

## Supplementary Materials

# Production of D-Tagatose by Whole-Cell Conversion of Recombinant *Bacillus subtilis* in the Absence of Antibiotics

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**Table S1.** Comparison of characteristics of L-Als from various microorganisms.

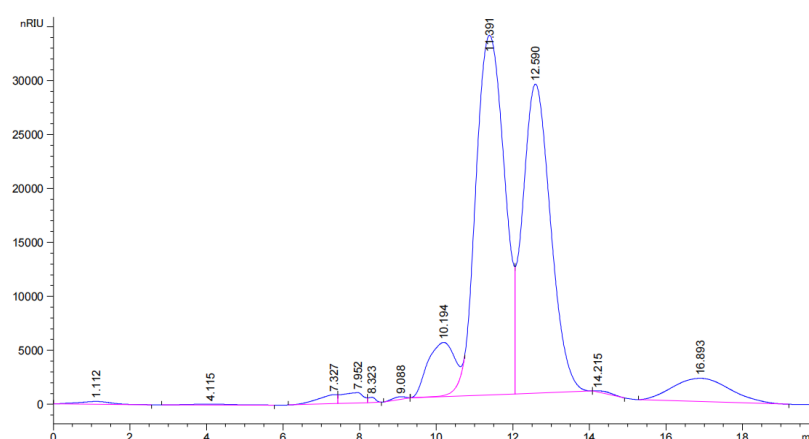
Source of L-arabinose isomerase	Reaction condition	Metal ion or boric acid requirement	concentration	concentration	Conversion yield(%)
<b>Mesophilic</b>					
<i>Lactobacillus sakei</i>	pH7.4,50°C	Mn <sup>2+</sup> (1.0)	54	45.9	85 (48 h)
<i>Lactobacillus brevis</i> PC16	pH6.5,67°C	Mn <sup>2+</sup> (1.0)	125	99.6	79.7(28 h)
<i>Lactobacillus fermentum</i> CGMCC 2971	pH6.5,65°C	Mn <sup>2+</sup> (1.0)	9	4.95	55(12 h)
<i>Lactobacillus fermentum</i> CGMCC 2971	pH6.5,70°C	Mn <sup>2+</sup> (1.0)	100	75	75(24 h)
<i>Pediococcus pentosaceus</i> PC-5	pH6.0,50°C	Mn <sup>2+</sup> (0.6) Co <sup>2+</sup> (0.8)	5	2.6	52(24 h)
<i>Enterococcus faecium</i> DBFIOE36	pH5.5,50°C	Mn <sup>2+</sup> (0.3)	90	40.5	45(48 h)
<i>Lactobacillus plantarum</i> WU14	pH7.17,50°C	Mn <sup>2+</sup> (10.0)	108	41.4	38.31(28 h)
<i>Bacillus coagulans</i> NL01	pH7.5,60°C	Mn <sup>2+</sup> (0.5) Co <sup>2+</sup> (0.5)	20	10.6	53(20 h)
<b>Thermophilic</b>					
<i>Geobacillus therodentitricans</i>	pH8.5,50°C	No	300	165	55(2 h)
<i>Geobacillus therodentitricans</i>	pH8.5-9.0, 60°C	boric acid (1110.0)	500	370	74(24 h)
<i>Bacillus thermoglucosidasius</i> KCTC 1828	pH7.4,40°C	Mn <sup>2+</sup> (1.0)	36	16.42	45.6(48 h)
<i>Acidothermus cellulolytic</i> ATCC 43068	pH7.5,75°C	Mn <sup>2+</sup> (1.0) Co <sup>2+</sup> (0.5)	9	4.8	53(12 h)
<i>Alicyclobacillus acidocaldarius</i>	pH6.0,60°C	Mn <sup>2+</sup> (1.0) Co <sup>2+</sup> (1.0)	18	7.9	44(12 h)
<i>Alicyclobacillus hesperidium</i> URH17-3-68	pH7.0,70°C	Co <sup>2+</sup> (1.0)	9	3.9	43(27 h)
<b>Hyperthermophilic</b>					
<i>Thermotoga neapolitana</i> 5068	pH7.0,80°C	Mn <sup>2+</sup> (1.0) Co <sup>2+</sup> (1.0)	1.8	1.2	68(20 h)
<i>Thermotoga Maritima</i>	pH7.5,70°C	Mn <sup>2+</sup> (5.0)	500	190	38(242 h)

Host expressing system	Reference
<i>Lactobacillus plantarum</i>	[16]
<i>B. subtilis</i> DB403	[15]
<i>E. coli</i>	[14]
<i>B. subtilis</i> 168	[13]
<i>E. coli</i>	[12]
<i>E. coli</i>	[11]
<i>Lactococcus lactis</i> NZ9000	[10]
<i>E. coli</i>	[9]
<i>Corynebacterium glutamicum</i>	[8]
<i>E. coli</i>	[7]
<i>E. coli</i>	[6]
<i>E. coli</i>	[5]
<i>E. coli</i>	[4]
<i>E. coli</i>	[3]
<i>E. coli</i>	[2]
<i>E. coli</i>	[1]

**Table S2.** List of bacterial strains and plasmids used in this study.

Strains and plasmids	Characteristics	Source and reference
<i>E. coli</i> K-12	$\beta$ -galactosidase and arabinose isomerase gene provider	Invitrogen
<i>B. subtilis</i> 168	trpC, expression host of <i>B. subtilis</i>	Takara
168 D1P1( <i>alrA</i> -, Zeo <sup>r</sup> )	168 derivative, $\Delta$ <i>alrA</i> , lox71-Zeo <sup>r</sup> -lox66	This study
168 D1P2( <i>alrA</i> -, Kan <sup>r</sup> )	168 derivative, $\Delta$ <i>alrA</i> , lox72, Kan <sup>r</sup> , pTSC	This study
168 D1( <i>alrA</i> -)	168 derivative, $\Delta$ <i>alrA</i>	This study
168 D1/pMA5- <i>alrA</i>	The recombinant 168 D1 strain harboring the plasmid pMA5- <i>alrA</i>	This study
168 D1/pMA5- <i>alrA-araA</i>	The recombinant 168 D1 strain harboring the plasmid pMA5- <i>alrA-araA</i>	This study
168 D1/pMA5- <i>alrA-lacZ</i>	The recombinant 168 D1 strain harboring the plasmid pMA5- <i>alrA-lacZ</i>	This study
168 D1/pMA5- <i>alrA-lacZ-araA</i>	The recombinant 168 D1 strain harboring the plasmid pMA5- <i>alrA-lacZ-araA</i>	This study
168 D2P1(Zeo <sup>r</sup> )	168 derivative, <i>lacZ</i> , $\Delta$ <i>alrA</i> , lox71-Zeo <sup>r</sup> -lox66	This study
168 D2P2(Kan <sup>r</sup> )	168 derivative, <i>lacZ</i> , $\Delta$ <i>alrA</i> , lox72, Kan <sup>r</sup> , pTSC	This study
168 D2( <i>alrA</i> -)	168 derivative, <i>lacZ</i> , $\Delta$ <i>alrA</i>	This study
168 D2/pMA5- <i>alrA</i>	The recombinant 168 D2 strain harboring the plasmid pMA5- <i>alrA</i>	This study
168 D2/pMA5- <i>alrA-araA</i>	The recombinant 168 D2 strain harboring the plasmid pMA5- <i>alrA-araA</i>	This study
<b>Plasmids</b>		
pMA5	Kan <sup>r</sup> , Zeo <sup>r</sup> , containing the <i>Hpa</i> II promoter	Takara
p7Z6	pMD18-T ligated with lox71-Zeo <sup>r</sup> -lox66 cassette	Takara
pTSC	Kan <sup>r</sup> , temperature sensitive in <i>B. subtilis</i> containing Pspac-cre expression cassette	Takara
pMA5- <i>alrA</i>	<i>B. subtilis</i> expression vector containing <i>alrA</i> gene	This study
pMA5- <i>alrA-araA</i>	<i>B. subtilis</i> expression vector containing <i>alrA</i> and <i>araA</i> genes	This study
pMA5- <i>alrA-lacZ</i>	<i>B. subtilis</i> expression vector containing <i>alrA</i> and <i>lacZ</i> genes	This study
pMA5- <i>araA</i>	<i>B. subtilis</i> expression vector containing <i>araA</i> gene	This study
pMA5- <i>alrA-lacZ-araA</i>	<i>B. subtilis</i> co-expression vector containing <i>alrA</i> , <i>araA</i> and <i>lacZ</i> gene	This study

Zeo<sup>r</sup>: bleomycin resistance; Kan<sup>r</sup>: kanamycin resistance;.



**Figure S1.** HPLC results of conversion products. The conversion using *B.subtilis* 168 D1/pMA5-*alrA-lacZ-araA* whole-cells. Bioconversion was performed at 40 °C, pH 8 (0.2 mol/L citric acid- $\text{Na}_2\text{HPO}_4$ ),  $\text{Mn}^{2+}$  concentration of 0.1 mol/L, and 0.1% TritonX-100, and cell concentration resulting in an OD of 40 at 600 nm. 10.194 min: lactose, 11.391 min: glucose, 12.590min: galactose, 16.893 min: D-tagatose.

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