

Supplemental Materials

S1. List of Transition Invariants

The network has 6 transition invariants (TI):

1. TI₁: $k_{\text{bind}}, k_{\text{phos}}, k_{\text{dephos,m}}$ (binding of insulin, phosphorylation, dephosphorylation on membrane)
2. TI₂: $k_{\text{bind}}, k_{\text{phos}}, k_{\text{in,p}}, k_{\text{dephos,c}}, k_{\text{out}}$, buffer (binding of insulin, phosphorylation, internalization, cytoplasmic dephosphorylation, translocation back to membrane)
3. TI₃: $k_{\text{in}}, k_{\text{out}}$ (internalization, translocation back to membrane)
4. TI₄: $k_{\text{in,p}}, k_{\text{out,p}}$ (internalization of phosphorylated insulin receptor (IR), translocation back to membrane)
5. TI₅: $k_{\text{bind}}, k_{\text{diss}}$ (extracellular binding of insulin, release of insulin)
6. TI₆: $k_{\text{syn}}, k_{\text{deg}}$ (synthesis, degradation of receptor)

Each transition is member of at least one TI, hence the network is covered by TI (CTI).

S2. Quasi-Steady-State Approximation for TI₁

TI₁ describes a cycle of reactions for the species IR, IRI, and IRIP. The corresponding dynamic system is given by

$$\frac{\partial \vec{c}}{\partial t} = \begin{pmatrix} -k_{\text{bind}} i_0 & k_{\text{diss}} & k_{\text{dephos,m}} \\ +k_{\text{bind}} i_0 & -k_{\text{diss}} - k_{\text{phos}} & 0 \\ 0 & k_{\text{phos}} & -k_{\text{dephos,m}} \end{pmatrix} \vec{c}, \tag{S1}$$

where $\vec{c} = (ir, iri, irip)^T$ denotes a vector of concentrations. The concentration of free insulin is assumed to be constant, i.e., $i = i_0$. Within the QSSA we solved the linear system

$$\frac{\partial \vec{c}}{\partial \tau} = 0 \tag{S2}$$

and obtained the steady state for the 3 concentrations

$$\begin{aligned} ir^* &= \left[1 - \frac{i_0}{i_0 + i_c} \right] ir_0, \\ iri^* &= \frac{k_{\text{dephos,m}}}{k_{\text{phos}}} \left(1 + \frac{k_{\text{dephos,m}}}{k_{\text{phos}}} \right)^{-1} \frac{i_0}{i_0 + i_c} ir_0, \text{ and} \\ irip^* &= \left[1 - \frac{k_{\text{dephos,m}}}{k_{\text{phos}}} \left(1 + \frac{k_{\text{dephos,m}}}{k_{\text{phos}}} \right)^{-1} \right] \frac{i_0}{i_0 + i_c} ir_0 \end{aligned} \tag{S3}$$

with the equilibrium constant

$$i_c = \frac{k_{\text{dephos,m}}}{k_{\text{bind}}} \left(1 + \frac{k_{\text{diss}}}{k_{\text{phos}}}\right) \left(1 + \frac{k_{\text{dephos,m}}}{k_{\text{phos}}}\right)^{-1}. \quad (\text{S4})$$

For our choice of kinetic rate constants, the insulin-binding equilibrium constant becomes $i_c = 3.33$ nM. Sedaghat *et al.* assume a fast process of phosphorylation (*i.e.*, $k_{\text{phos}} \gg k_{\text{diss}}$ and $k_{\text{phos}} \gg k_{\text{dephos,m}}$). In this case the equation

$$i_c \approx \frac{k_{\text{dephos,m}}}{k_{\text{bind}}} \quad (\text{S5})$$

is a reasonable approximation. Since the ratio $k_{\text{dephos,m}}/k_{\text{phos}}$ is less than 0.1 %, we may neglect iri^* , and the formula

$$irip^* \approx \frac{i_0}{i_0 + i_c} ir_0$$

is sufficiently precise for practical applications.

S3. Quasi-Steady-State Approximation for TI_2

The steady-state concentrations ir^* , iri^* , and $irip^*$ completely ignore the process of translocation of receptor into the cytoplasm and are a justifiable approximation only for a short reaction time compared to the time scale of the translocation process. The process of translocation of the activated IR into the cytoplasm ($k_{\text{in,p}}$) is member of the subnetwork defined by TI_2 . The ODE system of the subnetwork reads

$$\frac{\partial \vec{c}}{\partial t} = \begin{pmatrix} 0 \\ 0 \\ 0 \\ k_{\text{syn}} \\ 0 \end{pmatrix} - \begin{pmatrix} k_{\text{bind}} i_0 + k_{\text{in}} & -k_{\text{diss}} & -k_{\text{dephos,m}} & -k_{\text{out}} & 0 \\ -k_{\text{bind}} i_0 & k_{\text{diss}} + k_{\text{phos}} & 0 & 0 & 0 \\ 0 & -k_{\text{phos}} & k_{\text{dephos,m}} + k_{\text{in,p}} & 0 & -k_{\text{out,p}} \\ -k_{\text{in}} & 0 & 0 & k_{\text{out}} + k_{\text{deg}} & -k_{\text{dephos,c}} \\ 0 & 0 & -k_{\text{in,p}} & 0 & k_{\text{dephos,c}} + k_{\text{out,p}} \end{pmatrix} \vec{c} \quad (\text{S6})$$

with the vector of concentrations, $\vec{c} = (ir, iri, irip, ir_{in}, irip_{in})^T$. The steady state is given by

$$\begin{aligned} ir^\dagger &= \frac{i_0}{i_c^\dagger + i_0} \left(1 + \frac{k_{\text{diss}}}{k_{\text{phos}}}\right) \left[\frac{k_{\text{dephos,m}}(k_{\text{out,p}} + k_{\text{dephos,c}})}{k_{\text{bind}}i_0 k_{\text{in,p}}} + \frac{k_{\text{dephos,c}}}{k_{\text{bind}}i_0} \right] \frac{k_{\text{out}}}{k_{\text{dephos,c}}} ir_{in}^\dagger, \\ iri^\dagger &= \frac{i_0}{i_c^\dagger + i_0} \left[\frac{k_{\text{dephos,m}}(k_{\text{out,p}} + k_{\text{dephos,c}})}{k_{\text{phos}}k_{\text{in,p}}} + \frac{k_{\text{dephos,c}}}{k_{\text{phos}}} \right] \frac{k_{\text{out}}}{k_{\text{dephos,c}}} ir_{in}^\dagger, \\ irip^\dagger &= \frac{i_0}{i_c^\dagger + i_0} \frac{k_{\text{out,p}} + k_{\text{dephos,c}}}{k_{\text{in,p}}} \frac{k_{\text{out}}}{k_{\text{dephos,c}}} ir_{in}^\dagger, \\ ir_{in}^\dagger &= \frac{k_{\text{syn}}}{k_{\text{deg}}}, \text{ and} \\ irip_{in}^\dagger &= \frac{i_0}{i_c^\dagger + i_0} \frac{k_{\text{out}}}{k_{\text{dephos,c}}} ir_{in}^\dagger \end{aligned} \quad (\text{S7})$$

with the constant

$$i_c^\dagger = \frac{k_{\text{in}}}{k_{\text{bind}}} \left[1 + \frac{k_{\text{dephos,m}}}{k_{\text{in,p}}} \left(1 + \frac{k_{\text{out,p}}}{k_{\text{dephos,c}}}\right) \right] \left(1 + \frac{k_{\text{diss}}}{k_{\text{phos}}}\right). \quad (\text{S8})$$

i_c^\dagger is the critical insulin concentration for the internalization of receptor. For a fast phosphorylation process as postulated by Sedaghat *et al.*, (*i.e.*, $k_{\text{phos}} = 2.500 \text{ min}^{-1}$) a simplification of equations (S7,S8) is feasible.

We considered nonzero degradation and nonzero synthesis of the receptor, *i.e.*, k_{syn} , k_{deg} , in the steady state (S7). However, the degradation and synthesis are not members of TI_2 but form the trivial TI_6 . For $k_{\text{syn}} = k_{\text{deg}} = 0$ (*i.e.* in the case of no degradation and no synthesis), the steady-state concentration, ir_{in}^\dagger , becomes a free parameter and has to be determined by a mass conservation equation for the amount of the receptor in the cell.

For our choice of kinetic constants, we get the numerical value, $i_c^\dagger = 0.535 \text{ nM}$, for the critical insulin concentration of internalization of the IR and the steady state concentrations (S7) become

$$\begin{aligned} ir^\dagger &= 0.9 \text{ pM} \times \left[1 - \frac{i_0}{i_c^\dagger + i_0} \right], \\ iri^\dagger &= 0.0116 \text{ fM} \times \frac{i_0}{i_c^\dagger + i_0}, \\ irip^\dagger &= 0.143 \text{ pM} \times \frac{i_0}{i_c^\dagger + i_0}, \\ ir_{in}^\dagger &= 0.1 \text{ pM}, \text{ and} \\ irip_{in}^\dagger &= 0.651 \text{ fM} \times \frac{i_0}{i_c^\dagger + i_0}. \end{aligned} \quad (\text{S9})$$

The steady state concentrations, iri^\dagger and $irip_{in}^\dagger$, of the transient complexes are below experimental detection limits. The steady state concentration, ir_{in}^\dagger , of free intracellular receptor is regulated by synthesis (k_{syn}) and degradation (k_{deg}), and hence remains constant for all values of i_0 . In the limit of small concentrations of insulin, $i_0 \rightarrow 0$, the function

$$f(i_0) = \frac{i_0}{i_c^\dagger + i_0} \quad (\text{S10})$$

approaches zero for vanishing concentration of external insulin, *i.e.*, $\lim_{i_0 \rightarrow 0} f(i_0) = 0$. For increasing concentrations of insulin, $i_0 \rightarrow \infty$, the function $f(i_0)$ converges to 1. Since the steady-state concentrations, iri^\dagger , $irip^\dagger$ and $irip_{in}^\dagger$, are proportional to $f(i_0)$, they are zero in the basal state of the cell, *i.e.*, in absence of extracellular insulin, $i_0 = 0$. In the process of down-regulation by insulin, the concentrations, iri^\dagger , $irip^\dagger$, and $irip_{in}^\dagger$, increase proportionally to the function $f(i_0)$ until they reach their maximal values for $i_0 \gg i_c^\dagger$. The steady-state concentration, ir^\dagger , of the surface receptor is proportional to $1 - f(i_0)$, and hence, ir^\dagger is maximal in the basal state and drops down to zero for $i_0 \gg i_c^\dagger$.

S4. Characteristic Eigenvalue for TI_1

The characteristic eigenvalue of ODE (S1) is given by

$$\lambda_1 = -\frac{k_{\text{bind}} i_0 + k_{\text{diss}} + k_{\text{phos}} + k_{\text{dephos,m}}}{2} \left[1 - \sqrt{1 - \frac{4(k_{\text{bind}} i_0 (k_{\text{phos}} + k_{\text{dephos,m}}) + (k_{\text{diss}} + k_{\text{phos}}) k_{\text{dephos,m}})}{(k_{\text{bind}} i_0 + k_{\text{diss}} + k_{\text{phos}} + k_{\text{dephos,m}})^2}} \right]. \quad (\text{S11})$$

The simplification

$$\lambda_1 \approx -\frac{k_{\text{phos}} (k_{\text{bind}} i_0 + k_{\text{dephos,m}})}{k_{\text{bind}} i_0 + k_{\text{phos}}} \quad (\text{S12})$$

approximates the eigenvalue, λ_1 , within a relative precision of 2×10^{-5} .

S5. Characteristic Eigenvalue for TI_2

The characteristic eigenvalue of ODE (S6) is given by

$$\lambda_2 = -\frac{L}{2} \left[1 - \sqrt{1 - \frac{4(k_{\text{out}}(k_{\text{out,p}} + k_{\text{dephos,c}}) + (k_{\text{out,p}} + k_{\text{dephos,c}})K_1 + (k_{\text{out}} + k_{\text{dephos,c}})K_2)}{L^2}} \right],$$

$$L = k_{\text{out,p}} + k_{\text{out}} + k_{\text{dephos,c}} + K_1 + K_2,$$

$$K_1 = k_{\text{in}} \frac{i_c}{i_0 + i_c}, \text{ and}$$

$$K_2 = \frac{k_{\text{phos}} k_{\text{in,p}}}{k_{\text{phos}} + k_{\text{dephos,m}}} \frac{i_0}{i_0 + i_c}.$$

S6. Drop of Insulin and the Lambert Function

We have abstained from discussing the development of insulin concentration with time based on the functional regimes of the Lambert function W . It is easy to see that for insulin concentrations well below the critical concentration of $i_c^\dagger = 0.535$ nM, the differential equation simplifies to

$$\frac{\partial i}{\partial t} = -\frac{i}{t_4}, \quad (\text{S13})$$

and the insulin concentration drops down exponentially in time

$$i(t) = i_0 e^{-t/t_4}. \quad (\text{S14})$$

In the case of a high concentration of insulin (*i.e.*, for $i \gg i_c^\dagger = 0.535$ nM), the cell is maximally down-regulated, and the differential equation is given by

$$\frac{\partial i}{\partial t} = -\frac{i_c}{t_4}. \quad (\text{S15})$$

Consequently, the consumption of insulin with constant maximal velocity leads to a linear diminishment of insulin:

$$i(t) = i_0 - i_c \frac{t}{t_4}. \quad (\text{S16})$$

The consumption of insulin by the cell leads to an exponential drop on the time scale of $t_4 = 5$ h 33 min, if the insulin concentration is below the critical insulin concentration, $i_c^\dagger = 0.535$ nM. For insulin given in excess (*i.e.*, for $i \gg i_c^\dagger$), the insulin concentration decreases linearly with a flat-angle slope of 0.535 nM/5 h 33 min.

S7. Phosphorylation Dynamics

Cedersund *et al.* [1] have discussed the short-term phosphorylation dynamics of the insulin receptor. They have measured a rapid transient overshoot in tyrosine phosphorylation for human adipocytes after a step increase from 0 to 0.1 μM in insulin concentration and have discussed the implication of such an “overshoot” on various model structures. Cedersund *et al.* [2] have rejected model structures based on the zeros and complex poles of the linearized transfer function, see also Brännmark *et al.* [3]. In terms of the Petri net formalism, the model structure requires a certain substructure to produce an overshoot behavior. For Sedaghat *et al.*’s model [4] such a substructure is defined by transition invariant TI_1 . The Petri net approach explains the overshoot by the high concentration of phosphorylated receptor, $irip^*$, of the meta-stable quasi-steady state associated with transition invariant TI_1 . Figure S1 shows the percentage of transient phosphorylated IR versus the concentration of insulin.

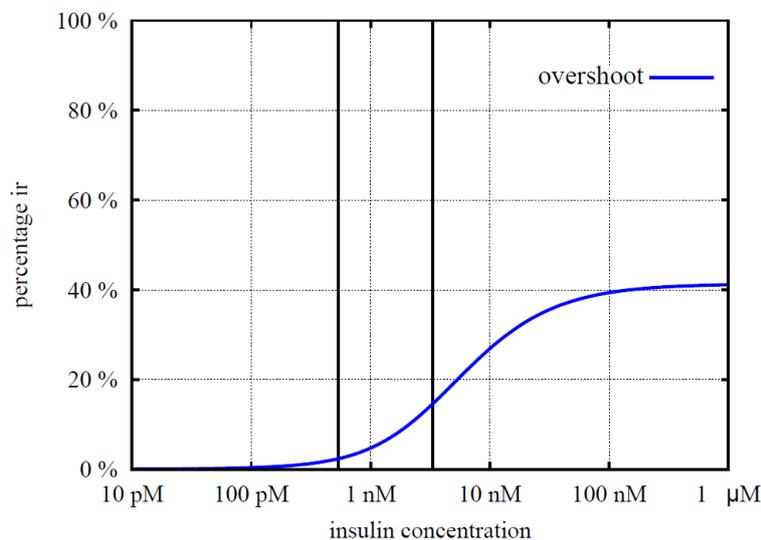


Figure S1. After a step increase in insulin concentration the concentration of phosphorylated IR approach the value $irip^*$ of meta-stable steady state (S3). This transient high value of phosphorylated IR drops to the value $\nu \times irip^\dagger$ of meta-stable steady state (S7) due to endocytosis and dephosphorylation of the internalized IR. Plotted is the percentage of transient phosphorylated $irip^* - \nu \times irip^\dagger$ versus the concentration of insulin. For 0.1 μM insulin concentration, Sedaghat *et al.*’s model estimates an “overshoot” at in round numbers 40% .

References

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