



Review

Classical and Unexpected Effects of Ultra-Micronized PEA in Neuromuscular Function

Pierangelo Cifelli ^{1,*}, Gabriele Ruffolo ^{2,3}, Marco Ceccanti ⁴, Chiara Cambieri ⁴, Laura Libonati ⁴, Eleonora Palma ² and Maurizio Inghilleri ^{4,*}

- ¹ Department of Applied Clinical and Biotechnological Sciences, University of L'Aquila, 67100 L'Aquila, Italy
- Department of Physiology and Pharmacology, Istituto Pasteur-Fondazione Cenci Bolognetti, University of Rome Sapienza, 00185 Rome, Italy; gabriele.ruffolo@uniroma1.it (G.R.); eleonora.palma@uniroma1.it (E.P.)
- ³ IRCCS San Raffaele Roma, 00163 Rome, Italy
- Department of Human Neuroscience, University of Rome Sapienza, 00185 Rome, Italy; marco.ceccanti@uniroma1.it (M.C.); chiara.cambieri@uniroma1.it (C.C.); laura.libonati@uniroma1.it (L.L.)
- * Correspondence: pierangelo.cifelli@univaq.it (P.C.); maurizio.inghilleri@uniroma1.it (M.I.)

Abstract: Recently, the endocannabinoid system has attracted growing attention from the scientific community for its involvement in homeostatic and pathological processes as they pertains to human physiology. Among the constituents of the endocannabinoid system, the molecule palmitoyl ethanolamide has particularly been studied for its ability to reduce several inflammatory processes involving the central nervous system. Here, we reviewed published literature and summarized the main targets of the palmitoyl ethanolamide, along with its unique possible mechanisms for restoring correct functioning of the central nervous system. Moreover, we have highlighted a less-known characteristic of palmitoyl ethanolamide, namely its ability to modulate the function of the neuromuscular junction by binding to acetylcholine receptors in different experimental conditions. Indeed, there are several studies that have highlighted how ultra-micronized palmitoyl ethanolamide is an interesting nutraceutical support for the treatment of pathological neuromuscular conditions, specifically when the normal activity of the acetylcholine receptor is altered. Although further multicentric clinical trials are needed to confirm the efficacy of ultra-micronized palmitoyl ethanolamide in improving symptoms of neuromuscular diseases, all the literature reviewed here strongly supports the ability of this endocannabinoid-like molecule to modulate the acetylcholine receptors thus resulting as a valid support for the treatment of human neuromuscular diseases.

Keywords: palmitoyl ethanolamide; neuromuscular junction; neurophysiology



Citation: Cifelli, P.; Ruffolo, G.; Ceccanti, M.; Cambieri, C.; Libonati, L.; Palma, E.; Inghilleri, M. Classical and Unexpected Effects of Ultra-Micronized PEA in Neuromuscular Function. *Biomolecules* 2022, 12, 758. https://doi.org/ 10.3390/biom12060758

Academic Editors: Salvatore Cuzzocrea and Rosalia Crupi

Received: 9 May 2022 Accepted: 27 May 2022 Published: 29 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Palmitoyl ethanolamide [PEA] represents the best-known endocannabinoid-like molecule of the "autacoid local injury antagonist amide" [ALIAmide] family. ALIAmides are endogenous N-acyl ethanol amines [NAEs] with homeostatic functions able to modulate different physiological pathways involved in inflammation, pain, and metabolism [1]. Among the different ALIAmides, discovered up to now, PEA, oleoyl ethanol amide (OEA), stearoyl ethanol amide (SEA), and anandamide (ANA) represent the most-studied compounds, while others such as eicosatrienoyl ethanol amide (ETEA) still need to be investigated further. In this review we will highlight the different mechanisms of action of PEA, an endocannabinoid [eCB]-like bioactive lipid mediator, and its involvement in different human pathologies. Finally, we will place under the spotlight PEA's ability to modulate the synaptic function in the neuromuscular system and at neuromuscular junction [NMJ].

2. What, Where and Why?

NAEs are endogenous bioactive lipids characterized by common precursors, namely the N-acylated ethanolaminephospholipids such as N-acylphosphatidylethanolamines Biomolecules **2022**, 12, 758 2 of 11

(NAPEs) and N-acylplasmenylethanolamines (pNAPEs). The biological targets and the specific functions of the different NAEs depend upon the origin of the N-acyl group, which can arise from a variety of different fatty acids [2]. The endogenous levels of NAEs in different tissues are mainly regulated by enzymatic reactions responsible for both their biosynthesis and degradation. Two main enzyme reactions are involved in NAEs biosynthesis. The first is the process of N-acylation of ethanolamine phospholipids mediated by a Ca²⁺-dependent N-acyltransferase, while the second is represented by the release of the different NAEs from NAPEs, mediated by NAPE-hydrolyzing phospholipase (NAPE-PLD) [3]. Catabolism of NAEs is rapidly put in effect by the action of the free acid amide hydroxylase [FAAH], an integral membrane enzyme able to hydrolyze them in their corresponding fatty acids and ethanolamine [4].

Among NAEs, PEA was first identified in mammalian tissues in 1965 [5]. In central nervous system [CNS] it is produced and released both from neurons and glial cells as an "on-demand molecule" to promote endogenous neuroprotection following tissue damage and the subsequent processes of neuroinflammation. PEA is part of the endocannabinoid system (ECS), a regulatory signaling network made up of lipidic molecules, the endocannabinoids (eCBs), their receptors, and different enzymes involved in the metabolism and catabolism of the lipidic signals. The ECS is strongly involved in different homeostatic processes of our body, both in physiological and pathological conditions [6]. Indeed, the ECS is implicated in the regulation of several functions, such as the cycle of sleep, the regulation of food intake, and the perception of painful stimuli and in high cognitive functions such as mood regulation, memory, and reward [7].

As mentioned, PEA can carry out a plethora of actions, and its tissue levels are often up regulated in several pathological conditions [8]. Its capacity to modulate divergent pathways is largely due to its ability to interact, directly or indirectly, with different targets [9]. The main target of PEA is represented by the nuclear peroxisome proliferator-activated receptor- α (PPAR- α) [10]. This receptor, part of the larger family of nuclear receptor proteins, has the capability, when activated, to modulate and regulate the expression of different genes involved in inflammation [11]. Indeed, the activation of PPAR- α receptors induces and promotes a cascade of events leading to inhibition and suppression of the release of pro-inflammatory cytokines, such as Interleukin-1β (IL-1β), Interleukin 6 (IL6), and the tumor-necrosis factor (TNF- α) [12]. All these actions were further confirmed in several pre-clinical studies, where the anti-inflammatory effect of PEA was totally abolished when applied on PPAR- α knockout models of disease [12]. Another target activated by PEA is represented by GPR55 and GPR119, two orphan receptors [13], considered up to now as novel cannabinoid receptors [14]. PEA is also able to activate and modulate the transient receptor potential vanilloid receptor 1 (TRPV1) channels, accounting thus, at least in part, for its anti-nociceptive effect [15]. Furthermore, PEA can indirectly activate the canonical cannabinoid receptors, namely CB1 and CB2, increasing the level of anandamide (AEA) by inhibiting its enzymatic-mediated hydrolysis carried out by fatty acid amide hydrolase (FAAH) [16].

One of the main targets of PEA is represented by the resident immune cells of CNS, namely microglial cells. These latter are responsible for CNS homeostasis, being able to modify their function following CNS injuries [17]. Indeed, after multiple insults, [e.g., inflammation, tumors, and traumas], they can switch their phenotype between the M1, considered unanimously the pro-inflammatory, and the M2, considered instead as anti-inflammatory [18]. The imbalance between the two states is considered as one of the main events underlying many neurogenerative diseases, where M1 phenotypes are found in a condition of continuous activation [19]. In this state, M1-activated microglia releases many inflammatory cytokines, such as t TNF- α , IL-1 β , and inducible reactive oxygen species (iROS), which lead the neuronal population to death [19]. Recent studies demonstrated that PEA can modulate microglia polarization, reducing the release of pro-inflammatory cytokines and promoting migration and phagocytic activity [20]. In detail, PEA can reduce the lipopolysaccharide (LPS)-induced microglial activation by switching their phenotypic

Biomolecules 2022, 12, 758 3 of 11

constructs from M1 to M2, thus inhibiting the release of pro-inflammatory elements in the extracellular milieu and reducing the hyperexcitability of cultured primary cortical cells together with LPS-activated microglial cells [21]. Notably, these effects were all mediated by cannabinoid receptor type 2 (CB2), thus confirming the strong involvement of the ECS during inflammatory processes [22].

3. Pharmacology of PEA

PEA is a lipophilic molecule with low solubility rate. Hence, specific formulations, such as micronized and ultra-micronized PEA, have been used to increase its solubility and enhance its bioavailability. Indeed, with these techniques PEA particles became smaller, thus increasing surface areas to improve absorption [23]. Different studies highlight that ultra-micronized PEA has greater absorbability than micronized and non-micronized PEA, also showing better results in terms of reducing pain perception [24–26] Another issue about PEA is that its bioavailability is related to its possible pre-systemic metabolism. Since enzymes involved in its metabolism are expressed both in the small intestine and in the liver [27], it is quite difficult to calculate its range of distribution and how this parameter may change in disparate individuals.

Due to its partially unclear mechanisms of absorption and pre-systemic metabolism, the distribution mechanism of PEA is still far from being elucidated. However, due to its lipophilic nature, several studies demonstrated that, even with its low level of bioavailability, PEA its able to reach tissues to target its receptors effectively [28,29]. Currently, the only way to characterise its absorption is by measuring its blood levels after administration [28].

Once PEA reaches the tissues, there is a fast cellular uptake in order to expose PEA to its intracellular PPAR- α receptors. This crucial step needs to be further deciphered, as research pertaining to its mechanism is still lacking. It appears that the intracellular process of uptake is strictly regulated by PEA hydrolysis, to balance its extracellular/intracellular ratio [30]. In contrast with data about absorption and availability, studies about PEA metabolism are abundant in literature. Indeed, PEA is hydrolysed in two compounds, namely palmitic acid and ethanolamine by the fatty acid amide hydrolase [FAAH] enzyme, located on endoplasmic reticulum membranes [31,32]. Subsequently, another enzyme involved in PEA hydrolysis was found and described: the FAAH-2 [33]. Notably, both FAAH and FAAH-2 are able to hydrolase AEA faster than PEA, while the third enzyme involved in PEA catabolism, namely N-acylethanolamine acid amidase (NAAA), is able to hydrolase more efficiently PEA than AEA [34,35].

The reason why three distinct enzymes are involved in PEA catabolism is still obscure. Nevertheless, recent works have highlighted the possibility that it may depend on the source of PEA, with differences between endogenous and exogenous PEA. Endogenous PEA is hydrolysed by means of both FAAH and NAAA, but several studies indicate that the prevalence of one or the other is strongly tissue-specific and may change also in different pathological conditions [36].

Alterations were described when catabolism of exogenous PEA was scrutinized; indeed, the fatty acid binding proteins do not deliver exogenous PEA to lysosomal NAAA, thus suggesting a prevalent role of FAAH for exogenous PEA metabolism [37].

The catabolism of the main metabolite of PEA, palmitic acid, was fully described by Carta et al. in 2017 [38]. However, data describing the excretion of unmetabolized PEA is still lacking, even though the kidney route seems to be prominent [5].

4. PEA, a Multi Target Drug for Different Clinical Applications

Since, as mentioned above, PEA can bind different receptor targets and is thus able to modulate different signalling pathways, its rationale for clinical uses embraces a wide variety of pathological conditions, from chronic inflammations to perseverant pain conditions. Here we will address the main applications in routinely clinical practice with a particular focus on neurological conditions.

Biomolecules 2022, 12, 758 4 of 11

5. PEA and Neuroinflammation

Nowadays, neuroinflammation is considered a key element in the pathogenesis of a broad spectrum of neurological diseases, from neurodegenerative conditions, such as Alzheimer's disease [39–43] and Parkinson's disease [44], to traumatic diseases, such as traumatic brain injuries [TBI] [45–49], strokes [50], and other conditions where neuronal excitability is increased, such as epilepsy [51]. Moreover, recently, attention has been called to the significant effect of PEA in clinical conditions involving the higher functions of CNS, such as cognitive impairment [52] and mood disorders [53]. In all these conditions, microglial cells, together with mast cells and astrocytes, play a crucial role in inducing and maintaining the inflammatory processes, leading to neuronal cell death, and thus impairing the CNS functions. PEA capacity to dampen neuroinflammation could represent a viable tool to preserve neuronal populations and to retain physiological brain functions [54,55]. Most of PEA's effects are due, as already mentioned, to its ability to modulate and modify microglial, astrocyte and mast cells activation [56,57]. Indeed, PEA can enhance microglial migration without switching their phenotype towards the pro-inflammatory M1 state, thus increasing the resistance to infections without activation of inflammatory cascade [57]. PEA is able to carry out its neuroprotective effects on CNS also with other mechanisms, like for example, inhibiting the apoptosis processes and modulating different pathways, such as the bax/bcl-2 and Akt/mTOR/p70S6K pathways [58]. PEA is also able to target and modulate NMDA receptors, thus protecting cells for glutamate toxicity [59]. PEA neuroprotective actions are also carried out by promoting and modulating synaptic homeostasis and favouring neurogenesis [60–62]. Notably, all these PEA-mediated actions were confirmed using different animal models of disease characterized by neuroinflammation. Chronic administration of PEA at different concentrations (ranging from 10 to 100 mg/Kg/day) was able to significantly reduce neuroinflammation [63–65], to protect neurons from death [21,66], to reduce iROS productions [66], and to improve behavioural, motor and cognitive deficits [67-69].

6. PEA in Central Nervous System Diseases

It is well known that both ECS and NAEs are strongly involved in regulation and modulation of behaviour, cognition and in mood regulation [70]. Plasmatic PEA levels are significantly altered in conditions as post-traumatic stress disease (PTSD) [71], depression [72,73], and in autism spectrum disorders (ASD) [74]. The mechanisms by which PEA can improve these conditions are different and include increased hippocampal neurogenesis [59–61] and increased maturation of oligodendrocyte precursor cells [59]. PEA was able to prevent the plasmatic reduction of the brain-derived neurotrophic factor (BDNF) and glial cell line-derived neurotrophic factor (GDNF) in a murine model of stroke [75], thus explaining, at least in part, its neuroprotective effects. Interestingly, PEA is also strongly involved in the neuromodulation of GABAergic transmission. As a matter of fact, PEA by binding GPR55 receptors is able to enhance GABAergic transmission in the corpus striatum and to increase the synthesis of the endocannabinoid 2-AG at postsynaptic level thus modulating GABA release through the stimulation of presynaptic CB1Rs [76]. These biological effects may partially explain the neuroprotective action of PEA and its potential use to treat these patients in clinical activities. To confirm the pre-clinical results obtained by using PEA, human studies were performed on different pathological diseases. In a yearlong study, PD patients used ultra-micronized PEA as add-on therapy along with levodopa, showing a significant improvement on both motor- and non-motor-symptoms, ameliorating the typical mood deficits of Parkinsonian patients as well as reducing fatigue and improving sleep-cycle coupled with improved responses to different mental tasks [77]. Notably, in a double-blind, randomized, placebo-controlled study performed on patients with major depressive disorder [MDD], 600 mg of ultra-micronized PEA used as add-on therapy with typical selective serotonin reuptake inhibitors [SSRI] for 6 weeks was able to significantly improve the depressive scores, ameliorating symptoms when compared to SSRI plus placebo group [78]. It is important to note that, also if further clinical studies are

Biomolecules **2022**, 12, 758 5 of 11

required in all the human studies performed to date, PEA was well tolerated and displayed only few and minor adverse effects [79].

7. Analgesic Properties of PEA

The direct link between inflammation and pain kicked off a series of studies with the aim to emphasize PEA analgesic properties. Taking advantage of pre-clinical models of inflammation and neuropathic pain, it was possible to demonstrate the strong analgesic properties of PEA [80-82]. When inflammation processes persist, PEA levels increase as a protective mechanism to inhibit inflammatory pathways leading to tissue damage and loss of functions. If the stimulus is protracted, PEA levels start to decrease, thus indicating that PEA's increased concentration is not enough to restore the physiological conditions when inflammation becomes chronic [83]. The PEA's analgesic properties are guaranteed by several mechanisms. For instance, its ability to directly bind to PPAR- α and GPR55 receptors or to indirectly bind to CB1, CB2, and TRPV1 receptors may partially account for its properties [15]. The analgesic properties of PEA also take into account its capacity to suppress inflammation by reducing mast cells activation and to reduce the production and the release of different inflammatory mediators such as nerve growth factor (NGF), cyclo-oxygenase-2 (COX-2), TNF- α , and iROS [84]. All these pathways are responsible for the preservation of peripheral nerve morphology and for reducing inflammation-coupled oedema and microglial activation [85,86]. In addition, several preclinical and clinical studies confirm the anti-nociceptive properties of PEA in neuropathic pain [87,88]. PEA treatment was also able to preserve Langerhans islets' morphology in the pancreatic gland of diabetic mice [89]. Orefice et al., in a randomized, double-blind, placebo-controlled study, demonstrated that ultra-micronized PEA as add-on therapy in patients afflicted by multiple sclerosis was able to significantly reduce the pain sensation and to improve their quality of life [90].

Notably, currently available drugs used to treat inflammation and pain-related conditions often display several adverse effects, especially when used chronically, such gastrointestinal, hepatic, renal, and cardiovascular disorders. PEA thus represents a promising alternative treatment, since it shows a high profile of safety and tolerability [78].

8. PEA in Neuromuscular Alterations and Diseases

The first study that hypothesized a potential use of ultra-micronized PEA in pathological conditions involving neuromuscular transmission was published in 2012 by Clemente who described the improvement of different vital parameters in a single case of a patient affected by amyotrophic lateral sclerosis (ALS), a neurodegenerative disorder able to disrupt the physiological function of the neuromuscular junction leading, in a few years, to muscle paralysis and death of the patients [85]. Up to now, there are several studies that highlight some of the possible mechanisms involved in this pathology, but unfortunately, ALS is still a disease with a poor prognosis lacking effective therapies and treatments in spite of recent efforts to develop new therapeutic strategies [91,92]. The degeneration of the second motor neuron is considered a hallmark of this disease, but different studies performed on animal models of ALS highlighted that the first alterations are localized at the neuromuscular junction (NMJ) [93,94]. Indeed, it was described that, even during the pre-symptomatic period, changes in NMJ function and muscular functions are present, thus leading to the "dying-back" theory [95]. According to this hypothesis, the first pathogenic step in ALS is represented by strong functional alterations at the NMJ level leading subsequently to the progression of the disease involving motor neuron degeneration [96], muscle atrophy, mitochondrial dysfunction, microgliosis, and neuronal degeneration [97]. To strengthen this hypothesis, it was demonstrated that muscular acetylcholine receptors (AChRs) obtained from muscle biopsy of ALS patients are functionally impaired when compared with AChRs obtained from non-ALS denervated muscle, showing also a reduction of sensitivity to ACh 98]. In detail, it is well known that after denervation, muscular AChRs subunits' composition undergoes modification, with a wide up-regulation of the $\alpha 1\beta 1\gamma \delta$ (γ -AChR) Biomolecules **2022**, 12, 758 6 of 11

on the muscular sarcolemma. Indeed, in ALS muscles where both denervation and aberrant reinnervation are present, both the γ -AChR and the ϵ -AChRs are expressed. It was demonstrated that PEA can significantly reduce the rate of AChR desensitization of receptors obtained from ALS patients, during repetitive application of ACh [98]. In detail PEA was able to reduce this functional property predominantly on the ε-AChRs form, while this effect was to a lesser extent on γ -AChR [98]. To further strengthen this observation in the same study, an observational double-blind clinical test was performed on a cohort of ALS patients recruited by the neurological unit at the Policlinico Umberto I in Rome headed by Inghilleri. In detail, 28 patients were co-treated with riluzole (at the time of the study, the only officially approved treatment) [99] plus ultra-micronized PEA twice a day (50 mg and 600 mg, respectively) and compared with 36 patients receiving only riluzole. The two groups of patients were monitored, and clinical and electrophysiological test were assessed. Interestingly, ultra-micronized PEA-treated patients showed a slow respiratory impairment and a delayed need of tracheotomy. This clinical improvement was associated with a reduced decay of the forced vital capacity (FVC) in comparison with patients treated only with riluzole [100]. Therefore, the authors suggested that the PEA's effect on FVC is due to its ability to reduce the muscle AChRs desensitization induced by repetitive stimulation, as in respiratory muscles. Taken together, data obtained from this study showed that PEA add-on therapy slows the disease progression, suggesting that PEA could represent a valid aid to slow respiratory impairment in these patients, thus increasing their life expectancy. In another open-label pilot study, also carried out by Inghilleri's group at the Policlinico Umberto I in Rome, oral ultra-micronized PEA was administered to a cohort of patients afflicted by myasthenia gravis (MG) [101]. This autoimmune disease is characterized by the presence of auto-antibodies able to disrupt the physiological function of NMJ. Symptoms are often variable, from mild deficits limited to single muscle groups to generalized weakness involving vital functions. However, the main symptom is muscular fatigue, which causes characteristic fluctuations in symptoms that worsen in the evening and improve after periods of rest. The progression of the disease can be sneaky, with long periods of spontaneous remission or rapid progression [102]. In the aforementioned study [101], after only one week of treatment, patients displayed a significant improvement of all the neurophysiological parameters taken in account. In detail, ultra-micronized PEA induced a statistically significant improvement of the quantitative myasthenia gravis score (QMG), thus reducing the level of disability of the treated patients, and also a significant effect in improving the pathological muscle responses (measured as "decremental muscle responses", RNS). Since these positive effects appeared after only one week of treatment, it is not surprising that they disappeared one week after the withdrawal of the ultra-micronized PEA. Notably, the antibody titre did not significantly change following PEA treatment [101], suggesting a possible direct effect of ultra-micronized PEA on nAChRs as already shown in ALS patients. The capacity of PEA to reduce the release of pro-inflammatory cytokines could also be exploited to treat diseases such as sarcopenia, a condition characterized by progressive and generalized loss of skeletal muscle mass and strength combined with low physical performance [103]. Recently, new efforts have been made to improve PEA bioavailability, in order to better reach peripheral cells such as muscle cells [104], opening thus new scenarios in the treatments of diseases characterized by strong inflammation. On the other hand, new insights on the use of ultra-micronized PEA on inflammatory pathways modulation came from studies using PEA in association with antioxidant agents, such as phycocyanin extract (PC) from spirulina algae [66], that further enhanced PEA's beneficial properties. Indeed, a multi-center, double-blinded, randomized placebo-controlled clinical trial showed that co-administration of PEA plus Luteolin was able to significantly improve the recovery of olfactory function in patients after SARS-CoV-2 infection [105].

Biomolecules **2022**, 12, 758 7 of 11

9. Conclusions

PEA signifies a valid support in different pathological conditions, since it displays a wide range of positive effects without noteworthy side effects. Until now, PEA has been extensively used to treat and ameliorate conditions characterized by neuroinflammation and to treat and reduce the perception of neuropathic pain. It is possible to find separate commercially available PEA formulations, (micronized, ultra-micronized), which increase its solubility and bioavailability, and in doses ranging from 200 mg to up to 1000 mg. While its neuroprotective effect is well known, its capacity to interact with the NMJs deserves further consideration, as it could pave the way for novel therapeutic implications. In detail, the ability to modulate the AChR function could open up new perspectives for its use in different pathological conditions characterized by alterations of NMJ, such as ALS, but also in other diseases such as MG where AChRs function is strongly altered. Further studies including larger double-blind multicentre clinical studies are required in order to find the best medical approach for any condition taken into consideration.

Author Contributions: Conceptualization, E.P. and M.I.; writing—original draft preparation, P.C., G.R., M.C., E.P. and M.I.; writing—review and editing, P.C., G.R., M.C., C.C., L.L., E.P. and M.I.; funding acquisition, P.C., E.P. and M.I. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the Fondazione Italiana di Ricerca per la Sclerosi Laterale Amiotrofica (AriSLA Grant "MUSALS-AChR") to E.P. and M.I. This research was supported by intramural "DISCAB" GRANT 2022 code 07_DG_2022_05 to P.C. G.R. was supported by the Italian Ministry of Health "Ricerca Corrente". The work was supported by grants from the Ateneo Project (Sapienza University)", grant n° RM11916B84D24429 to E.P.

Acknowledgments: The authors thank Epitech group SpA for the ongoing support to this research. Authors also thank Anwesha Ghosh for English proofreading.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Aloe, L.; Leon, A.; Levi-Montalcini, R. A proposed autacoid mechanism controlling mastocyte behaviour. Agents Actions 1993, 39, C145–C147. [CrossRef] [PubMed]
- 2. Wellner, N.; Diep, T.A.; Janfelt, C.; Hansen, H.S. N-acylation of phosphatidylethanolamine and its biological functions in mammals. *Biochim. Biophys Acta.* **2013**, *1831*, 652–662. [CrossRef] [PubMed]
- 3. Rahman, I.A.S.; Tsuboi, K.; Uyama, T.; Ueda, N. New players in the fatty acyl ethanolamide metabolism. *Pharmacol. Res.* **2014**, *86*, 1–10. [CrossRef] [PubMed]
- 4. Deutsch, D.G.; Ueda, N.; Yamamoto, S. The fatty acid amide hydrolase (FAAH). *Prostaglandins Leukot Essent Fat. Acids* **2002**, *66*, 201–210. [CrossRef] [PubMed]
- 5. Rankin, L.; Fowler, C.J. The Basal Pharmacology of Palmitoylethanolamide. Int. J. Mol. Sci. 2020, 21, 7942. [CrossRef]
- 6. Di Marzo, V.; Piscitelli, F. The endocannabinoid system and its modulation by phytocannabinoids. *Neurotherapeutics* **2015**, 12, 692–698. [CrossRef]
- 7. PPacher, P.; Bátkai, S.; Kunos, G. The endocannabinoid system as an emerging target of pharmacotherapy. *Pharmacol. Rev.* **2006**, *58*, 389–462. [CrossRef]
- 8. Raso, G.M.; Russo, R.; Calignano, A.; Meli, R. Palmitoylethanolamide in CNS health and disease. *Pharmacol. Res.* **2014**, *86*, 32–41. [CrossRef]
- 9. Di Marzo, V.; Melck, D.; Bisogno, T.; de Petrocellis, L. Endocannabinoids: Endogenous cannabinoid receptor ligands with neuromodulatory action. *Trends Neurosci.* **1998**, *21*, 521–528. [CrossRef]
- 10. Verme, J.L.; Fu, J.; Astarita, G.; la Rana, G.; Russo, R.; Calignano, A.; Piomelli, D. The Nuclear Receptor Peroxisome Proliferator-Activated Receptor-α Mediates the Anti-Inflammatory Actions of Palmitoylethanolamide. *Mol. Pharmacol.* **2004**, *67*, 15–19. [CrossRef]
- 11. Bougarne, N.; Weyers, B.; Desmet, S.J.; Deckers, J.; Ray, D.W.; Staels, B.; de Bosscher, K. Molecular actions of PPARα in lipid metabolism and inflammation. *Endocr. Rev.* **2018**, *39*, 760–802. [CrossRef] [PubMed]
- 12. Verme, J.L.; Russo, R.; la Rana, G.; Fu, J.; Farthing, J.; Mattace-Raso, G.; Meli, R.; Hohmann, A.; Calignano, A.; Piomelli, D. Rapid broad-spectrum analgesia through activation of peroxisome proliferator-activated receptor-alpha. *J. Pharmacol. Exp. Ther.* **2006**, 319, 1051–1061.
- 13. Ryberg, E.; Larsson, N.; Sjögren, S.; Hjorth, S.; Hermansson, N.-O.; Leonova, J.; Elebring, T.; Nilsson, K.; Drmota, T.; Greasley, P.J. The orphan receptor GPR55 is a novel cannabinoid receptor. *Br. J. Pharmacol.* **2009**, *152*, 1092–1101. [CrossRef] [PubMed]

Biomolecules 2022, 12, 758 8 of 11

- 14. Brown, A.J. Novel cannabinoid receptors. Br. J. Pharmacol. 2007, 152, 567–575. [CrossRef]
- 15. Costa, B.; Comelli, F.; Bettoni, I.; Colleoni, M.; Giagnoni, G. The endogenous fatty acid amide, palmitoylethanolamide, has antiallodynic and anti-hyperalgesic effects in a murine model of neuropathic pain: Involvement of CB1, TRPV1 and PPARγ receptors and neurotrophic factors. *Pain* **2008**, *139*, 541–550. [CrossRef]
- 16. Deutsch, D.; Glaser, S.; Howell, J.; Kunz, J.; Puffenbarger, R.; Hillard, C.; Abumrad, N. The cellular uptake of anandamide is coupled to its breakdown by fatty-acid amide hydrolase. *J. Biol. Chem.* **2001**, *276*, 6967–6973. [CrossRef]
- 17. Kettenmann, H.; Hanisch, U.-K.; Noda, M.; Verkhratsky, A. Physiology of microglia. Physiol. Rev. 2011, 91, 461–553. [CrossRef]
- 18. Kigerl, K.A.; Gensel, J.C.; Ankeny, D.P.; Alexander, J.K.; Donnelly, D.J.; Popovich, P.G. Identification of two distinct macrophage subsets with divergent effects causing either neurotoxicity or regeneration in the injured mouse spinal cord. *J. Neurosci. Off. J. Soc. Neurosci.* 2009, 29, 13435–13444. [CrossRef]
- 19. Ransohoff, R.M.; Perry, V.H. Microglial physiology: Unique stimuli, specialized responses. *Annu Rev. Immunol.* **2009**, 27, 119–145. [CrossRef]
- Cordaro, M.; Cuzzocrea, S.; Crupi, R. An Update of Palmitoylethanolamide and Luteolin Effects in Preclinical and Clinical Studies
 of Neuroinflammatory Events. Antioxidants 2020, 9, 216. [CrossRef]
- 21. D'Aloia, A.; Molteni, L.; Gullo, F.; Bresciani, E.; Artusa, V.; Rizzi, L.; Ceriani, M.; Meanti, R.; Lecchi, M.; Coco, S.; et al. Palmitoylethanolamide Modulation of Microglia Activation: Characterization of Mechanisms of Action and Implication for Its Neuroprotective Effects. *Int. J. Mol. Sci.* 2021, 22, 3054. [CrossRef] [PubMed]
- 22. Guida, F.; Luongo, L.; Boccella, S.; Giordano, M.E.; Romano, R.; Bellini, G.; Manzo, I.; Furiano, A.; Rizzo, A.; Imperatore, R.; et al. Palmitoylethanolamide induces microglia changes associated with increased migration and phagocytic activity: Involvement of the CB2 receptor. *Sci. Rep.* 2017, 7, 375. [CrossRef] [PubMed]
- 23. Petrosino, S.; di Marzo, V. The pharmacology of palmitoylethanolamide and first data on the therapeutic efficacy of some of its new formulations. *Br. J. Pharmacol.* **2017**, *174*, 1349–1365. [CrossRef] [PubMed]
- 24. Petrosino, S.; Cordaro, M.; Verde, R.; Moriello, A.S.; Marcolongo, G.; Schievano, C.; Siracusa, R.; Piscitelli, F.; Peritore, A.; Crupi, R.; et al. Oral Ultramicronized Palmitoylethanolamide: Plasma and Tissue Levels and Spinal Anti-hyperalgesic Effect. *Front Pharmacol.* **2018**, *9*, 249. [CrossRef]
- 25. Impellizzeri, D.; Bruschetta, G.; Cordaro, M.; Crupi, R.; Siracusa, R.; Esposito, E.; Cuzzocrea, S. Micronized/ultramicronized palmitoylethanolamide displays superior oral efficacy compared to nonmicronized palmitoylethanolamide in a rat model of inflammatory pain. *J. Neuroinflamm.* 2014, 11, 136. [CrossRef]
- 26. Fusco, R.; Gugliandolo, E.; Campolo, M.; Evangelista, M.; di Paola, R.; Cuzzocrea, S. Effect of a new formulation of micronized and ultramicronized N-palmitoylethanolamine in a tibia fracture mouse model of complex regional pain syndrome. *PLoS ONE* **2017**, 12, e0178553. [CrossRef]
- Schmid, P.; Zuzarte-Augustin, M.; Schmid, H. Properties of rat liver N-acylethanolamine amidohydrolase. J. Biol. Chem. 1985, 260, 14145–14149. [CrossRef]
- 28. Vacondio, F.; Bassi, M.; Silva, C.; Castelli, R.; Carmi, C.; Scalvini, L.; Lodola, A.; Vivo, V.; Flammini, L.; Barocelli, E.; et al. Amino acid derivatives as palmitoylethanolamide prodrugs: Synthesis, in vitro metabolism and in vivo plasma profile in rats. *PLoS ONE* **2015**, *10*, e0128699. [CrossRef]
- 29. Bracey, M.; Hanson, M.; Masuda, K.; Stevens, R.; Cravatt, B. Structural adaptions in a membrane enzyme that terminates endocannabinoid signaling. *Science* **2002**, *298*, 1793–1796. [CrossRef]
- 30. Boger, D.; Fecik, R.; Patterson, J.; Miyauchi, H.; Patricelli, M.; Cravatt, B. Fatty acid amide hydrolase substrate specificity. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 2613–2616. [CrossRef]
- 31. Tsuboi, K.; Sun, Y.-X.; Okamoto, Y.; Araki, N.; Tonai, T.; Ueda, N. Molecular characterization of N-acylethanolamine-hydro lyzing acid amidase, a novel member of the choloylglycine hydrolase family with structural and functional similarity to acid ceramidase. *J. Biol. Chem.* 2005, 280, 11082–11092. [CrossRef] [PubMed]
- 32. Ueda, N.; Yamanaka, K.; Yamamoto, S. Purification and characterization of an acid amidase selective for N-palmitoylethan olamine, a putative endogenous anti-inflammatory substance. *J. Biol. Chem.* **2001**, *276*, 35552–35557. [CrossRef] [PubMed]
- 33. Wei, B.; Mikkelsen, T.; McKinney, M.; Lander, E.; Cravatt, B. A second fatty acid amide hydrolase with variable distribution among placental mammals. *J. Biol. Chem.* **2006**, *281*, 36569–36578. [CrossRef] [PubMed]
- 34. Bonezzi, F.T.; Sasso, O.; Pontis, S.; Realini, N.; Romeo, E.; Ponzano, S.; Nuzzi, A.; Fiasella, A.; Bertozzi, F.; Piomelli, D. An important role for N-acylethanolamine acid amidase in the complete Freund's adjuvant rat model of arthritis. *J. Pharmacol. Exp. Ther.* **2016**, *356*, 656–663. [CrossRef]
- 35. Artamonov, M.; Zhukov, O.; Shuba, I.; Storozhuk, L.; Khmel, T.; Klimashevsky, V.; Mikosha, A.; Gula, N. Incorporation of labelled N-acylethanolamine (NAE) into rat brain regions in vivo and adaptive properties of saturated NAE under X-ray irradiation (1999). *Ukr. Biokhimicheskii Zhurnal* **2005**, 77, 51–62.
- 36. Jhaveri, M.; Richardson, D.; Robinson, I.; Garle, M.; Patel, A.; Sun, Y.; Sagar, D.; Bennett, A.; Alexander, S.; Kendall, D.; et al. Inhibition of fatty acid amide hydrolase and cyclooxygenase-2 increases levels of endocannabinoid related molecules and pro duces analgesia via peroxisome proliferator-activated receptor-alpha in a model of inflammatory pain. Neuropharmacology 2008, 55, 85–93. [CrossRef]
- 37. Kaczocha, M.; Glaser, S.T.; Chae, J.; Brown, D.A.; Deutsch, D.G. Lipid droplets are novel sites of N-acylethanolamine inacti vation by fatty acid amide hydrolase-2. *J. Biol. Chem.* **2010**, *285*, 2796–2806. [CrossRef]

Biomolecules **2022**, 12, 758 9 of 11

38. Carta, G.; Murru, E.; Bani, S.; Manca, C. Palmitic acid: Physiological role, metabolism and nutritional implications. *Front. Physiol.* **2017**, *8*, 902. [CrossRef]

- 39. Scuderi, C.; Stecca, C.; Valenza, M.; Ratano, P.; Bronzuoli, M.R.; Bartoli, S.; Steardo, L.; Pompili, E.; Fumagalli, L.; Campolongo, P.; et al. Palmitoylethanolamide controls reactive gliosis and exerts neuroprotective functions in a rat model of Alzheimer's dis ease. *Cell Death Dis.* **2014**, *11*, e1419. [CrossRef]
- 40. Bronzuoli, M.R.; Facchinetti, R.; Steardo, L.; Romano, A.; Stecca, C.; Passarella, S.; Cassano, T.; Scuderi, C. Palmitoylethanola mide Dampens Reactive Astrogliosis and Improves Neuronal Trophic Support in a Triple Transgenic Model of Alzheimer's Disease: In Vitro and In Vivo Evidence. *Oxid Med. Cell Longev.* 2018, 2018, 4720532. [CrossRef]
- 41. Beggiato, S.; Tomasini, M.C.; Ferraro, L. Palmitoylethanolamide (PEA) as a Potential Therapeutic Agent in Alzheimer's Disease. *Front. Pharmacol.* **2019**, 24, 821. [CrossRef] [PubMed]
- 42. D'Agostino, G.; Russo, R.; Avagliano, C.; Cristiano, C.; Meli, R.; Calignano, A. Palmitoylethanolamide protects against the amyloidβ25-35-induced learning and memory impairment in mice, an experimental model of Alzheimer disease. *Neuropsy Chopharmacol.* **2012**, *37*, 1784–1792. [CrossRef] [PubMed]
- 43. Scuderi, C.; Steardo, L. Neuroglial roots of neurodegenerative diseases: Therapeutic potential of palmitoylethanolamide in models of Alzheimer's disease. CNS. Neurol. Disord. Drug. Targets 2013, 12, 62–69. [CrossRef] [PubMed]
- 44. Esposito, E.; Impellizzeri, D.; Mazzon, E.; Paterniti, I.; Cuzzocrea, S. Neuroprotective activities of palmitoylethanolamide in an animal model of Parkinson's disease. *PLoS ONE* **2012**, *7*, e41880. [CrossRef]
- 45. Esposito, E.; Cuzzocrea, S. Palmitoylethanolamide in homeostatic and traumatic central nervous system injuries. *CNS Neu. rol. Disord. Drug Targets* **2013**, *12*, 55–61. [CrossRef]
- 46. De Palma, L.; Santamato, A.; Montillo, V.; Fiore, P. Co-ultra micronized palmitoylethanolamide/luteolin treatment as add- on to intensive neuro-rehabilitation in a young patient with traumatic brain injury. *Gazz Med. Ital. Arch. Sci. Med.* **2018**, 177, 237–242. [CrossRef]
- 47. Ordaro, M.; Impellizzeri, D.; Paterniti, I.; Bruschetta, G.; Siracusa, R.; de Stefano, D.; Cuzzocrea, S.; Esposito, E. Neuroprotective Effects of Co-UltraPEALut on Secondary Inflammatory Process and Autophagy Involved in Traumatic Brain Injury. *J. Neuro Trauma* 2016, 33, 132–146.
- 48. Ahmad, A.; Crupi, R.; Impellizzeri, D.; Campolo, M.; Marino, A.; Esposito, E.; Cuzzocrea, S. Administration of palmitoyleth anolamide (PEA) protects the neurovascular unit and reduces secondary injury after traumatic brain injury in mice. *Brain. Behav. Immun.* 2012, 26, 1310–1321. [CrossRef]
- 49. Guida, F.; Boccella, S.; Iannotta, M.; de Gregorio, D.; Giordano, C.; Belardo, C.; Romano, R.; Palazzo, E.; Scafuro, M.A.; Serra, N.; et al. Palmitoylethanolamide Reduces Neuropsychiatric Behaviors by Restoring Cortical Electrophysiological Activity in a Mouse Model of Mild Traumatic Brain Injury. *Front. Pharmcol.* **2017**, *6*, 95. [CrossRef]
- 50. Caltagirone, C.; Cisari, C.; Schievano, C.; di Paola, R.; Cordaro, M.; Bruschetta, G.; Esposito, E.; Cuzzocrea, S. Stroke Study Group. Co-ultramicronized Palmitoylethanolamide/Luteolin in the Treatment of Cerebral Ischemia: From Rodent to Man. *Transl. Stroke. Res.* 2016, 7, 54–69. [CrossRef]
- 51. Citraro, R.; Russo, E.; Scicchitano, F.; van Rijn, C.M.; Cosco, D.; Avagliano, C.; Russo, R.; D'Agostino, G.; Petrosino, S.; Guida, F.; et al. Antiepileptic action of N-palmitoylethanolamine through CB1 and PPAR-α receptor activation in a genetic model of absence epilepsy. *Neuropharmacology* **2013**, *69*, 115–126. [CrossRef] [PubMed]
- 52. Kruk-Slomka, M.; Dzik, A.; Budzynska, B.; Biala, G. Endocannabinoid System: The Direct and Indirect Involvement in the Memory and Learning Processes—A Short Review. *Mol. Neurobiol.* **2017**, *54*, 8332–8347. [CrossRef] [PubMed]
- 53. Hou, R.; Garner, M.; Holmes, C.; Osmond, C.; Teeling, J.; Lau, L.; Baldwin, D.S. Peripheral inflammatory cytokines and im mune balance in generalised anxiety disorder: Case-controlled study. *Brain Behav. Immun.* **2017**, *62*, 212–218. [CrossRef] [PubMed]
- 54. Klein, T.W. Cannabinoid-based drugs as anti-inflammatory therapeutics. Nat. Rev. Immunol. 2005, 5, 400–411. [CrossRef]
- 55. Skaper, S.D.; Facci, L. Mast cell-glia axis in neuroinflammation and therapeutic potential of the anandamide congener pal mitoylethanolamide. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2012**, *367*, 3312–3325. [CrossRef]
- 56. Skaper, S.D.; Facci, L.; Giusti, P. Mast cells, glia and neuroinflammation: Partners in crime? *Immunology* **2014**, 141, 314–327. [CrossRef]
- 57. Nau, R.; Ribes, S.; Djukic, M.; Eiffert, H. Strategies to increase the activity of microglia as efficient protectors of the brain against infections. *Front. Cell Neurosci.* **2014**, 22, 138. [CrossRef]
- 58. D'Agostino, G.; la Rana, G.; Russo, R.; Sasso, O.; Iacono, A.; Esposito, E.; Raso, G.M.; Cuzzocrea, S.; Verme, J.L.; Peiomelli, D. Central administration of palmitoylethanolamide reduces hyperalgesia in mice via inhibition of NF-κB nuclear signalling in dorsal root ganglia. *Eur. J. Pharmcol.* **2009**, *613*, 54–59. [CrossRef]
- 59. Zeise, M.; Lehmann, A. Phosphoethanolamine transiently enhances excitability of rat hippocampal neurons in vitro. *J. Comp. Physiol. A.* **1987**, *161*, 461–467. [CrossRef]
- 60. Barbierato, M.; Facci, L.; Marinelli, C.; Zusso, M.; Argentini, C.; Skaper, S.D.; Giusti, P. Co-ultramicronized Palmitoylethano lamide/Luteolin Promotes the Maturation of Oligodendrocyte Precursor Cells. *Sci. Rep.* **2015**, *5*, 16676. [CrossRef]
- 61. Fidaleo, M.; Fanelli, F.; Ceru, M.P.; Moreno, S. Neuroprotective properties of peroxisome proliferator-activated receptor alpha (PPARα) and its lipid ligands. *Curr. Med. Chem.* **2014**, *21*, 2803–2821. [CrossRef] [PubMed]

Biomolecules **2022**, 12, 758

62. Boccella, S.; Cristiano, C.; Romano, R.; Iannotta, M.; Belardo, C.; Farina, A.; Guida, F.; Piscitelli, F.; Palazzo, E.; Mazzitelli, M.; et al. Ultra-micronized pal mitoylethanolamide rescues the cognitive decline-associated loss of neural plasticity in the neuropathic mouse entorhinal cor tex-dentate gyrus pathway. *Neurobiol. Dis.* 2019, 121, 106–119. [CrossRef] [PubMed]

- 63. Scuderi, C.; Esposito, G.; Blasio, A.; Valenza, M.; Arietti, P.L.; Steardo, J.; Carnuccio, R.; de Filippis, D.; Petrosino, S.; Iuvone, T.; et al. Palmitoylethanolamide counteracts reactive astrogliosis induced by β-amyloid peptide. *J. Cell Mol. Med.* **2011**, *15*, 2664–2674. [CrossRef] [PubMed]
- 64. Keppel Hesselink, J.M.R. Levi-Montalcini on Nerve Growth Factor, mast cells and palmitoylethanolamide, an endogenous anti-inflammatory and analgesic compound. *Pain Relief.* **2013**, 2, 1. [CrossRef]
- 65. Beggiato, S.; Cassano, T.; Ferraro, L.; Tomasini, M.C. Astrocytic palmitoylethanolamide pre-exposure exerts neuroprotective effects in astrocyte-neuron co-cultures from a triple transgenic mouse model of Alzheimer's disease. *Life Sci.* **2020**, 257, 118037. [CrossRef]
- 66. Bergandi, L.; Apprato, G.; Silvagno, F. Antioxidant and Anti-Inflammatory Activity of Combined Phycocyanin and Pal mitoylethanolamide in Human Lung and Prostate Epithelial Cells. *Antioxidants* **2022**, *11*, 201. [CrossRef]
- 67. Antonucci, N.; Cirillo, A.; Siniscalco, D. Beneficial Effects of Palmitoylethanolamide on Expressive Language, Cognition, and Behaviors in Autism: A Report of Two Cases. *Case Rep. Psychiatry* **2015**, 2015, 325061. [CrossRef]
- 68. Müller-Vahl, K.R.; Bindila, L.; Lutz, B.; Musshoff, F.; Skripuletz, T.; Baumgaertel, C.; Sühs, K.W. Cerebrospinal fluid endocan nabinoid levels in Gilles de la Tourette syndrome. *Neuropsychopharmacology* **2020**, 45, 1323–1329. [CrossRef]
- 69. Altamura, C.; Ventriglia, M.; Martini, M.G.; Montesano, D.; Errante, Y.; Piscitelli, F.; Scrascia, F.; Quattrocchi, C.; Palazzo, P.; Seccia, S.; et al. Elevation of Plasma 2-Arachidonoylglycerol Levels in Alzheimer's Disease Patients as a Po tential Protective Mechanism against Neurodegenerative Decline. *J. Alzheimers Dis.* 2015, 46, 497–506. [CrossRef]
- 70. Alcaraz-Silva, J.; Feingold, D.; Viana-Torre, G.; Budde, H.; Imperatori, C.; Machado, S.; Murillo-Rodríguez, E. The Endocanna binoid System as a Biomarker for Diagnostic and Therapeutic Applications in Depression and Anxiety. CNS Neurol. Disord. Drug Targets 2022. [CrossRef]
- 71. Hauer, D.; Schelling, G.; Gola, H.; Campolongo, P.; Morath, J.; Roozendaal, B.; Hamuni, G.; Karabatsiakis, A.; Atsak, P.; Vogeser, M.; et al. Plasma concentrations of endocannabinoids and related primary fatty acid amides in patients with post-trau matic stress disorder. *PLoS ONE* **2013**, *8*, e62741. [CrossRef] [PubMed]
- 72. Hill, M.N.; Miller, G.E.; Carrier, E.J.; Gorzalka, B.B.; Hillard, C.J. Circulating endocannabinoids and N-acyl ethanolamines are differentially regulated in major depression and following exposure to social stress. *Psychoneuroendocrinology* **2009**, *34*, 1257–1262. [CrossRef] [PubMed]
- 73. Hill, M.N.; Miller, G.E.; Ho, W.S.; Gorzalka, B.B.; Hillard, C.J. Serum endocannabinoid content is altered in females with depressive disorders: A preliminary report. *Pharmacopsychiatry* **2008**, *41*, 48–53. [CrossRef] [PubMed]
- 74. Pietropaolo, S.; Marsicano, G. The role of the endocannabinoid system as a therapeutic target for autism spectrum disorder: Lessons from behavioral studies on mouse models. *Neurosci. Biobehav. Rev.* **2022**, *132*, 664–678. [CrossRef]
- 75. Impellizzeri, D.; Cordaro, M.; Bruschetta, G.; Siracusa, R.; Crupi, R.; Esposito, E.; Cuzzocrea, S. *N*-Palmitoylethanolamine-Oxazoline as a New Therapeutic Strategy to Control Neuroinflammation: Neuroprotective Effects in Experimental Models of Spinal Cord and Brain Injury. *J. Neurotrauma*. **2017**, *34*, 2609–2623. [CrossRef]
- 76. Musella, A.; Fresegna, D.; Rizzo, F.R.; Gentile, A.; Bullitta, S.; de Vito, F.; Guadalupi, L.; Centonze, D.; Mandolesi, G. A novel crosstalk within the endocannabinoid system controls GABA transmission in the striatum. *Sci. Rep.* **2017**, 7, 7363. [CrossRef]
- 77. Ghazizadeh-Hashemi, M.; GhajarBrotini, S.; Schievano, C.; Guidi, L. Ultra-micronized Palmitoylethanolamide: An Efficacious Adjuvant Therapy for Parkinson's Disease. *CNS Neurol. Disord Drug Targets* **2017**, *16*, 705–713.
- 78. Ghazizadeh-Hashemi, M.; Ghajar, A.; Shalbafan, M.R.; Ghazizadeh-Hashemi, F.; Afarideh, M.; Malekpour, F.; Ghaleiha, A.; Ardebili, M.E.; Akhondzadeh, S. Palmitoylethanolamide as adjunctive therapy in major depressive disorder: A double-blind, randomized and placebo-controlled trial. *J. Affect. Disord.* 2018, 232, 127–133. [CrossRef]
- 79. Gabrielsson, L.; Mattson, S.; Fowler, C.J. Palmitoylethanolamide for the treatment of pain: Pharmacokinetics, safety and efficacy. *Br. J. Clin. Pharmacol.* **2016**, *82*, 932–942. [CrossRef]
- 80. Bettoni, I.; Comelli, F.; Colombo, A.; Bonfanti, P.; Costa, B. Non-neuronal cell modulation relieves neuropathic pain: Efficacy of the endogenous lipid palmitoylethanolamide. *CNS Neurol. Disord. Drug Targets* **2013**, *12*, 34–44. [CrossRef]
- 81. Mannelli, L.D.C.; D'Agostino, G.; Pacini, A.; Russo, R.; Zanardelli, M.; Ghelardini, C.; Calignano, A. Palmitoylethanolamide is a disease-modifying agent in peripheral neuropathy: Pain relief and neuroprotection share a PPAR-alpha-mediated mecha nism. *Mediat. Inflamm.* **2013**, 2013, 328797.
- 82. Petrosino, S.; Moriello, A.S.; Cerrato, S.; Fusco, M.; Puigdemont, A.; de Petrocellis, L.; di Marzo, V. The anti-inflamma tory mediator palmitoylethanolamide enhances the levels of 2-arachidonoyl-glycerol and potentiates its actions at TRPV1 cat ion channels. *Br. J. Pharmacol.* **2016**, *173*, 1154–1162. [CrossRef] [PubMed]
- 83. Cerrato, S.; Brazis, P.; della Valle, M.F.; Miolo, A.; Puigdemont, A. Effects of palmitoylethanolamide on immunologically induced histamine, PGD2 and TNFα release from canine skin mast cells. *Vet. Immunol. Immunopathol.* **2010**, *133*, 9–15. [CrossRef] [PubMed]
- 84. Davis, A. The dangers of NAIDS: Look both ways. Br. J. Gen. Pract. 2016, 66, 172–173. [CrossRef] [PubMed]
- 85. Clemente, S. Amyotrophic lateral sclerosis treatment with ultramicronized palmitoylethanolamide: A case report. CNS Neurol Disord Drug Targets 2012, 11, 933–936. [CrossRef]

Biomolecules **2022**, 12, 758

86. Impellizzeri, D.; Peritore, A.F.; Cordaro, M.; Gugliandolo, E.; Siracusa, R.; Crupi, R.; D'Amico, R.; Fusco, R.; Evangelista, M.; Cuzzocrea, S.; et al. The neuroprotective effects of micronized PEA (PEA-m) formulation on diabetic peripheral neuropa thy in mice. *FASEB J.* **2019**, *33*, 11364–11380. [CrossRef]

- 87. Hesselink, J.M.K.; Hekker, T.A. Therapeutic utility of palmitoylethanolamide in the treatment of neuropathic pain associated with various pathological conditions: A case series. *J. Pain Res.* **2012**, *5*, 437–442. [CrossRef]
- 88. Schifilliti, C.; Cucinotta, L.; Fedele, V.; Ingegnosi, C.; Luca, S.; Leotta, C. Micronized palmitoylethanolamide reduces the symp toms of neuropathic pain in diabetic patients. *Pain Res. Treat* **2014**, 2014, 849623. [CrossRef]
- 89. Donvito, G.; Bettoni, I.; Comelli, F.; Colombo, A.; Costa, B. Palmitoylethanolamide relieves pain and preserves pancreatic islet cells in a murine model of diabetes. CNS Neurol. Disord Drug Targets. 2015, 14, 452–462. [CrossRef]
- 90. Orefice, N.S.; Alhouayek, M.; Carotenuto, A.; Montella, S.; Barbato, F.; Comelli, A.; Calignano, A.; Muccioli, G.G.; Orefice, G. Oral Palmitoylethanolamide Treatment Is Associated with Reduced Cutaneous Adverse Effects of Interferon-β1a and Circu lating Proinflammatory Cytokines in Relapsing-Remitting Multiple Sclerosis. *Neurotherapeutics* **2016**, *13*, 428–438. [CrossRef]
- 91. Taylor, J.P.; Brown, R.H.; Cleveland, D.W. Decoding ALS: From genes to mechanism. *Nature* **2016**, *539*, 197–206. [CrossRef] [PubMed]
- 92. Musarò, A.; Dobrowolny, G.; Cambieri, C.; Onesti, E.; Ceccanti, M.; Frasca, V.; Pisano, A.; Cerbelli, B.; Lepore, E.; Ruffolo, G.; et al. Neuromuscular magnetic stimulation counteracts muscle decline in ALS patients: Results of a randomized, double-blind, controlled study. *Sci. Rep.* **2019**, *9*, 2837. [CrossRef]
- 93. Dupuis, L.; Loefer, J.-P. Neuromuscular junction destruction during amyotrophic lateral sclerosis: Insights from trans genic models. *Curr. Opin. Pharmacol.* **2009**, *9*, 341–346. [CrossRef] [PubMed]
- 94. Rocha, M.C.; Pousinha, P.A.; Correia, A.M.; Sebastião, A.M.; Ribeiro, J.A. Early changes of neuromuscular transmission in the SOD1(G93A) mice model of ALS start long before motor symptoms onset. *PLoS ONE* **2013**, *8*, e73846. [CrossRef] [PubMed]
- 95. Dadon-Nachum, M.; Melamed, E.; Ofen, D. The "Dying-Back" Phenomenon of Motor Neurons in ALS. *J. Mol. Neurosci.* **2011**, 43, 470–477. [CrossRef] [PubMed]
- Wong, M.; Martin, L.J. Skeletal muscle-restricted expression of human SOD1 causes motor neuron degeneration in trans genic mice. Hum. Mol. Genet. 2010, 19, 2284–2302. [CrossRef] [PubMed]
- 97. Musarò, A. Understanding ALS: New therapeutic approaches. FEBS J. 2013, 280, 4315–4322. [CrossRef]
- 98. Palma, E.; Inghilleri, M.; Conti, L.; Deflorio, C.; Frasca, V.; Manteca, A.; Pichiorri, F.; Roseti, C.; Torchia, G.; Limatola, C.; et al. Physiological characterization of human muscle acetylcholine receptors from ALS patients. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 20184–20188. [CrossRef]
- 99. Deflorio, C.; Palma, E.; Conti, L.; Roseti, C.; Manteca, A.; Giacomelli, E.; Catalano, M.; Limatola, C.; Inghilleri, M.; Grassi, F. Riluzole blocks human muscle acetylcholine receptors. *J. Physiol.* **2012**, *590*, 2519–2528. [CrossRef]
- 100. Palma, E.; Reyes-Ruiz, J.M.; Lopergolo, D.; Roseti, C.; Bertollini, C.; Ruffolo, G.; Cifelli, P.; Onesti, E.; Limatola, C.; Miledi, R.; et al. Acetylcholine receptors from human muscle as pharmacological target for ALS therapy. *Proc. Natl. Acad. Sci. USA* **2016**, 113, 3060–3065. [CrossRef]
- 101. Onesti, E.; Frasca, V.; Ceccanti, M.; Tartaglia, G.; Gori, M.C.; Cambieri, C.; Libonati, L.; Palma, E.; Inghilleri, M. Short-Term Ultramicronized Palmitoylethanolamide Therapy in Patients with Myasthenia Gravis: A Pilot Study to Possible Future Impli cations of Treatment. CNS Neurol. Disord. Drug Targets. 2019, 18, 232–238. [CrossRef] [PubMed]
- 102. Hehir, M.K.; Silvestri, N.J. Generalized Myasthenia Gravis: Classification, Clinical Presentation, Natural History, and Ep idemiology. *Neurol. Clin.* **2018**, *36*, 253–260. [CrossRef] [PubMed]
- 103. Cruz-Jentoft, A.J.; Bahat, G.; Bauer, J.; Boirie, Y.; Bruyère, O.; Cederholm, T.; Cooper, C.; Landi, F.; Rolland, Y.; Sayer, A.A.; et al. Sarcopenia: Revised European consensus on definition and diagnosis. *Age Ageing*. **2019**, *48*, 16–31. [CrossRef] [PubMed]
- 104. Maretti, E.; Molinari, S.; Battini, R.; Rustichelli, C.; Truzzi, E.; Iannuccelli, V.; Leo, E. Design, Characterization, and In Vitro Assays on Muscle Cells of Endocannabinoid-like Molecule Loaded Lipid Nanoparticles for a Therapeutic Anti-Inflammatory Approach to Sarcopenia. *Pharmaceutics* 2022, 14, 648. [CrossRef]
- 105. Di Stadio, A.; D'Ascanio, L.; Vaira, L.A.; Cantone, E.; de Luca, P.; Cingolani, C.; Motta, G.; de Riu, G.; Vitelli, F.; Spriano, G.; et al. Ultramicronized Palmitoylethanolamide and Luteolin Supplement Combined with Olfactory Training to Treat Post-COVID-19 Olfactory Impairment: A Multi-Center Double- Blinded Randomized Placebo-Controlled Clinical Trial. *Curr. Neuropharmacol.* 2022, 20, 1. [CrossRef]