

# Identification of Quinone Degradation as a Triggering Event for Intense Pulsed Light-Elicited Metabolic Changes in *Escherichia coli* by Metabolomic Fingerprinting

Qingqing Mao<sup>1</sup>, Juer Liu<sup>1</sup>, Justin R. Wiertzema<sup>1</sup>, Dongjie Chen<sup>1</sup>, Paul Chen<sup>2</sup>, David J. Baumler<sup>1</sup>, Roger Ruan<sup>2</sup>, Chi Chen<sup>1,\*</sup>

<sup>1</sup> Department of Food Science and Nutrition, University of Minnesota, 1334 Eckles Ave, Saint Paul, MN 55108, USA; maoux113@umn.edu (Q.M.); liux3514@umn.com (J.L.); wiert006@umn.edu (J.R.W.); chen5166@umn.edu (D.C.); dbaumler@umn.edu (D.J.B.)

<sup>2</sup> Department of Bioproducts and Biosystems Engineering, University of Minnesota, 1390 Eckles Ave., Saint Paul, MN 55108, USA; chenx088@umn.edu (P.C.); ruanx001@umn.edu (R.R.)

\* Correspondence: chichen@umn.edu; Tel.: +1-612-624-7704; Fax: +1-612-625-5272

## Supplementary Data

**Table S1.** MSMS fragments of selective metabolite markers

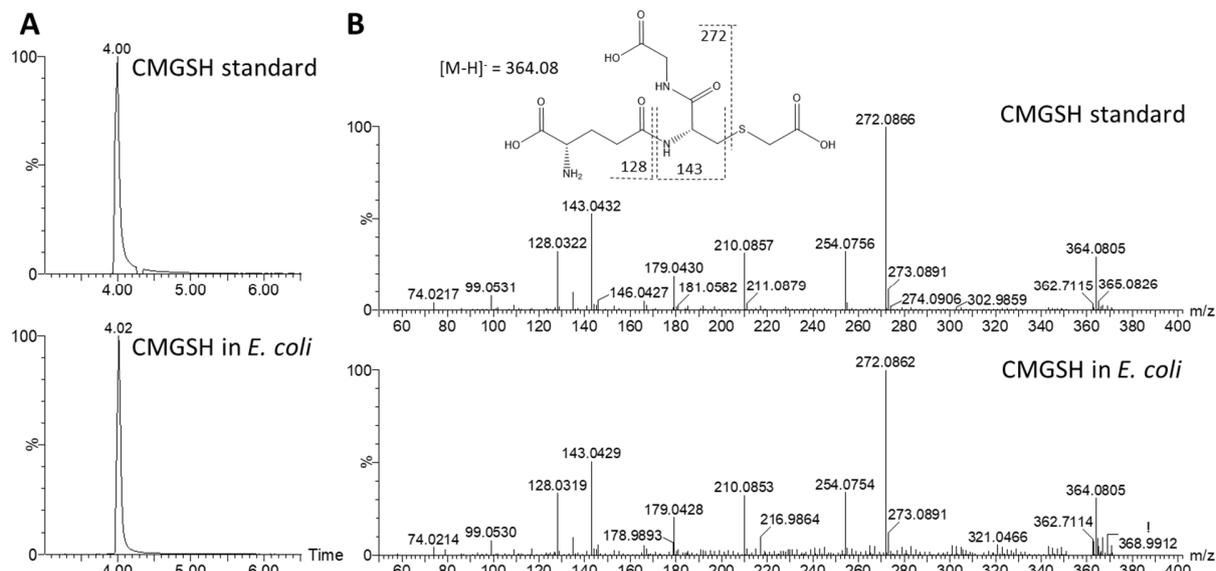
Ions	Detected Ion Adduct	m/z	Identity	Formula	m/z of major MS/MS fragments
V	[M+H] <sup>+</sup>	704.5238	PE(16:0/17:0Cyclo)#	C <sub>38</sub> H <sub>74</sub> NO <sub>8</sub> P	563 in ESI <sup>+</sup> ; 255, 267 in ESI <sup>-</sup>
VIII	[M+NH <sub>4</sub> ] <sup>+</sup>	746.6103	Ubiquinol-8#	C <sub>49</sub> H <sub>76</sub> O <sub>4</sub>	197
IX	[M+NH <sub>4</sub> ] <sup>+</sup>	734.5888	Menaquinone-8#	C <sub>51</sub> H <sub>72</sub> O <sub>2</sub>	187
X	[M+H] <sup>+</sup>	727.5684	Ubiquinone-8#	C <sub>49</sub> H <sub>74</sub> O <sub>4</sub>	197
XI	[M+H] <sup>+</sup>	732.5553	PE(16:0/19:0Cyclo)#	C <sub>40</sub> H <sub>78</sub> NO <sub>8</sub> P	593 in ESI <sup>+</sup> ; 255, 295 in ESI <sup>-</sup>
XII	[M+H] <sup>+</sup>	664.4928	PE(14:0/16:0)#	C <sub>35</sub> H <sub>70</sub> NO <sub>8</sub> P	523 in ESI <sup>+</sup> ; 227, 255 in ESI <sup>-</sup>
XVII	[M+H] <sup>+</sup>	188.1761	N-Acetylspermidine#	C <sub>9</sub> H <sub>21</sub> N <sub>3</sub> O	171, 114, 84
XIX	[M+DC] <sup>+</sup>	378.1845	N-Acetylcadaverine#	C <sub>7</sub> H <sub>16</sub> N <sub>2</sub> O	170, 86

**Table S2.** Respective mobile phase gradients for 10-min LC runs of *E. coli* extracts

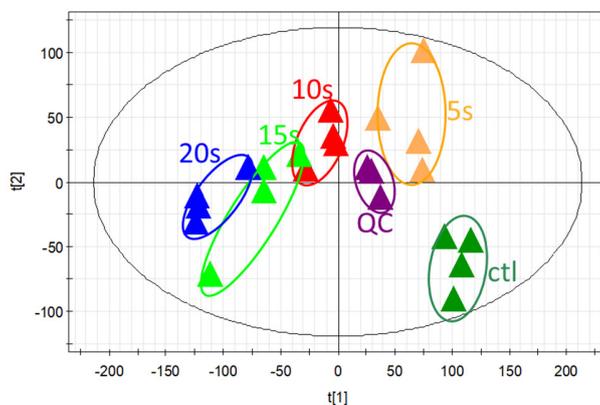
Column	BEH C18	BEH Amide	BEH C8
<b>Solvent A</b>	0.1% formic acid in water	0.1% formic acid in water	0.1% formic acid and 10 mM NH <sub>4</sub> OAc in 40% aqueous acetonitrile
<b>Solvent B</b>	0.1% formic acid in acetonitrile	0.1% formic acid in acetonitrile	0.1% formic acid and 10 mM NH <sub>4</sub> OAc in methanol
<b>Gradient</b>	0.5 min-99.5% A, 4 min-80% A, 8 min-5% A, 8.1 min-0% A, 9.0 min-0% A, 9.1 min-99.5% A, 10 min-99.5% A	0.5 min-0.5% A, 2 min-20% A, 8 min-50% A, 9.1 min-0.5% A, 10 min-0.5% A	0.5 min-55.5% A, 2.5 min-20% A, 5 min-15% A, 8 min-5% A, 8.1 min-0% A, 9.0 min-0% A, 9.1 min-55% A, 10 min-55% A

**Table S3.** MS settings in the ESI detection

<b>Capillary voltage</b>	3 kV for ESI <sup>+</sup> , -3 kV for ESI <sup>-</sup>
<b>Cone voltage</b>	40 V for ESI <sup>+</sup> , -35 V for ESI <sup>-</sup>
<b>Gas flow</b>	Nitrogen, cone gas 50 L/h, desolvation gas 600 L/h



**Figure S1.** Confirmation of S-carboxymethyl-glutathione (CMGSH) in *E. coli* by a comparison with its standard. The CMGSH standard was synthesized by a reaction between GSH and iodoacetic acid as described in the Materials and Methods. (A) Extracted chromatographs of CMGSH standard and CMGSH in the polar extract of IPL-treated *E. coli*. (B) MSMS fragmentograms of CMGSH standard and CMGSH in the polar extract of IPL-treated *E. coli*. The fragmentation was conducted at the negative ionization mode and interpreted in the inlaid structure diagram.



**Figure S2.** Scores plot with quality control (QC) samples. A pooled sample was injected for a total of three times (beginning, middle, and end) as the QC in each run for monitoring the LC-MS performance.