₂O₃ nanomaterials activated by a change in pH [68]. They found that the discharge frequency at pH 6.0 was approximately 20% higher than at pH 7.4, suggesting the nanocarrier's capacity for cancer treatment. Various proportions of oxidized pectin/chitosan were loaded in nano γ -Fe₂O₃ to improve the antitumor properties of the 5-FU medication by Li et al. [69] pH levels below 7 and temperatures greater than 36.5 °C demonstrated the system's pH and thermo-sensitivity by showing a larger swelling frequency. The MMT test against L929 and MCF-7 cell cultures was used to determine the synthesized composite's biocompatibility and cancer-killing abilities. Due to this, 5-FU may be more effectively targeted, and its anticancer capabilities are enhanced when the oxidized pectin/chitosan/ γ -Fe₂O₃ NPs are used.

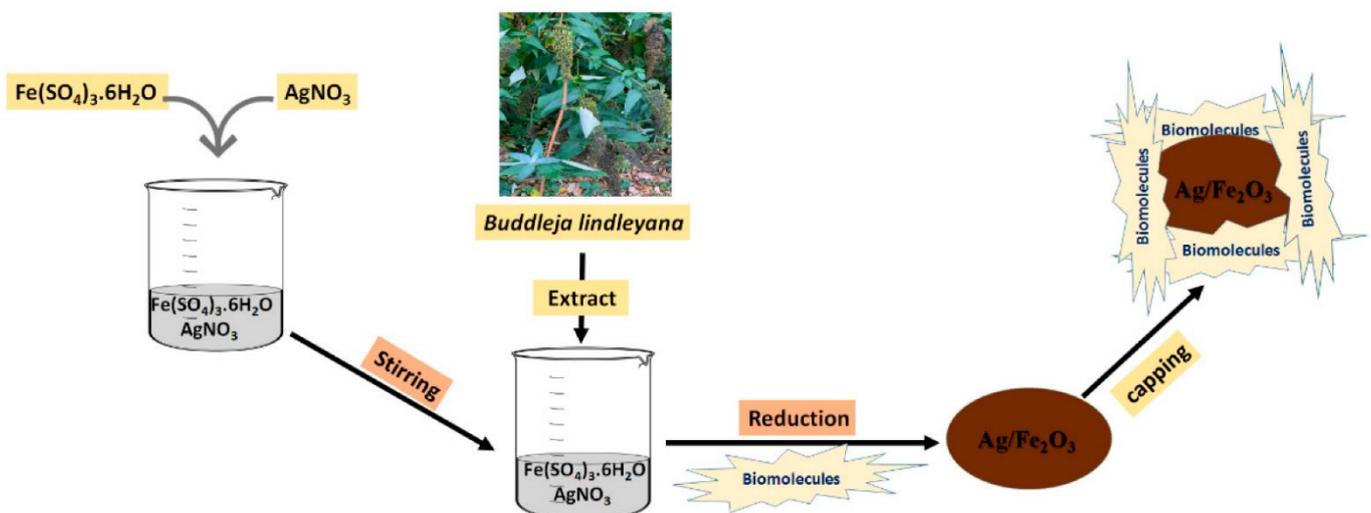


Figure 2. The reduction procedures of the Ag/Fe₂O₃ nanocarrier for loading biomolecules [66].

The exceptional electrochemical redox capabilities of magnetic nanoparticles may be attributed to their substantial electrocatalytic characteristics in a single structure and their

composites. It may be formed by optimizing factors, including the transmission of ions by solid NPs because of their high penetration, which accelerates internal dispersion's phase changes [32]. With the help of their redox capabilities, magnetic nanomaterials can gather and concentrate the desired electrochemical data for quick reading [70].

The most often employed NPs in this sector are γ -Fe₂O₃ (oxidized state, Fe³⁺) and Fe₃O₄ (two oxidized states, Fe²⁺ and Fe³⁺) in MNMs-based composites, respectively [71, 72]. Magnetic nanoparticles and targets discovered by cyclic voltammetry influence the electrodes' predicted redox potential [73].

Poly (ethylene glycol) and poly(ϵ -caprolactone) disulfide bonds (PEG SS PCL) were used to create biodegradable reduction-responsive micelles for the administration of superparamagnetic iron oxide (SP IO) nanoparticles and doxorubicin (a chemotherapeutic drug). In the main chain of disulfide bonds, the amphiphilic deblock copolymer has redox reaction characteristics. Doxorubicin (DOX) was loaded 32% throughout magnetic nanomicelles. Consolidation of self-assembled PEG-PCL micelles with oleic acid and water was achieved using coarse-grained molecular dynamics (CG-MD) modeling. In comparison, the hydrophobic and hydrophilic proportions of each copolymer block were identical, and each oleic acid was similarly attached to magnetic nanomaterials [74].

To modulate camptothecin pharmaceutical discharge from the mesoporous silica pathways and achieve a dynamic dual-mode MRI contrast, researchers employed superparamagnetic iron oxide nanoparticles (SPION) coated by acid, oxidation stress, and redox-sensitive manganese oxide (MnOx). The efficiency of responsive therapy has been tested throughout pancreatic tumor cells and tumor-bearing animals, and results have supported the performance of Non-vehicle MnOx-SPION@MSn@CPT fighting cells [75]. To further improve biodistribution and efficacy for convection-enhanced delivery (CED) of BG to GBM, iron oxide superparamagnetic nanomaterials were developed. The nanomaterials (NPCP-BG-CTX) consist of a magnetic core surrounded by a surface coating of redox-responsive chitosan-PEG copolymer altered by BG covalent attachment and peptide chlorotoxin tumor (CTX). NPAC-BG-CTX demonstrated precise in vitro BG transport in human GBM cells under decreased intracellular circumstances, allowing for controlled and targeted BG discharge. MGMT activity and TMZ toxicity were significantly reduced in cells administered with NPCP-BG-CTX. Animal studies in mice with primary human GBM xenografts showed that CED from NPCP-BGCTX had good in vivo distribution volume (Vd). When NPCP-BG-CTX and NPCP-CTX were used in conjunction, the mean survival rate in both medicated and unprotected animals increased by three times [76].

Enzymes are widely used by internal stimulus-responsive nanocarriers since they are involved in a wide range of biochemical and physiological mechanisms. Enzyme transcription at specific places is critical in a variety of pathological processes. In addition to indicating a sick condition, it may also be modified to transport medications with more safety to the appropriate region and serves as a possible screening tool [33]. In contrast to other stimuli, enzymatic processes are more powerful, efficient, and accurate, providing extra benefits by intracellularly participating in various metabolic reactions. The transcription of enzymes, including proteases, phospholipases, hydrolases, and glucosidase, increases in a variety of malignancies that use growth factors to increase cell populations. These specific enzymes detect and split intelligent nanocarriers comprised of substances such as Fe₂O₃ to deliver the payload, namely the medicine, to the targeted place. Multifunctioning is typically supported by adding more than one molecule to the surface of a material [77]. The overexpression of MMP-14 in MMTV-PyMT cells was shown to be much more hazardous than the downregulation of MMP-14 throughout fibroblasts. CLIO-ICT (a platform for iron oxide nanocarrier) was produced by Ansari et al. [78], to be used as theragnostic nanomaterials for magnetic resonance imaging (MRI) and medication administration. CLIO-ICT was created by linking an azademethylcolchicine (ICT) conjugator to an MMP-14-cleavable region. MMP-14 overexpression caused considerable toxicity among MMTV-PyMT cells, as seen by comparison to fibroblasts with modest frequencies of MMP-14 transcription.

Furthermore, in MR imaging of MMTV-PyMT tumor-bearing mice given by CLIO-ICT, preferential accumulation of nanomaterials in the cancerous site was detected. An enzyme-response, multifunctional DOX-SMNP combination was produced employing clicking chemical properties. Due to the upregulation of a specific enzyme in these live cells, the chemotherapeutic drugs released by this SMNPs-combined medication are selectively controlled. For the detection of intracellular drug delivery and tumor cell scanning, this drug-loaded nanoparticle combination demonstrates a potential mixture of fluorescence and magnetic resonance imaging methods performed in real time. Prospective uses of the multipurpose drug-coated nanostructures include tumor-targeted medication delivery and concurrent diagnostic or detection of therapeutic effects [79].

4. Fe₂O₃ in Magnetic-Responsive Drug Delivery Systems

Site-specific drug distribution may be achieved by directing IONs caused by a confined external magnetic field, which makes use of the magnetic characteristics of iron oxide nanomaterials. Several illnesses, including tumors and inflammation, have been shown to benefit from this strategy [80]. To govern the nanomaterials' passage through the circulation and their concentration at the intended places, the magnetic reaction of IONs is highly associated with their physicochemical qualities; more precisely, the saturation magnetization of the manufactured nanostructures should be significant [81]. Some scientific teams are creating magnetic nanocarriers with unique structural properties that have examined this site-directed function. There was a publication in 2012 by the group of Wagstaff et al. [82] about how to make gold-covered iron oxide nanomaterials loaded with a platinum-containing chemotherapeutic agent (cisplatin). Coprecipitation and oxidation of iron oxide nanocrystals resulted in the formation of maghemite during this investigation. A technique known as "incremental hydroxylamine seeding" was used to cover the nanomaterials with gold [83]. Subsequently, thiolation was used to coat the particles. Earlier, the same group discovered substances such as thiolated polyethylene glycol (PEG) linkers [84,85]. Eventually, robust coordination linkages involving the PEG linker were used to load cisplatin onto the magnetic nanocarriers. Using a naked magnet, the nanocarriers were tested in vitro and shown to have a 110-fold enhancement in cytotoxic effects on human ovarian cancer cell lines A2780, as well as particular cell development suppression.

Unterweger et al. [86] created IONs coated with dextran/hyaluronic acid, which have cisplatin-carrying IONs66. The loading of medications, including cisplatin, and the targeting of overexpressed CD44 receptors throughout malignant cells, were made possible by applying hyaluronic acid (HA) [87]. Following the dextran-coated IONs amination, they were covered by lower-molar mass HA (acquired via enzymatic breakdown of HA) [88]. After forming a polymer-metal combination with HA, cisplatin was successfully incorporated into the resultant nanocarriers, which were noted for their effective drug entrapment (43.2 ± 0.2%). A beginning 30-minute explosive discharge was accompanied by an ongoing discharge for 48 h, which was described by the researchers as being typical for surface-bound drugs in their study. A substantial enhancement in drug delivery has also been evidenced in the availability of hyaluronidases, which are extensively discovered in tumors and make these nanostructures especially encouraging for tumor-targeted drug delivery [89]. This is made even more promising by their regulation through magnetic power illustrated by the uptake of nanomaterials containing a neodymium magnet. PVA-coated iron oxide nanostructures loaded with doxorubicin have also been explored by Nadeem et al. [90], who investigated the magnetic manipulation of IONs for specific drug delivery purposes. These nanocarriers exhibited good management for magnetically directed medication delivery by providing a magnetic flux. The antineoplastic medication mitoxantrone was adsorbed on the HSA shell of iron oxide nanostructures (average inorganic diameter of around 7 nm) synthesized by Zaloga et al. [91] using lauric acid (LA) and HSA coatings. Over 72 h, such nanocarriers showed improved durability and linear discharge pharmacokinetics. Using an in vitro magneto-guided approach, researchers were able to convincingly demonstrate the site-specific curative impact of these nanocarriers [92].

In a recent study, Jeon et al. [93] demonstrated the synthesis of polymerized β -cyclodextrin covered IONs that, via host-guest association, were capable of loading polymerized paclitaxel (a chemotherapeutic medication renowned for its limited solubility in water). Due to the strong magnetism of the produced nano assemblies, they may be used as magnetically directed drug delivery platforms. The improved antitumor efficacy of these platforms was shown in vivo throughout CT26-bearing mice due to magnetically-mediated targeting.

Normal cells suffer adverse effects when the body's temperature is elevated during tumor hyperthermia therapy. An external magnetic field would allow for targeted hyperthermia, which reduces the damage to healthy tissues and reduces the needed medication dosage [94,95]. External signals may act as a guide for transporting the medicine to the tumor location and initiating pharmaceutical discharge [96]. Since malignant cells are less resistant to temperature rise than healthy ones, utilizing MNPs in the carrier generates a thermal flow that aids in the destruction of diseased cells [97,98]. An external source's proximity to the targeted region influences the magnetic-responsive drug delivery platform's precision; as tissue profundity increases, the required intensity magnetic flux decreases [99]. As a consequence, this technique is confined to surface organs and so cannot be used to treat pulmonary or hepatic tumors [100]. Aside from that, frequencies greater than $8\text{--}16\text{ kA}\cdot\text{m}^{-1}$ generate eddy currents that are damaging to normal organs [101,102]. Owing to their super magnetic capabilities, biocompatibility, high specialized region, appropriate nano-sized particulates, and lower toxic effects, Fe_2O_3 nanomaterials are one of the most promising alternatives for magnetic-responsive devices according to the researchers [31,103,104] (Figure 3). Improved drug loading potential, increased carrier durability, and opportunity for future functionalization might be achieved by coating iron oxide NPs using appropriate polymers [67,105–108]. In vitro experiments using human hepatocarcinoma SMMC-7721 cells, Yan et al. [109] employed a combination of Fe_2O_3 nanomaterials and magnetic fluid hyperthermia (MFH) and found that apoptosis was dose-dependent, and proliferation was inhibited. Due to their absorption capabilities, Fe_2O_3 nanomaterials have magnetic sensitivity, which causes the magnetic fluid temperature to rise by around $7\text{ }^\circ\text{C}$. An AC magnetic field was used to study the emission of heat from the nanohydroxyapatite matrix-encased $\gamma\text{-Fe}_2\text{O}_3$ NPs. Investigations on the cytotoxic effects of human (sarcoma osteogenic) SAOS-2 cell lines in vitro demonstrated minimal toxicity, indicating that they may be employed in magnetic hyperthermia [110].

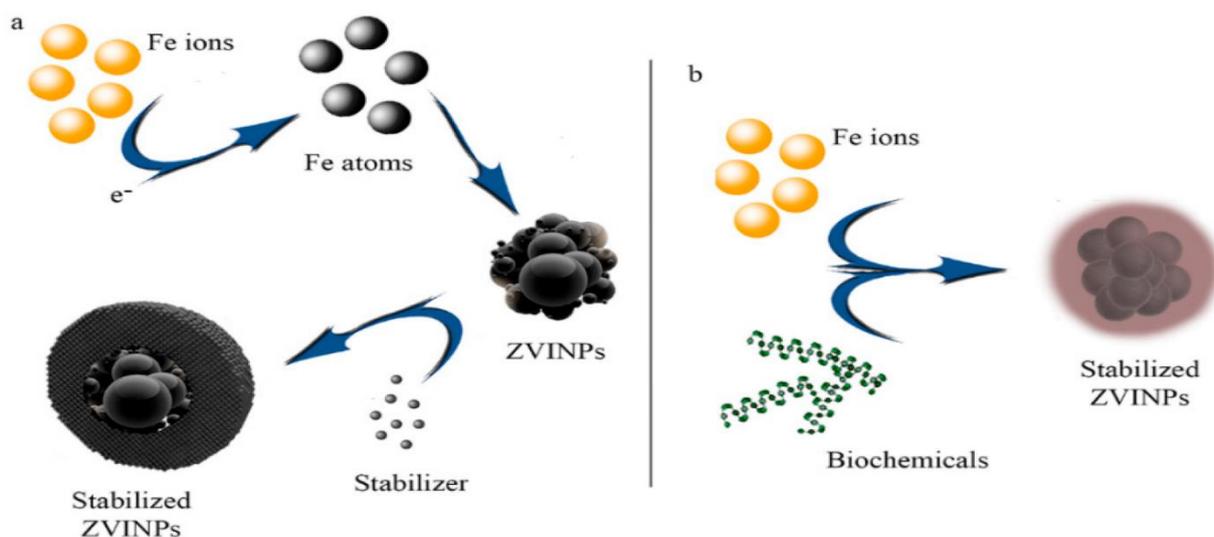


Figure 3. Chemical (a) and biological (b) reduction of iron ions to zero-valent iron nanoparticles (ZVINPs) and their stabilization [104].

Due to low viral concentration in the targeted tissue, gene therapy faces a key drawback: delayed vector deposition [111]. The specificity can be improved by attaching targeted ligands to viral particles; however, this is insufficient [112]. The difficulty may be solved by introducing an exterior alternating current magnetic field to gene vectors. For this technique of gene delivery, Scherer et al. [113] attached gene vectors to superparamagnetic nanoparticles. Transduction effectiveness was shown to be great in vivo, and the delivery time of genes was lowered to minutes, supporting the idea that magnetic field guidance may be used to overcome fundamental obstacles. Fe₂O₃ uses in magnetic-responsive biological devices are listed in Table 1.

Table 1. Some few biological uses of the newly developed magnetic-responsive devices containing Fe₂O₃.

Nanoparticles	Usage	Target	Ref.
Fe ₂ O ₃	Magnetic hyperthermia	human hepatocarcinoma SMMC-7721 cells in vitro and xenograft liver cancer in nude mice	[109]
γ-Fe ₂ O ₃ NPs (NPs) embedded in a nanohydroxyapatite matrix	Magnetic hyperthermia	Human (Sarcoma osteogenic) SAOS-2 line-cells	[110]
Phosphatidylcholine coated γ-Fe ₂ O ₃	Magnetically-induced cell mobility	colon cell lines	[114]
α-Fe ₂ O ₃	Drug carrier	human lung fibroblasts (MRC5) cell line	[115]
SiO ₂ /γ-Fe ₂ O ₃	Magnetic hyperthermia	BRL-3A cells	[116]
γ-Fe ₂ O ₃ nanosheets surface-modified containing polyethylene glycol	Ibuprofen delivery		[117]
polyethyleneimine coated with iron oxide	gene delivery		[113]
hyaluronic acid-Fe ₂ O ₃	Peptides delivery		[118]

5. Biocompatibility and Toxicity of Fe₂O₃-Related Drug Carriers

The immunological system reaction after delivery and the inherent toxicity of the carrier and/or biodegradation of metabolites are all variables that impact the biocompatibility of a drug nanocarrier. Due to alterations in cell/tissue bioavailability and clearance/metabolization related to the nanocarrier, the toxic effects pattern of the medication may be raised or lowered. In physiological enclaves where the medication cannot disperse on its own, aggregation may develop.

After drug delivery, there must be a rapid removal of drug-related colloids and their breakdown products to minimize toxicological effects. Dosage, chemical constitution, size, shape, solubility, surface chemistry, mode of prescription, biodegradability, pharmacokinetics, and bioavailability are a variety of parameters affecting magnetic particle toxicity [119].

The structure of the NP surface has a major role in the NP–cell association, as previously stated. The effectiveness of IONP cell absorption and internalization is a precondition for enhanced medication administration, enhanced diagnostic efficiency, and other usages of IONPs. When nanoparticles are incubated in a cell culture environment, it is possible that they could change the pattern of cell attachment, which will have consequences for the cell's shape and cytoskeleton and its multiplication, differentiation, motility, and longevity (Figure 4) [120–122].

When exposed to physiological circumstances, bare Fe₂O₃ nanocrystals create damaging free radicals [123,124]. The cores of the magnets should be coated with additional particles to circumvent this limitation. In this method, not only would the colloidal durability of the nanocomposite improve, but the performance probability of the experiment would also rise. Throughout drug delivery platforms, low-toxicity coated iron oxides may have an EPR impact on certain cells [125]. Particle accumulation may also be prevented by using an appropriate covering [126].

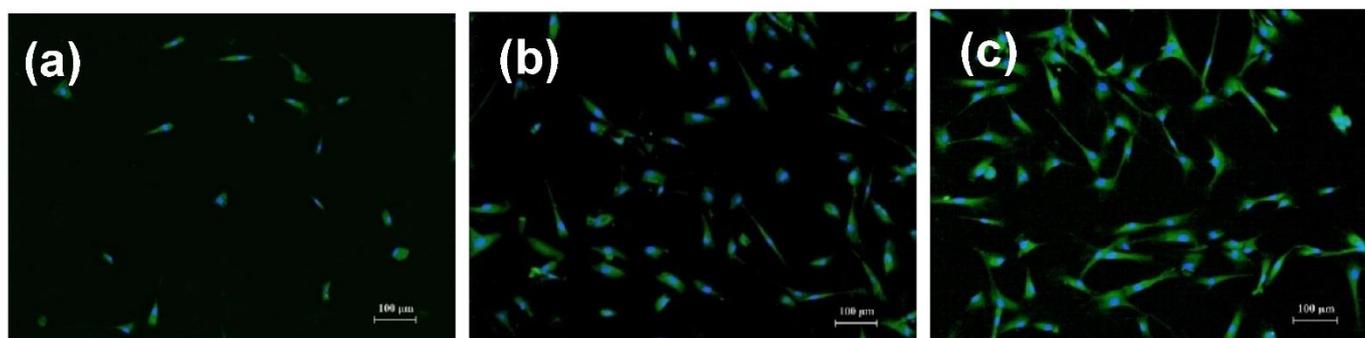


Figure 4. Fibroblasts were cultured on the surface of gel-based substrate (a), with 1.2% Al_2O_3 (b), and with 1.2% Fe_2O_3 NPs (c). Cell nuclei and the cytoplasm were stained with DAPI and pyrazolone yellow, respectively [122].

Magnetic Fe_2O_3 -based nanomaterials were manufactured by Dumitrache et al. [127] and contrasted to l-DOPA-coated nanostructures. Thermal conductance and distribution in biological medium (physiologic pH) were much enhanced following coating. Dopamine was used as a biocompatible surfactant in the research of Li et al. [128] to boost the durability of a nanocomposite with a nucleus of $\alpha\text{-Fe}_2\text{O}_3$. $\alpha\text{-Fe}_2\text{O}_3$ @dopamine was administered to a mouse and was shown to be non-toxic to live cells when incubated with NIH3T3 cells. Furthermore, the PA indication of the mouse's cerebral vascular system increased when the created theragnostic platform was administered, compared to commercialized contrast compounds. In vitro grown primary human fibroblasts (hTERT-Bj1) were used to test the biocompatibility of IONPs with and without a PEG surface coating [129]. Without PEG coatings, relative to the controls, the 50 nm diameter uncoated IONPs decreased cell adherence by 64%; however, a significant alteration was not seen with PEG-coated nanoparticles. Several cytoplasmic vacuoles with the disordered cytoskeleton and defective and disrupted cell membrane were seen throughout the NP-incubated cells without coating.

Even though that they exhibited a greater cell absorption relative to uncoated NPs, PEGylated MNPs, on the other hand, did not disrupt the fibroblast cytoskeleton. If PEG is present on the particle surface, the NP/cell interaction will be altered, and this might be due to variations in the kinetics of NP metabolic activities.

Comparable apoptosis and cytotoxicity were seen in hTERT-BJ1 primary human fibroblasts incubated with dextran-coated IONPs relative to those incubated with uncoated IONPs [130]. There was no difference in cell absorption between dextran-coated and uncoated NPs; however, the dextran-coated NPs were more likely to produce cellular mobility and membrane rupture. In contrast to dextran-coated and uncoated NPs, albumin-coated IONPs did not affect cell survival [131,132]. Since albumin interacts with membrane phospholipids/fatty acids, the albumin-coated NPs just marginally disrupted the membrane. Nonetheless, in comparison to the uncoated and dextran-coated nanoparticles, the toxicity of such NPs was much reduced totally. A remarkable finding was that the albumin-coated nanoparticles were internalized at a lesser rate compared to the uncoated and dextran-coated particulates. This raises the possibility that the NP toxicity outlined above may be influenced by the various amounts of intracellular particulate. In vitro studies have shown that various particle sizes may have diverse impacts on cells. For example, relative to 0.5- μm -sized IONPs, 30-nm-sized types had much larger harmful impacts [133]. An effect on cellular destruction was detected depending on cell size and dosage whenever the A549 alveolar epithelial cells were incubated.

Some of the changes in cell culture caused by NP incubation may be responsible for the in vitro cytotoxic effects of MNPs M [134]. For example, the absorption of OH^- ions onto the surface of naked (uncoated) iron oxides causes these materials to be negatively charged throughout the aqueous solution. Counterions may be attracted to the resultant electric field, which may aid in protein adhesion. Furthermore, the presence of Cl^- ions throughout the cell culture media may potentially compete with the Fe, resulting in a change in the

pH of the medium and the surficial properties of the nanoparticles. As a consequence of these connections with the cell culture medium, which result in modifications, including fluctuations in the ionic concentration and protein activity, cells may become detached from the medium and die as a result. MNPs coated with a surface layer, on the other hand, exhibit low interaction with their surrounding environment in the cell culture medium, which is most likely since there are fewer sites available on the particle surface for protein attachment [134].

The form and behavior of cells may be influenced by the IONP surficial charge or other nanomaterials [135]. Endocytosis by multiple cell lines was boosted through the coating of 10-nm-sized IONPs containing DMSA (anionic charge) [136,137], which decreased cell adherence and survival of the PC12 pheochromocytoma cell line in a concentration-dependent manner [138]. PC12 spherical cells start to develop into neuronal cells following 48 h of treatment with nerve growth factor (NGF), resulting in mature neuritis that reaches the peripheral areas. The disruption of the cytoskeleton caused by anionic NPs, on the other hand, prevented neurite development and resulted in a significant reduction in the frequency of neurites formed per cell, as well as in the neuritis extent. Toxicities associated with these anionic MNPs may be depending on concentration, as seen by a reduction in GAP-43 protein production, which is linked to neuronal budding and neuronal functioning. Besides, Figure 5 represents the scratch morphology altering with time for a Fe_2O_3 based nanocomposite and shows the production impairment of endothelial cell caused by PSC- Fe_2O_3 .

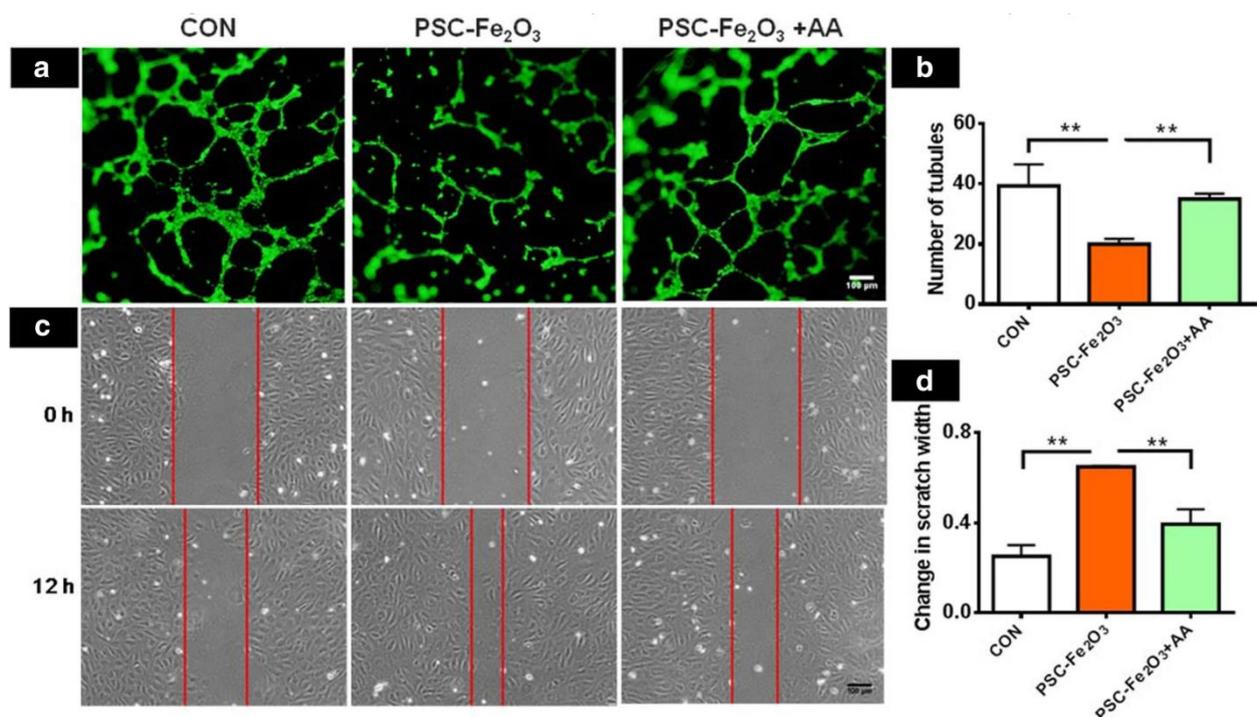


Figure 5. ROS scavenger AA converted the function of endothelial cells happening EndMT. (a) AA converted tubule production impairment of endothelial cell caused by PSC- Fe_2O_3 . (b) measurement of endothelial meshes ($n = 3$). (c) pictures of scratch morphology altering with time. (d) Relative scratch width alteration ($n = 2$). Data are mean \pm SD, one-way ANOVA with LSD-t, ** $p < 0.01$ [139].

On the other hand, it has been shown that the anionic IONPs' cytotoxicity varies depending on the kind of surface coating used. Cell lines from the SK-MEL-37 melanoma were cytotoxic to DMSA, citric acid, or lauric acid in distinct ways. DMSA-coated, citric acid-coated, and lauric acid-coated nanoparticles had IC50 (half-maximal inhibitory concentration) values of 2260, 433, and 254 g Fe/mL, correspondingly. Cellular death was seen when lauric acid-coated NPs were utilized at higher levels, including 800 g Fe/mL,

as shown by morphological characteristics of the apoptosis profile, notably surficial blebbing, severe vacuolization, and chromatin contraction [140]. Therefore, even though the generated NPs have a comparable surface charge, the surface coatings may significantly modify their toxicity dose-dependently. Due to the variable nature of the intended use, it is necessary to conduct an individual cytotoxicity test for each sample.

Although early data on the security of MNPs may be revealed using *in vitro* toxicity assessments on different cell types; however, these tests may have restricted applicability since they do not match the real-life situations. Since under cell culture conditions, the NPs and their breakdown products (which might impair cell survival) stay in near the cells and may operate as a repository with a continual influence, the cytotoxic activity may be more severe *in vitro* compared to *in vivo*. On the contrary, if they are biodegradable, NPs are continually removed from the body [141]. Regarding pharmacological purposes, it is clear that the evaluation of *in vivo* security of MNPs is critical, as shown above.

Macrophages in the reticuloendothelial tissues remove intravenously administered IONPs from the systemic circulation, as previously stated and generally nanoparticles can induce diverse effects on cells (Figure 6) [142]. A lower pH, hydrolytic enzymes, and proteins involved in iron metabolic processes are among the features that allow these NPs to be picked by cells through receptor-mediated endocytosis and probably digested in the lysosomal compartment [143]. The liberated free iron is subsequently integrated into the body's iron storage, where it is eventually discovered as hemoglobin and in part attached to transferrin before being removed mostly via the feces [144,145]. For an average adult male, the entire bodily iron storage is about 4000 mg. Small doses of iron infusion may not cause iron-related toxicity complications; however, larger concentrations may cause a rise in plasmatic iron levels due to the exceeded transferrin iron-binding capability. Oxidative stress and different toxicities, such as heart and liver damage, might be the result [146]. As a result, the cytotoxicity of MNPs has been investigated using a variety of delivery methods, including intravenous, intraperitoneal, and subcutaneous injection.

The security of IONPs stabilized using OA and Pluronic F-127 coatings after administration to rats at a dosage of 10 mg Fe/kg was studied [147]. Alanine aminotransferase, aspartate aminotransferase, and alkaline phosphatase concentrations in the blood did not change significantly or for a long period after the iron administration, although 55% of the administered iron had been deposited in the hepatocytes 6 h after the infusion. IONPs may produce oxidative harm to the phospholipidic cell membrane. Therefore, it was decided to assess the amounts of lipid hydroperoxide to determine the extent of the oxidative injury. The tissue-specific variations in lipid hydroperoxide concentrations were discovered. They were somewhat greater for the hepatic, splenic, and renal tissues compared to the other sites, including pulmonary, cerebral, and cardiac tissues. Nevertheless, even after one-week post injection, the elevated lipid hydroperoxide concentrations did not affect tissue shape or cellular stability, showing that the oxidative stress was modest and negligible at the administered dosage. MNP dosage and dosing duration are critical to ensuring the safety of this medication, as will be addressed more below.

To be sure, in several animal species, the MNPs exhibited an acceptable level of safety when supplied at various doses. As an example, rats given Ferumoxtran-10 at the 2.6 mg Fe/kg *i.v.* dosage was found to be protected, with no alterations in hemodynamic variables, whereas rats given 13 mg Fe/kg dosage had a gentle enhancement in aortic blood circulation; however, there were no arrhythmias or adjustments in the electrocardiogram, and no breathing toxicities were identified. Just very high single dosages of Ferumoxtran-10, including 126 mg Fe/kg and 400 mg Fe/kg, were treatment-related clinical symptoms in rats found to remain for 24 h post administration. There were modest alterations in body weight and hematological markers throughout rats after continuous intravenous treatment of 17.9 mg Fe/kg Ferumoxtran-10 (3–5 dosages). Ferumoxtran-10 was neither mutagenic nor hazardous to the maternal rats or rabbits given doses as high as 15 mg Fe/kg/day or repetitive administrations (up to < 50 mg Fe/kg/day throughout rabbits) [148]. Up to a dosage of 20 mg Fe/kg *i.v.* administration throughout dogs, Ferumoxtran-10 showed

no treatment-related heart or kidney toxicities, but at elevated dosages, including 200 mg Fe/kg (75 times greater than the level employed in humans as an MRI contrast agent), modest cardiac toxic effects, hypotension, increased respiratory level, increased heart rate, as well as a significant rise in urinary discharge and salt excretion in the urine, were seen. Singular dosages of up to 25 mg Fe/kg throughout monkeys did not result in any significant evidence of cardiac toxicity or changes in blood parameters. Once treatment with iron infusion, blood iron concentrations were shown to be significantly elevated among rats, dogs, and monkeys; however, normal concentrations were restored following the therapy was stopped. It was shown that the dose-dependent toxicity of IONPs could be characterized whereas, at a higher dosage, the toxicity of the toxicological reactions was identical throughout all animal groups.

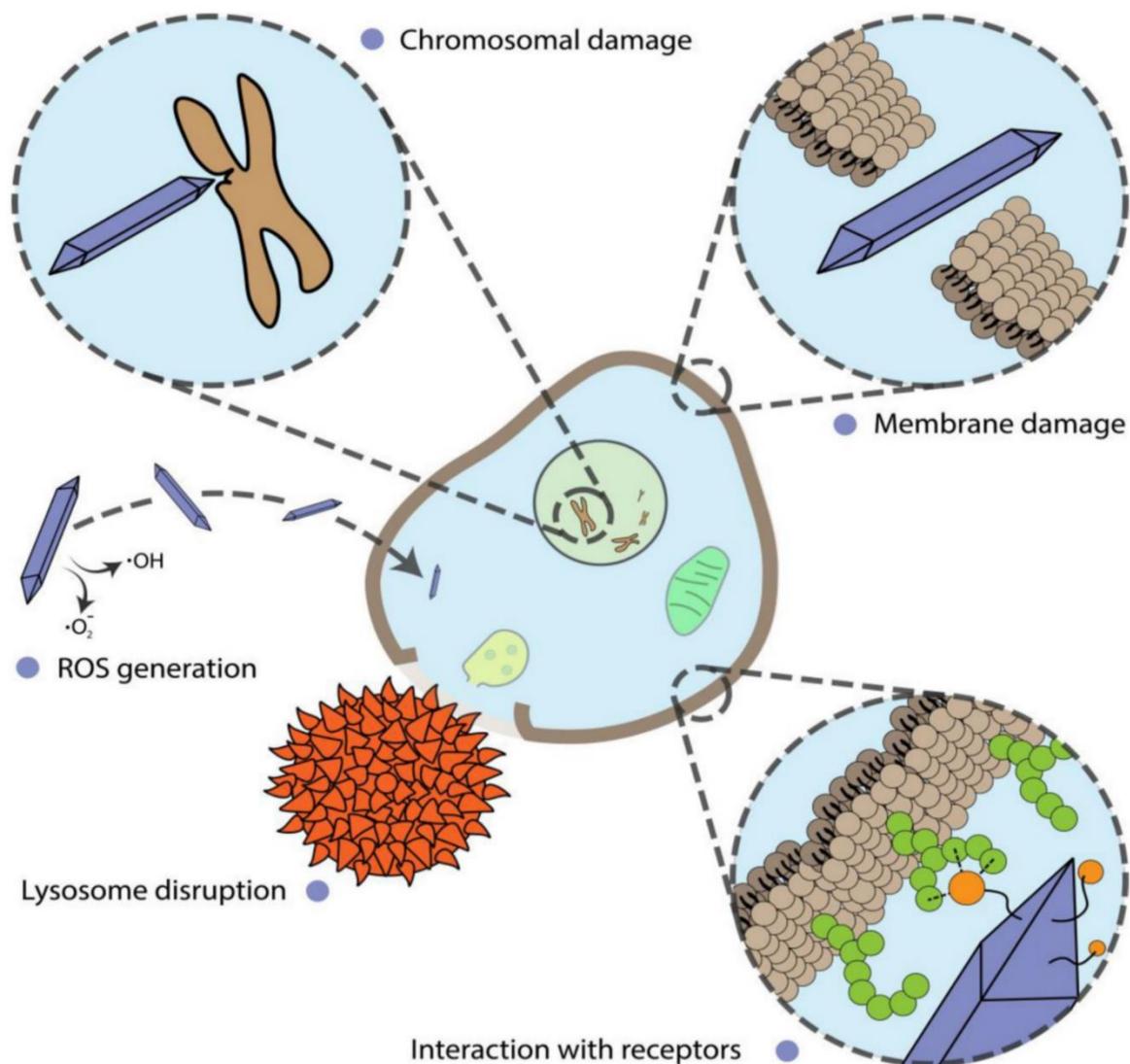


Figure 6. Various mechanisms of nanoparticles influence on cells [135].

According to the results of comparative acute toxicity research (intravenous singular dosage) conducted in mice utilizing IONPs that were either uncoated or coated with various materials,

Including dextran, polyethylene glycol (PEG), albumin, and polylactic acid (PLA), the IC50 values were found to be 355, 355, 232, and 559 mg/kg, correspondingly. As a consequence of these tests, it seems that PLA coatings are more tolerable for NPs than the other coating substances. Pulmonary toxicity, cardiac black material, and moderate

splenomegaly were found during the pathological examination of the deceased mice [149]. There was no information provided in this paper on the various NPs' particle size and coating degree, and this information is critical to understanding the toxic effects of NPs. It was discovered that the mean lethal dosage (LD50) of the aforementioned NPs was more than 2000 mg/kg following oral administration, indicating that oral consumption is more tolerable than a single intravenous infusion of the same dose. When administered intravenously, anhydroglucose polymer (50–150 nm diameter) was shown to be biocompatible since no clinical symptoms of toxicity were noticed throughout rats or mice following the delivery of the IONPs [150]. Dextran stabilized IONPs (AMI-25, Feridex) showed excellent endurance throughout rats and dogs up to 3000 mol Fe/kg, which is about 150 times greater than the dosage necessary for diagnostic hepatic MRI [142]. This suggests that these NPs have a wide security window, which is consistent with previous studies.

Intravenous injections of IONPs (100-nm-sized nanoparticles covered by anhydroglucose polymer) throughout human tumor individuals were proven to be safe and effective, also IONPs can be applied efficiently for regenerative tissue engineering (Figure 7). NPs administered using doxorubicin (Dox) (overall iron infused per therapeutic session, 132 to 468 mg) were used to treat cancer. The serum iron concentrations were temporarily elevated; however, no hazardous manifestations were noticed, and the quantities of iron in the urine were confirmed to be within the acceptable limit. Toxicological events were not seen in this clinical study, where a magnetic field was given to the upper layer of the skin, and only a small cutaneous discoloration was noticed [151,152]. On the other hand, intravenously administered Ferucarbotran (between 5- and 40-mM Fe/kg) was shown to be safe, except for a transitory reduction in platelet clotting factor XI function, which did not result in any medically meaningful complications. It was also discovered that there were no alterations in cardiac parameters such as blood pressure and cardiac pulse [153–157]. Several mild complications involving the central nervous system (CNS), such as diminished or enhanced voluntary locomotion, rearing, exophthalmos, or mydriasis, have been identified in another research investigation [158]. Human chronic iron toxicity (cirrhosis or hepatocellular cancer) has been reported throughout the publications when large dosages of iron have been administered, with hepatic iron levels above 4 mg Fe/g liver wet weight being seen in certain cases [159].

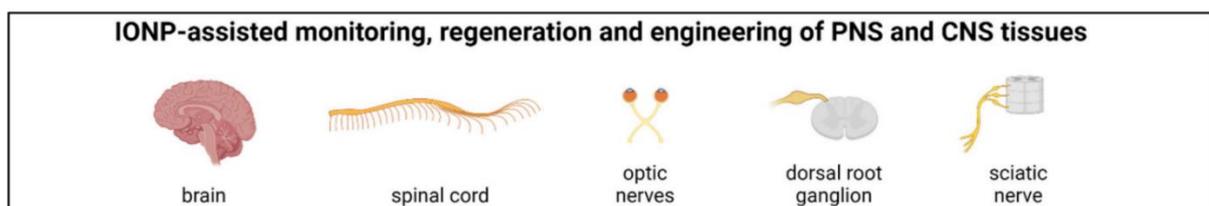


Figure 7. Probable targets for iron oxide nanoparticles-assisted tissue engineering and regenerative tissue engineering for both peripheral and central nervous system [157].

The connections of NPs with the physiological system and, therefore, their security is influenced by the surface features of NPs, including surficial charge. Nonspecific binding may occur, and cells can be destroyed when neutral or negatively charged NPs bind less effectively with blood constituents and plasma polypeptides than positively charged NPs. The fast elimination from the circulatory system of positively charged NPs also results in nonspecific tissue absorption, as previously stated [160,161].

6. Tissue Engineering Applications of Fe₂O₃-Based Nanocomposite

Creating substitutes to maintain, restore, or augment the function of injured or damaged tissues is the main goal of tissue engineering [162]. To induce and guide tissue growth, tissue engineering has attempted to utilize scaffolds that mechanically support structure and cells that drive the regeneration process, and bioactive molecules that regulate cell differentiation and functionality [163,164]. Among the polymorphic forms of iron oxides,

including $\gamma\text{-Fe}_2\text{O}_3$ (maghemite), Fe_3O_4 (magnetite), and FeO (wustite), while $\gamma\text{-Fe}_2\text{O}_3$ and Fe_3O_4 nanoparticles are the most studied ones in tissue engineering due to their exceptional properties, such as paramagnetic, high specific surface area, biocompatibility, nontoxicity, etc., at the nanoscale. Due to these materials' magnetic response abilities, research in biodegradable magnetic nanocomposites is a rapidly growing topic for biomedical applications (Figure 8), particularly tissue regeneration and musculoskeletal issues such as osteoarthritis [165–168].

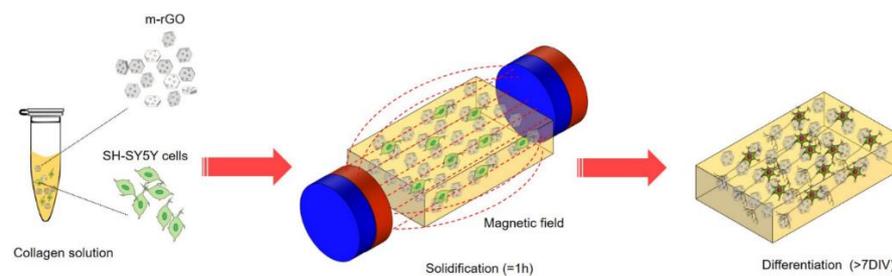


Figure 8. Magnetic-assisted cell alignment for a magnetic-responsive nanocomposite of rGO/collagen hydrogel [168].

Fe₂O₃-Based Nanocomposites in Bone Tissue Regeneration

The Bone has a special architecture; a more detailed look at the bone structure reveals that it comes into two main parts: compact bone (includes the osteon structures and osteoblast) and spongy bone (the highly porous part) [169]. The bone regeneration procedure begins with the migration and recruitment of osteoprogenitor cells to a flawed section, where they differentiate into osteoblasts and form bone's ECM mineral part, also known as hydroxyapatite (HA). Magnetic nanoparticles solely or in combination with a magnetic field can help improve and accelerate this process by affecting the three main components of tissue regeneration: (1) Scaffolds (2) Stem cells and (3) Growth factors.

Aside from blood cells, most, if not all, normal cells in human tissues are anchorage-dependent and reside in a solid matrix known as extracellular matrix (ECM). One strategy for bone regeneration is using scaffolds to mimic the natural bone tissue condition. An applicable scaffold must possess a surface that is appropriate for cell attachment, and also should be made with biocompatible substances that act as temporary support to tolerate mechanical forces in the injured zone until the host tissue is completely recuperated. In particular, the biodegradation rate should be matched to the rate of bone formation operation, and the interconnectivity of the porous structure for vascularization is crucial for scaffold design [170].

Figure 9 describes how a bone tissue scaffold works in the bone regeneration process. The particular scaffold that has been represented in the Figure 9 is made of carbon nanotube as the base component with combination of polymers. Afterward, mesenchymal stem cell (MSC) with accompanying of appropriate growth factor, antibiotic, and organic molecules was cultured and its effects on bone healing process demonstrated step by step.

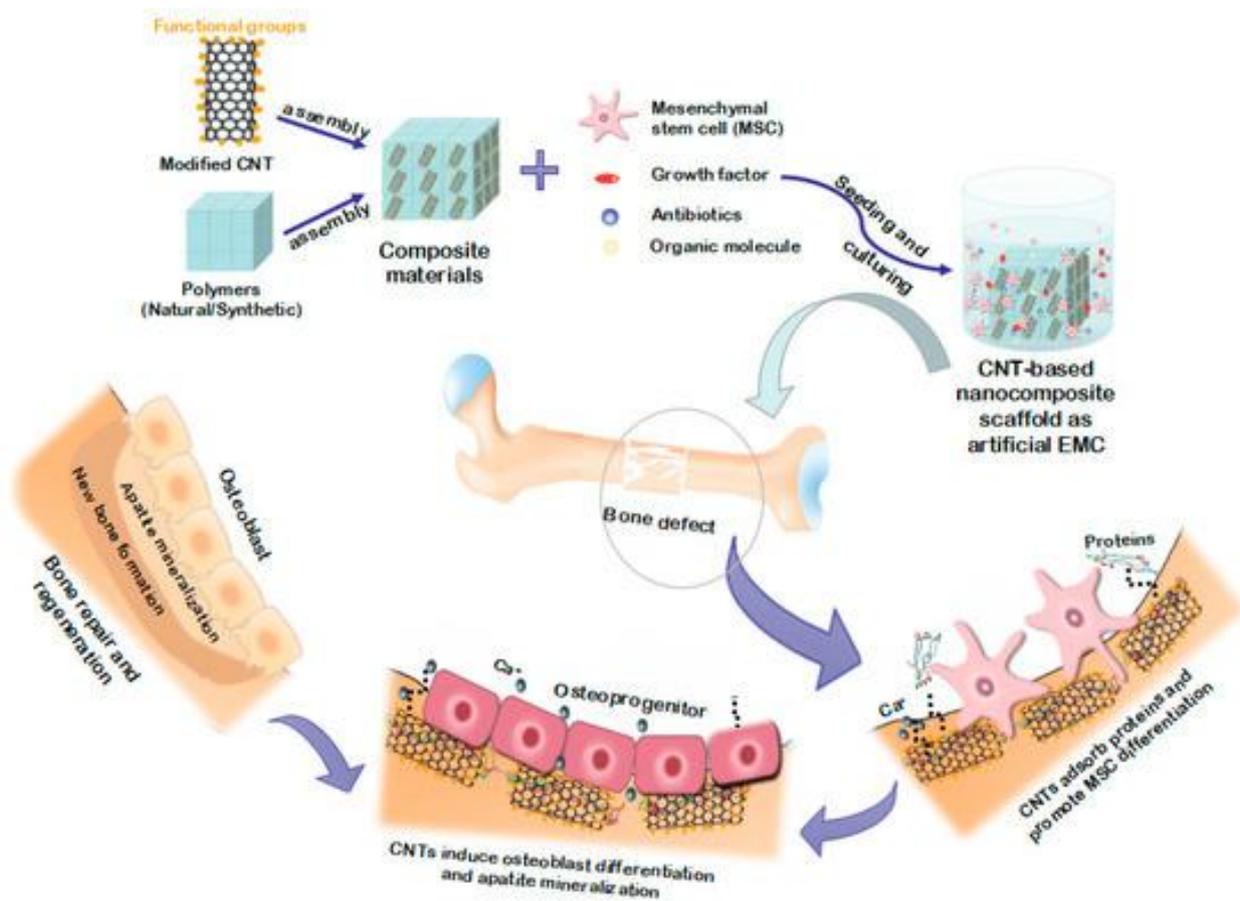


Figure 9. Describes the process of fabricating tissue scaffold and its role in tissue regeneration. Reproduced with permission from [171].

The use of magnetic nanoparticles in bone tissue engineering is a growing approach to designing a scaffold. Indeed, magnetic fields have been shown to improve implant integration, and the mineral density of Neotissue, and fasten the defect healing process [172]. These properties associated with the potential of magnetic nanoparticles serving as drug delivery and gene transfection vessels have compelled scientists to conduct extensive research on them in recent years [173]. In a study by Saber-Samandari et al. [174] a combination of these properties was shown. They synthesized bifunctional nanocomposite scaffolds for photothermal therapy and tissue engineering using magnetic nanoparticles as photothermal conversion agents. The incorporation of carboxyl-functionalized multiwalled carbon nanotubes and magnetic iron oxide nanoparticles into the porous matrix of the scaffold caused an increase in BSA (Bovine Serum Albumin) surface adsorption. They also discovered that the presence of magnetic nanoparticles enhanced biocompatibility, as demonstrated by an osteoblastic cell line, as well as the compressive strength of the scaffolds. Świątek et al. [175] created a multifunctional system that allows for multiway cell stimulation by combining hydroxylated MWCNTs with magnetic iron oxide nanoparticles embedded into a porous PCL matrix. Coprecipitation was used to create these hybrid nanoparticles, and the solvent casting/porogen leaching method was used to create the scaffolds. The carbon nanotubes positively affected cell adhesion, according to preliminary research conducted with an osteoblastic cell line. In another study, Hu et al. [176] used poly (lactic-co-glycolic acid)/polycaprolactone/beta-tricalcium phosphate (PPT) scaffolds in another study with γ -Fe₂O₃ encapsulation. The γ -Fe₂O₃ scaffolds were coated with polyglucose sorbitol carboxymethyl ether. PPT-Fe scaffolds exhibited homogeneous iron distribution and long-term iron release while encapsulated within resorbable PPT scaffolds. Wettability, superparamagnetic, hardness, tensile strength, and elasticity modulus were

also observed in that scaffold. Experiments with rat adipose-derived mesenchymal stem cells (rADSCs) in scaffolds revealed an increase in the expression of integrin 1, alkaline phosphatase, and osteogenesis-related genes. Furthermore, improved *in vivo* bone regeneration was observed after implanting PPT-Fe scaffolds in rat calvarial bone defects. In addition, Meng et al. [177] also developed a novel magnetic nanofibrous composite scaffold by combining γ -Fe₂O₃ nanoparticles, hydroxyapatite nanoparticles, and polymer (lactide acid). Under the influence of a static magnetic field, this magnetic scaffold accelerated the formation of new bone tissue in a rabbit model.

A growing number of studies have revealed that IONPs influence BMSC differentiation. On the one hand, the magnetic scaffold itself can induce osteogenesis, as evidenced by data that polymer/IONP (iron oxide nanoparticles) scaffolds can enhance bone regeneration in the presence and absence of a constant magnetic field (CMF) [178–181]. Cells in interaction with magnetic scaffolds can experience nanoscale stresses analogous to mechanical forces, stretching cell membranes and activating channels and receptors [182]. Furthermore, under cytoplasmic neutral circumstances, IONPs display catalase-like activity, breaking H₂O₂ to produce H₂O with no cytotoxicity [183]. This distinct feature of IONPs increases cell proliferation and accelerates cell cycle progression in MSCs by stimulating the p38 mitogen-activated protein kinase (MAPK) signaling pathway [184,185]. In the acidic environment of endosomes or lysosomes, IONPs break down into Fe³⁺ and PSC molecules. Since these two components will not increase MSC development into osteoblasts, the promotion effect needs the presence of integral IONPs. The studies conducted in this regard are not limited to BMSC differentiation. For instance, Ishmukhametov et al. studied differentiation of adipose-derived MSCs (hMSCs) into chondrocytes, adipocytes and osteoblasts on DNA/Magnetic nanocomposites [186].

Magnetic fields, on the other hand, may be capable of stimulating membrane phospholipid rearrangement and activating the material sensing system of cells, for instance, by activating Ca²⁺ channels on the cell membrane and stimulating various signaling pathways such as the BMP-2, integrin, and MAPK pathways [185].

Kim et al. [187] investigated the effects of electromagnetic fields (45 Hz, 1 mT intensity) and MNPs (50 mg/mL Fe₃O₄) on hBMSCs alone or in combination and discovered that both electromagnetic fields and MNPs can enhance osteogenic differentiation, but that using MNPs in combination with electromagnetic field exposure is more operative for enhancing osteogenic differentiation.

In addition, a CMF can impact the shape and orientation of ECM proteins. It has been reported that a unique protein complex (magnetoreceptor, MagR) with an inherent magnetic moment may detect the direction of the magnetic field and deviate from it. MagR is made up of two proteins: iron-sulfur cluster assembly protein (ISCA1) and cryptochrome (human homologous gene Cry1) [188]. According to research, the ISCA1 protein alone is insufficient to cause the cell to react to a magnetic field, hence the protein complex's stability is critical [189]. The actin cytoskeleton, focal adhesions, integrin binding, stress fibers, and cell responses to mechanical stimulation are all associated with magnetically sensitive proteins. As the MagR protein compound in the cell lines up in the direction of a magnetic field, torque or pressure produce toward the mechanical force receptors such as actin fibers, adhesion sites, and integrins that go along with actin redispersion or integrin-mediated MAPK signaling pathway activation. Thereby, mechanical signals can be transferred to the nucleus to influence the expression of osteogenic genes [190].

The findings of Wang et al. [191] indicated that magnetic IONPs encouraged osteogenic differentiation of MSCs as whole particles and that the IONP-driven magnetogenetic response resulted in gene deregulation in their experimental system. The lncRNA INZEB2 is important for osteogenic differentiation because it controls ZEB2 expression as well as the BMP/Smads pathway. Their findings showed how IONPs influence MSC osteogenic differentiation and how lncRNAs contribute to the regulation of magnetic IONP cellular effects.

7. Wound Healing Applications of Fe₂O₃-Based Nanocomposite

Wound healing is a highly systematic procedure of repairing defected tissue, which is consisted of four consecutive, but overlapping, biological phases: hemostasis, inflammation, proliferation, and remodeling [192,193]. Any disruption by both extrinsic and intrinsic factors in any of the aforementioned phases may cause each phase to be prolonged and result in a dissatisfying result, leading to chronic wound status. The colonization of contaminating pathogens at the spot of damage during the natural wound-healing process is the most commonly occurring issue with complete wound healing [194]. At the spot of a wound, the bacteria that constituted the skin microbiota protect against pathogen colonization. When pathogenic bacteria hit the critical level and generate a large amount of biofilm, the healing process slowed. *Staphylococcus aureus* is the most widely recognized colonizing pathogen influencing the early stages of wound healing, while *Pseudomonas aeruginosa* and *Escherichia coli* are commonly seen in chronic wounds and impact downward layers of the skin [195]. This pathogen-associated bare skin infection may cause acute inflammatory reactions and delayed wound healing. To prevent bacterial infections and assist in the natural wound-healing process, a suitable antimicrobial wound dressing must be used.

However, the proliferation of antimicrobial-resistant bacteria slows the healing process, entailing the development of new wound-dressing substances that are non-toxic or resistant to employment and ameliorating the efficiency of the wound-healing procedure.

In other words, the primary goal of wound dressing is to keep the wound free of extrinsic contamination. It also keeps the wound hydrated, promoting regeneration and preventing the wound's origin from being exposed [196]. As a result, wound-healing materials must be biocompatible, semi-permeable to water and oxygen, hypoallergenic, and economical. As a result, wound dressing necessitates the use of technologically progressed dressing substances instead of traditional wound-dressing materials such as cotton and wool.

The new materials can preserve the wound environment and also transfer active compounds to facilitate the wound-healing process [197].

Diverse wound-dressing products, including ointments, hydrogels, and antibacterial agents combined with polymers, are recently accessible and are made primarily of biodegradable substances including chitosan, hyaluronic acid, collagen, silicon, cellulose, and gelatin [198–203].

Due to their capability to prevent bacterial proliferation, quinolones [204], cephalosporins [205], polymyxin B [206], neomycin [206], and tetracyclines [207] are the most typically utilized antibiotics in wound dressing.

However, repeated and ineffective antibiotic administration would hasten the emergence of antibacterial resistance. To prevent such resistance, treatments based on alternative antibiotics, non-antibiotic materials, and a combination of antibiotics and non-antibiotic materials such as essential oils, honey, and nanomaterials (e.g., Ag and Au) have been recommended for wound dressings [207].

The usage of nanomaterials in wound healing is growing quickly and is being studied in clinical trials [192,208–210] (Figure 10). The primary reason for this increase is their physiochemical characteristics, which include nano size, and extended surface with a high specific area. Furthermore, because nanomaterials play a role in active drug delivery, penetrability, and cellular responses, their size and shape are favorable to their use in wound healing [197]. In wound therapy, two types of nanomaterials are widely being used (Table 2): (1) nanomaterials with inherent characteristics that typically boost wound treatment, and (2) nanomaterials as carriers for drug delivery.

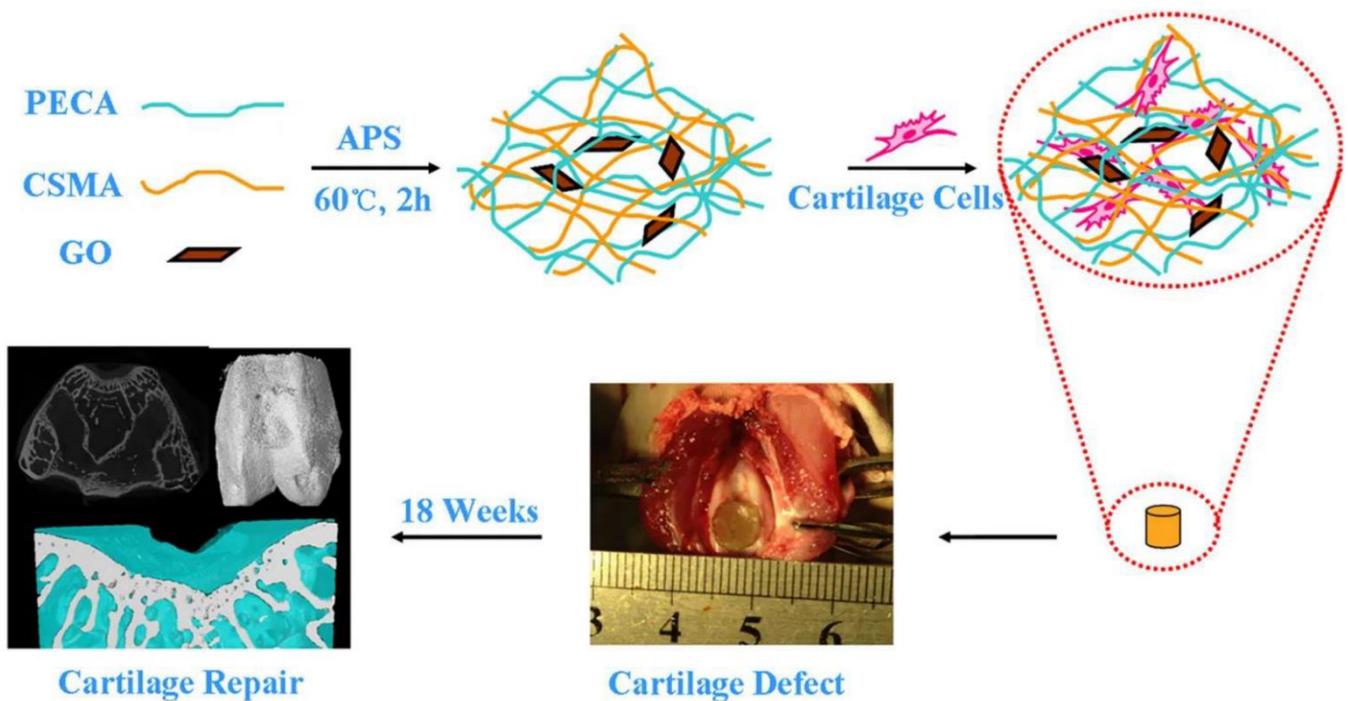


Figure 10. CSMA/PECA/GO hybrid scaffold in bone tissue engineering application (cartilage repairing) [210].

Table 2. The scaffolds fabrication methodology, their characteristics and the effect of the presence of IONPs on their properties, and potential clinical translations have been discussed.

Scaffold Type	Scaffold Fabrication Method	Effects of IONPs on the Scaffold	Ref.
Magnetic hydrogels	nanohydroxyapatite-coated γ -Fe ₂ O ₃ (around 10 wt%)/PVA composite hydrogels	remarkable influence on the porous structures average pore diameter of: $1.6 \pm 0.3 \mu\text{m}$ enhancing compressive strength: $29.6 \pm 6.5 \text{ MPa}$ positive impact on osteoblasts adhesion and proliferation	[211]
Magnetic hydrogels	hyaluronic acid/chondroitin sulfate/Fe ₂ O ₃ /nHAP/PVA hydrogels	Promotion of chondrocyte adhesion, proliferation, and growth	[212]
Electrospinning	electrospun PCL incorporated by dendrimerized superparamagnetic nanoparticles	Significantly decreases the PCL nanofibers size to $495 \pm 144 \text{ nm}$ and improves cell attachment and growth	[213]
Electrospinning	γ -Fe ₂ O ₃ nanoparticles filled polyvinyl alcohol	higher fiber diameter and surface roughness higher cells proliferation rate	[214]
Electrospinning	A novel nanofibrous composite scaffold composed of super-paramagnetic γ -Fe ₂ O ₃ nanoparticles (MNP), hydroxyapatite nanoparticles (nHA) and poly lactide acid (PLA)	MNPs accelerates new bone tissue formation and remodeling in the rabbit defect.	[177]
Electrospinning	poly(vinyl alcohol) filled by γ -Fe ₂ O ₃ nanoparticles	maximum Young's modulus (273.51 MPa) cell viability and cell growth rate Enhancing scaffold's mechanical properties	[215]
Magnetic Hydrogel	Poly(vinyl alcohol)/nano-hydroxyapatite (n-HA)/magnetic nanoparticles (Fe ₂ O ₃) fibers	Uniform and enhanced growth of BMSCs on the surface High rates of proliferation Significant simulated chondrocyte-related gene expression	[216]

Due to their inherent nature, metal and metal oxide nanomaterials are more effective in the wound-healing treatment and antibacterial action than traditional materials. Metallic nanoparticles' size, structure, surface modification, zeta potential, porosity, and thermal

stability typically influence their efficacy in biological applications [217–220]. Considering their favorable physiochemical properties and antibacterial activity, Silver (Ag) [83], gold (Au) [84], and zinc oxide (ZnO) [85] are the most investigated metallic nanoparticles. Other metals and metal oxides, such as iron oxide (Fe_2O_3), are also being studied in this regard [195].

Harandi et al. [45] studied the antimicrobial activity of Fe_2O_3 nanoparticles as coating for polyvinyl alcohol-prebiotic gum arabic-polycaprolactone (PVA-GA-PCL) electrospun nanofibers for wound dressing applications. Their findings revealed spherical Fe_2O_3 NPs with a mean size diameter of 100 nm that contained LAB and demonstrated significant antimicrobial potential against pathogens such as *E. coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Candida albicans*. These results demonstrated both the biocompatibility of coated nanofibers with mouse embryonic fibroblast cell lines and their antimicrobial and antibiofilm efficacy against pathogens. *C. Albicans* exhibited the greatest growth inhibition by Fe_2O_3 NPs-LAB@PVA-GA-PCL. Intriguingly, combining probiotic *Lactobacillus* and prebiotic GA as a symbiotic hybrid revealed synergistic antimicrobial effects, implying the therapeutic potential for wound healing applications.

Another study by Raisi et al. [46] fabricated a nanocomposite dressing (NCD) made up of carboxymethyl chitosan (CMC) with various content of Fe_2O_3 nanoparticles (0, 2.5, 5, and 7.5 wt%) by a method called freeze-drying (FD) technique to study its potential for wound healing applications. The results showed that the wound dress was porous with micron-sized interconnections. Indeed, the results show that as the amount of Fe_2O_3 nanoparticles increases, so does the porosity. Tensile strength was 0.32 MPa for the pure sample and 0.85 MPa for the sample with the highest percentage of magnetic nanoparticles, respectively.

Furthermore, the cytotoxicity of this nanocomposite revealed no cytotoxicity toward fibroblast cell growth and suitable biocompatibility. The results indicated that NCD had exceptional biodegradability, biocompatibility, and mechanical properties. As a result, NCD made of CMC and Fe_2O_3 nanoparticles were proposed as a promising candidate for wound healing applications.

The positive effects of Fe_2O_3 NPs on the rapid blood coagulation in wounds with excessive bleeding were proved by a study by Rubtsov et al. [221] in 2019. They synthesized a novel nanoparticle-hydrogel composite as a potential hemostatic wound-healing agent composed of Poly (sodium acrylate) cross-linked by aluminum ions and nanoscale boehmite ($\gamma\text{-AlOOH}$) and Fe_2O_3 as fillers.

8. Conclusions

In advanced biomedical systems, scientists are looking for some special features such as good mechanical and thermal stability, biocompatibility, porosity, and biodegradability. These specific characteristics have been fulfilled by employing Fe_2O_3 in nanocomposite structures. Thus, IONPs increased drug loading capacity in novel drug delivery and efficiency of tissue engineering applications due to their extended surface area and porosity. In addition, in wound dressings, IONPs have enhanced the permeability of water and oxygen in a more controllable manner.

Due to their incredible superparamagnetic properties, IONPs have been used widely in biomedical engineering applications, especially novel targeted drug delivery systems. This review extensively was devoted to the structural characteristics, and potential applications of IONPs in the areas such as drug delivery, tissue engineering, and wound healing. Different methods of utilizing IONPs in each of those areas have also been discussed and supplied outcomes from recent studies on Fe_2O_3 -based nanocomposites revealed the applicability of this metal oxide for advanced biomedical purposes.

Author Contributions: Conceptualization, M.P.; writing—original draft preparation, E.R., A.S., S.M., M.A.; writing—review and editing, A.R., A.M.D.-P.; supervision, A.R., A.M.D.-P. All authors have read and agreed to the published version of the manuscript.

Funding: Financial support from the Community of Madrid within the framework of the Multiyear Agreement with the University of Alcalá in the line of action “Stimulus to Excellence for Permanent University Professors”, Ref. EPU-INV/2020/012, is gratefully acknowledged.

Data Availability Statement: Data sharing is not applicable as no new data was created or analyzed in this study.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Iqbal, M.Z.; Wu, A.; Shi, D.; Liu, Q. Magnetic Nanohybrids for Magnetic Resonance Imaging and Phototherapy Applications. In *Tissue Engineering and Nanotheranostics*; World Scientific: Singapore, 2018; pp. 101–149. [[CrossRef](#)]
2. Pourmadadi, M.; Ahmadi, M.; Abdouss, M.; Yazdian, F.; Rashedi, H.; Navaei-Nigjeh, M.; Hesari, Y. The synthesis and characterization of double nanoemulsion for targeted Co-Delivery of 5-fluorouracil and curcumin using pH-sensitive agarose/chitosan nanocarrier. *J. Drug Deliv. Sci. Technol.* **2022**, *70*, 102849. [[CrossRef](#)]
3. Brero, F.; Albino, M.; Antocchia, A.; Arosio, P.; Avolio, M.; Berardinelli, F.; Bettega, D.; Calzolari, P.; Ciocca, M.; Corti, M.; et al. Hadron Therapy, Magnetic Nanoparticles and Hyperthermia: A Promising Combined Tool for Pancreatic Cancer Treatment. *Nanomaterials* **2020**, *10*, 1919. [[CrossRef](#)] [[PubMed](#)]
4. Ferreira, L.P.; Reis, C.P.; Robalo, T.T.; Jorge, M.E.M.; Ferreira, P.; Gonçalves, J.; Hajalilou, A.; Cruz, M.M. Assisted Synthesis of Coated Iron Oxide Nanoparticles for Magnetic Hyperthermia. *Nanomaterials* **2022**, *12*, 1870. [[CrossRef](#)] [[PubMed](#)]
5. Pourmadadi, M.; Farokh, A.; Rahmani, E.; Shamsabadipour, A.; Eshaghi, M.M.; Rahdar, A.; Ferreira, L.F.R. Porous alumina as potential nanostructures for drug delivery applications, synthesis and characteristics. *J. Drug Deliv. Sci. Technol.* **2022**, *77*, 103877. [[CrossRef](#)]
6. Samadi, A.; Haseli, S.; Pourmadadi, M.; Rashedi, H.; Yazdian, F.; Navaei-Nigjeh, M. Curcumin-loaded chitosan-agarose-montmorillonite hydrogel nanocomposite for the treatment of breast cancer. In Proceedings of the 2020 27th National and 5th International Iranian Conference on Biomedical Engineering (ICBME), Tehran, Iran, 26–27 November 2020. [[CrossRef](#)]
7. Mirzaie, Z.; Barati, M.; Tokmedash, M.A. Anticancer Drug Delivery Systems Based on Curcumin Nanostructures: A Review. *Pharm. Chem. J.* **2020**, *54*, 353–360. [[CrossRef](#)]
8. Tokmedash, M.A.; Zadeh, E.S.; Balouchi, E.N.; Salehi, Z.; Ardestani, M.S. Synthesis of smart carriers based on tryptophan-functionalized magnetic nanoparticles and its application in 5-fluorouracil delivery. *Biomed. Mater.* **2022**, *17*, 045026. [[CrossRef](#)]
9. Fusser, M.; Øverbye, A.; Pandya, A.D.; Mørch, Y.; Borgos, S.E.; Kildal, W.; Snipstad, S.; Sulheim, E.; Fleten, K.G.; Askautrud, H.A.; et al. Cabazitaxel-loaded Poly(2-ethylbutyl cyanoacrylate) nanoparticles improve treatment efficacy in a patient derived breast cancer xenograft. *J. Control. Release* **2019**, *293*, 183–192. [[CrossRef](#)]
10. Samadi, A.; Pourmadadi, M.; Yazdian, F.; Rashedi, H.; Navaei-Nigjeh, M.; Eufrazio-da-Silva, T. Ameliorating quercetin constraints in cancer therapy with pH-responsive agarose-polyvinylpyrrolidone-hydroxyapatite nanocomposite encapsulated in double nanoemulsion. *Int. J. Biol. Macromol.* **2021**, *182*, 11–25. [[CrossRef](#)]
11. Gao, X.; Wang, B.; Wei, X.; Men, K.; Zheng, F.; Zhou, Y.; Zheng, Y.; Gou, M.; Huang, M.; Guo, G.; et al. Anticancer effect and mechanism of polymer micelle-encapsulated quercetin on ovarian cancer. *Nanoscale* **2012**, *4*, 7021–7030. [[CrossRef](#)]
12. Haseli, S.; Pourmadadi, M.; Samadi, A.; Yazdian, F.; Abdouss, M.; Rashedi, H.; Navaei-Nigjeh, M. A novel pH-responsive nanoniosomal emulsion for sustained release of curcumin from a chitosan-based nanocarrier: Emphasis on the concurrent improvement of loading, sustained release, and apoptosis induction. *Biotechnol. Prog.* **2022**, *38*, e3280. [[CrossRef](#)]
13. Foroushani, P.H.; Rahmani, E.; Alemzadeh, I.; Vossoughi, M.; Pourmadadi, M.; Rahdar, A.; Díez-Pascual, A.M. Curcumin Sustained Release with a Hybrid Chitosan-Silk Fibroin Nanofiber Containing Silver Nanoparticles as a Novel Highly Efficient Antibacterial Wound Dressing. *Nanomaterials* **2022**, *12*, 3426. [[CrossRef](#)] [[PubMed](#)]
14. Rahmani, E.; Pourmadadi, M.; Ghorbanian, S.A.; Yazdian, F.; Rashedi, H.; Navaee, M. Preparation of a pH-responsive chitosan-montmorillonite-nitrogen-doped carbon quantum dots nanocarrier for attenuating doxorubicin limitations in cancer therapy. *Eng. Life Sci.* **2022**, *22*, 634–649. [[CrossRef](#)] [[PubMed](#)]
15. Lee, A.L.; Wang, Y.; Cheng, H.Y.; Pervaiz, S.; Yang, Y.Y. The co-delivery of paclitaxel and Herceptin using cationic micellar nanoparticles. *Biomaterials* **2009**, *30*, 919–927. [[CrossRef](#)] [[PubMed](#)]
16. Sun, C.; Lee, J.S.H.; Zhang, M. Magnetic nanoparticles in MR imaging and drug delivery. *Adv. Drug Deliv. Rev.* **2008**, *60*, 1252–1265. [[CrossRef](#)] [[PubMed](#)]
17. Darbandi, M.; Stromberg, F.; Landers, J.; Reckers, N.; Sanyal, B.; Keune, W.; Wende, H. Nanoscale size effect on surface spin canting in iron oxide nanoparticles synthesized by the microemulsion method. *J. Phys. D Appl. Phys.* **2012**, *45*, 195001. [[CrossRef](#)]
18. Yu, M.K.; Park, J.; Jon, S. Targeting Strategies for Multifunctional Nanoparticles in Cancer Imaging and Therapy. *Theranostics* **2012**, *2*, 3–44. [[CrossRef](#)]
19. Kanwar, J.R.; Roy, K.; Kanwar, R.K. Chimeric aptamers in cancer cell-targeted drug delivery. *Crit. Rev. Biochem. Mol. Biol.* **2011**, *46*, 459–477. [[CrossRef](#)]
20. Rahdar, S.; Rahdar, A.; Ahmadi, S.; Trant, J. Adsorption of bovine serum albumin (BSA) by bare magnetite nanoparticles with surface oxidative impurities that prevent aggregation. *Can. J. Chem.* **2019**, *97*, 577–583. [[CrossRef](#)]

21. Davarpanah, A.M.; Rahdar, A.; Dastnae, M.A.; Zeybek, O.; Beyzaei, H. (1-x) BaFe₁₂O₁₉/xCoFe₂O₄ hard/soft magnetic nanocomposites: Synthesis, physical characterization, and antibacterial activities study. *J. Mol. Struct.* **2019**, *1175*, 445–449. [[CrossRef](#)]
22. Taimoory, S.M.; Rahdar, A.; Aliahmad, M.; Sadeghfhar, F.; Hajinezhad, M.R.; Jahantigh, M.; Shahbazi, P.; Trant, J.F. The synthesis and characterization of a magnetite nanoparticle with potent antibacterial activity and low mammalian toxicity. *J. Mol. Liq.* **2018**, *265*, 96–104. [[CrossRef](#)]
23. Nouri Hajbaba, M.; Pourmadadi, M.; Yazdian, F.; Rashedi, H.; Abdouss, M.; Zhohrabi, D.S. The Function of Chitosan/Agarose Biopolymer on Fe₂O₃ Nanoparticles and Evaluation of Their Effects on MCF-7 Breast Cancer Cell Line and Expression of BCL2 and BAX Genes. *Biotechnol. Prog.* **2022**, e3305. [[CrossRef](#)] [[PubMed](#)]
24. Pourmadadi, M.; Ahmadi, M.J.; Dinani, H.S.; Ajalli, N.; Dorkoosh, F. Theranostic applications of stimulus-responsive systems based on Fe₂O₃. *Pharm. Nanotechnol.* **2022**, *10*, 90–112. [[CrossRef](#)] [[PubMed](#)]
25. Qiao, Z.; Shi, X. Dendrimer-based molecular imaging contrast agents. *Prog. Polym. Sci.* **2015**, *44*, 1–27. [[CrossRef](#)]
26. Rahdar, A.; Taboada, P.; Aliahmad, M.; Hajinezhad, M.R.; Sadeghfhar, F. Iron oxide nanoparticles: Synthesis, physical characterization, and intraperitoneal biochemical studies in Rattus norvegicus. *J. Mol. Struct.* **2018**, *1173*, 240–245. [[CrossRef](#)]
27. Ahmadi, M.; Pourmadadi, M.; Ghorbanian, S.A.; Yazdian, F.; Rashedi, H. Ultra pH-sensitive nanocarrier based on Fe₂O₃/chitosan/montmorillonite for quercetin delivery. *Int. J. Biol. Macromol.* **2021**, *191*, 738–745. [[CrossRef](#)] [[PubMed](#)]
28. Mishra, M.; Chun, D.-M. α -Fe₂O₃ as a photocatalytic material: A review. *Appl. Catal. A Gen.* **2015**, *498*, 126–141. [[CrossRef](#)]
29. Debnath, N.C.; Anderson, A.B. Optical Spectra of Ferrous and Ferric Oxides and the Passive Film: A Molecular Orbital Study. *J. Electrochem. Soc.* **1982**, *129*, 2169–2174. [[CrossRef](#)]
30. Xing, R.; Lin, H.; Jiang, P.; Qu, F. Biofunctional mesoporous silica nanoparticles for magnetically oriented target and pH-responsive controlled release of ibuprofen. *Colloids Surf. A Physicochem. Eng. Asp.* **2012**, *403*, 7–14. [[CrossRef](#)]
31. Ye, Y.; Chen, H.; Zou, Y.; Ye, Y.; Zhao, H. Corrosion protective mechanism of smart graphene-based self-healing coating on carbon steel. *Corros. Sci.* **2020**, *174*, 108825. [[CrossRef](#)]
32. Korkut, S.E.; Akyüz, D.; Özdoğan, K.; Yerli, Y.; Koca, A.; Şener, M.K. TEMPO-functionalized zinc phthalocyanine: Synthesis, magnetic properties, and its utility for electrochemical sensing of ascorbic acid. *Dalton Trans.* **2016**, *45*, 3086–3092. [[CrossRef](#)]
33. Shahriari, M.; Zahiri, M.; Abnous, K.; Taghdisi, S.M.; Ramezani, M.; Alibolandi, M. Enzyme responsive drug delivery systems in cancer treatment. *J. Control. Release* **2019**, *308*, 172–189. [[CrossRef](#)] [[PubMed](#)]
34. Li, R.; Peng, F.; Cai, J.; Yang, D.; Zhang, P. Redox dual-stimuli responsive drug delivery systems for improving tumor-targeting ability and reducing adverse side effects. *Asian J. Pharm. Sci.* **2020**, *15*, 311–325. [[CrossRef](#)] [[PubMed](#)]
35. Hasan, A.; Morshed, M.; Memic, A.; Hassan, S.; Webster, T.J.; Marei, H. Nanoparticles in tissue engineering: Applications, challenges and prospects. *Int. J. Nanomed.* **2018**, *13*, 5637–5655. [[CrossRef](#)]
36. Xu, Y.; Xiao, L.; Chang, Y.; Cao, Y.; Chen, C.; Wang, D. pH and redox dual-responsive MSN-SS-CS as a drug delivery system in cancer therapy. *Materials* **2020**, *13*, 1279. [[CrossRef](#)]
37. Pourmadadi, M.; Eshaghi, M.M.; Ostovar, S.; Shamsabadipour, A.; Safakhah, S.; Mousavi, M.S.; Rahdar, A.; Pandey, S. UiO-66 metal-organic framework nanoparticles as gifted MOFs to the biomedical application: A comprehensive review. *J. Drug Deliv. Sci. Technol.* **2022**, *76*, 103758. [[CrossRef](#)]
38. Abdullah, N.H.; Shameli, K.; Abdullah, E.C.; Abdullah, L.C. Solid matrices for fabrication of magnetic iron oxide nanocomposites: Synthesis, properties, and application for the adsorption of heavy metal ions and dyes. *Compos. Part B Eng.* **2019**, *162*, 538–568. [[CrossRef](#)]
39. Eivazzadeh-Keihan, R.; Bahojb Noruzi, E.; Khanmohammadi Chenab, K.; Jafari, A.; Radinekiyan, F.; Hashemi, S.M.; Ahmadpour, F.; Behboudi, A.; Mosafar, J.; Mokhtarzadeh, A.; et al. Metal-based nanoparticles for bone tissue engineering. *J. Tissue Eng. Regen. Med.* **2020**, *14*, 1687–1714. [[CrossRef](#)] [[PubMed](#)]
40. Kim, K.; Fisher, J.P. Nanoparticle technology in bone tissue engineering. *J. Drug Target.* **2007**, *15*, 241–252. [[CrossRef](#)]
41. Zhang, S.; Vijayavenkataraman, S.; Lu, W.F.; Fuh, J.Y. A review on the use of computational methods to characterize, design, and optimize tissue engineering scaffolds, with a potential in 3D printing fabrication. *J. Biomed. Mater. Res. Part B Appl. Biomater.* **2019**, *107*, 1329–1351. [[CrossRef](#)]
42. Eltom, A.; Zhong, G.; Muhammad, A. Scaffold Techniques and Designs in Tissue Engineering Functions and Purposes: A Review. *Adv. Mater. Sci. Eng.* **2019**, *2019*, 3429527. [[CrossRef](#)]
43. Rajendran, N.K.; Kumar, S.S.D.; Houreld, N.N.; Abrahamse, H. A review on nanoparticle based treatment for wound healing. *J. Drug Deliv. Sci. Technol.* **2018**, *44*, 421–430. [[CrossRef](#)]
44. Chai, M.; Tong, W.; Wang, Z.; Chen, Z.; An, Y.; Zhang, Y. Piezoelectric-Fenton degradation and mechanism study of Fe₂O₃/PVDF-HFP porous film drove by flowing water. *J. Hazard. Mater.* **2022**, *430*, 128446. [[CrossRef](#)] [[PubMed](#)]
45. Harandi, F.N.; Khorasani, A.C.; Shojaosadati, S.A.; Hashemi-Najafabadi, S. Surface modification of electrospun wound dressing material by Fe₂O₃ nanoparticles incorporating Lactobacillus strains for enhanced antimicrobial and antibiofilm activity. *Surfaces Interfaces* **2022**, *28*, 101592. [[CrossRef](#)]
46. Raisi, A.; Asefnejad, A.; Shahali, M.; Doozandeh, Z.; Kamyab Moghadas, B.; Saber-Samandari, S.; Khandan, A. A soft tissue fabricated using a freeze-drying technique with carboxymethyl chitosan and nanoparticles for promoting effects on wound healing. *J. Nanoanalysis* **2020**, *7*, 262–274.

47. Tadic, M.; Kopanja, L.; Panjan, M.; Lazovic, J.; Tadic, B.V.; Stanojevic, B.; Motte, L. Rhombohedron and plate-like hematite (α -Fe₂O₃) nanoparticles: Synthesis, structure, morphology, magnetic properties and potential biomedical applications for MRI. *Mater. Res. Bull.* **2021**, *133*, 111055. [[CrossRef](#)]
48. Dash, P.; Raut, S.; Jena, M.; Nayak, B. Harnessing the biomedical properties of ferromagnetic α -Fe₂O₃ NPs with a plausible formation mechanism. *Ceram. Int.* **2020**, *46*, 26190–26204. [[CrossRef](#)]
49. Shahrousvand, M.; Hoseinian, M.S.; Ghollasi, M.; Karbalaemahdi, A.; Salimi, A.; Tabar, F.A. Flexible magnetic polyurethane/Fe₂O₃ nanoparticles as organic-inorganic nanocomposites for biomedical applications: Properties and cell behavior. *Mater. Sci. Eng. C* **2017**, *74*, 556–567. [[CrossRef](#)]
50. Zhao, G.; Wang, J.; Peng, X.; Li, Y.; Yuan, X.; Ma, Y. Facile solvothermal synthesis of mesostructured Fe₃O₄/chitosan nanoparticles as delivery vehicles for pH-responsive drug delivery and magnetic resonance imaging contrast agents. *Chem.–Asian J.* **2014**, *9*, 546–553. [[CrossRef](#)]
51. Gao, W.; Chan, J.M.; Farokhzad, O.C. pH-responsive nanoparticles for drug delivery. *Mol. Pharm.* **2010**, *7*, 1913–1920. [[CrossRef](#)]
52. Colombo, P.; Sonvico, F.; Colombo, G.; Bettini, R. Novel Platforms for Oral Drug Delivery. *Pharm. Res.* **2009**, *26*, 601–611. [[CrossRef](#)]
53. Wang, B.; Xu, C.; Xie, J.; Yang, Z.; Sun, S. pH controlled release of chromone from chromone-Fe₃O₄ nanoparticles. *J. Am. Chem. Soc.* **2008**, *130*, 14436–14437. [[CrossRef](#)] [[PubMed](#)]
54. Zhang, Q.; Liu, F.; Nguyen, K.T.; Ma, X.; Wang, X.; Xing, B.; Zhao, Y. Multifunctional Mesoporous Silica Nanoparticles for Cancer-Targeted and Controlled Drug Delivery. *Adv. Funct. Mater.* **2012**, *22*, 5144–5156. [[CrossRef](#)]
55. Murphy, R.F.; Powers, S.; Cantor, C.R. Endosome pH measured in single cells by dual fluorescence flow cytometry: Rapid acidification of insulin to pH 6. *J. Cell Biol.* **1984**, *98*, 1757–1762. [[CrossRef](#)] [[PubMed](#)]
56. Majewski, A.P.; Schallon, A.; Jérôme, V.; Freitag, R.; Müller, A.H.E.; Schmalz, H. Dual-Responsive Magnetic Core–Shell Nanoparticles for Nonviral Gene Delivery and Cell Separation. *Biomacromolecules* **2012**, *13*, 857–866. [[CrossRef](#)] [[PubMed](#)]
57. Shoaib, M.; Bahadur, A.; Saeed, A.; Rahman, M.S.U.; Naseer, M.M. Biocompatible, pH-responsive, and biodegradable polyurethanes as smart anti-cancer drug delivery carriers. *React. Funct. Polym.* **2018**, *127*, 153–160. [[CrossRef](#)]
58. Medina-Reyes, E.-I.; Garcia-Viacobo, D.; Martinez, F.A.C.; Chirino, Y.I. Applications and Risks of Nanomaterials Used in Regenerative Medicine, Delivery Systems, Theranostics, and Therapy. *Crit. Rev. Ther. Drug Carr. Syst.* **2017**, *34*, 35–61. [[CrossRef](#)]
59. Nabid, M.R.; Omrani, I. Facile preparation of pH-responsive polyurethane nanocarrier for oral delivery. *Mater. Sci. Eng. C* **2016**, *69*, 532–537. [[CrossRef](#)]
60. Wang, H.; Liu, G.; Gao, H.; Wang, Y. A pH-responsive drug delivery system with an aggregation-induced emission feature for cell imaging and intracellular drug delivery. *Polym. Chem.* **2015**, *6*, 4715–4718. [[CrossRef](#)]
61. Yan, L.; Chen, X.; Wang, Z.; Zhang, X.; Zhu, X.; Zhou, M.; Chen, W.; Huang, L.; Roy, V.A.L.; Yu, P.K.N.; et al. Size Controllable and Surface Tunable Zeolitic Imidazolate Framework-8–Poly(acrylic acid sodium salt) Nanocomposites for pH Responsive Drug Release and Enhanced in Vivo Cancer Treatment. *ACS Appl. Mater. Interfaces* **2017**, *9*, 32990–33000. [[CrossRef](#)]
62. Polat, T.G.; Topel, S.D. pH-responsive carboxymethyl cellulose conjugated superparamagnetic iron oxide nanocarriers. *J. Sci. Perspect.* **2019**, *3*, 99–110.
63. Maiti, D.; Mukhopadhyay, S.; Mohanta, S.C.; Saha, A.; Devi, P.S. A multifunctional nanocomposite of magnetic γ -Fe₂O₃ and mesoporous fluorescent ZnO. *J. Alloys Compd.* **2015**, *653*, 187–194. [[CrossRef](#)]
64. Zhao, C.; Qiao, X.; Shao, Q.; Hassan, M.; Ma, Z. Evolution of the Lignin Chemical Structure during the Bioethanol Production Process and Its Inhibition to Enzymatic Hydrolysis. *Energy Fuels* **2020**, *34*, 5938–5947. [[CrossRef](#)]
65. Gerami, S.E.; Pourmadadi, M.; Fatoorehchi, H.; Yazdian, F.; Rashedi, H.; Nigjeh, M.N. Preparation of pH-sensitive chitosan/polyvinylpyrrolidone/ α -Fe₂O₃ nanocomposite for drug delivery application: Emphasis on ameliorating restrictions. *Int. J. Biol. Macromol.* **2021**, *173*, 409–420. [[CrossRef](#)] [[PubMed](#)]
66. Al-Zahrani, F.A.; Salem, S.S.; Al-Ghamdi, H.A.; Nhari, L.M.; Lin, L.; El-Shishtawy, R.M. Green Synthesis and Antibacterial Activity of Ag/Fe₂O₃ Nanocomposite Using Buddleja lindleyana Extract. *Bioengineering* **2022**, *9*, 452. [[CrossRef](#)]
67. Li, S.; Zhang, R.; Wang, D.; Feng, L.; Cui, K. Synthesis of hollow maghemite (γ -Fe₂O₃) particles for magnetic field and pH-responsive drug delivery and lung cancer treatment. *Ceram. Int.* **2021**, *47*, 7457–7464. [[CrossRef](#)]
68. Patil, P.; Parit, S.; Waifalkar, P.; Patil, S.; Dongale, T.; Sahoo, S.C.; Kollu, P.; Nimbalkar, M.; Chougale, A. pH triggered curcumin release and antioxidant activity of curcumin loaded γ -Fe₂O₃ magnetic nanoparticles. *Mater. Lett.* **2018**, *223*, 178–181. [[CrossRef](#)]
69. Li, D.Q.; Wang, S.Y.; Meng, Y.J.; Li, J.F.; Li, J. An injectable, self-healing hydrogel system from oxidized pectin/chitosan/ γ -Fe₂O₃. *Int. J. Biol. Macromol.* **2020**, *164*, 4566–4574. [[CrossRef](#)]
70. Li, Y.; Liu, Y.; Kim, E.; Song, Y.; Tsao, C.-Y.; Teng, Z.; Gao, T.; Mei, L.; Bentley, W.E.; Payne, G.F.; et al. Electrodeposition of a magnetic and redox-active chitosan film for capturing and sensing metabolic active bacteria. *Carbohydr. Polym.* **2018**, *195*, 505–514. [[CrossRef](#)]
71. Arai, R.; Li, M.; Toyoda, R.; Maeda, H.; Nishihara, H. Redox-active, luminescent coordination nanosheet capsules containing magnetite. *Sci. Rep.* **2020**, *10*, 13818. [[CrossRef](#)]
72. Akhtar, H.; Pourmadadi, M.; Yazdian, F.; Rashedi, H. Kosmotropic and chaotropic effect of biocompatible Fe₃O₄ nanoparticles on egg white lysozyme; the key role of nanoparticle-protein corona formation. *J. Mol. Struct.* **2022**, *1253*, 132016. [[CrossRef](#)]
73. Gong, T.; Yang, X.; Fang, J.-J.; Sui, Q.; Xi, F.-G.; Gao, E.-Q. Distinct Chromic and Magnetic Properties of Metal–Organic Frameworks with a Redox Ligand. *ACS Appl. Mater. Interfaces* **2017**, *9*, 5503–5512. [[CrossRef](#)] [[PubMed](#)]

74. Mousavi, S.D.; Maghsoodi, F.; Panahandeh, F.; Yazdian-Robati, R.; Reisi-Vanani, A.; Tafaghodi, M. Doxorubicin delivery via magnetic nanomicelles comprising from reduction-responsive poly (ethylene glycol)-b-poly (ϵ -caprolactone)(PEG-SS-PCL) and loaded with superparamagnetic iron oxide (SPIO) nanoparticles: Preparation, characterization and simulation. *Mater. Sci. Eng. C* **2018**, *92*, 631–643. [[CrossRef](#)] [[PubMed](#)]
75. Ren, S.; Yang, J.; Ma, L.; Li, X.; Wu, W.; Liu, C.; He, J.; Miao, L. Ternary-Responsive Drug Delivery with Activatable Dual Mode Contrast-Enhanced in Vivo Imaging. *ACS Appl. Mater. Interfaces* **2018**, *10*, 31947–31958. [[CrossRef](#)]
76. Stephen, Z.R.; Kievit, F.M.; Veisoh, O.; Chiarelli, P.A.; Fang, C.; Wang, K.; Hatzinger, S.J.; Ellenbogen, R.G.; Silber, J.R.; Zhang, M. Redox-Responsive Magnetic Nanoparticle for Targeted Convection-Enhanced Delivery of O⁶-Benzylguanine to Brain Tumors. *ACS Nano* **2014**, *8*, 10383–10395. [[CrossRef](#)]
77. Irshad, S.; Siddiqui, B.; ur-Rehman, A.; Farooq, R.K.; Ahmed, N. Recent trends and development in targeted delivery of therapeutics through enzyme responsive intelligent nanoplatfrom. *Int. J. Polym. Mater. Polym. Biomater.* **2020**, *71*, 403–413. [[CrossRef](#)]
78. Ansari, C.; Tikhomirov, G.; Hong, S.H.; Falconer, R.; Loadman, P.; Gill, J.; Castaneda, R.; Hazard, F.K.; Tong, L.; Lenkov, O.D.; et al. Development of Novel Tumor-Targeted Theranostic Nanoparticles Activated by Membrane-Type Matrix Metalloproteinases for Combined Cancer Magnetic Resonance Imaging and Therapy. *Small* **2014**, *10*, 566–575. [[CrossRef](#)] [[PubMed](#)]
79. Yang, Y.; Aw, J.; Chen, K.; Liu, F.; Padmanabhan, P.; Hou, Y.; Cheng, Z.; Xing, B. Enzyme-Responsive Multifunctional Magnetic Nanoparticles for Tumor Intracellular Drug Delivery and Imaging. *Chem.-Asian J.* **2011**, *6*, 1381–1389. [[CrossRef](#)] [[PubMed](#)]
80. Estelrich, J.; Escribano, E.; Queralt, J.; Busquets, M.A. Iron Oxide Nanoparticles for Magnetically-Guided and Magnetically-Responsive Drug Delivery. *Int. J. Mol. Sci.* **2015**, *16*, 8070–8101. [[CrossRef](#)]
81. Issa, B.; Obaidat, I.M.; Albiss, B.A.; Haik, Y. Magnetic Nanoparticles: Surface Effects and Properties Related to Biomedicine Applications. *Int. J. Mol. Sci.* **2013**, *14*, 21266–21305. [[CrossRef](#)]
82. Wagstaff, A.J.; Brown, S.D.; Holden, M.R.; Craig, G.E.; Plumb, J.A.; Brown, R.E.; Schreiter, N.; Chrzanowski, W.; Wheate, N.J. Cisplatin drug delivery using gold-coated iron oxide nanoparticles for enhanced tumour targeting with external magnetic fields. *Inorganica Chim. Acta* **2012**, *393*, 328–333. [[CrossRef](#)]
83. Lyon, J.L.; Fleming, D.A.; Stone, M.B.; Schiffer, A.P.; Williams, M.E. Synthesis of Fe Oxide Core/Au Shell Nanoparticles by Iterative Hydroxylamine Seeding. *Nano Lett.* **2004**, *4*, 719–723. [[CrossRef](#)]
84. Brown, S.D.; Nativo, P.; Smith, J.-A.; Stirling, D.; Edwards, P.R.; Venugopal, B.; Flint, D.J.; Plumb, J.A.; Graham, D.; Wheate, N.J. Gold Nanoparticles for the Improved Anticancer Drug Delivery of the Active Component of Oxaliplatin. *J. Am. Chem. Soc.* **2010**, *132*, 4678–4684. [[CrossRef](#)] [[PubMed](#)]
85. Craig, G.E.; Brown, S.D.; Lamprou, D.A.; Graham, D.; Wheate, N.J. Cisplatin-tethered gold nanoparticles that exhibit enhanced reproducibility, drug loading, and stability: A step closer to pharmaceutical approval? *Inorg. Chem.* **2012**, *51*, 3490–3497. [[CrossRef](#)] [[PubMed](#)]
86. Unterweger, H.; Tietze, R.; Janko, C.; Zaloga, J.; Lyer, S.; Taccardi, N.; Goudouri, M.; Hoppe, A.; Eberbeck, D.; Schubert, D.; et al. Development and characterization of magnetic iron oxide nanoparticles with a cisplatin-bearing polymer coating for targeted drug delivery. *Int. J. Nanomed.* **2014**, *9*, 3659–3676. [[CrossRef](#)] [[PubMed](#)]
87. Mattheolabakis, G.; Milane, L.; Singh, A.; Amiji, M.M. Hyaluronic acid targeting of CD44 for cancer therapy: From receptor biology to nanomedicine. *J. Drug Target.* **2015**, *23*, 605–618. [[CrossRef](#)]
88. Jeong, Y.I.; Kim, S.T.; Jin, S.G.; Ryu, H.H.; Jin, Y.H.; Jung, T.Y.; Kim, I.Y.; Jung, S. Cisplatin-incorporated hyaluronic acid nanoparticles based on ion-complex formation. *J. Pharm. Sci.* **2008**, *97*, 1268–1276. [[CrossRef](#)]
89. Sugahara, K.N.; Hirata, T.; Hayasaka, H.; Stern, R.; Murai, T.; Miyasaka, M. Tumor Cells Enhance Their Own CD44 Cleavage and Motility by Generating Hyaluronan Fragments. *J. Biol. Chem.* **2006**, *281*, 5861–5868. [[CrossRef](#)]
90. Nadeem, M.; Ahmad, M.; Akhtar, M.S.; Shaari, A.; Riaz, S.; Naseem, S.; Masood, M.; Saeed, M.A. Magnetic Properties of Polyvinyl Alcohol and Doxorubicine Loaded Iron Oxide Nanoparticles for Anticancer Drug Delivery Applications. *PLoS ONE* **2016**, *11*, e0158084. [[CrossRef](#)]
91. Zaloga, J.; Janko, C.; Nowak, J.; Matuszak, J.; Knaup, S.; Eberbeck, D.; Tietze, R.; Unterweger, H.; Friedrich, R.P.; Heimke-Brinck, R.; et al. Development of a lauric acid/albumin hybrid iron oxide nanoparticle system with improved biocompatibility. *Int. J. Nanomed.* **2014**, *9*, 4847–4866. [[CrossRef](#)]
92. Zaloga, J.; Pöttler, M.; Leitinger, G.; Friedrich, R.P.; Almer, G.; Lyer, S.; Baum, E.; Tietze, R.; Heimke-Brinck, R.; Mangege, H.; et al. Pharmaceutical formulation of HSA hybrid coated iron oxide nanoparticles for magnetic drug targeting. *Eur. J. Pharm. Biopharm.* **2016**, *101*, 152–162. [[CrossRef](#)]
93. Jeon, H.; Kim, J.; Lee, Y.M.; Kim, J.; Choi, H.W.; Lee, J.; Park, H.; Kang, Y.; Kim, I.-S.; Lee, B.-H.; et al. Poly-paclitaxel/cyclodextrin-SPIO nano-assembly for magnetically guided drug delivery system. *J. Control. Release* **2016**, *231*, 68–76. [[CrossRef](#)] [[PubMed](#)]
94. Chicheł, A.; Skowronek, J.; Kubaszewska, M.; Kanikowski, M. Hyperthermia—description of a method and a review of clinical applications. *Rep. Pract. Oncol. Radiother.* **2007**, *12*, 267–275. [[CrossRef](#)]
95. Yang, J.; Park, S.-B.; Yoon, H.-G.; Huh, Y.-M.; Haam, S. Preparation of poly ϵ -caprolactone nanoparticles containing magnetite for magnetic drug carrier. *Int. J. Pharm.* **2006**, *324*, 185–190. [[CrossRef](#)] [[PubMed](#)]
96. Islam, M.S. Development and Evaluation of Magnetic, Photocatalytic and Photothermal Nanoparticles and their Application to Cancer Therapy. Ph.D. Thesis, Kagoshima University, Kagoshima, Japan, 2012.

97. Laurent, S.; Forge, D.; Port, M.; Roch, A.; Robic, C.; Vander Elst, L.; Muller, R.N. Magnetic Iron Oxide Nanoparticles: Synthesis, Stabilization, Vectorization, Physicochemical Characterizations, and Biological Applications. *Chem. Rev.* **2008**, *108*, 2064–2110. [[CrossRef](#)] [[PubMed](#)]
98. Brazel, C.S. Magnetothermally-responsive Nanomaterials: Combining Magnetic Nanostructures and Thermally-Sensitive Polymers for Triggered Drug Release. *Pharm. Res.* **2009**, *26*, 644–656. [[CrossRef](#)]
99. Mitra, S.; Maitra, A. Inorganic Nanoparticles for Therapeutics, Drug and Gene Delivery. CENTERA. In *Advances in Nanotechnology and Applications*; C.E.N.T.E.R.A.: Louisville, KY, USA, 2009.
100. Nagel, S. *Theoretische und Experimentelle Untersuchungen zum Magnetischen Drug Targeting*; Ernst-Moritz-Arndt-Universität: Greifswald, Germany, 2004.
101. Batista, S.O.S.; Morales, M.A.; dos Santos, W.C.; Iglesias, C.A.; Baggio-Saitovitch, E.; Carriço, A.S.; Bohn, F.; de Medeiros, S.N. Mechano-synthesis, structural and magnetic characterization, and heat release of α -Fe nanoparticles embedded in a wüstite matrix. *J. Magn. Magn. Mater.* **2015**, *391*, 83–88. [[CrossRef](#)]
102. Araújo-Neto, R.P.; Silva-Freitas, E.L.; Carvalho, J.F.; Pontes, T.R.F.; Silva, K.L.; Damasceno, I.H.M.; Egito, E.S.T.; Dantas, A.L.; Morales, M.A.; Carriço, A.S. Monodisperse sodium oleate coated magnetite high susceptibility nanoparticles for hyperthermia applications. *J. Magn. Magn. Mater.* **2014**, *364*, 72–79. [[CrossRef](#)]
103. Alomari, M.; Almohazey, D.; Almoftay, S.; Alhibshi, A.; Almansour, I.; Kaewsaneha, C.; Badri, W.; Fessi, H.; Elaissari, A. Magnetic-responsive polysaccharide-inorganic composite materials for cancer therapeutics. In *Polysaccharide Carriers for Drug Delivery*; Elsevier: Amsterdam, The Netherlands, 2019; pp. 179–216. [[CrossRef](#)]
104. Taghizadeh, S.-M.; Berenjian, A.; Zare, M.; Ebrahiminezhad, A. New Perspectives on Iron-Based Nanostructures. *Processes* **2020**, *8*, 1128. [[CrossRef](#)]
105. Viswanath, B.; Kim, S.; Lee, K. Recent insights into nanotechnology development for detection and treatment of colorectal cancer. *Int. J. Nanomed.* **2016**, *11*, 2491–2504. [[CrossRef](#)]
106. Rosengart, A.J.; Kaminski, M.D.; Chen, H.; Caviness, P.L.; Ebner, A.D.; Ritter, J.A. Magnetizable implants and functionalized magnetic carriers: A novel approach for noninvasive yet targeted drug delivery. *J. Magn. Magn. Mater.* **2005**, *293*, 633–638. [[CrossRef](#)]
107. Rahban, D.; Doostan, M.; Salimi, A. Cancer Therapy; Prospects for Application of Nanoparticles for Magnetic-Based Hyperthermia. *Cancer Investig.* **2020**, *38*, 507–521. [[CrossRef](#)] [[PubMed](#)]
108. Magro, M.; Vianello, F. Bare Iron Oxide Nanoparticles: Surface Tunability for Biomedical, Sensing and Environmental Applications. *Nanomaterials* **2019**, *9*, 1608. [[CrossRef](#)] [[PubMed](#)]
109. Yan, S.; Zhang, D.; Gu, N.; Zheng, J.; Ding, A.; Wang, Z.; Xing, B.; Ma, M.; Zhang, Y. Therapeutic Effect of Fe₂O₃ Nanoparticles Combined with Magnetic Fluid Hyperthermia on Cultured Liver Cancer Cells and Xenograft Liver Cancers. *J. Nanosci. Nanotechnol.* **2005**, *5*, 1185–1192. [[CrossRef](#)] [[PubMed](#)]
110. Ramos-Guivar, J.A.; Morales, M.A.; Litterst, F.J. γ -Fe₂O₃ nanoparticles embedded in nanohydroxyapatite matrix for magnetic hyperthermia and in vitro osteoblast cell studies. *Ceram. Int.* **2020**, *46*, 10658–10666. [[CrossRef](#)]
111. Luo, D.; Saltzman, W.M. Enhancement of transfection by physical concentration of DNA at the cell surface. *Nat. Biotechnol.* **2000**, *18*, 893–895. [[CrossRef](#)] [[PubMed](#)]
112. Curiel, D.T. Strategies to Adapt Adenoviral Vectors for Targeted Delivery. *Ann. N. Y. Acad. Sci.* **1999**, *886*, 158–171. [[CrossRef](#)]
113. Scherer, F.; Anton, M.; Schillinger, U.; Henke, J.; Bergemann, C.; Krüger, A.; Gänsbacher, B.; Plank, C. Magnetofection: Enhancing and targeting gene delivery by magnetic force in vitro and in vivo. *Gene Ther.* **2002**, *9*, 102–109. [[CrossRef](#)]
114. Lorente, C.; Cabeza, L.; Clares, B.; Ortiz, R.; Halbaut, L.; Delgado, V.; Perazzoli, G.; Prados, J.; Arias, J.L.; Melguizo, C. Formulation and in vitro evaluation of magnetoliposomes as a potential nanotool in colorectal cancer therapy. *Colloids Surf. B Biointerfaces* **2018**, *171*, 553–565. [[CrossRef](#)]
115. Tadic, M.; Trpkov, D.; Kopanja, L.; Vojnovic, S.; Panjan, M. Hydrothermal synthesis of hematite (α -Fe₂O₃) nanoparticle forms: Synthesis conditions, structure, particle shape analysis, cytotoxicity and magnetic properties. *J. Alloys Compd.* **2019**, *792*, 599–609. [[CrossRef](#)]
116. Wei, W.; Ding, Y.; Zhao, A.; Ge, K.; Zhang, C.; Li, Y.; Zhang, J.; Jia, G. Monodisperse and mesoporous walnut kernel-like SiO₂/ γ -Fe₂O₃ nanocomposite: Synthesis, magnetic properties, and application in drug delivery. *J. Alloys Compd.* **2017**, *728*, 585–591. [[CrossRef](#)]
117. Cao, S.W.; Zhu, Y.J.; Ma, M.Y.; Li, L.; Zhang, L. Hierarchically nanostructured magnetic hollow spheres of Fe₃O₄ and γ -Fe₂O₃: Preparation and potential application in drug delivery. *J. Phys. Chem. C* **2008**, *112*, 1851–1856. [[CrossRef](#)]
118. Kumar, A.; Sahoo, B.; Montpetit, A.; Behera, S.; Lockey, R.F.; Mohapatra, S.S. Development of hyaluronic acid-Fe₂O₃ hybrid magnetic nanoparticles for targeted delivery of peptides. *Nanomed. Nanotechnol. Biol. Med.* **2007**, *3*, 132–137. [[CrossRef](#)]
119. Arruebo, M.; Fernández-Pacheco, R.; Ibarra, M.R.; Santamaría, J. Magnetic nanoparticles for drug delivery. *Nano Today* **2007**, *2*, 22–32. [[CrossRef](#)]
120. Absolom, D.R.; Zingg, W.; Neumann, A.W. Protein adsorption to polymer particles: Role of surface properties. *J. Biomed. Mater. Res.* **1987**, *21*, 161–171. [[CrossRef](#)]
121. Haas, T.A.; Plow, E.F. Integrin-ligand interactions: A year in review. *Curr. Opin. Cell Biol.* **1994**, *6*, 656–662. [[CrossRef](#)]

122. Blyakhman, F.; Safronov, A.; Makarova, E.; Fadeyev, F.; Shklyar, T.; Shabadrov, P.; Armas, S.; Kurlyandskaya, G. Magnetic Properties of Iron Oxide Nanoparticles Do Not Essentially Contribute to Ferrogel Biocompatibility. *Nanomaterials* **2021**, *11*, 1041. [[CrossRef](#)] [[PubMed](#)]
123. Sabale, S.; Kandesar, P.; Jadhav, V.; Komorek, R.; Motkuri, R.K.; Yu, X.-Y. Recent developments in the synthesis, properties, and biomedical applications of core/shell superparamagnetic iron oxide nanoparticles with gold. *Biomater. Sci.* **2017**, *5*, 2212–2225. [[CrossRef](#)]
124. Piñeiro, Y.; Vargas, Z.; Rivas, J.; López-Quintela, M.A. Iron Oxide Based Nanoparticles for Magnetic Hyperthermia Strategies in Biological Applications. *Eur. J. Inorg. Chem.* **2015**, *2015*, 4495–4509. [[CrossRef](#)]
125. Ansari, M.O.; Ahmad, F.; Shadab, G.; Siddique, H.R. Superparamagnetic iron oxide nanoparticles based cancer theranostics: A double edge sword to fight against cancer. *J. Drug Deliv. Sci. Technol.* **2018**, *45*, 177–183. [[CrossRef](#)]
126. Park, K.; Liang, G.; Ji, X.; Luo, Z.P.; Li, C.; Croft, M.C.; Markert, J.T. Structural and magnetic properties of gold and silica doubly coated γ -Fe₂O₃ nanoparticles. *J. Phys. Chem. C* **2007**, *111*, 18512–18519. [[CrossRef](#)]
127. Dumitrache, F.; Morjan, I.; Fleaca, C.; Badoi, A.; Manda, G.; Pop, S.; Marta, D.; Humnic, G.; Vekas, L.; Daia, C.; et al. Highly magnetic Fe₂O₃ nanoparticles synthesized by laser pyrolysis used for biological and heat transfer applications. *Appl. Surf. Sci.* **2015**, *336*, 297–303. [[CrossRef](#)]
128. Li, P.; He, Z.; Luo, C.; Xiao, Y.; Wang, Y.; Hu, J.; Li, G.; Jiang, H.; Zhang, W. α -Fe₂O₃@dopamine core-shell nanocomposites and their highly enhanced photoacoustic performance. *Appl. Surf. Sci.* **2018**, *466*, 185–192. [[CrossRef](#)]
129. Gupta, A.K.; Curtis, A.S.G. Surface modified superparamagnetic nanoparticles for drug delivery: Interaction studies with human fibroblasts in culture. *J. Mater. Sci. Mater. Med.* **2004**, *15*, 493–496. [[CrossRef](#)] [[PubMed](#)]
130. Berry, C.C.; Wells, S.; Charles, S.; Aitchison, G.; Curtis, A.S. Cell response to dextran-derivatised iron oxide nanoparticles post internalisation. *Biomaterials* **2004**, *25*, 5405–5413. [[CrossRef](#)] [[PubMed](#)]
131. Berry, C.C.; Wells, S.; Charles, S.; Curtis, A.S. Dextran and albumin derivatised iron oxide nanoparticles: Influence on fibroblasts in vitro. *Biomaterials* **2003**, *24*, 4551–4557. [[CrossRef](#)]
132. Simioni, A.R.; Martins, O.P.; Lacava, Z.G.M.; Azevedo, R.B.; Lima, E.C.D.; Lacava, B.M.; Morais, P.C.; Tedesco, A. Cell toxicity studies of albumin-based nanosized magnetic beads. *J. Nanosci. Nanotechnol.* **2006**, *6*, 2413–2415. [[CrossRef](#)]
133. Karlsson, H.L.; Gustafsson, J.; Cronholm, P.; Möller, L. Size-dependent toxicity of metal oxide particles—A comparison between nano- and micrometer size. *Toxicol. Lett.* **2009**, *188*, 112–118. [[CrossRef](#)]
134. Mahmoudi, M.; Simchi, A.; Imani, M.; Shokrgozar, M.A.; Milani, A.S.; Häfeli, U.O.; Stroeve, P. A new approach for the in vitro identification of the cytotoxicity of superparamagnetic iron oxide nanoparticles. *Colloids Surf. B Biointerfaces* **2010**, *75*, 300–309. [[CrossRef](#)]
135. Kladko, D.; Falchevskaya, A.; Serov, N.; Prilepskii, A. Nanomaterial Shape Influence on Cell Behavior. *Int. J. Mol. Sci.* **2021**, *22*, 5266. [[CrossRef](#)]
136. Billotey, C.; Wilhelm, C.; Devaud, M.; Bacri, J.C.; Bittoun, J.; Gazeau, F. Cell internalization of anionic maghemite nanoparticles: Quantitative effect on magnetic resonance imaging. *Magn. Reson. Med. Off. J. Int. Soc. Magn. Reson. Med.* **2003**, *49*, 646–654. [[CrossRef](#)]
137. Wilhelm, C.; Billotey, C.; Roger, J.; Pons, J.; Bacri, J.-C.; Gazeau, F. Intracellular uptake of anionic superparamagnetic nanoparticles as a function of their surface coating. *Biomaterials* **2003**, *24*, 1001–1011. [[CrossRef](#)]
138. Pisanic, T.R.; Blackwell, J.D.; Shubayev, V.I.; Fiñones, R.R.; Jin, S. Nanotoxicity of iron oxide nanoparticle internalization in growing neurons. *Biomaterials* **2007**, *28*, 2572–2581. [[CrossRef](#)] [[PubMed](#)]
139. Wen, T.; Du, L.; Chen, B.; Yan, D.; Yang, A.; Liu, J.; Gu, N.; Meng, J.; Xu, H. Iron oxide nanoparticles induce reversible endothelial-to-mesenchymal transition in vascular endothelial cells at acutely non-cytotoxic concentrations. *Part. Fibre Toxicol.* **2019**, *16*, 30. [[CrossRef](#)] [[PubMed](#)]
140. De Freitas, E.R.L.; Soares, P.R.O.; de Paula Santos, R.; dos Santos, R.L.; da Silva, J.R.; Porfirio, E.P.; Bão, S.N.; de Oliveira Lima, E.C.; Morais, P.C.; Guillo, L.A. In vitro biological activities of anionic γ -Fe₂O₃ nanoparticles on human melanoma cells. *J. Nanosci. Nanotechnol.* **2008**, *8*, 2385–2391. [[CrossRef](#)] [[PubMed](#)]
141. Maassen, S.; Fattal, E.; Müller, R.H.; Couvreur, P. Cell cultures for the assessment of toxicity and uptake of polymeric particulate drug carriers. *STP Pharma Sci.* **1993**, *3*, 11–22.
142. Weissleder, R.; Stark, D.D.; Engelstad, B.L.; Bacon, B.R.; Compton, C.C.; White, D.L.; Jacobs, P.; Lewis, J. Superparamagnetic iron oxide: Pharmacokinetics and toxicity. *Am. J. Roentgenol.* **1989**, *152*, 167–173. [[CrossRef](#)]
143. Klausner, R.D.; Rouault, T.A.; Harford, J.B. Regulating the fate of mRNA: The control of cellular iron metabolism. *Cell* **1993**, *72*, 19–28. [[CrossRef](#)]
144. Schulze, E.; Ferrucci, J.T.; Poss, K.; Lapointe, L.; Bogdanova, A.; Weissleder, R. Cellular Uptake and Trafficking of a Prototypical Magnetic Iron Oxide Label In Vitro. *Investig. Radiol.* **1995**, *30*, 604–610. [[CrossRef](#)]
145. Elias, A.; Tsourkas, A. Imaging circulating cells and lymphoid tissues with iron oxide nanoparticles. *Hematology* **2009**, *2009*, 720–726. [[CrossRef](#)]
146. Glickstein, H.; Ben El, R.; Link, G.; Breuer, W.; Konijn, A.M.; Hershko, C.; Nick, H.; Cabantchik, Z.I.; El, R.B. Action of chelators in iron-loaded cardiac cells: Accessibility to intracellular labile iron and functional consequences. *Blood* **2006**, *108*, 3195–3203. [[CrossRef](#)]

147. Jain, T.K.; Reddy, M.K.; Morales, M.A.; Leslie-Pelecky, D.L.; Labhassetwar, V. Biodistribution, Clearance, and Biocompatibility of Iron Oxide Magnetic Nanoparticles in Rats. *Mol. Pharm.* **2008**, *5*, 316–327. [[CrossRef](#)] [[PubMed](#)]
148. Bourrinet, P.; Bengel, H.H.; Bonnemain, B.; Dencausse, A.; Idee, J.-M.; Jacobs, P.M.; Lewis, J.M. Preclinical Safety and Pharmacokinetic Profile of Ferumoxtran-10, an Ultrasmall Superparamagnetic Iron Oxide Magnetic Resonance Contrast Agent. *Investig. Radiol.* **2006**, *41*, 313–324. [[CrossRef](#)] [[PubMed](#)]
149. Gajdosíková, A.; Gajdosik, A.; Koneracká, M.; Závistová, V.; Stvrtina, S.; Krchnárová, V.; Kopcanský, P.; Tomasovicová, N.; Stolc, S.; Timko, M. Acute toxicity of magnetic nanoparticles in mice. *Neuro Endocrinol. Lett.* **2006**, *27*, 96–99. [[PubMed](#)]
150. Lübbe, A.S.; Bergemann, C.; Huhnt, W.; Fricke, T.; Riess, H.; Brock, J.W.; Huhn, D. Preclinical experiences with magnetic drug targeting: Tolerance and efficacy. *Cancer Res.* **1996**, *56*, 4694–4701.
151. Lübbe, A.S.; Bergemann, C.; Riess, H.; Schriever, F.; Reichardt, P.; Possinger, K.; Matthias, M.; Dörken, B.; Herrmann, F.; Gürtler, R.; et al. Clinical experiences with magnetic drug targeting: A phase I study with 4'-epidoxorubicin in 14 patients with advanced solid tumors. *Cancer Res.* **1996**, *56*, 4686–4693.
152. Lemke, A.-J.; Von Pilsach, M.-I.S.; Bergemann, C.; Riess, H.; Felix, R. MRI after magnetic drug targeting in patients with advanced solid malignant tumors. *Eur. Radiol.* **2004**, *14*, 1949–1955. [[CrossRef](#)]
153. Reimer, P.; Balzer, T. Ferucarbotran (Resovist): A new clinically approved RES-specific contrast agent for contrast-enhanced MRI of the liver: Properties, clinical development, and applications. *Eur. Radiol.* **2003**, *13*, 1266–1276. [[CrossRef](#)]
154. Reimer, P.; Marx, C.; Rummeny, E.J.; Müller, M.; Lentschig, M.; Balzer, T.; Diel, K.-H.; Sulkowski, U.; Berns, T.; Shamsi, K.; et al. SPIO-enhanced 2D-TOF MR angiography of the portal venous system: Results of an intraindividual comparison. *J. Magn. Reson. Imaging* **1997**, *7*, 945–949. [[CrossRef](#)]
155. Kopp, A.F.; Laniado, M.; Dammann, F.; Stern, W.; Grönwäller, E.; Balzer, T.; Schimpfky, C.; Claussen, C.D. MR imaging of the liver with Resovist: Safety, efficacy, and pharmacodynamic properties. *Radiology* **1997**, *204*, 749–756. [[CrossRef](#)]
156. Kehagias, D.T.; Gouliamos, A.D.; Smyrniotis, V.; Vlahos, L.J. Diagnostic efficacy and safety of MRI of the liver with superparamagnetic iron oxide particles (SH U 555 A). *J. Magn. Reson. Imaging Off. J. Int. Soc. Magn. Reson. Med.* **2001**, *14*, 595–601. [[CrossRef](#)]
157. Friedrich, R.P.; Cicha, I.; Alexiou, C. Iron Oxide Nanoparticles in Regenerative Medicine and Tissue Engineering. *Nanomaterials* **2021**, *11*, 2337. [[CrossRef](#)] [[PubMed](#)]
158. Muldoon, L.L.; Sándor, M.; Pinkston, K.E.; Neuwelt, E.A. Imaging, distribution, and toxicity of superparamagnetic iron oxide magnetic resonance nanoparticles in the rat brain and intracerebral tumor. *Neurosurgery* **2005**, *57*, 785–796. [[CrossRef](#)] [[PubMed](#)]
159. Weissleder, R.; Bogdanov, A.; Neuwelt, E.A.; Papisov, M. Long-circulating iron oxides for MR imaging. *Adv. Drug Deliv. Rev.* **1995**, *16*, 321–334. [[CrossRef](#)]
160. Aillon, K.L.; Xie, Y.; El-Gendy, N.; Berkland, C.J.; Forrest, M.L. Effects of nanomaterial physicochemical properties on in vivo toxicity. *Adv. Drug Deliv. Rev.* **2009**, *61*, 457–466. [[CrossRef](#)] [[PubMed](#)]
161. Alexis, F.; Pridgen, E.; Molnar, L.K.; Farokhzad, O.C. Factors Affecting the Clearance and Biodistribution of Polymeric Nanoparticles. *Mol. Pharm.* **2008**, *5*, 505–515. [[CrossRef](#)] [[PubMed](#)]
162. O'Connor, C.; Brady, E.; Zheng, Y.; Moore, E.; Stevens, K.R. Engineering the multiscale complexity of vascular networks. *Nat. Rev. Mater.* **2022**, *7*, 702–716. [[CrossRef](#)] [[PubMed](#)]
163. Koons, G.L.; Diba, M.; Mikos, A.G. Materials design for bone-tissue engineering. *Nat. Rev. Mater.* **2020**, *5*, 584–603. [[CrossRef](#)]
164. Wang, J.; Tian, L.; He, L.; Chen, N.; Ramakrishna, S.; So, K.-F.; Mo, X. Lycium barbarum polysaccharide encapsulated Poly lactic-co-glycolic acid Nanofibers: Cost effective herbal medicine for potential application in peripheral nerve tissue engineering. *Sci. Rep.* **2018**, *8*, 8669. [[CrossRef](#)]
165. Giannelli, M.; Barbalinardo, M.; Riminucci, A.; Belvedere, K.; Boccalon, E.; Sotgiu, G.; Corticelli, F.; Ruani, G.; Zamboni, R.; Aluigi, A.; et al. Magnetic keratin/hydroxylapatite sponges as potential scaffolds for tissue regeneration. *Appl. Clay Sci.* **2021**, *207*, 106090. [[CrossRef](#)]
166. Goranov, V.; Shelyakova, T.; De Santis, R.; Haranava, Y.; Makhaniok, A.; Gloria, A.; Tampieri, A.; Russo, A.; Kon, E.; Marcacci, M.; et al. 3D Patterning of cells in Magnetic Scaffolds for Tissue Engineering. *Sci. Rep.* **2020**, *10*, 2289. [[CrossRef](#)]
167. García, R.S.; Stafford, S.; Gun'Ko, Y.K. Recent Progress in Synthesis and Functionalization of Multimodal Fluorescent-Magnetic Nanoparticles for Biological Applications. *Appl. Sci.* **2018**, *8*, 172. [[CrossRef](#)]
168. Santhosh, M.; Choi, J.-H.; Choi, J.-W. Magnetic-assisted cell alignment within a magnetic nanoparticle-decorated reduced graphene oxide/collagen 3D nanocomposite hydrogel. *Nanomaterials* **2019**, *9*, 1293. [[CrossRef](#)] [[PubMed](#)]
169. Crolet, J.; Aoubiza, B.; Meunier, A. Compact bone: Numerical simulation of mechanical characteristics. *J. Biomech.* **1993**, *26*, 677–687. [[CrossRef](#)]
170. Luo, Y.; Zhang, T.; Lin, X. 3D printed hydrogel scaffolds with macro pores and interconnected microchannel networks for tissue engineering vascularization. *Chem. Eng. J.* **2022**, *430*, 132926. [[CrossRef](#)]
171. Pei, B.; Wang, W.; Dunne, N.; Li, X. Applications of Carbon Nanotubes in Bone Tissue Regeneration and Engineering: Superiority, Concerns, Current Advancements, and Prospects. *Nanomaterials* **2019**, *9*, 1501. [[CrossRef](#)]
172. Xia, Y.; Sun, J.; Zhao, L.; Zhang, F.; Liang, X.-J.; Guo, Y.; Weir, M.D.; Reynolds, M.A.; Gu, N.; Xu, H.H.K. Magnetic field and nano-scaffolds with stem cells to enhance bone regeneration. *Biomaterials* **2018**, *183*, 151–170. [[CrossRef](#)]
173. Li, X.; Wei, J.; Aifantis, K.E.; Fan, Y.; Feng, Q.; Cui, F.Z.; Watari, F. Current investigations into magnetic nanoparticles for biomedical applications. *J. Biomed. Mater. Res. Part A* **2016**, *104*, 1285–1296. [[CrossRef](#)]

174. Saber-Samandari, S.; Saber-Samandari, S. Biocompatible nanocomposite scaffolds based on copolymer-grafted chitosan for bone tissue engineering with drug delivery capability. *Mater. Sci. Eng. C* **2017**, *75*, 721–732. [[CrossRef](#)]
175. Świątek, M.; Broż, A.; Tarasiuk, J.; Wroński, S.; Tokarz, W.; Kozieł, A.; Błażewicz, M.; Bačáková, L. Carbon nanotube/iron oxide hybrid particles and their PCL-based 3D composites for potential bone regeneration. *Mater. Sci. Eng. C* **2019**, *104*, 109913. [[CrossRef](#)]
176. Hu, S.; Chen, H.; Zhou, F.; Liu, J.; Qian, Y.; Hu, K.; Yan, J.; Gu, Z.; Guo, Z.; Zhang, F.; et al. Superparamagnetic core-shell electrospun scaffolds with sustained release of IONPs facilitating in vitro and in vivo bone regeneration. *J. Mater. Chem. B* **2021**, *9*, 8980–8993. [[CrossRef](#)]
177. Meng, J.; Xiao, B.; Zhang, Y.; Liu, J.; Xue, H.; Lei, J.; Kong, H.; Huang, Y.; Jin, Z.; Gu, N.; et al. Super-paramagnetic responsive nanofibrous scaffolds under static magnetic field enhance osteogenesis for bone repair in vivo. *Sci. Rep.* **2013**, *3*, 2655. [[CrossRef](#)] [[PubMed](#)]
178. Liao, J.; Han, R.; Wu, Y.; Qian, Z. Review of a new bone tumor therapy strategy based on bifunctional biomaterials. *Bone Res.* **2021**, *9*, 18. [[CrossRef](#)]
179. Montoya, C.; Du, Y.; Gianforcaro, A.L.; Orrego, S.; Yang, M.; Lelkes, P.I. On the road to smart biomaterials for bone research: Definitions, concepts, advances, and outlook. *Bone Res.* **2021**, *9*, 12. [[CrossRef](#)] [[PubMed](#)]
180. Yuan, Z.; Memarzadeh, K.; Stephen, A.S.; Allaker, R.P.; Brown, R.A.; Huang, J. Development of a 3D Collagen Model for the In Vitro Evaluation of Magnetic-assisted Osteogenesis. *Sci. Rep.* **2018**, *8*, 16270. [[CrossRef](#)] [[PubMed](#)]
181. Zhu, L.; Luo, D.; Liu, Y. Effect of the nano/microscale structure of biomaterial scaffolds on bone regeneration. *Int. J. Oral Sci.* **2020**, *12*, 6. [[CrossRef](#)] [[PubMed](#)]
182. Wang, X.; Law, J.; Luo, M.; Gong, Z.; Yu, J.; Tang, W.; Zhang, Z.; Mei, X.; Huang, Z.; You, L.; et al. Magnetic Measurement and Stimulation of Cellular and Intracellular Structures. *ACS Nano* **2020**, *14*, 3805–3821. [[CrossRef](#)]
183. Huo, M.; Wang, L.; Chen, Y.; Shi, J. Tumor-selective catalytic nanomedicine by nanocatalyst delivery. *Nat. Commun.* **2017**, *8*, 357. [[CrossRef](#)]
184. Abdal Dayem, A.; Lee, S.B.; Cho, S.-G. The Impact of Metallic Nanoparticles on Stem Cell Proliferation and Differentiation. *Nanomaterials* **2018**, *8*, 761. [[CrossRef](#)]
185. Wang, Q.; Chen, B.; Cao, M.; Sun, J.; Wu, H.; Zhao, P.; Xing, J.; Yang, Y.; Zhang, X.; Ji, M.; et al. Response of MAPK pathway to iron oxide nanoparticles in vitro treatment promotes osteogenic differentiation of hBMSCs. *Biomaterials* **2016**, *86*, 11–20. [[CrossRef](#)]
186. Ishmukhametov, I.; Batasheva, S.; Rozhina, E.; Akhatova, F.; Mingaleeva, R.; Rozhin, A.; Fakhruллин, R. DNA/Magnetic Nanoparticles Composite to Attenuate Glass Surface Nanotopography for Enhanced Mesenchymal Stem Cell Differentiation. *Polymers* **2022**, *14*, 344. [[CrossRef](#)]
187. Kim, J.H.; Liu, X.; Wang, J.; Chen, X.; Zhang, H.; Kim, S.H.; Cui, J.; Li, R.; Zhang, W.; Kong, Y.; et al. Wnt signaling in bone formation and its therapeutic potential for bone diseases. *Ther. Adv. Musculoskelet. Dis.* **2013**, *5*, 13–31. [[CrossRef](#)] [[PubMed](#)]
188. Zhang, X.; Huang, P.; Jiang, G.; Zhang, M.; Yu, F.; Dong, X.; Wang, L.; Chen, Y.; Zhang, W.; Qi, Y.; et al. A novel magnesium ion-incorporating dual-crosslinked hydrogel to improve bone scaffold-mediated osteogenesis and angiogenesis. *Mater. Sci. Eng. C* **2021**, *121*, 111868. [[CrossRef](#)] [[PubMed](#)]
189. Abdollahiyan, P.; Oroojalian, F.; Hejazi, M.; de la Guardia, M.; Mokhtarzadeh, A. Nanotechnology, and scaffold implantation for the effective repair of injured organs: An overview on hard tissue engineering. *J. Control. Release* **2021**, *333*, 391–417. [[CrossRef](#)]
190. Pan, Y.; Du, X.; Zhao, F.; Xu, B. Magnetic nanoparticles for the manipulation of proteins and cells. *Chem. Soc. Rev.* **2012**, *41*, 2912–2942. [[CrossRef](#)] [[PubMed](#)]
191. Wang, Q.; Chen, B.; Ma, F.; Lin, S.; Cao, M.; Li, Y.; Gu, N. Magnetic iron oxide nanoparticles accelerate osteogenic differentiation of mesenchymal stem cells via modulation of long noncoding RNA INZEB2. *Nano Res.* **2017**, *10*, 626–642. [[CrossRef](#)]
192. Henry, G.; Garner, W.L. Inflammatory mediators in wound healing. *Surg. Clin. N. Am.* **2003**, *83*, 483–507. [[CrossRef](#)]
193. Serhan, C.N.; Chiang, N. Novel endogenous small molecules as the checkpoint controllers in inflammation and resolution: Entrée for resolomics. *Rheum. Dis. Clin. N. Am.* **2004**, *30*, 69–95. [[CrossRef](#)]
194. Lawrence, W.T.; Diegelmann, R.F. Growth factors in wound healing. *Clin. Dermatol.* **1994**, *12*, 157–169. [[CrossRef](#)]
195. Witte, M.B.; Barbul, A. GENERAL PRINCIPLES OF WOUND HEALING. *Surg. Clin. N. Am.* **1997**, *77*, 509–528. [[CrossRef](#)]
196. Allen, D.B.; Maguire, J.J.; Mahdavian, M.; Wicke, C.; Marcocci, L.; Scheuenstuhl, H.; Chang, M.; Le, A.X.; Hopf, H.; Hunt, T.K. Wound Hypoxia and Acidosis Limit Neutrophil Bacterial Killing Mechanisms. *Arch. Surg.* **1997**, *132*, 991–996. [[CrossRef](#)]
197. Jimenez, P.A.; Rampy, M.A. Keratinocyte Growth Factor-2 Accelerates Wound Healing in Incisional Wounds. *J. Surg. Res.* **1999**, *81*, 238–242. [[CrossRef](#)]
198. Bankey, P.; Fiegel, V.; Singh, R.; Knighton, D.; Cerra, F. Hypoxia and Endotoxin Induce Macrophage-mediated Suppression of Fibroblast Proliferation. *J. Trauma Inj. Infect. Crit. Care* **1989**, *29*, 972–980. [[CrossRef](#)] [[PubMed](#)]
199. Messina, A.; Knight, K.R.; Dowsing, B.J.; Zhang, B.; Phan, L.H.; Hurley, J.V.; A Morrison, W.; Stewart, A. Localization of Inducible Nitric Oxide Synthase to Mast Cells During Ischemia/Reperfusion Injury of Skeletal Muscle. *Lab. Investig.* **2000**, *80*, 423–431. [[CrossRef](#)] [[PubMed](#)]
200. Robson, M.C. WOUND INFECTION: A Failure of Wound Healing Caused by an Imbalance of Bacteria. *Surg. Clin. N. Am.* **1997**, *77*, 637–650. [[CrossRef](#)]
201. Simman, R.; Alani, H.; Williams, F. Effect of Mitomycin C on Keloid Fibroblasts: An In Vitro Study. *Ann. Plast. Surg.* **2003**, *50*, 71–76. [[CrossRef](#)]

202. Steed, D.L. Debridement. *Am. J. Surg.* **2004**, *187*, S71–S74. [[CrossRef](#)]
203. Steinbrech, D.S.; Longaker, M.T.; Mehrara, B.J.; Saadeh, P.; Chin, G.S.; Gerrets, R.P.; Chau, D.C.; Rowe, N.M.; Gittes, G.K. Fibroblast Response to Hypoxia: The Relationship between Angiogenesis and Matrix Regulation. *J. Surg. Res.* **1999**, *84*, 127–133. [[CrossRef](#)]
204. Riou, J.-P.A.; Cohen, J.R.; Johnson, H. Factors influencing wound dehiscence. *Am. J. Surg.* **1992**, *163*, 324–330. [[CrossRef](#)]
205. Chang, C.-H.; Song, J.-Y.; Park, J.-H.; Seo, S.-W. The Efficacy of Magnetic Disks for the Treatment of Earlobe Hypertrophic Scar. *Ann. Plast. Surg.* **2005**, *54*, 566–569. [[CrossRef](#)]
206. Farrar, M.A.; Schreiber, R.D. The molecular cell biology of interferon-gamma and its receptor. *Annu. Rev. Immunol.* **1993**, *11*, 571–611. [[CrossRef](#)]
207. Lingen, M.W. Role of leukocytes and endothelial cells in the development of angiogenesis in inflammation and wound healing. *Arch. Pathol. Lab. Med.* **2001**, *125*, 67–71. [[CrossRef](#)] [[PubMed](#)]
208. Ashcroft, G.S.; Greenwell-Wild, T.; Horan, M.A.; Wahl, S.M.; Ferguson, M.W. Topical Estrogen Accelerates Cutaneous Wound Healing in Aged Humans Associated with an Altered Inflammatory Response. *Am. J. Pathol.* **1999**, *155*, 1137–1146. [[CrossRef](#)]
209. Dalgleish, A.G.; O’Byrne, K.J. Chronic immune activation and inflammation in the pathogenesis of aids and cancer. *Adv. Cancer Res.* **2002**, *84*, 231–276. [[CrossRef](#)]
210. Eftekhari, A.; Dizaj, S.M.; Sharifi, S.; Salatin, S.; Saadat, Y.R.; Vahed, S.Z.; Samiei, M.; Ardalan, M.; Rameshrad, M.; Ahmadian, E.; et al. The Use of Nanomaterials in Tissue Engineering for Cartilage Regeneration; Current Approaches and Future Perspectives. *Int. J. Mol. Sci.* **2020**, *21*, 536. [[CrossRef](#)] [[PubMed](#)]
211. Hou, R.; Zhang, G.; Du, G.; Zhan, D.; Cong, Y.; Cheng, Y.; Fu, J. Magnetic nanohydroxyapatite/PVA composite hydrogels for promoted osteoblast adhesion and proliferation. *Colloids Surf. B Biointerfaces* **2013**, *103*, 318–325. [[CrossRef](#)]
212. Hou, R.; Nie, L.; Du, G.; Xiong, X.; Fu, J. Natural polysaccharides promote chondrocyte adhesion and proliferation on magnetic nanoparticle/PVA composite hydrogels. *Colloids Surf. B Biointerfaces* **2015**, *132*, 146–154. [[CrossRef](#)]
213. Khalili, M.; Keshvari, H.; Imani, R.; Sohi, A.N.; Esmaeili, E.; Tajabadi, M. Study of osteogenic potential of electrospun PCL incorporated by dendrimerized superparamagnetic nanoparticles as a bone tissue engineering scaffold. *Polym. Adv. Technol.* **2021**, *33*, 782–794. [[CrossRef](#)]
214. Ngadiman, N.H.A.; Idris, A.; Irfan, M.; Kurniawan, D.; Yusof, N.M.; Nasiri, R. γ -Fe₂O₃ nanoparticles filled polyvinyl alcohol as potential biomaterial for tissue engineering scaffold. *J. Mech. Behav. Biomed. Mater.* **2015**, *49*, 90–104. [[CrossRef](#)]
215. Ngadiman, N.H.A.; Yusof, N.M.; Idris, A.; Kurniawan, D.; Fallahiarezoudar, E. Fabricating high mechanical strength γ -Fe₂O₃ nanoparticles filled poly(vinyl alcohol) nanofiber using electrospinning process potentially for tissue engineering scaffold. *J. Bioact. Compat. Polym.* **2016**, *32*, 411–428. [[CrossRef](#)]
216. Huang, J.; Liang, Y.; Jia, Z.; Chen, J.; Duan, L.; Liu, W.; Zhu, F.; Liang, Q.; Zhu, W.; You, W.; et al. Development of Magnetic Nanocomposite Hydrogel with Potential Cartilage Tissue Engineering. *ACS Omega* **2018**, *3*, 6182–6189. [[CrossRef](#)]
217. Desmoulière, A.; Geinoz, A.; Gabbiani, F. Transforming growth factor-beta 1 induces alpha-smooth muscle actin expression in granulation tissue myofibroblasts and in quiescent and growing cultured fibroblasts. *J. Cell Biol.* **1993**, *122*, 103–111. [[CrossRef](#)] [[PubMed](#)]
218. Diegelmann, R.F. Analysis of Collagen Synthesis. In *Wound Healing*; Humana Press: Totowa, NJ, USA, 2003; Volume 78, pp. 349–358. [[CrossRef](#)]
219. Ehrlich, H.P.; Krummel, T.M. Regulation of wound healing from a connective tissue perspective. *Wound Repair Regen.* **1996**, *4*, 203–210. [[CrossRef](#)] [[PubMed](#)]
220. Pierce, G.F.; Mustoe, T.A.; Altmann, B.W.; Deuel, T.F.; Thomason, A. Role of platelet-derived growth factor in wound healing. *J. Cell. Biochem.* **1991**, *45*, 319–326. [[CrossRef](#)] [[PubMed](#)]
221. Rubtsov, K.V.; Kondranova, A.M.; Lozhkomoiev, A.S.; Kazantsev, S.O.; Mamonova, E.V.; Gavrilova, L.O.; Sadovoy, M.A. Synthesis of nanoparticle-hydrogel composites based on crosslinked by Al ions poly(sodium acrylate), Fe₂O₃ and γ -AlOOH. *AIP Conf. Proc.* **2019**, *2167*, 020298. [[CrossRef](#)]