

Supplementary Material

Portability of a Small Molecule Binding Site Between Disordered Proteins

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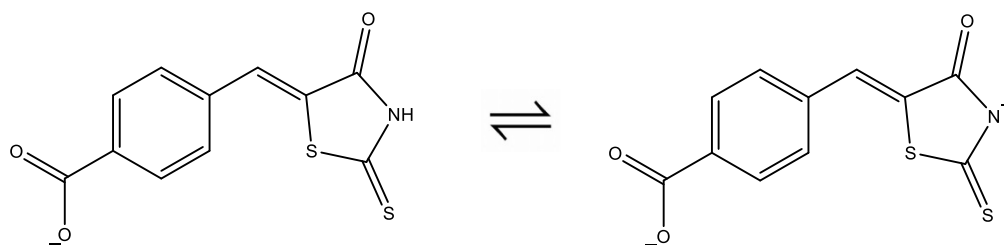
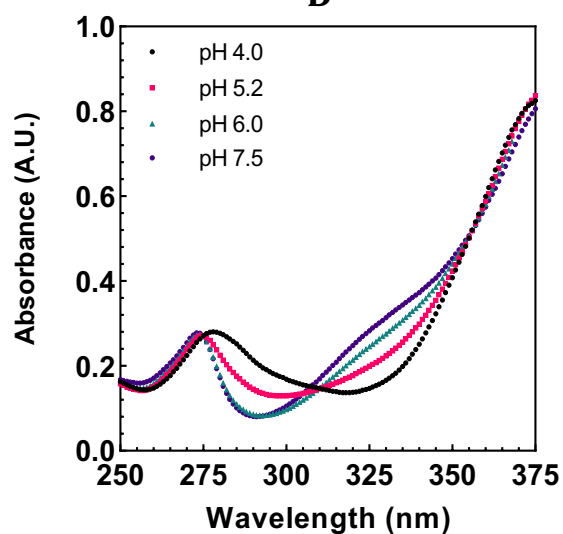
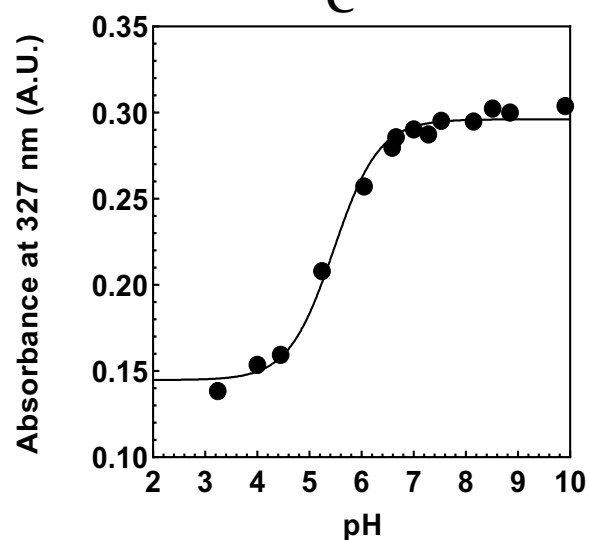
A**B****C**

Fig S2. UV/Vis characterization of 34RH. (A) pH dependent deprotonation of 34RH. (B) UV/Vis absorbance of 10 μ M 34RH in 1xPBS, 5% DMSO, 25 $^{\circ}$ C, at pH 4.0 (black), pH 5.2 (dark pink), pH 6.0 (teal), and pH 7.5 (purple). (C) 34RH absorbance at 327 nm at various pH values showing a pH dependent titration fit to the Henderson-Hasselbalch equation [45].

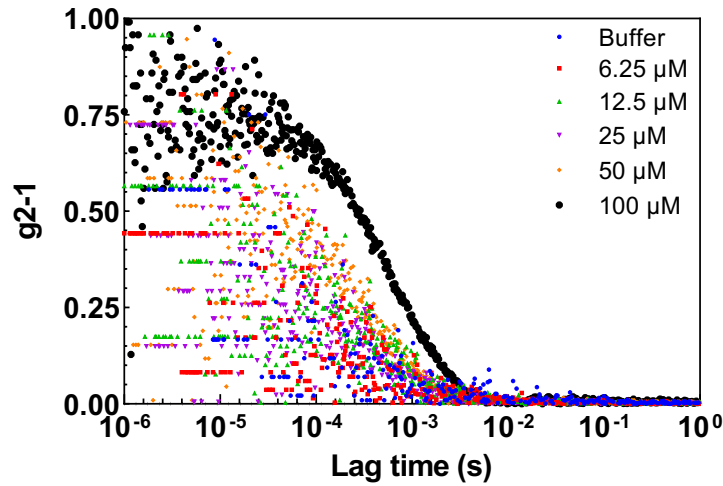


Fig S3. Dynamic Light Scattering of 34RH in 1xPBS and 5% DMSO (buffer) at pH 7.4. Concentrations of 34RH are shown in the upper right corner. Concentrations below 50 μM show no correlation function indicating that there are none to minimal nano-sized particles in the samples. At 100 μM , a correlation was observed indicating that nano-sized particles may exist in solution. All experiments were carried out at 50 μM and below to avoid any potential particle interference in the data.

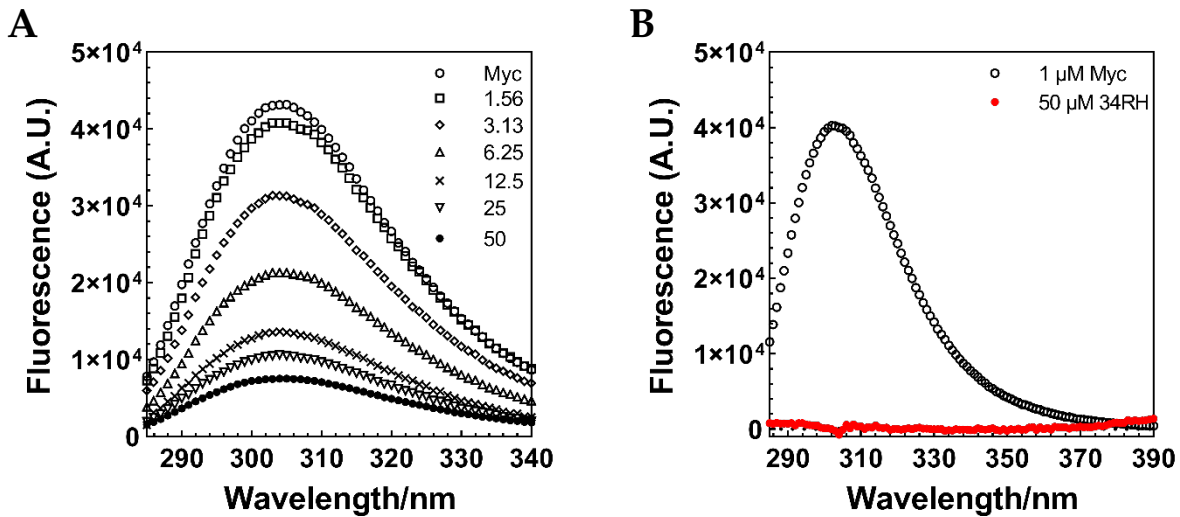


Fig S4. Myc₃₅₃₋₄₃₇ and 34RH interaction monitored via tyrosine fluorescence quenching and free fluorescence spectra of Myc₃₅₃₋₄₃₇ and 34RH. (A) Inner filter corrected fluorescence emission spectra of 1 μM Myc₃₅₃₋₄₃₇ (open circles) and 1 μM Myc₃₅₃₋₄₃₇ with various concentrations of 34RH in 1xPBS, 5% DMSO at 25 $^{\circ}\text{C}$, pH 7.4. Concentrations of 34RH (in micromolar) are shown in the upper right corner. (B) Comparison of inner filter corrected emission spectra of 1 μM Myc₃₅₃₋₄₃₇ (open circles) and 50 μM 34RH without Myc (closed circles) demonstrating no fluorescence contribution from 34RH in this wavelength range.

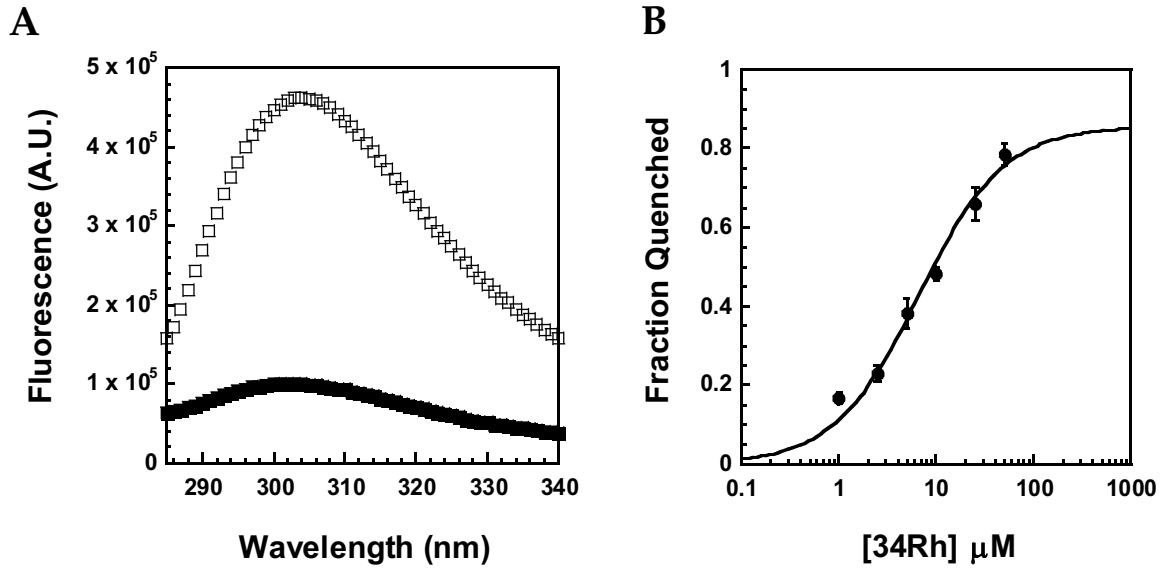


Fig S5. Equimolar Myc₃₅₃₋₄₃₇ and 34RH interaction monitored via tyrosine fluorescence quenching. **(A)** 50 μ M Myc₃₅₃₋₄₃₇ in the presence (filled black squares) and absence (open black squares) equimolar 34RH in 1x PBS at 25 °C, pH 7.4. **(B)** Equilibrium quenching of an equimolar titration of Myc₃₅₃₋₄₃₇ and 34RH fit to a 1:1 binding model, K_D of 5.9 ± 0.8 μ M. Error bars represent the standard error of three independent trials. Representative fluorescence signals were inner filter corrected to account for any fluorescence suppression due the absorbance of protein and 34RH [46]. The quenching data was fitted to a binding model shown below described in Dobrev et al.

$$\text{Fraction Quenched} = Q_{\max} \times \frac{2 + \frac{K_D}{[C]_{\text{tot}}} - \sqrt{\left(-2 - \frac{K_D}{[C]_{\text{tot}}}\right)^2 - 4}}{2}$$

Here, Q_{\max} is the fraction quenched for the formation of the complete complex between Myc₃₅₃₋₄₃₇ and 34RH. $[C]_{\text{tot}}$ is the total concentration of Myc₃₅₃₋₄₃₇ or 34RH and K_D the dissociation constant. [46,49]

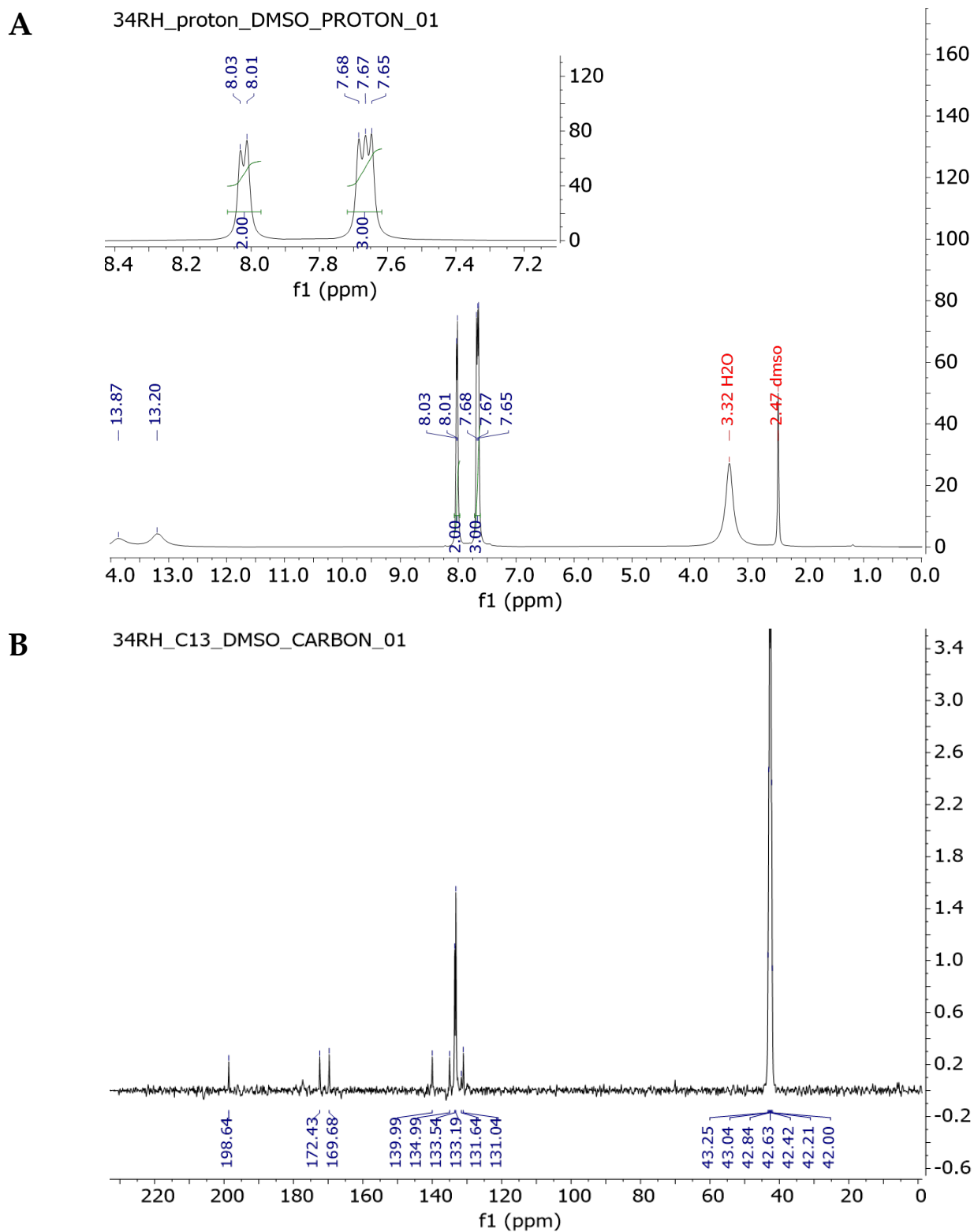


Fig S6. NMR characterization of 34RH **A)** ¹H NMR of 34RH in DMSO-d₆. **B)** ¹³C NMR of 34RH in DMSO-d₆.

References

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