Supplementary Materials: Mutated Human P-Selectin Glycoprotein Ligand-1 and Viral Protein-1 of Enterovirus 71 Interactions on Au Nanoplasmonic Substrate for Specific Recognition by Surface-Enhanced Raman Spectroscopy

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1. Experimental Section



Figure S1. Schematic illustration of fabrication of Au nanoporous substrate.

2. Characterization of Au Nanoporous Substrate



Figure S2. HR-FETEM image of optimized Au nanoporous substrate and EDS mappings of areas marked in HR-FETEM image.

| Sample - | Nanopore S | Size (±2 nm) | Contact Angle (°) | | |
|----------|------------|--------------|-------------------|---------|--|
| | 0.3 (Å/s) | 1 (Å/s) | 0.3 (Å/s) | 1 (Å/s) | |
| Alloy | - | - | 80.23 | 80.23 | |
| Aup_30s | 39 | 22 | 98.12 | 97.58 | |
| Aup_60s | 46.5 | 41.3 | 76.08 | 75.88 | |
| Aup_90s | 50 | 44.7 | 76.00 | 75.13 | |
| Aup_120s | 60.7 | 50.7 | 75.61 | 74.65 | |
| Aup_150s | 66.6 | 57 | 75.43 | 74.41 | |
| Aup-180s | 73.7 | 61.8 | 75.27 | 74.19 | |
| Aup_210s | 85.8 | 66.3 | 75.15 | 74.5 | |

Table S1. Nanopore size and contact angle of as-fabricated Au nanoporous substrates.

3. Optical Properties of Au Nanoporous Substrate

The COMSOL calculated Au nanoporous cavity width and length were 60 and 90 nm, respectively, with and without roughness of surface. Figure S(3c) shows the simulation results of the SPR shift and local field distributions on the Au nanoporous cavity surface. The strong SPR peaks obtained at 605 nm calculated for the nanoporous substrate are due to high plasmonic coupling generated in the porous cavity environment. In addition, the EF increased with increasing roughness of porous cavities.



Figure S3. SPR spectra of as-fabricated Au nanoporous substrate created at deposition rates of (**a**) 0.3 Å/s and (**b**) 1 Å/s with optimized dealloying times (120 and 150 s, respectively). (**c**) COMSOL calculation of smooth and roughed Au nanocavity.

4. SERS Characterization of PPIs on Au Nanoporous Substrate

| 3APTES | 3APTES + Glu | GST-PSGL-1 | S-GST-PSGL-1 | Raman Assignment |
|---------------|--------------|-------------|------------------|---|
| 629, 677 | 670 | 671 | 647 | υ(C–C), υ(C–S), amino acid (tyrosine) |
| 751 | 784 | 778, 742 | 694, 753 | u(C–S) |
| 813 | - | - | 805 | (C–O–C) weak |
| - | - | 827 | 851 | NH2, CH2 wag |
| 861 | 878 | - | 873, 888 | QCH₂, δCH₃ |
| 948 | 903 | 974 | 950 | CH ₂ , Carboxylic acid |
| | - | - | 984 | SO_4^{2-} |
| 1007 | 1005 | - | - | Si–O–Si, Si–O–C |
| | _ | - | 1030 | -SO3 |
| | 1046 | 1035 | | C ₄ N ⁺ stretching |
| | _ | _ | 1077 | C–C in-plane bending, –SO3 |
| 1101 | 1113, 1162 | 1132, 1172 | 1121, 1150 | U(C=S), (C–C) stretching, CH ₃ , CH ₂ |
| | , | , | , | twisting, –SO ₃ |
| 1188 | - | - | 1192 | -SO3 |
| 1240 | 1206 | 1222 | 1237 | C–C stretching, CO in ring stretching CH of ring, Amide III |
| 1288 | _ | 1261 | _ | C–C stretching, C–N |
| | | 1001 | | Stretching, S=O |
| 1312 | - | 1304 | - | δ(CH ₂) twisting or wagging of protein |
| 1336 | 1326 | 1348 | 1325, 1345 | Nitro, Amide III, and CH2 wagging vibrations from glycine backbone |
| 1379 | _ | _ | 1380 | C–CH3, δCH3 symmetric (lipid |
| 1577 | | | 1500 | assignment) |
| 1486 | 1438 | 1419, 1449, | 1409 1451 1477 | CH ₂ /CH ₃ deformation of lipids and |
| 1100 | 1100 | 1498 | 1107, 1101, 1177 | proteins |
| 1534 | 1501, 1528 | _ | 1530, 1548, 1572 | C–N, Nitro, Amide |
| | _ | 1584 | 1589 | C=C aromatic |
| 1601 | 1593, 1657 | 1619 | 1627 | O–C=O stretching |

 Table S2. Raman assignment for protein immobilized on Au nanoporous substrate [1–12].

Table S3. Raman assignment for protein mutants on Au nanoporous substrate [1–12].

| M0 | M1 | M2 | M3 | M4 | M5 | M6 | M7 | Raman Assignment |
|---------------------|------|------|------|----------|------------|------------------|------------------|--------------------------------|
| 647, 693, 752 | 674 | 729 | _ | 622, 794 | 778 | 637, 677, 771 | 621, 689, 728 | υ(C–C), υ(C– S), amino acid |
| 824 | 812 | _ | _ | _ | _ | _ | _ | NH2, CH2 wag |
| 870 | _ | - | _ | _ | 886 | _ | 891 | QCH2, δCH3 |
| 950 | _ | - | _ | _ | _ | 949 | 914 | CH2, carboxylic acid |
| 986 | - | 982 | _ | _ | _ | _ | _ | SO4 ²⁻ |
| _ | 1004 | 1002 | 999 | 998 | 999 | 998 | 1001 | Phenylalanine |
| 1030 | - | _ | _ | _ | _ | _ | _ | –SO3 |
| 1084 | _ | - | _ | _ | _ | 1067 | 1082 | C4N+ stretching, – SO3 |
| 1122, 1151, 1194 | | 1148 | 1148 | | 1118, 1148 | 1124 | 1103, 1166 | -SO3 |
| 1239 | 1216 | 1270 | _ | 1261 | 1278 | _ | 1261 | S=O |
| _ | _ | - | _ | - | _ | - | 1302 | δ(CH2) twisting or |

| S4 | of | S1 | 0 |
|----|----|----|---|
| | | | |

| | | | | | | | | wagging of protein | |
|-------------|-------|-----------|-------|------|------------|------------|------------|----------------------------------|------------|
| | | | | | | | | Nitro, amide | |
| | | | | | | | 100(| III, and CH ₂ | |
| 1323 | - | - | - | - | 1328 | - | 1326, | wagging | |
| | | | | | | | 1347 | vibrations | |
| | | | | | | | | from glycine | |
| | | | | | | | | backbone | |
| | | | | | | | | С–СН3, бСН3 | |
| - | 1361 | 1387 | 1390 | 1385 | 1397 | 1388 | - | symmetric | |
| | | | | | | | | (lipid | |
| | | | | | | | | assignment) | |
| | | | | | | | | CH ₂ /CH ₃ | |
| 1412, | 1450 | _ | - | - | 1452, 1496 | 1475 | 1410, | deformation | |
| 1449 | 1452 | | | | | | 1499 | of lipids and | |
| | | | | | | | | proteins | |
| 1532, 1548, | 1531, | 1570 | | 1515 | 1526 1554 | 1510, | 1550 | C–N, nitro, | |
| 1574 | 1576 | 1570 | - | 1515 | 1536, 1554 | 1543, 1570 | 1552 | amide | |
| _ | - | - | | 1585 | - | _ | - | C=C aromatic | |
| 1501 1/0/ | 1(20 | 1622, | 1595, | 1(0) | 1(02 1(22 | 1502 1(21 | 1744 | O-C=O | |
| 1591, 1626 | 1638 | 1638 1649 | 1649 | 1622 | 1626 | 1602, 1632 | 1392, 1631 | 1044 | stretching |

Samples are denoted as M0 (46, 48, 51), M1 (46F), M2 (48F), M3 (51F), M4 (46F, 48F), M5 (46F, 51F), M6 (48F, 51F), and M7 (46F, 48F, 51F).

Table S4. Raman assignment for protein-antibody interaction on Au nanoporous substrate [1–13].

| Anti-Sulfotyrosine | S-GST-PSGL-1 | Raman Assignment | | | |
|--------------------|----------------|---|--|--|--|
| 649, 789 | 659, 743, 799 | U(C–S) | | | |
| 888 | 840, 878 | QCH2, δCH3, NH2 | | | |
| _ | 901 | CH ₂ , carboxylic acid | | | |
| 989 | 978 | SO_42^- | | | |
| 1025 | _ | -SO3 | | | |
| 1057 | 1057 | C ₄ N ⁺ stretching | | | |
| 1112,1135 | 1115,1194 | -SO3 | | | |
| 1248, 1285 | 1223, 1282 | -SO3, S=O | | | |
| 1000 1000 | 1228 1206 | Nitro, amide III, CH2 wagging, δCH3 | | | |
| 1555, 1577 | 1556, 1596 | symmetric (lipid assignment) | | | |
| 1401, 1452,1483 | 1449 | CH ₂ /CH ₃ deformation of lipids and proteins | | | |
| 1534 | 1531, 1556 | C–N, nitro, amide | | | |
| | 1597 | C=C aromatic | | | |
| 1642, 1684 | 1701,1764,1801 | H–C=O, O–C=O stretching | | | |



Figure S4. Relationship of relative Raman intensities for various mutants and antibody-sulfotyrosine interactions on Au nanoporous substrate determined using *I*_{SERS} peak regions at 990–1010 and 1601–1636 cm⁻¹.



Figure S5. VP1 of EV71 on Au nanoporous substrate examined at Raman laser wavelength of 633 nm.

| Fable 5. Raman assignmen | for PPIs on Au nano | porous substrate [3–13] |
|--------------------------|---------------------|-------------------------|
|--------------------------|---------------------|-------------------------|

| V 0 | V1 | V2 | V3 | V4 | V 5 | V6 | V7 | Raman Assignment |
|------------|-----|------|------|-----------|------------|------|-----------|--|
| (5) | 665 | 675 | 629, | 648, 681, | 650 | 645, | 709 | v(C–C), v(C–S), amino |
| 652 | 663 | 675 | 757 | 793 | 639 | 676 | 708 | acid |
| _ | 813 | 826 | 822 | _ | _ | 838 | _ | NH2, CH2 wag |
| _ | _ | 886 | 886 | 870 | _ | 858 | 882 | QCH2, δ CH3 |
| _ | _ | 948 | | 931 | 954 | _ | | CH2, carboxylic acid |
| _ | _ | _ | 983 | _ | _ | _ | 978 | SO4 ²⁻ |
| 1014 | _ | 993 | - | _ | _ | _ | | Phenylalanine |
| | _ | 1028 | _ | 1036 | _ | _ | 1028 | –SO3, phenylalanine |
| 1099 | _ | 1070 | _ | 1087 | _ | 1078 | _ | C ₄ N ⁺ stretching, –SO ₃ |

S6 of S1 0

| _ | 1121 | 1114 <i>,</i> 1154 | 1168 | 1105, 1139, 1183 | _ | 1158, 1174 | 1131 | –SO3 |
|---------------|---------------|-------------------------|---------------|------------------------|---------------|------------------------|---------------|--|
| 1237 | 1262 | 1215, 1240, 1288, | 1239 | 1219 | _ | 1254 | 1248 | S=O |
| _ | _ | _ | 1316 | _ | _ | 1329 | _ | Nitro, amide III, and CH2 wagging vibrations from glycine backbone |
| 1361 | _ | 1349 | _ | 1353 <i>,</i> 1391 | 1350 | _ | 1370 | C–CH3, δCH3 symmetric (lipid assignment) |
| 1407, 1489 | _ | 1425 <i>,</i> 1495 | 1402 | 1451 | _ | 1420, 1454, 1495 | 1474 | CH2/CH3 deformation of lipids and proteins (tyrosine) |
| 1531, 1555 | 1520, 1535 | 1562 | 1532 | 1543 | 1550, 1567 | 1523, 1550, 1579 | 1531, 1574 | C–N, nitro, amide |
| 1592 | _ | _ | 1580, | 1589 | _ | _ | _ | C=C aromatic |
| 1600, 1618 | 1600 | 1601, 1621 | 1601, 1628 | 1600, 1614, 1643 | 1600, 1633 | 1602, 1644 | 1600, 1618 | O–C=O stretching |

Samples are denoted as V0 (46, 48, 51), V1 (46F), V2 (48F), V3 (51F), V4 (46F, 48F), V5 (46F, 51F), V6 (48F, 51F), and V7 (46F, 48F, 51F).

References

- 1. Fischer, W.B.; Eysel, H.H. Polarized Raman spectra and intensities of aromatic amino acids phenylalanine, tyrosine and tryptophan. *Spectrochim. Acta A: Mol. Spectrosc.* **1992**, *48*, 725–732.
- Ashton, L.; Brewster, V.L.; Correa, E.; Goodacre, R. Detection of glycosylation and iron-binding protein modifications using Raman spectroscopy. *Analyst* 2017, 142, 808–814.
- 3. Li, Y.T.; Li, D.W.; Cao, Y.; Long, Y.T. Label-free in-situ monitoring of protein tyrosine nitration in blood by surface-enhanced Raman spectroscopy. *Biosens. Bioelectron.* **2015**, *69*, 1–7.
- 4. Ravikumar, B.; Rajaram, R.K.; Ramakrishnan, V. Raman and IR spectral studies of L-phenylalanine L-phenylalaninium dihydrogenphosphate and DL-phenylalaninium dihydrogenphosphate. *J. Raman Spectrosc.* **2006**, *37*, 597–605.
- 5. Singho, N.D.; Johan, M.R. Complex impedance spectroscopy study of silica nanoparticles via sol-gel method. *Int. J. Spectrosc.* **2012**, *2012*, 7.
- 6. Brewster, V.L.; Ashton, L.; Goodacre, R. Monitoring the glycosylation status of proteins using Raman spectroscopy. *Anal. Chem.* **2011**, *83*, 6074–6081.
- Davies, H.S.; Singh, P.; Deckert-Gaudig, T.; Deckert, V.; Rousseau, K.; Ridley, C.E.; Dowd, S.E.; Doig, A.J.; Pudney, P.D.A.; Thornton, D.J.; et al. Secondary structure and glycosylation of mucus glycoproteins by Raman spectroscopies. *Anal. Chem.* 2016, *88*, 11609–11615.
- 8. Madzharova, F.; Heiner, Z.; Kneipp, J. Surface enhanced hyper-Raman scattering of the amino acids tryptophan, Histidine, Phenylalanine, and Tyrosine. *J. Phys. Chem. C* **2017**, *121*, 1235–1242.
- 9. Arp, Z.; Autrey, D.; Laane, J.; Overman, S.A.; Thomas, G.J. Tyrosine Raman signatures of the filamentous virus Ff are diagnostic of non-hydrogen-bonded phenoxyls: demonstration by Raman and infrared spectroscopy of p-cresol vapor. *Biochemistry* **2001**, *40*, 2522–2529.
- 10. Hernández, B.; Pflüger, F.; Kruglik, S.G.; Ghomi, M. Characteristic Raman lines of phenylalanine analyzed by a multiconformational approach. *J. Raman Spectrosc.* **2013**, *44*, 827–833.
- 11. Li, X.; Martin, S.J.H.; Chinoy, Z.S.; Liu, L.; Rittgers, B.; Dluhy, R.A.; Boons, G.-J. Fast production of selfassembled hierarchical α-Fe₂O₃ nanoarchitectures. *Chem.A Eur. J.* **2016**, *22*, 11180–11185.

- 12. Lu, X.; Liu, Q.; Benavides-Montano, J.A.; Nicola, A.V.; Aston, D.E.; Rasco, B.A.; Aguilar, H.C. Detection of receptor-induced glycoprotein conformational changes on enveloped virions by using confocal micro-Raman spectroscopy. *J. Virol.* **2013**, *87*, 3130–3142.
- 13. Sivashanmugan, K.; Liu, P.-C.; Tsai, K.-W.; Chou, Y.-N.; Lin, C.-H.; Chang Y.; Wen, T.-C. An anti-fouling nanoplasmonic SERS substrate for trapping and releasing a cationic fluorescent tag from human blood solution. *Nanoscale* **2017**, *9*, 2865–2874.



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