

Review

Carbon Nanotubes and Chronic Granulomatous Disease

Barbara P. Barna ¹, Marc A. Judson ² and Mary Jane Thomassen ^{1,*}

- Division of Pulmonary, Critical Care and Sleep Medicine, East Carolina University, Brody Medical Sciences Building, 600 Moye Blvd. Rm. 3E-149, Greenville, NC 27834, USA; E-Mail: barnab@ecu.edu
- Division of Pulmonary and Critical Care Medicine, Albany Medical College, MC-91, 47 New Scotland Avenue, Albany, NY 12208, USA; E-Mail: judsonm@mail.amc.edu
- * Author to whom correspondence should be addressed; E-Mail: thomassenm@ecu.edu; Tel.: +1-252-744-1117; Fax: +1-252-744-4887.

Received: 4 April 2014; in revised form: 15 May 2014 / Accepted: 12 June 2014 /

Published: 23 June 2014

Abstract: Use of nanomaterials in manufactured consumer products is a rapidly expanding industry and potential toxicities are just beginning to be explored. Combustion-generated multiwall carbon nanotubes (MWCNT) or nanoparticles are ubiquitous in non-manufacturing environments and detectable in vapors from diesel fuel, methane, propane, and natural gas. In experimental animal models, carbon nanotubes have been shown to induce granulomas or other inflammatory changes. Evidence suggesting potential involvement of carbon nanomaterials in human granulomatous disease, has been gathered from analyses of dusts generated in the World Trade Center disaster combined with epidemiological data showing a subsequent increase in granulomatous disease of first responders. In this review we will discuss evidence for similarities in the pathophysiology of carbon nanotube-induced pulmonary disease in experimental animals with that of the human granulomatous disease, sarcoidosis.

Keywords: carbon nanotubes; sarcoidosis; alveolar macrophages

1. Granulomatous Lung Disease

1.1. Background

Granulomatous diseases represent a world-wide health problem. Granuloma formation is a host response designed to sequester and destroy an injurious substance or infectious organism (reviewed in [1]).

The granuloma itself is a complex, multicellular structure generated by components of the innate immune system (reviewed in [2]). Granulomas may form in the lung in response to environmental stimuli of a broad nature, including intracellular pathogens (mycobacteria, fungi), chemically inert materials such as silica, beryllium, or talc, and organic antigens as in hypersensitivity pneumonitis. Initial formation of the granuloma involves release of a large repertoire of pro-inflammatory mediators designed to recruit neutrophils and other cells into the area. If neutrophils cannot destroy the substance, it is phagocytized by macrophages which release additional pro-inflammatory cytokines and chemokines. This cytokine storm recruits T cells, natural killer (NK) cells, and dendritic cells so that any digested antigenic components of the substance can be presented to activate an adaptive immune response [3]. Failure of macrophages to destroy the substance within the granuloma results in a chronic inflammatory pulmonary condition that can have significant morbidity. Chronic granulomatous disease can occur in the presence or absence of a detectible adaptive immune response.

1.2. Sarcoidosis: Characterization

Sarcoidosis is a multi-organ, inflammatory granulomatous disease thought to occur as a result of exposure to an unidentified, poorly soluble environmental antigen(s). The antigenic substance triggers a host response which is related to genetic/epigenetic susceptibility factors [4]. As discussed below, many potential putative sarcoidosis "antigens" have been identified, but none has been clearly confirmed as a direct cause of the disease. The diagnosis of sarcoidosis is usually established by clinical history, a biopsy revealing non-caseating granulomas with adjacent inflammatory cell infiltrates, and negative studies for active infection. Currently there are no definitive biomarkers available for therapeutic targets or for assistance in the clinical evaluation of sarcoidosis patient prognosis, responses to therapy, or disease management.

1.3. Sarcoidosis: Epidemiology and Potential Etiology

The prevalence and incidence of sarcoidosis varies by ethnicity and sex. In the United States, the adjusted annual incidence is roughly three times higher in African-Americans than in white Americans [5,6]. Sarcoidosis is more common in females than in males by a ratio of less than two to one [7]. The prevalence of sarcoidosis also appears to vary in different geographic locations [6,8].

Several studies have reported microbial antigens (e.g., mycobacterium, propionibacterium acnes) in sarcoidosis tissue or lung, and blood cell reactivity to antigen challenge *in vitro* [9–15]. However, some studies have found similar carriage rates of organisms suspected to cause sarcoidosis in control subjects without disease [16]. In contrast, other studies have not confirmed the presence of potential sarcoidosis pathogens in sarcoidosis subjects [17,18].

Multiple environmental risk factors have been linked to sarcoidosis, including dose-related exposure to wood-burning stoves, fireplaces, and firefighting [4,19–22]. The atmosphere of such environmental conditions might well harbor combustion-generated contaminants composed of carbon nanotubes. Related to these findings, the incidence of "sarcoidosis-like" pulmonary granulomatous disease was reported to be increased in New York City Fire Department workers who were involved in the September 2001 World Trade Center (WTC) disaster where inhalation exposure to combustion materials was "intense" [23,24]. More recent follow-ups of some 20,000 WTC responders have

detected sarcoidosis-like granulomatous disease at higher levels than expected in non-firefighting individuals as well [25]. Of interest was the observation that carbon nanotubes of various sizes and lengths were present in WTC dust samples and in lung tissues of affected exposed individuals [26]. Extensive analyses of dusts/aerosols generated by the WTC collapse resulted in four major categories of components: (1) particulate matter (calcium carbonate and silica) and fibers (asbestos, glass, gypsum); (2) organic pollutants including polycyclic aromatic hydrocarbons; (3) gases (carbon dioxide, hydrogen sulfide, diesel exhaust fumes, combustion byproducts), and (4) heavy metals [24,27]. Many of these components such as particulate matter, fibers, and combustion byproducts have been previously associated with cases of sarcoidosis-like disease [24]. Other materials known to produce granulomatous reactions such as beryllium, zirconium, or tungsten, were not detectable in WTC dusts [27]. Thus, while WTC dust exposure has been clearly linked to increased incidence of respiratory disease [28], the identity of WTC dust component(s) responsible for increased cases of sarcoidosis-like granulomatous disease remains unknown. Despite the WTC studies and other large epidemiologic and genetic studies, no definitive etiology for sarcoidosis has emerged [29–31].

2. Animal Models of Carbon Nanotube-Mediated Lung Disease

2.1. Effects of Carbon Nanotubes

While nanomaterials such as carbon nanotubes have great potential in fields such as drug delivery and disease imaging, standardized means to determine potential toxicities have not yet been established (reviewed in [32,33]). Carbon nanotubes represent an arrangement of C60 atoms in an elongated cylindrical structure [34]. Interestingly, the history of carbon nanotube discovery is a complex tale which may go back well over a century. As colorfully described by Monthioux and Kuznetsov [35] the first mention of such structures may have appeared in Russia, in a 1952 volume of the Journal of Physical Chemistry. Not until 1993, however, were preparations of single wall carbon nanotubes (SWCNT) first detailed by two different investigative groups [35]. SWCNT can be massed into multi-wall carbon nanotubes (MWCNT) [34]. Outside of manufacturing, combustion-generated MWCNT and other carbon nanoparticles are ubiquitous within the environment, and have been detected in vapors from diesel fuel, methane, propane, and natural gas (reviewed in [36]). Thus, the importance and impact of carbon particles in our environment might be underestimated as a causative agent and/or factor involved in the development of respiratory illnesses.

Granulomatous inflammation and fibrosis have been reported in rodent models as a response to intratracheal administration of SWCNT or MWCNT [37–39]. Several factors have been cited as contributors to these findings. Pulmonary granulomatous changes have been associated with large agglomerates or aggregates of carbon nanotubes [40–42]. A comparative study of lung tissues in inhalation versus instillation reported smaller particle size and diminished pulmonary inflammation with inhalation [43]. The authors suggested that differences might be due to the reduced size and aggregation of inhaled carbon nanotubes. However, investigations carried out to determine long-term effects of inhalation versus instillation have demonstrated similar results with both techniques [44,45]. In animals receiving dispersed carbon nanotubes, granulomatous structures have not been observed, although increased collagen disposition was detected indicative of a fibrotic response [41,46].

Carbon nanotubes have also been implicated in triggering inflammasome formation [47–49]. Inflammasomes are part of innate immunity and consist of large intracellular protein complexes that regulate proteolytic activation of the proinflammatory cytokines, IL-1β and IL-18 [50,51]. These structures can be activated by microbial triggers as well as non-microbial materials such as asbestos and silica [52]. Inflammasome activation is now recognized as a contributor to a number of chronic lung diseases such as idiopathic pulmonary fibrosis, chronic obstructive lung disease, and asthma [51].

The physical and chemical properties of carbon nanotubes strongly influence pulmonary toxicity. Metal contaminants in commercial carbon nanotube preparations have been implicated in nanotube toxicity [53]. Hamilton *et al.* [47] presented both *in vitro* and *in vivo* results suggesting that inflammatory responses increased with MWCNT diameter and length. This group also found that nickel contamination of nanotubes activated inflammasomes, exacerbated lung pathology and promoted release of pro-inflammatory cytokines from alveolar macrophages [47]. Other properties of carbon nanotubes associated with inflammasome formation and toxicity included double-walled [49] and long-needle-like structures [48]. In contrast, modification of carbon nanotubes with polymer coatings [54] or other biocompatible materials has been reported to reduce pulmonary toxicity (summarized in [55,56]).

Currently, there is ample evidence that carbon nanotubes provoke pulmonary innate immune and inflammatory responses [37–39,48,49], but not adaptive immune responses (reviewed in [57]). Results of MWCNT interaction with existing immune system adaptive responses, however, are controversial. Inhalation of MWCNT (0.3–5 mg/m³) did not cause significant pulmonary changes but did increase TGFβ levels in bronchoalveolar lavage fluid which when applied *in vitro* to splenocytes from untreated mice, suppressed T cell activity [58]. In another inhalation model, Staal *et al.* [59] observed decreased allergen-specific IgE responses in sensitized animals. In contrast, Ronzani *et al.* [60], reported that instilled MWCNT elevated Th2 responses in a house dust mite allergy model. Finally, Ryman-Rasmussen *et al.* [61] found no significant pulmonary changes after MWCNT inhalation (100 mg/m³) except in ovalbumin-sensitized animals challenged with antigen. MWCNT treatment then increased pulmonary fibrosis and levels of the fibroblast growth factor, PDGF-AA, but had no effects on TGFβ or IL-13. These studies suggest that complications in both innate and adaptive immune functions may occur after exposure to carbon nanotubes.

2.2. The MWCNT-Induced Chronic Granuloma Model

Our major goal has been to improve understanding of sarcoidosis pathophysiology. The relatively recent reports of carbon nanotubes in environments favoring sarcoidosis incidence has been the impetus for us to develop a model of MWCNT-mediated lung disease that might have applicability to sarcoidosis. To our knowledge, no other MWCNT models have been used to study sarcoidosis. This model has proven to be extremely interesting with many pathologic changes identical to those found in sarcoidosis lung. Because most granulomatous diseases are chronic, and inciting factors may be unknown, there are no ideal animal models with which to investigate mechanisms of persistent granulomatous inflammation. Our first report described a model in which instilled MWCNT led to granulomatous lung inflammation persisting up to 90 days [62]. The long-term chronicity of this carbon nanotube granuloma model is relevant to the exploration of mechanisms in human chronic

granulomatous disease. Previously, major findings related to granulomatous inflammation were derived from studies using an acute murine granuloma model in which antigen-bound sepharose beads were injected intravenously into pre-sensitized animals [63,64]. However, such models are limited in that granulomatous inflammation forms and resolves within 14–20 days. Also of consideration was the delivery route, a tail vein injection of antigen bound-sepharose beads that lodged in the pulmonary circulation rather than directly in the alveolar space.

Dosage of reagent is problematic in all animal disease models particularly in the case of sarcoidosis where the inciting antigen(s), mode of induction, and minimum dose exposure have not been established for the human disease. Moreover, although murine models are ubiquitously studied, many basic biological differences exist in both the innate and adaptive immune responses of mice and man [65]. Further, pulmonary disease models must also take into account differences in murine and human bronchoalveolar lavage fluid proteins [66]. Despite these problems, murine models of pulmonary disease such as the sepharose bead granuloma model mentioned above, have provided valuable data on inflammatory granuloma formation [63,67].

The MWCNT preparation used in our model exhibited a dominant range of nanotube diameters from 20 to 40 nm and a net metal catalyst (Fe) content of less than 1.0% [62]. MWCNT (25–100 μg) were administered as a bolus in a 35% surfactant-saline solution via oropharyngeal instillation. The dosage we selected was within the range inducing granulomas in a study by Lam *et al.* [68] using intratracheally administered SWCNT. Inhalation studies have demonstrated that exposure of rats to aerosols containing 0.1 mg/m³ MWCNT can yield pulmonary granulomatous inflammation [69]. When compared to an aerosol delivery method, however, retropharyngeal instillation may be superior as it achieves full and consistent delivery, with decreased variability between exposures [43,67]. In our model, small granulomatous reactions surrounding MWCNT aggregates were detectable by 10 days post instillation with larger reactions persisting for up to 90 days. Bronchoalveolar lavage (BAL) of mice revealed MWCNT within alveolar macrophages [62].

The major goal in the initial studies of our chronic MWCNT model was to document pathways involved in granuloma formation by analyzing potential mediators in granuloma/non-granuloma lung tissues and BAL cells. Granulomatous foci and non-granulomatous tissues were collected by laser capture microdissection (LCM) for mRNA quantitation. Protein expression in lung tissues or BAL cells was evaluated by immunohistochemistry. Results showed elevated expression of osteopontin, a cytokine reported to be critical for granuloma formation [70], in granulomatous tissues and in BAL cells. Other granuloma-promoting factors found to be elevated were cell adhesion molecules (integrins) which are necessary for the fusion of macrophages into multinucleated giant cells within the granuloma structure [71]. Similarly, matrix metalloproteinases (MMPs) such as MMP-12 were also elevated in granulomatous tissue and in BAL cells. MMP-12 is a contributor to lung remodeling and targets elastin [72]. Overexpression has been reported to increase granuloma size in experimental animals [73]. Elevated MMP12 was also noted in a SWCNT model showing granulomas [74]. In our model, we also observed a concomitant decrease in Tissue Inhibitor of MMPs (TIMP)-3, an inhibitor of MMP-12 [75], suggesting that MMP-12 activity was unopposed by tissue inhibitors. These findings documented the formation of well-organized granulomas with time-related increases in granuloma size and mediator expression.

2.3. Parallel Findings in the MWCNT Model and Sarcoidosis

A review of selective findings from the chronic MWCNT granuloma model reveals many pathophysiologic similarities to human sarcoidosis (Table 1). With respect to granuloma formation, osteopontin and MMP-12 elevations noted in the MWCNT model have also been reported as characteristics of sarcoidosis granulomatous tissues [76,77].

Table 1. Comparative findings in the multiwall carbon nanotube (MWCNT) chronic granuloma model and human sarcoidosis.

Mediators/Regulators of	Pulmonary Findings	
Inflammation or Damage	MWCNT Model	Sarcoidosis
Osteopontin	Granuloma tissue; BAL * cells and fluids, [62]	Granuloma tissue [76]
MMP-12	Granuloma tissue, BAL * cells [62].	Granuloma tissue [77]
	Lung tissue [74]	
CCL2 (MCP-1)	Granuloma tissue [62]. Lung tissue [74]	BAL * fluids [78,79]
CCL5 (Rantes)	BAL * cells (unpublished data)	BAL * fluids [78,79]
TNFα	Granuloma tissue [62]	Trans-bronchial biopsy tissues [80]
IFN-γ	Granuloma tissue; BAL * cells [81]	Granuloma tissue [82]; BAL * cells [83]
PPARγ	Alveolar macrophages [81]	Alveolar macrophages [84]
Twist1	Alveolar macrophages [85]	Alveolar macrophages [85]

Note: * Bronchoalveolar Lavage.

A survey of pro-inflammatory cytokines and chemokines in the MWCNT model also demonstrated close similarities with sarcoidosis lung (Table 1). Elevated TNFα and the chemokine CCL2 (MCP-1) were noted in murine granulomatous tissue at 60 and 90 days post MWCNT instillation [62]. More recent studies in progress have also detected CCL5 (Rantes) production at 60 days after MWCNT instillation (unpublished data). All three of these pro-inflammatory mediators are known to be elevated in sarcoidosis granulomatous tissue and BAL cells [78,80] and are indicators of NF-κB activation which we have documented previously in sarcoidosis [84].

In our next study of the MWCNT model, we examined the role of the transcription factor, peroxisome proliferator-activated receptor gamma (PPARγ). While this transcription factor is a critical regulator of lipid and glucose metabolism, PPARγ also exerts negative effects on many pro-inflammatory genes [86]. Our previous studies had established that PPARγ binding activity and expression were deficient in alveolar macrophages from sarcoidosis patients [84]. Our first aim in the MWCNT model was to determine whether MWCNT instillation would also cause a reduction in alveolar macrophage PPARγ from wild-type mice. All studies were terminated at 60 days after MWCNT instillation. Results from wild-type mice proved to be virtually identical to those we observed in sarcoidosis, with alveolar macrophages showing significantly depressed PPARγ activity and expression [81]. Our second aim in the study was to determine whether pre-existing PPARγ deficiency would affect production of granuloma structures and inflammatory mediators. To this end, we utilized macrophage-specific PPARγ KO conditional mice. Results from MWCNT-instilled PPARγ KO mice demonstrated a more extensive granuloma formation and higher levels of osteopontin and pro-inflammatory cytokines (IFN-γ, CCL2) in granulomatous tissues and BAL cells than in similarly treated wild-type mice [81].

Thus results suggested that PPAR γ functions as a negative regulator of chronic granulomatous inflammation in the lung.

In our latest publication, we utilized the MWCNT model to explore expression of the Twist1 transcription factor in alveolar macrophages. Initially, we had carried out gene array analysis of BAL cells in sarcoidosis patients and healthy human controls to ascertain patterns of macrophage activation phenotypes M1 and M2 [85]. Findings showed a predominantly M1 phenotype in sarcoidosis alveolar macrophages together with a significant increase in Twist1 gene expression. Twist1 has been shown to be a regulator of embryonic mesenchymal development [87] but its possible function in alveolar macrophages has not been ascertained. Gene array data were confirmed by quantitative mRNA analysis of sarcoidosis BAL cells and protein expression was found to be localized to alveolar macrophages. To determine whether Twist1 expression could be a general marker of inflammatory alveolar macrophages, wild-type and macrophage-specific PPARy KO mice received instillations of MWCNT and were analyzed 60 days later. Results were similar to findings in sarcoidosis: Twist1 protein was elevated in both wild-type and PPARy KO alveolar macrophages after MWCNT instillation. Levels of Twist1 expression overall were higher in PPARy KO mice than wild-type, an observation that coincided with previous findings showing higher levels of inflammatory mediators in PPARy KO versus wild-type mice [81]. While functional studies of Twist1 are still in progress, the current data confirm that Twist1 may coincide with an inflammatory M1 alveolar macrophage phenotype in both mouse and man.

3. Summary and Conclusions

Human granulomatous diseases are chronic in nature and comprise a world-wide health problem. In many cases such as sarcoidosis, the etiology may be unknown. Efforts to study mechanisms of granulomatous disease have been hampered by the lack of a chronic pulmonary disease model. In studies of carbon nanotube toxicity, investigators have frequently reported persistent granulomatous structures within the lungs of experimental animals after instillation of carbon nanotubes. Based upon such studies, we developed a MWCNT model of chronic granulomatous lung disease which has proved to exhibit numerous pathophysiologic characteristics similar to those of human sarcoidosis. The possibility of carbon nanotube contribution to human granulomatous diseases such as sarcoidosis is an important environmental issue but has not been addressed previously. Overall, data from the chronic MWCNT model suggest that this model has a broad range of applicability for probing mechanisms of inflammation and granuloma persistence, as well as the potential for exploring novel therapeutic approaches to chronic granulomatous disease.

Acknowledgments

This work was supported in part by HL 077652 and ES 022462 to MJT.

Author Contributions

BPB and MJT wrote the manuscript. MAJ made suggestions and edited the manuscript.

Conflicts of Interest

MJT receives contract support from MedImmune. MAJ is consultant for Questcor, Janssen, Celgene, Novartis, and Mitsubishi-Tanabe. MAJ receives university grant support from T2-Biosystems. BPB declares no conflict of interest.

References

- 1. Perez, R.L.; Rivera-Marrero, C.A.; Roman, J. Pulmonary granulomatous inflammation: From sarcoidosis to tuberculosis. *Semin. Respir. Infect.* **2003**, *18*, 23–32.
- 2. Petersen, H.J.; Smith, A.M. The role of the innate immune system in granulomatous disorders. *Front. Immunol.* **2013**, *4*, doi:10.3389/fimmu.2013.00120.eCollection 2013.
- 3. Ito, T.; Connett, J.M.; Kunkel, S.L.; Matsukawa, A. The linkage of innate and adaptive immune response during granulomatous development. *Front. Immunol.* **2013**, *4*, doi:10.3389/fimmu.2013.00010.
- 4. Newman, L.S.; Rose, C.S.; Bresnitz, E.A.; Rossman, M.D.; Barnard, J.; Frederick, M.; Terrin, M.L.; Weinberger, S.E.; Moller, D.R.; McLennan, G.; *et al.* A case control etiologic study of sarcoidosis: Environmental and occupational risk factors. *Am. J. Respir. Crit. Care Med.* **2004**, *170*, 1324–1330.
- 5. Rybicki, B.A.; Major, M.; Popovich, J.J.; Maliarik, M.; Iannuzzi, M.C. Racial differences in sarcoidosis incidence: A five year study in a health maintenance organization. *Am. J. Epidemiol.* **1997**, *145*, 234–241.
- 6. Iannuzzi, M.C.; Rybicki, B.A.; Teirstein, A.S. Sarcoidosis. N. Engl. J. Med. 2007, 357, 2153–2165.
- 7. Judson, M.A.; Boan, A.D.; Lackland, D.T. The clinical course of sarcoidosis: Presentation, diagnosis, and treatment in a large white and black cohort in the United States. *Sarcoidosis Vasc. Diffus. Lung Dis.* **2012**, *29*, 119–127.
- 8. Kajdasz, D.K.; Judson, M.A.; Mohr, L.C.; Lackland, D.T., Jr. Geographic variation in sarcoidosis in South Carolina: Its relation to socioeconomic status and health care indicators. *Am. J. Epidemiol.* **1999**, *150*, 271–278.
- 9. Li, N.; Bajoghli, A.; Kubba, A.; Bhawan, J. Identification of mycobacterial DNA in cutaneous lesions of sarcoidosis. *J. Cutan. Pathol.* **1999**, *26*, 271–278.
- 10. Drake, W.P.; Dhason, M.S.; Nadaf, M.; Shepherd, B.E.; Vadivelu, S.; Hajizadeh, R. Cellular recognition of *Mycobacterium tuberculosis* ESAT-6 and KatG peptides in systemic sarcoidosis. *Infect. Immun.* **2007**, *75*, 527–530.
- 11. Song, Z.; Marzilli, L.; Greenlee, B.M.; Chen, E.S.; Silver, R.F.; Askin, F.B. Mycobacterial catalase-peroxidase is a tissue antigen and target of the adaptive immune response in systemic sarcoidosis. *J. Exp. Med.* **2005**, *201*, 755–767.
- 12. Dubaniewicz, A.; Dubaniewicz-Wybieralska, M.; Sternau, A.; Zwolska, Z.; Izycka-Swieszewska, E.; Augustynowicz-Kopec, E.; Skokowski, J.; Singh, M.; Zimnoch, L. *Mycobacterium tuberculosis* complex and mycobacterial heat shock proteins in lymph node tissue from patients with pulmonary sarcoidosis. *J. Clin. Microbiol.* **2006**, *44*, 3448–3451.

13. Chen, E.S.; Wahlstrom, J.; Song, Z.; Willett, M.H.; Wiken, M.; Yung, R.C.; Wahlström, J.; Song, Z.; West, E.E.; McDyer, J.F.; *et al.* T cell responses to mycobacterial catalase-peroxidase profile a pathogenic antigen in systemic sarcoidosis. *J. Immunol.* **2008**, *181*, 8784–8796.

- 14. Oswald-Richter, K.A.; Culver, D.A.; Hawkins, C.; Hajizadeh, R.; Abraham, S.; Shepherd, B.E.; Jenkins, C.A.; Judson, M.A.; Drake, W.P. Cellular responses to mycobacterial antigens are present in bronchoalveolar lavage fluid used in the diagnosis of sarcoidosis. *Infect. Immun.* **2009**, 77, 3740–3748.
- 15. Eishi, Y.; Suga, M.; Ishige, I.; Kobayashi, D.; Yamada, T.; Takemura, T.; Takizawa, T.; Koike, M.; Kudoh, S.; Costabel, U.; *et al.* Quantitative analysis of mycobacterial and propionibacterial DNA in lymph nodes of Japanese and European patients with sarcoidosis. *J. Clin. Microbiol.* **2002**, *40*, 198–204.
- 16. Ishige, I.; Eishi, Y.; Takemura, T.; Kobayashi, I.; Nakata, K.; Tanaka, I.; Nagaoka, S.; Iwai, K.; Watanabe, K.; Takizawa, T.; *et al.* Propionibacterium acnes is the most common bacterium commensal in peripheral lung tissue and mediastinal lymph nodes from subjects without sarcoidosis. *Sarcoidosis Vasc. Diffus. Lung Dis.* **2005**, *22*, 33–42.
- 17. Gerdes, J.; Richter, E.; Rusch-Gerdes, S.; Greinert, V.; Galle, J.; Schlaak, M.; Flad, H.D.; Magnussen, H. Mycobacterial nucleic acids in sarcoid lesions. *Lancet* **1992**, 339, 1536–1537.
- 18. Ghossein, R.A.; Ross, D.G.; Salomon, R.N.; Rabson, A.R. A search for mycobacterial DNA in sarcoidosis using the polymerase chain reaction. *Am. J. Clin. Pathol.* **1994**, *101*, 733–737.
- 19. Westney, G.E.; Judson, M.A. Racial and ethnic disparities in sarcoidosis: From genetics to socioeconomics. *Clin. Chest Med.* **2006**, *27*, 453–462.
- 20. Bardinas, F.; Morera, J.; Fite, E.; Plasencia, A. Seasonal clustering of sarcoidosis. *Lancet* **1989**, *2*, 455–456.
- 21. Kern, D.G.; Neill, M.A.; Wrenn, D.S.; Varone, J.C. Investigation of a unique time-space cluster of sarcoidosis in firefighters. *Am. Rev. Respir. Dis.* **1993**, *148*, 974–980.
- 22. Prezant, D.J.; Dhala, A.; Goldstein, A.; Janus, D.; Ortiz, F.; Aldrich, T.K.; Kelly, K.J. The incidence, prevalence, and severity of sarcoidosis in New York City firefighters. *Chest* **1999**, *116*, 1183–1193.
- 23. Miller, A. Sarcoidosis, firefighters sarcoidosis, and World Trade Center "sarcoid-like" granulomatous pulmonary disease. *Chest* **2007**, *132*, 2053.
- 24. Izbicki, G.; Chavko, R.; Banauch, G.I.; Weiden, M.D.; Berger, K.I.; Aldrich, T.K.; Hall, C.; Kelly, K.J.; Prezant, D.J. World Trade Center "sarcoid-like" granulomatous pulmonary disease in New York City fire department rescue workers. *Chest* **2007**, *131*, 1414–1423.
- 25. Crowley, L.E.; Herbert, R.; Moline, J.M.; Wallenstein, S.; Shukla, G.; Schechter, C.; Udasin, I.; Luft, B.J.; Harrison, D.; Shapiro, M.; *et al.* "Sarcoid like" granulomatous pulmonary disease in World Trade Center disaster responders. *Am. J. Ind. Med.* **2011**, *54*, 175–184.
- 26. Wu, M.; Gordon, R.E.; Herbert, R.; Padilla, M.; Moline, J.; Mendelson, D.; Litle, V.; Travis, W.D.; Gil, J. Case report: Lung disease in World Trade Center responders exposed to dust and smoke: Carbon nanotubes found in the lungs of World Trade Center patients and dust samples. *Environ. Health Perspect.* **2010**, *118*, 499–504.

27. Lioy, P.J.; Weisel, C.P.; Millette, J.R.; Eisenreich, S.; Vallero, D.; Offenberg, J.; Buckley, B.; Turpin, B.; Zhong, M.; Cohen, M.D.; *et al.* Characterization of the dust/smoke aerosol that settled east of the World Trade Center (WTC) in lower Manhattan after the collapse of the WTC 11 September 2001. *Environ. Health Perspect.* 2002, *110*, 703–714.

- 28. Banauch, G.I.; Dhala, A.; Prezant, D.J. Pulmonary disease in rescue workers at the World Trade Center site. *Curr. Opin. Pulm. Med.* **2005**, *11*, 160–168.
- 29. Rybicki, B.A.; Hirst, K.; Iyengar, S.K.; Barnard, J.G.; Judson, M.A.; Rose, C.S.; Donohue, J.F.; Kavuru, M.S.; Rabin, D.L.; Rossman, M.D.; *et al.* A sarcoidosis genetic linkage consortium: The sarcoidosis genetic analysis (SAGA) study. *Sarcoidosis Vasc. Diffus. Lung Dis.* **2005**, *22*, 115–122.
- 30. Iannuzzi, M.C.; Iyengar, S.K.; Gray-McGuire, C.; Elston, R.C.; Baughman, R.P.; Donohue, J.F.; Hirst, K.; Judson, M.A.; Kavuru, M.S.; Maliarik, M.J.; *et al.* Genome-wide search for sarcoidosis susceptibility genes in African Americans. *Genes Immun.* **2005**, *6*, 509–518.
- 31. Judson, M.A.; Baughman, R.P.; Thompson, B.W.; Teirstein, A.S.; Terrin, M.L.; Rossman, M.D.; Yeager, H., Jr.; McLennan, G.; Bresnitz, E.A.; DePalo, L.; *et al.* Two year prognosis of sarcoidosis: The ACCESS experience. *Sarcoidosis Vasc. Diffus. Lung Dis.* **2003**, *20*, 204–211.
- 32. De Volder, M.F.; Tawfick, S.H.; Baughman, R.H.; Hart, A.J. Carbon nanotubes: Present and future commercial applications. *Science* **2013**, *339*, 535–539.
- 33. Hirano, S. A current overview of health effect research on nanoparticles. *Environ. Health Prev. Med.* **2009**, *14*, 223–225.
- 34. Gordon, A.T.; Lutz, G.E.; Boninger, M.L.; Cooper, R.A. Introduction to nanotechnology: Potential applications in physical medicine and rehabilitation. *Am. J. Phys. Med. Rehabil.* **2007**, *86*, 225–241.
- 35. Monthioux, M.; Kuznetsov, V.L. Who should be given the credit for the discovery of carbon nanotubes? *Carbon* **2006**, *44*, 1621–1623.
- 36. Lam, C.W.; James, J.T.; McCluskey, R.; Arepalli, S.; Hunter, R.L. A review of carbon nanotube toxicity and assessment of potential occupational and environmental health risks. *Crit. Rev. Toxicol.* **2006**, *36*, 189–217.
- 37. Shvedova, A.A.; Kisin, E.R.; Porter, D.; Schulte, P.; Kagan, V.E.; Fadeel, B.; Castranova, V. Mechanisms of pulmonary toxicity and medical applications of carbon nanotubes: Two faces of Janus? *Pharmacol. Ther.* **2009**, *121*, 192–204.
- 38. Wang, X.; Katwa, P.; Podila, R.; Chen, P.; Ke, P.C.; Rao, A.M.; Walters, D.M.; Wingard, C.J.; Brown, J.M. Multi-walled carbon nanotube instillation impairs pulmonary function in C57BL/6 mice. *Part. Fibre Toxicol.* **2011**, *8*, 24.
- 39. Mercer, R.R.; Hubbs, A.F.; Scabilloni, J.F.; Wang, L.; Battelli, L.A.; Friend, S.; Castranova, V.; Porter, D.W. Pulmonary fibrotic response to aspiration of multi-walled carbon nanotubes. *Part. Fibre Toxicol.* **2011**, *8*, 21.
- 40. Elgrabli, D.; Abella-Gallart, S.; Robidel, F.; Rogerieux, F.; Boczkowski, J.; Lacroix, G. Induction of apoptosis and absence of inflammation in rat lung after intratracheal instillation of multiwalled carbon nanotubes. *Toxicology* **2008**, *253*, 131–136.
- 41. Mutlu, G.M.; Budinger, G.R.S.; Green, A.A.; Urich, D.; Soberanes, S.; Chiarella, S.E.; Alheid, G.F.; McCrimmon, D.R.; Szleifer, I.; Hersam, M.C. Biocompatible nanoscale dispersion of single-walled carbon nanotubes minimizes *in vivo* pulmonary toxicity. *Nano Lett.* **2010**, *10*, 1664–1670.

42. Murray, A.R.; Kisin, E.R.; Tkach, A.V.; Yanamala, N.; Mercer, R.; Young, S.-H.; Fadeel, B.; Kagan, V.E.; Shvedova, A.A. Factoring-in agglomeration of carbon nanotubes and nanofibers for better prediction of their toxicity *versus* asbestos. *Part. Fibre Toxicol.* **2012**, *9*, 10.

- 43. Li, J.-G.; Li, W.-L.; Xu, J.-Y.; Cai, X.-Q.; Liu, R.-L.; Li, Y.-J.; Zhao, Q.-F.; Li, Q.-N. Comparative study of pathological lesions induced by multiwalled carbon nanotubes in lungs of mice by intrtracheal instillation and inhalation. *Environ. Toxicol.* **2007**, *22*, 415–421.
- 44. Costa, D.L.; Lehmann, J.R.; Winsett, D.; Richards, J.; Ledbetter, A.D.; Dreher, K.L. Comparative pulmonary toxicological assessment of oil combustion particles following inhalation or instillation exposure. *Toxicol. Sci.* **2006**, *91*, 237–246.
- 45. Driscoll, K.E.; Costa, D.L.; Hatch, G.; Henderson, R.; Oberdorster, G.; Salem, H.; Schlesinger, R.B. Intratracheal instillation as an exposure technique for the evaluation of respiratory tract toxicity: Uses and limitations. *Toxicol. Sci.* **2000**, *55*, 24–35.
- 46. Mercer, R.R.; Scabilloni, J.; Wang, L.; Kisin, E.; Murray, A.R.; Schwegler-Berry, D.; Shvedova, A.A.; Castranova, V. Alteration of deposition pattern and pulmonary response as a result of improved dispersion of aspirated single-walled carbon nanotubes in a mouse model. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2008**, *294*, L87–L97.
- 47. Hamilton, R.F., Jr.; Wu, Z.; Mitra, S.; Shaw, P.K.; Holian, A. Effect of MWCNT size, carboxylation, and purification on *in vitro* and *in vivo* toxicity, inflammation and lung pathology. *Part. Fibre Toxicol.* **2013**, *10*, 57.
- 48. Palomaki, J.; Valimaki, E.; Sund, J.; Vippola, M.; Clausen, P.A.; Jensen, K.A.; Savolainen, K.; Matikainen, S.; Alenius, H. Long, needle-like carbon nanotubes and asbestos activate the NLRP3 inflammasome through a similar mechanism. *ACS Nano* **2011**, *5*, 6861–6870.
- 49. Meunier, E.; Coste, A.; Olagnier, D.; Authier, H.; Lefevre, L.; Dardenne, C.; Bemad, J.; Beraud, M.; Flahaut, E.; Pipy, B. Double-walled carbon nanotubes trigger IL-1β release in human monocytes through Nlrp3 inflammasome activation. *Nanomedicine* **2012**, *8*, 987–995.
- 50. Martinon, F.; Burns, K.; Tschopp, J. The inflammasome: A molecular platform triggering activation of inflammatory caspases and processing of proIL-β. *Mol. Cell* **2002**, *10*, 417–426.
- 51. De Nardo, D.; De Nardo, C.M.; Latz, E. New insights into mechanisms controlling the NLRP3 inflammasome and its role in lung disease. *Am. J. Pathol.* **2014**, *184*, 42–54.
- 52. Dostert, C.; Petrilli, V.; van Bruggen, R.; Steele, C.; Mossman, B.T.; Tschopp, J. Innate immune activation through Nalp3 inflammasome sensing of asbestos and silica. *Science* **2008**, *320*, 674–677.
- 53. Pulskamp, K.; Diabate, S.; Krug, H.F. Carbon nanotubes show no sign of acute toxicity but induce intracellular reactive oxygen species in dependence on contaminants. *Toxicol. Lett.* **2007**, *168*, 58–74.
- 54. Tabet, L.; Bussy, C.; Setyan, A.; Simon-Deckers, A.; Rossi, M.J.; Boczkowski, J.; Lanone, S. Coating carbon nanotubes with a polystyrene-based polymer protects against pulmonary toxicity. *Part. Fibre Toxicol.* **2011**, *8*, doi:10.1186/1743-8977-8-3..
- 55. Luo, E.; Song, G.; Li, Y.; Shi, P.; Hu, J.; Lin, Y. The toxicity and pharmacokinetics of carbon nanotubes as an effective drug carrier. *Curr. Drug Metab.* **2013**, *14*, 879–890.
- 56. Bonner, J.C. Carbon nanotubes as delivery systems for respiratory disease: Do the dangers outweigh the potential benefits? *Expert Rev. Respir. Med.* **2011**, *5*, 779–787.

57. Dumortier, H. When carbon nanotubes encounter the immune system: Desirable and undesirable effects. *Adv. Drug Deliv. Rev.* **2013**, *65*, 2120–2126.

- 58. Mitchell, L.A.; Lauer, F.T.; Burchiel, S.W.; McDonald, J.D. Mechanisms for how inhaled multiwalled carbon nanotubes suppress systemic immune function in mice. *Nat. Nanotechnol.* **2009**, *4*, 451–456.
- 59. Staal, Y.C.; van Triel, J.J.; Maarschalkerweerd, T.V.; Arts, J.H.; Duistermaat, E.; Muijser, H.; van de Sandt, J.J.; Kuper, C.F. Inhaled multiwalled carbon nanotubes modulate the immune response of trimellitic anhydride-induced chemical respiratory allergy in brown Norway rats. *Toxicol. Pathol.* **2014**, in press.
- 60. Ronzani, C.; Casset, A.; Pons, F. Exposure to multi-walled carbon nanotubes results in aggravation of airway inflammation and remodeling and in increased production of epithelium-derived innate cytokines in a mouse model of asthma. *Arch. Toxicol.* **2014**, *88*, 489–499.
- 61. Ryman-Rasmussen, J.P.; Tewksbury, E.W.; Moss, O.R.; Cesta, M.F.; Wong, B.A.; Bonner, J.C. Inhaled multiwalled carbon nanotubes potentiate airway fibrosis in murine allergic asthma. *Am. J. Respir. Cell Mol. Biol.* **2009**, *40*, 349–358.
- 62. Huizar, I.; Malur, A.; Midgette, Y.A.; Kukoly, C.; Chen, P.; Ke, P.C.; Podila, R.; Rao, A.M.; Wingard, C.J.; Dobbs, L.; *et al.* Novel murine model of chronic granulomatous lung inflammation elicited by carbon nanotubes. *Am. J. Respir. Cell Mol. Biol.* **2011**, *45*, 858–866.
- 63. Kunkel, S.; Lukacs, N.W.; Strieter, R.M.; Chensue, S.W. Animal models of granulomatous inflammation. *Semin. Respir. Infect.* **1998**, *13*, 221–228.
- 64. Chensue, S.W.; Otterness, I.G.; Higashi, G.I.; Forsch, C.S.; Kunkel, S.L. Monokine production by hypersensitivity (*Schistosoma mansoni* egg) and foreign body (Sephadex bead)-type granuloma macrophages. Evidence for sequential production of IL-1 and tumor necrosis factor. *J. Immunol.* 1989, 142, 1281–1286.
- 65. Mestas, J.; Hughes, C.C. Of mice and not men: Differences between mouse and human immunology. *J. Immunol.* **2004**, *172*, 2731–2738.
- 66. Gharib, S.A.; Nguyen, E.; Altemeier, W.A.; Shaffer, S.A.; Doneanu, C.E.; Goodlett, D.R.; Schnapp, L.M. Of mice and men: Comparative proteomics of bronchoalveolar fluid. *Eur. Respir. J.* **2010**, *35*, 1388–1395.
- 67. Su, X.; Looney, M.; Robriquet, L.; Fang, X.; Matthay, M.A. Direct visual instillation as a method for efficient delivery of fluid into the distal airspaces of anesthetized mice. *Exp. Lung Res.* **2004**, *30*, 479–493.
- 68. Lam, C.W.; James, J.T.; McCluskey, R.; Hunter, R.L. Pulmonary toxicity of single-wall carbon nanotubes in mice 7 and 90 days after intratracheal instillation. *Toxicol. Sci.* **2004**, *77*, 126–134.
- 69. Ma-Hock, L.; Treumann, S.; Strauss, V.; Brill, S.; Luizi, F.; Mertler, M.; Wiench, K.; Gamer, A.O.; van Ravenzwaay, B.; Landsiedel, R. Inhalation toxicity of multiwall carbon nanotubes in rats exposed for 3 months. *Toxicol. Sci.* **2009**, *112*, 468–481.
- 70. O'Regan, A.W.; Hayden, J.M.; Body, S.; Liaw, L.; Mulligan, N.; Goetschkes, M.; Berman, J.S. Abnormal pulmonary granuloma formation in osteopontin-deficient mice. *Am. J. Respir. Crit. Care Med.* **2001**, *164*, 2243–2247.
- 71. Helming, L.; Gordon, S. The molecular basis of macrophage fusion. *Immunobiology* **2007**, *212*, 785–793.

72. Lanone, S.; Zheng, T.; Zhu, Z.; Liu, W.; Lee, C.G.; Ma, B.; Chen, Q.; Homer, R.J.; Wang, J.; Rabach, L.A.; *et al.* Overlapping and enzyme-specific contributions of matrix metalloproteinases-9 and -12 in IL-13-induced inflammation and remodeling. *J. Clin. Investig.* **2002**, *110*, 463–474.

- 73. Fan, J.; Wang, X.; Wu, L.; Matsumoto, S.-I.; Liang, J.; Koike, T.; Ichikawa, T.; Sun, H.; Shikama, H.; Sasaguri, Y.; *et al.* Macrophage-specific overexpression of human matrix metalloproteinase-12 in transgenic rabbits. *Transgenic Res.* **2004**, *13*, 261–269.
- 74. Hsieh, W.Y.; Chou, C.C.; Ho, C.C.; Yu, S.L.; Chen, H.Y.; Chou, H.Y.; Chen, J.J.; Chen, H.W.; Yang, P.C. Single-walled carbon nanotubes induce airway hyperreactivity and parenchymal injury in mice. *Am. J. Respir. Cell Mol. Biol.* **2012**, *46*, 257–267.
- 75. Qi, J.H.; Ebrahem, Q.; Moore, N.; Murphy, G.; Claesson-Welsh, L.; Bond, M.; Baker, A.; Anand-Apte, B. A novel function for tissue inhibitor of metalloproteinases-3 (TIMP3): Inhibition of angiogenesis by blockage of VEGF binding to VEGF receptor-2. *Nat. Med.* **2003**, *9*, 407–415.
- 76. O'Regan, A.W.; Chupp, G.L.; Lowry, J.A.; Goetschkes, M.; Mulligan, N.; Berman, J.S. Osteopontin is associated with T cells in sacroid granulomas and has T cell adhesive and cytokin-like properties *in vitro*. *J. Immunol.* **1999**, *162*, 1024–1031.
- 77. Crouser, E.D.; Culver, D.A.; Knox, K.S.; Julian, M.W.; Shao, G.; Abraham, S.; Liyanarachchi, S.; Macre, J.E.; Wewers, M.D.; Gavrilin, M.A.; *et al.* Gene expression profiling identifies MMP-12 and ADAMDEC1 as potential pathogenic mediators of pulmonary sarcoidosis. *Am. J. Respir. Crit. Care Med.* **2009**, *179*, 929–938.
- 78. Iyonaga, K.; Suga, M.; Ichiyasu, H.; Yamamoto, T.; Hiraga, Y.; Ando, M. Measurement of serum monocyte chemoattractant protein-1 and its clinical application for estimating the activity of granuloma formation in sarcoidosis. *Sarcoidosis Vasc. Diffus. Lung Dis.* **1998**, *15*, 165–172.
- 79. Palchevskiy, V.; Hashemi, N.; Weigt, S.S.; Xue, Y.Y.; Derhovanessian, A.; Keane, M.P.; Strieter, R.M.; Fishbein, M.C.; Deng, J.C.; Lynch, C.P., 3rd; *et al.* Immune response CC chemokines CCL2 and CCL5 are associated with pulmonary sarcoidosis. *Fibrogenesis Tissue Repair* **2011**, *4*, doi:10.1186/1755-1536-4-10.
- 80. Fehrenbach, H.; Zissel, G.; Goldmann, T.; Tschernig, T.; Vollmer, E.; Pabst, R.; Muller-Quemheim, J. Alveolar macrophages are the main source for tumour necrosis factor-alpha in patients with sarcoidosis. *Eur. Respir. J.* **2003**, *21*, 421–428.
- 81. Huizar, I.; Malur, A.; Patel, J.; McPeek, M.; Dobbs, L.; Wingard, C.; Barna, B.P.; Thomassen, M.J. The role of PPARγ in carbon nanotube-elicited granulomatous lung inflammation. *Respir. Res.* **2013**, *14*, doi:10.1186/1465-9921-14-7.
- 82. Bergeron, A.; Bonay, M.; Kambouchner, M.; Lecossier, D.; Riquet, M.; Soler, P.; Hance, A.; Tazi, A. Cytokine patterns in tuberculous and sarcoid granulomas. *J. Immunol.* **1997**, *159*, 3034–3043.
- 83. Robinson, B.W.; McLemore, T.L.; Crystal, R.G. Gamma interferon is spontaneously released by alveolar macrophages and lung T lymphocytes in patients with pulmonary sarcoidosis. *J. Clin. Investig.* **1985**, *75*, 1488–1495.
- 84. Culver, D.A.; Barna, B.P.; Raychaudhuri, B.; Bonfield, T.L.; Abraham, S.; Malur, A.; Farver, C.F.; Kavuru, M.S.; Thomassen, M.J. Peroxisome proliferator-activated receptor gamma activity is deficient in alveolar macrophages in pulmonary sarcoidosis. *Am. J. Respir. Cell Mol. Biol.* **2004**, *30*, 1–5.

85. Barna, B.P.; Huizar, I.; Malur, A.; McPeek, M.; Marshall, I.; Jacob, M.; Dobbs, L.; Kavuru, M.S.; Thomassen, M.J. Carbon nanotube-induced pulmonary granulomatous disease: Twist1 and alveolar macrophage M1 activation. *Int. J. Mol. Sci.* **2013**, *14*, 23858–23871.

- 86. Straus, D.S.; Glass, C.K. Anti-inflammatory actions of PPAR ligands: New insights on cellular and molecular mechanisms. *Trends Immunol.* **2007**, *28*, 551–558.
- 87. Franco, H.L.; Casasnovas, J.; Rodriguez-Medina, J.R.; Cadilla, C.L. Redundant or separate entities?—Roles of Twist1 and Twist2 as molecular switches during gene transcription. *Nucleic Acids Res.* **2011**, *39*, 1177–1186.
- © 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).