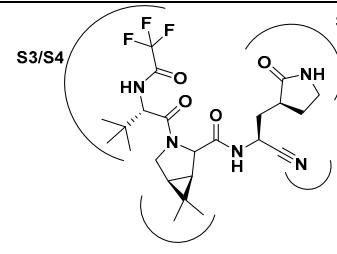
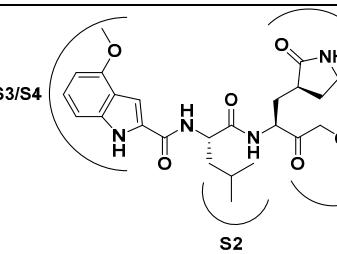
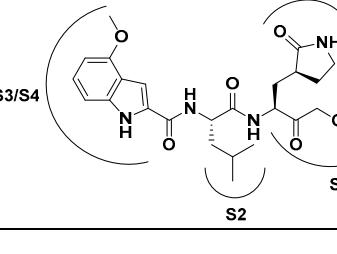
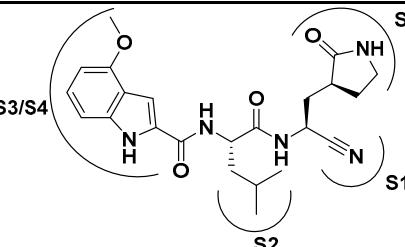
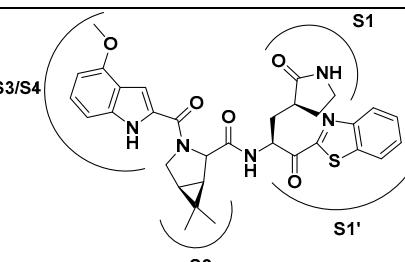
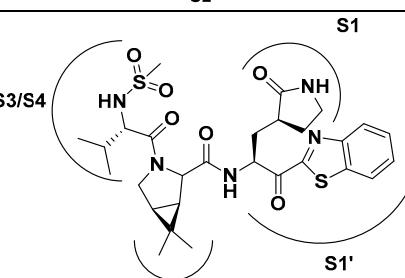
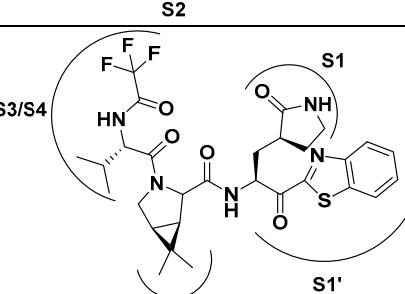


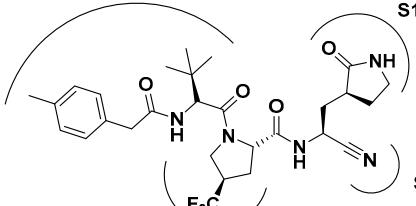
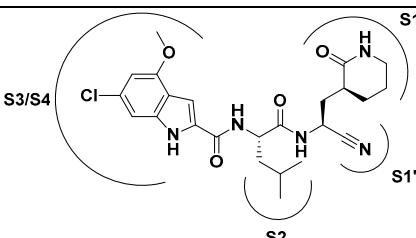
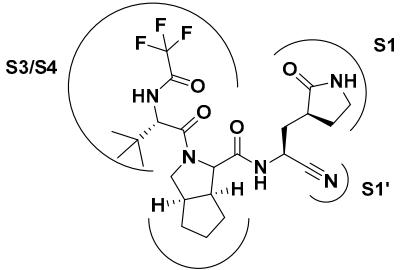
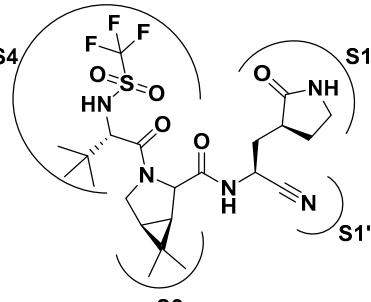
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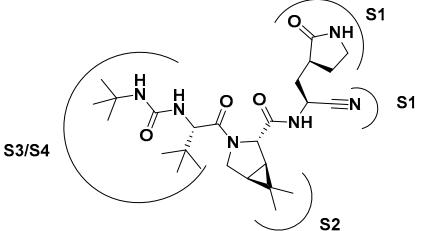
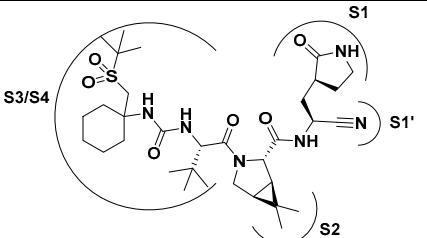
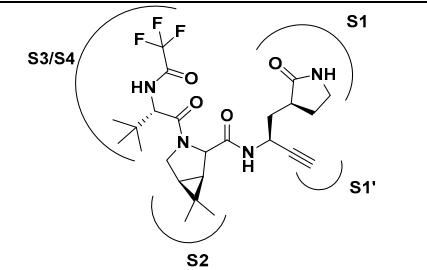
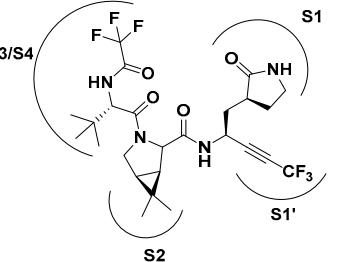
Recent Advances in SARS-CoV-2 Main Protease Inhibitors: From Nirmatrelvir to Future Perspectives

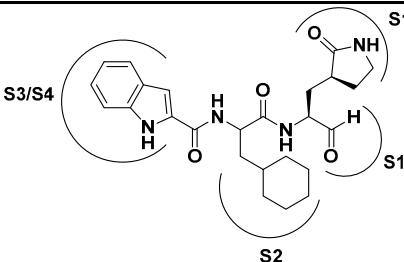
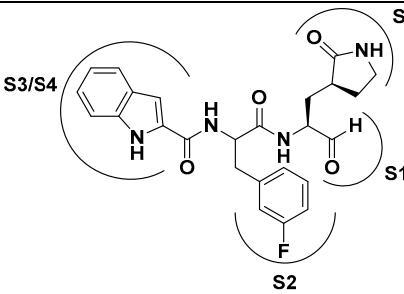
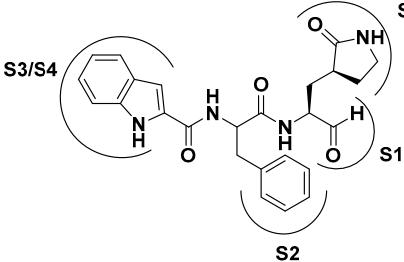
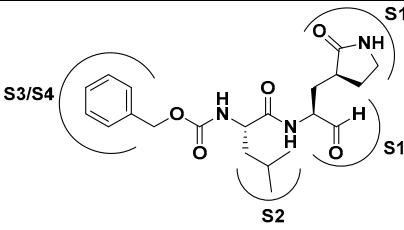
Table S1. Most relevant SARS-CoV-2 M^{Pro} inhibitors discovered so far.

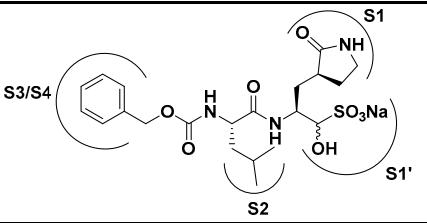
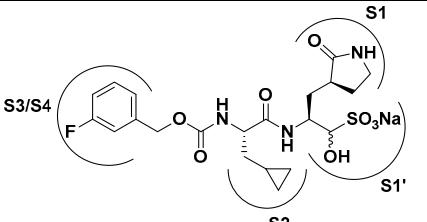
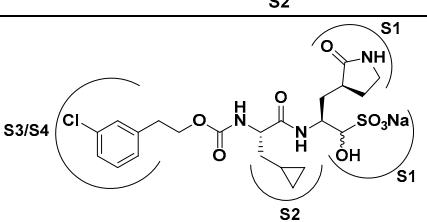
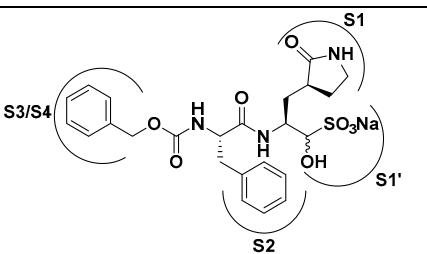
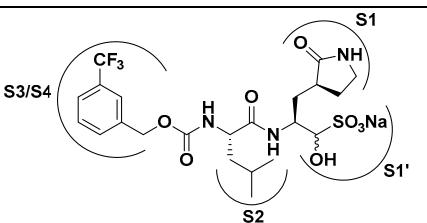
Compound	Chemical class	Structure	IC ₅₀ (nM) ^a	Mechanism	EC ₅₀ (μ M) ^c	Discovery method	References
1 NIRMATRELVIR	Nitriles		3.11 ^b	Covalent Reversible	0.075	Lead optimization	[1]
2 PF-00835231	Hydroxy ketones		4 ^b	Covalent Reversible	0.23	Drug repurposing	[1]
3 PF-07304814	Hydroxy ketones (phosphate prodrug)		27.9 ^b	Covalent Reversible	1.4	Drug repurposing	[1]

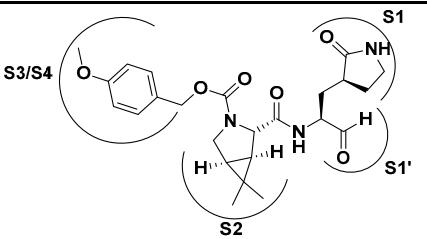
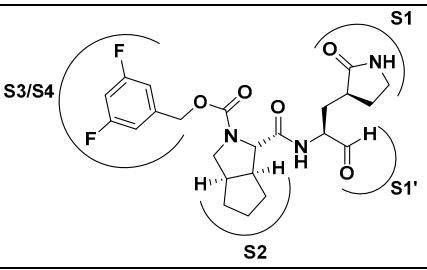
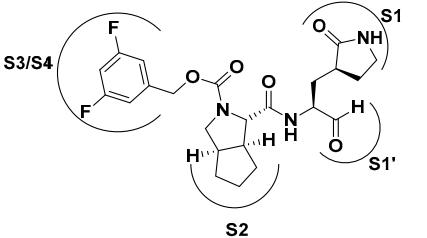
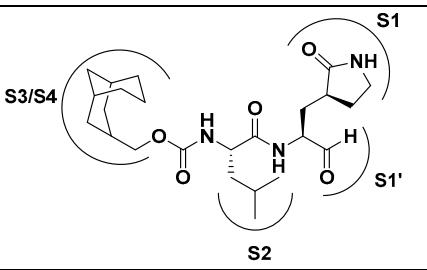
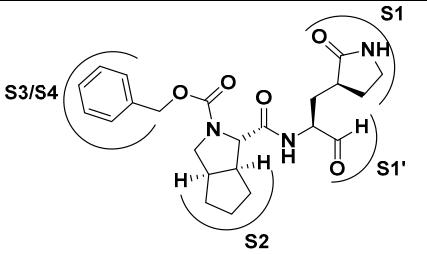
4	Nitriles		27.7 ^b	Covalent Reversible	1.4	Lead optimization	[1]
5	Ketones		230 ^b	Covalent Reversible	5.6	Lead optimization	[1]
6	Ketones		7.93 ^b	Covalent Reversible	0.9	Lead optimization	[1]
7	Ketones		12.1 ^b	Covalent Reversible	0.85	Lead optimization	[1]

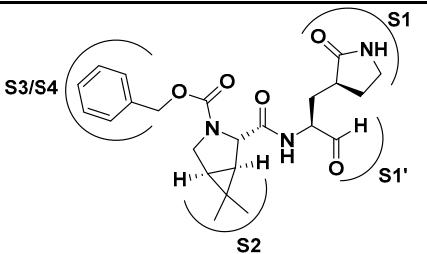
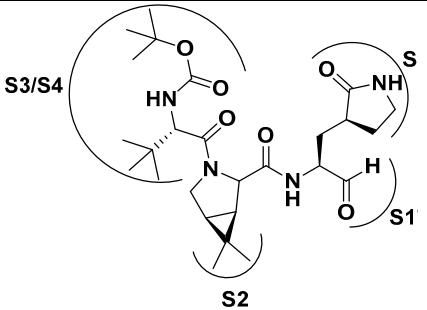
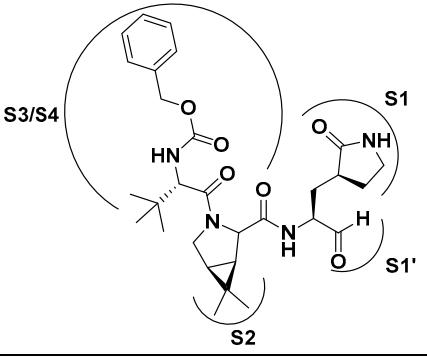
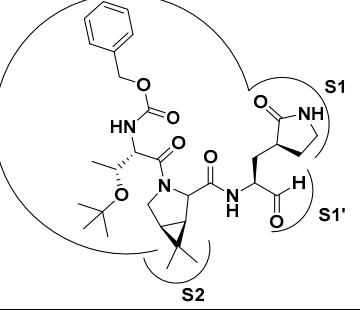
		S3/S4					
8	Nitriles		4 ^b	Covalent Reversible	0.019	Lead optimization	[2]
9	Nitriles		9 ^b	Covalent Reversible	2.2	Lead optimization	[3]
10	Nitriles		18	Covalent Reversible	0.31	Lead optimization	[4]
11	Nitriles		22	Covalent Reversible	0.17	Lead optimization	[4]

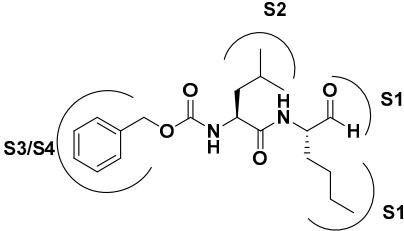
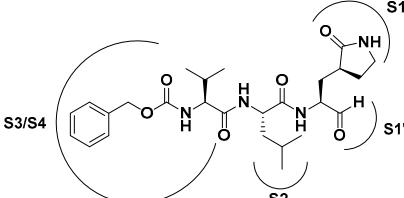
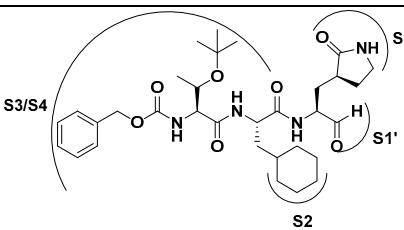
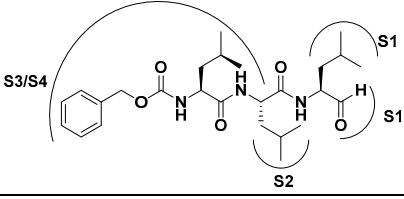
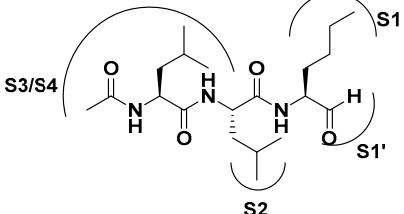
12	BBH-2	Nitriles		26 ^d	Covalent Reversible	0.88	Lead optimization	[5]
13	NBH-2	Nitriles		30 ^d	Covalent Reversible	1.82	Lead optimization	[5]
14		Alkynes		140	Covalent Irreversible	25.7	Lead optimization	[6]
15		Alkynes		230	Covalent Irreversible	5.1	Lead optimization	[6]

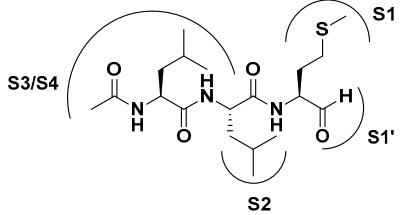
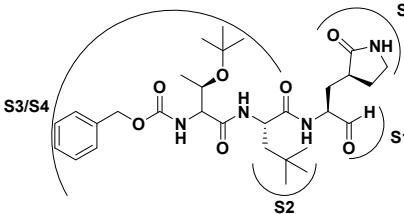
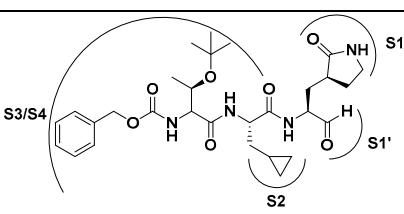
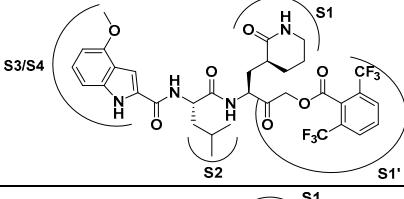
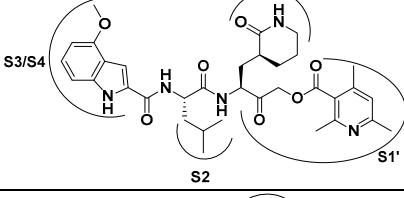
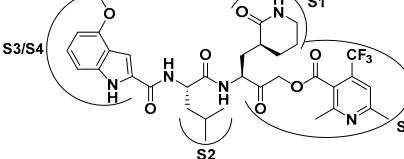
16	Aldehydes		53	Covalent Reversible	0.53	Structure-based design	[7]
17	Aldehydes		40	Covalent Reversible	0.72	Structure-based design	[7]
18	Aldehydes		34	Covalent Reversible	0.29	Lead optimization	[8]
19 GC-373	Aldehydes		400	Covalent Reversible	1.5	Drug repurposing	[9]

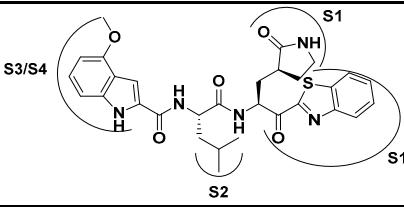
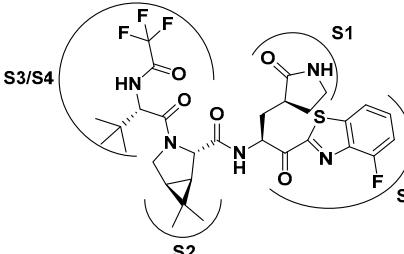
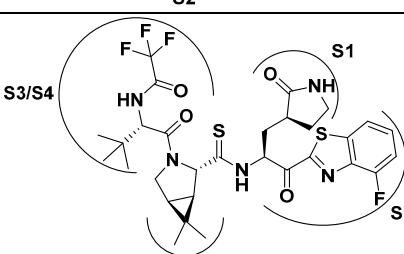
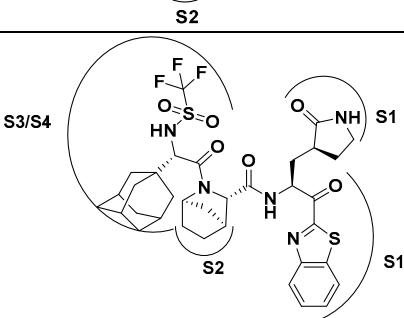
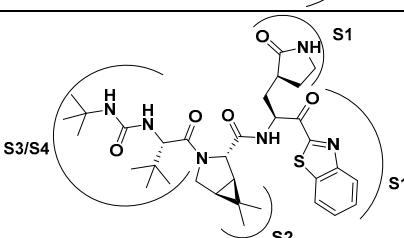
20 GC-376	Aldehydes (phosphate prodrug)		190	Covalent Reversible	0.9	Drug repurposing	[9]
21	Aldehydes (phosphate prodrug)		70	Covalent Reversible	0.57	Lead optimization	[10]
22	Aldehydes (phosphate prodrug)		80	Covalent Reversible	0.7	Lead optimization	[10]
23 UAWJ247	Aldehydes (phosphate prodrug)		45	Covalent Reversible	6.8	Lead optimization	[11]
24 Coronastat	Aldehydes (phosphate prodrug)		16	Covalent Reversible	0.006	Lead optimization	[12]

25 MI-09	Aldehydes		15.2	Covalent Reversible	0.86	Structure-based design	[13]
26 Mi-23	Aldehydes		7.6	Covalent Reversible	n.d.	Structure-based design	[13]
27 MI-30	Aldehydes		17.2	Covalent Reversible	0.54	Structure-based design	[13]
28	Aldehydes		180	Covalent Reversible	0.035	Lead optimization	[14]
29 UAWJ9-36-1	Aldehydes		51	Covalent Reversible	2.6	Rational design	[15]

30 UAWJ9-36-3	Aldehydes		54	Covalent Reversible	0.37	Rational design	[15]
31 MPI143	Aldehydes		45	Covalent Reversible	0.4 – 1.0 ^e	Structure-based design	[16]
32 MPI144	Aldehydes		59	Covalent Reversible	0.9 – 2.9 ^e	Structure-based design	[16]
33 MPI146	Aldehydes		120	Covalent Reversible	0.8 – 2.3 ^e	Structure-based design	[16]

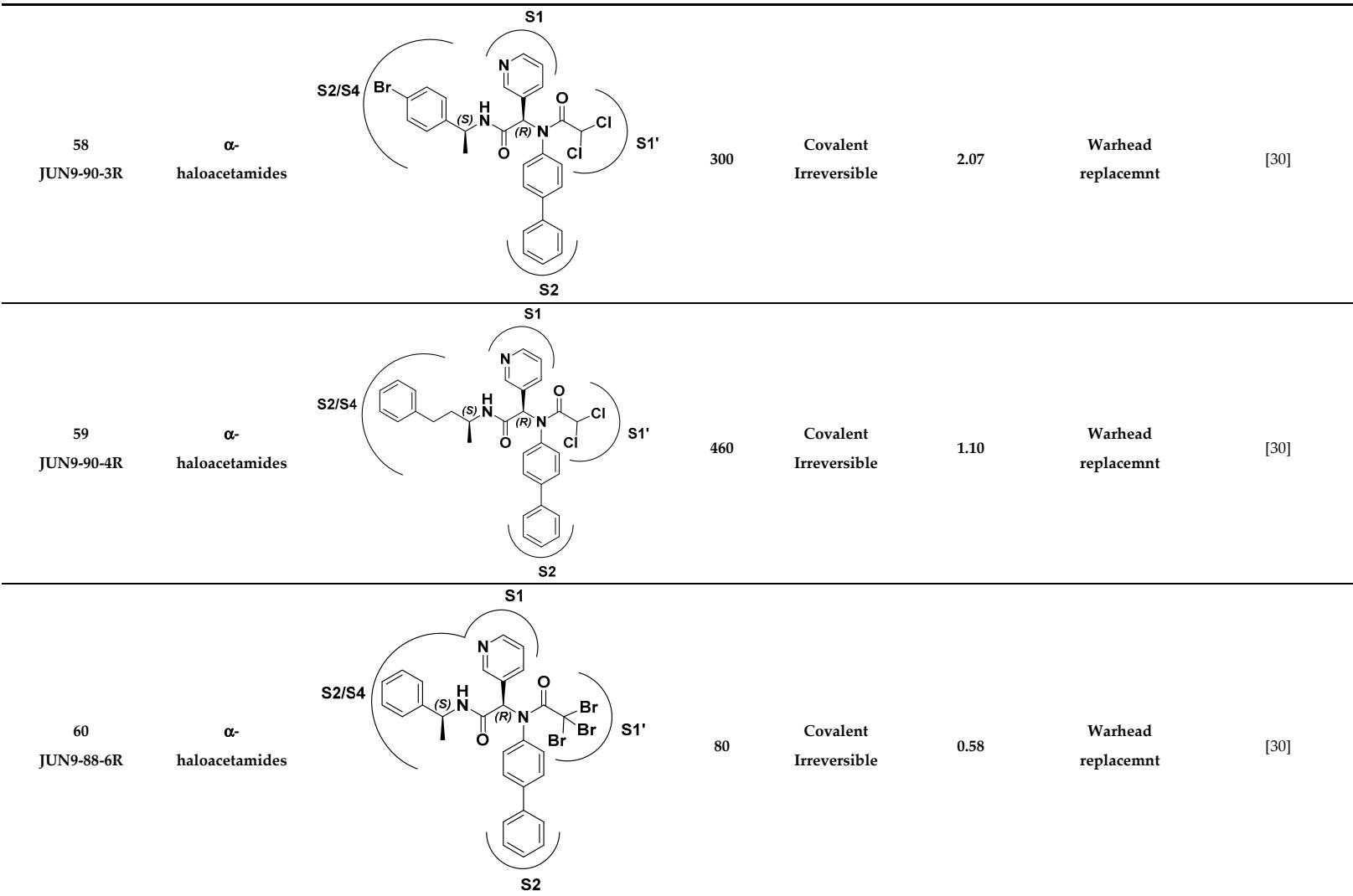
			S2				
34 Calpeptin	Aldehydes		10700	Covalent Reversible	0.072	X-ray screening	[17]
35 MPI3	Aldehydes		8.5	Covalent Reversible	n.d.	Structure-based design	[18]
36 MPI8	Aldehydes		108	Covalent Reversible	2.5	Structure-based design	[18]
37 MGI132	Aldehydes		7500	Covalent Reversible	n.d.	Drug repurposing	[19]
38 Calpain inhibitor I	Aldehydes		970	Covalent Reversible	n.d.	Drug repurposing	[11]

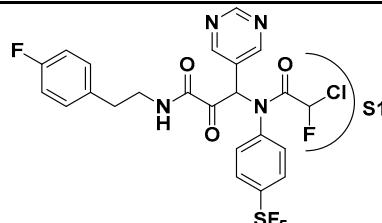
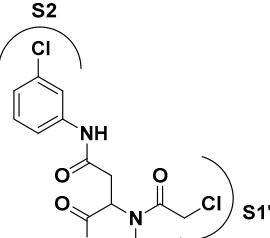
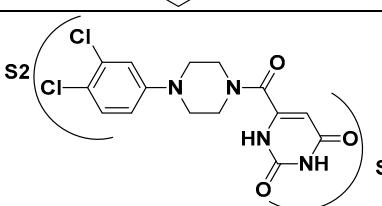
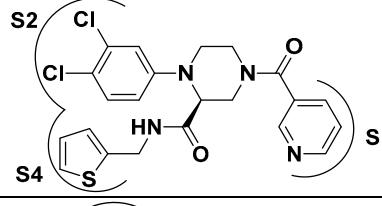
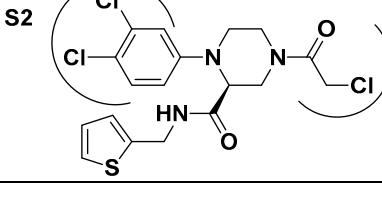
39	Calpain inhibitor II	Aldehydes		8600	Covalent Reversible	2.07	Drug repurposing	[11]
40	MPI16	Aldehydes		105	Covalent Reversible	0.056	Lead optimization	[20]
41	MPI17	Aldehydes		60	Covalent Reversible	0.097	Lead optimization	[20]
42		Ketones		1.0	Covalent Reversible	0.16	Structure-based design	[21]
43		Ketones		19.0	Covalent Reversible	0.3	Structure-based design	[21]
44		Ketones		14.0	Covalent Reversible	0.47	Structure-based design	[21]

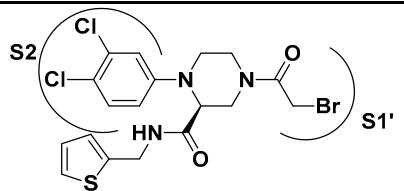
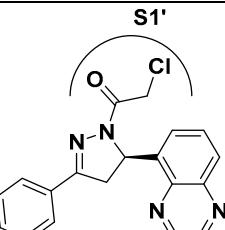
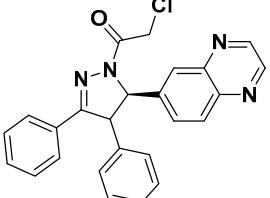
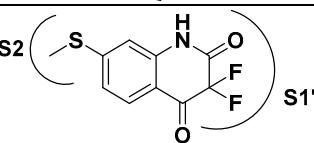
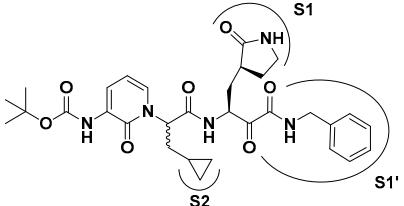
45	YH-53	Ketones		130	Covalent Reversible	2.6	Drug repurposing	[22]
46	TKB245	Ketones		7	Covalent Reversible	0.03	Lead optimization	[23]
47	TKB248	Ketones		74	Covalent Reversible	0.22	Lead optimization	[23]
48		Ketones		1650	Covalent Reversible	0.18	Lead optimization	[24]
49	BBH1	Ketones		n.d.	Covalent Reversible	16.1	Lead optimization	[5]

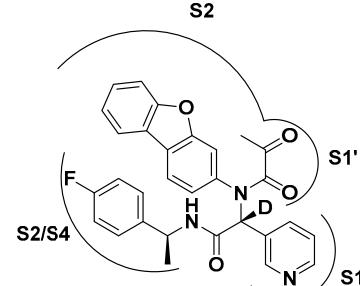
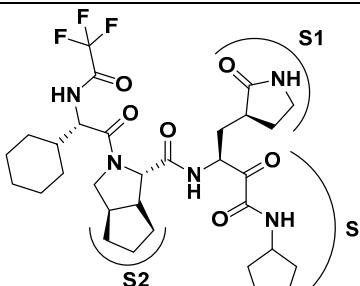
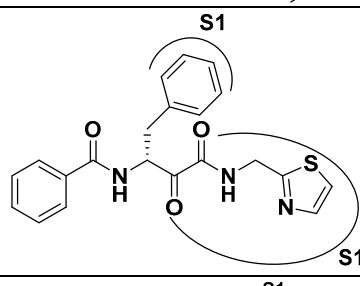
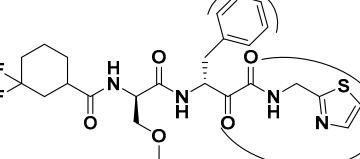
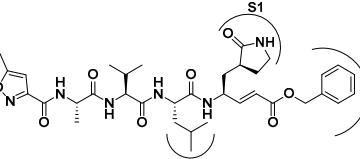
50 Z-FA-FMK	Ketones	S3/S4		11400	Covalent Irreversible	0.13	High throughput screening	[25]
52	Ketones	S3/S4		n.d.	Covalent Reversible	12.9 ^f	Warhead modification	[26]
53	Ketones	S1'		750	Covalent Reversible	n.d.	Structure-based design	[27]
54 X77	Miscellaneous	S2		4100	Non Covalent	n.d.	n.d.	[28]

55	α -haloacetamides		410	Covalent Irreversible	n.d.	Covalent docking	[28]
56 23R	Miscellaneous		200	Non Covalent	1.4	Structure-based design	[29]
57 JUN9-62-2R	α -haloacetamides		430	Covalent Irreversible	0.90	Warhead replacement	[30]

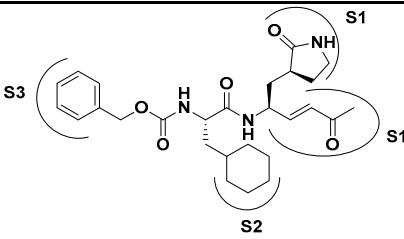
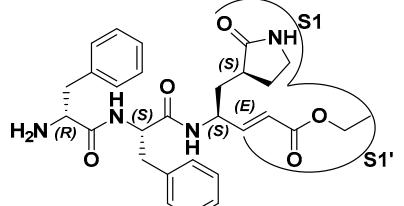
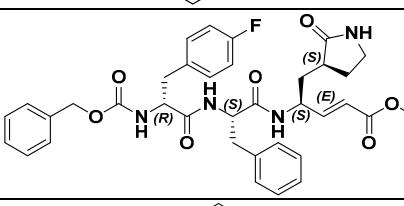
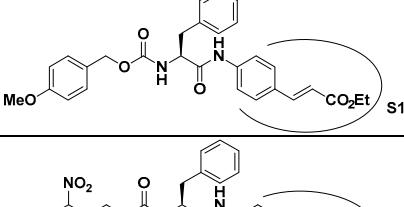
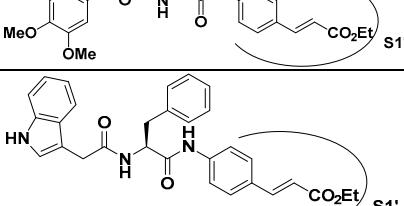
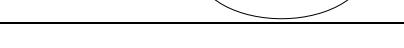


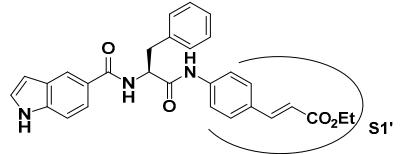
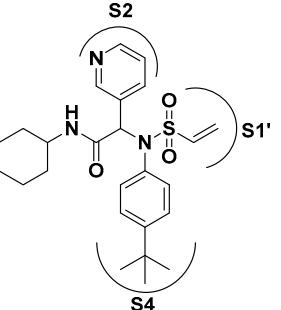
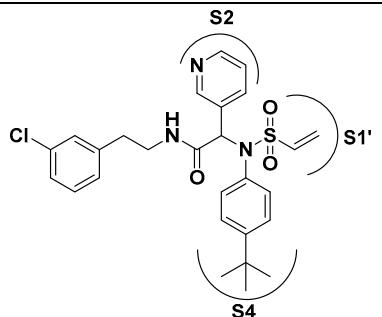
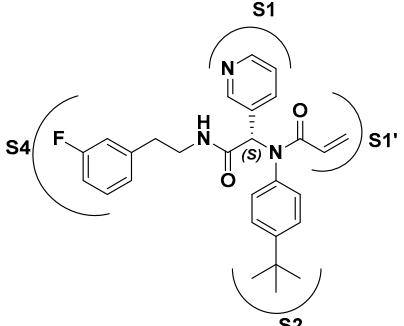
61	α -haloacetamides		56	Covalent Reversible	n.d.	Lead optimization	[31]
62	α -haloacetamides		8500	Covalent Irreversible	n.d.	In silico screening and covalent docking	[32]
63	Miscellaneous		4200	Non Covalent	n.d.	High throughput screening	[33]
64	Miscellaneous		400	Non Covalent	1.1	Structure-based design	[34]
65	α -haloacetamides		180	Covalent Irreversible	2.6	Warhead replacement	[35]

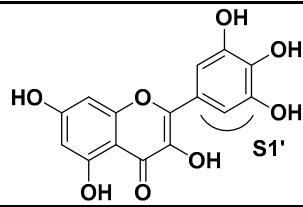
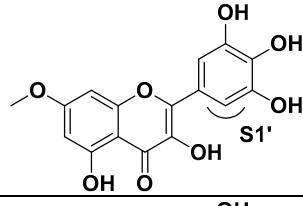
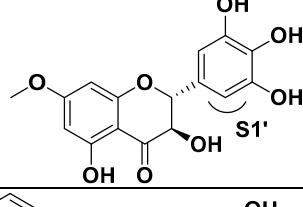
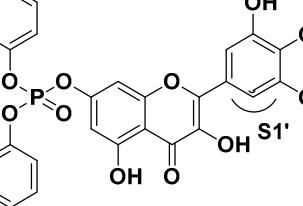
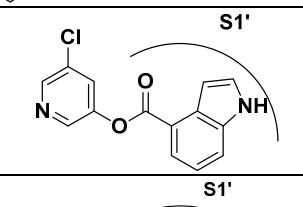
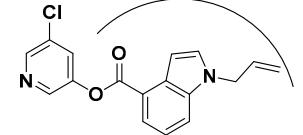
66 GD13	α -haloacetamides		310	Covalent Irreversible	4.2	Warhead replacement	[35]
67 (R)-EN82	α -haloacetamides		530	Covalent Irreversible	n.d.	Covalent chemoproteomic strategies and structure-based design	[36]
68 HW-2-010b	α -haloacetamides		14	Covalent Irreversible	n.d.	Covalent chemoproteomic strategies and structure-based design	[36]
69 QUB-00006-Int-07	α -haloacetamides		830	Covalent	n.d.	In silico strategies	[37]
70	α -ketoamides		670	Covalent Reversible	4-5	Lead optimization	[38]

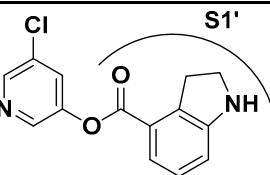
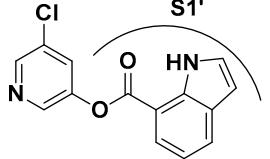
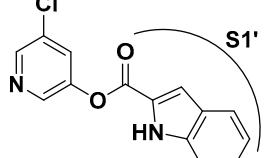
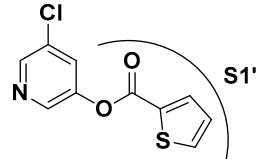
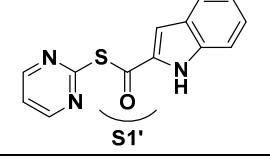
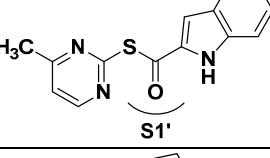
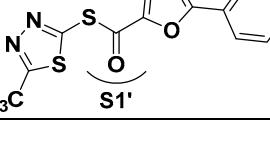
71 Y180	α -ketoamides		8.1	Covalent Reversible	0.011	Structure-based design	[39]
72 RAY1216	α -ketoamides		8.4 ^b	Covalent Reversible	0.13	Lead optimization	[40]
73	α -ketoamides		1300	Covalent Reversible	n.d.	In vitro screening	[41]
74 SY110	α -ketoamides		14.4	Covalent Reversible	0.38 - 2.8 ^c	Lead optimization	[41]
75 N3	Michael Acceptors		n.d.	Covalent Irreversible	16.8	Computer- aided drug design	[42]

76	Michael Acceptors		125000	Covalent Irreversible	20.6	Virtual screening	[42]
77	Michael Acceptors		151	Covalent Irreversible	2.9	<i>In</i> <i>vitro</i> screening	[43]
78	Michael Acceptors		47200	Covalent Irreversible	n.d.	Virtual screening	[44]
79	Michael Acceptors		157000	Covalent Irreversible	n.d.	Virtual screening	[44]
80	Michael Acceptors		260 ^b	Covalent Irreversible	18.5	Lead optimization	[45]
81	Michael Acceptors		250 ^b	Covalent Irreversible	1.5	Lead optimization	[45]

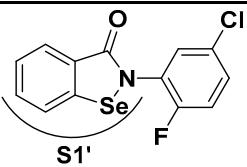
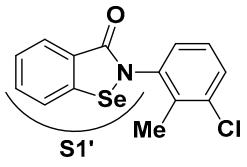
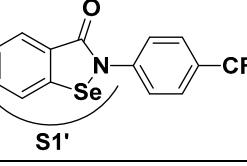
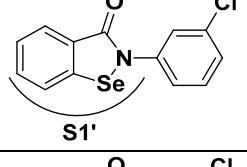
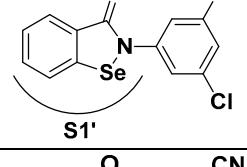
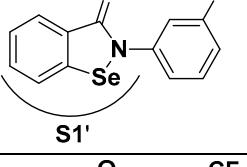
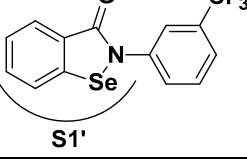
82	Michael Acceptors		180 ^b	Covalent Irreversible	1.8	Lead optimization	[45]
83	Michael Acceptors		900	Covalent Irreversible	0.0082	Lead optimization	[46]
84	Michael Acceptors		1800	Covalent Irreversible	0.0147	Lead optimization	[46]
85	Michael Acceptors		1900	Covalent Irreversible	n.d.	Warhead replacement	[47]
86	Michael Acceptors		14000	Covalent Irreversible	5.3 ^f	Warhead replacement	[47]
87	Michael Acceptors		12400	Covalent Irreversible	9.1 ^g	Warhead replacement	[47]

88	Michael Acceptors		10100	Covalent Irreversible	10.1s	Warhead replacement	[47]
89	Michael Acceptors		420	Covalent Irreversible	n.d.	Covalent docking	[28]
90	Michael Acceptors		170	Covalent Irreversible	n.d.	Covalent docking	[28]
92	Michael Acceptors		2860	Covalent Irreversible	n.d.	Automatic pipeline	[48]

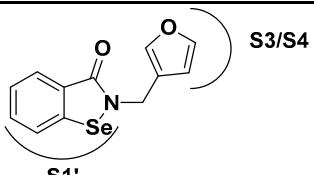
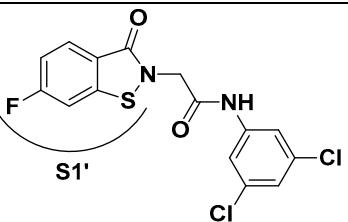
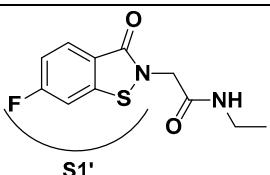
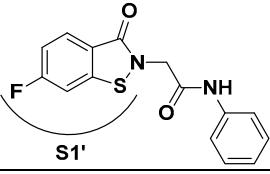
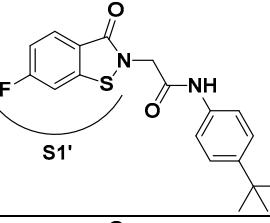
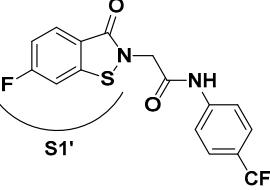
93 MYRECITIN	Michael Acceptors		200 - 600	Covalent Irreversible	8	in Vitro Repurposing Screen	[49]
94	Michael Acceptors		300	Covalent Irreversible	12.6	Structure-based optimization	[50]
95	Michael Acceptors		260	Covalent Irreversible	11.5	Structure-based optimization	[50]
96	Michael Acceptors		3100	Covalent Irreversible	3.2	Structure-based optimization	[50]
97 GRL-0920	Esters		250	Covalent Irreversible	2.8	Drug repurposing	[51]
98 GRL-0820	Esters		73	Covalent Irreversible	15	Structure-based design	[51]

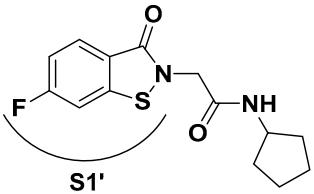
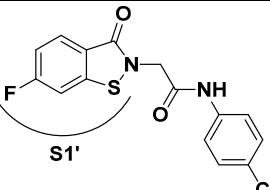
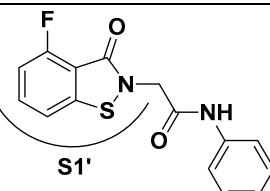
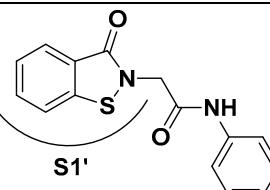
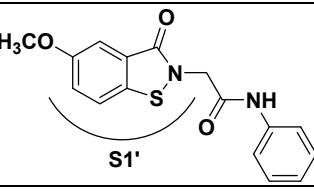
99 GRL-1720	Esters		320	Covalent Irreversible	15	Lead optimization	[52]
100	Esters		55	Covalent Irreversible	n.d.	Lead optimization	[53]
101	Esters		34	Covalent Irreversible	n.d.	Lead optimization	[53]
102	Esters		81	Covalent Irreversible	n.d.	<i>In vitro</i> screening	[43]
103	Thioesters		11	Covalent Irreversible	0.11	Virtual screening Structure-based design	[54]
104	Thioesters		88	Covalent Irreversible	0.73	Virtual screening Structure-based design	[54]
105	Thioesters		n.d.	n.d.	0.1	Virtual screening Structure-based design	[54]

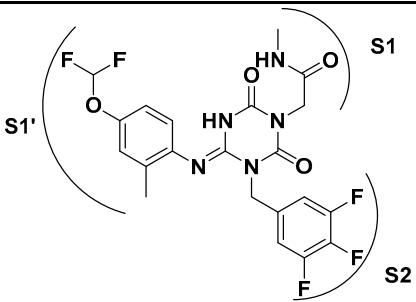
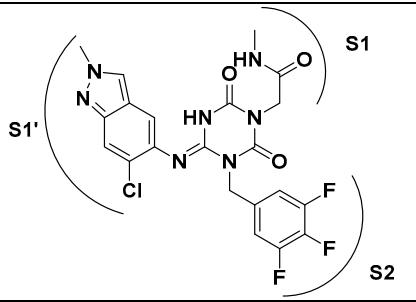
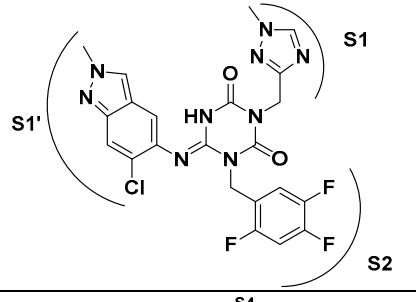
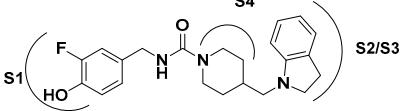
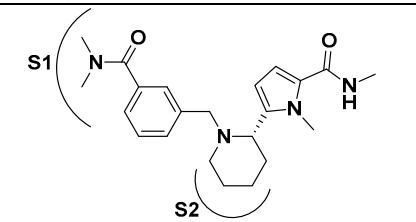
106	Thioesters		n.d.	n.d.	0.44	Virtual screening Structure-based design	[54]
107	Thioesters		n.d.	n.d.	0.66	Virtual screening Structure-based design	[54]
108	Thioesters		n.d.	n.d.	0.038	Virtual screening Structure-based design	[54]
109	Thioesters		n.d.	n.d.	0.045	Virtual screening Structure-based design	[54]
110 EBSELEN	Selenium-based compounds		670	Covalent	4.67	Drug repurposing	[42]
111	Selenium-based compounds		25.7	Covalent	n.d.	In vitro screening	[55]
112	Selenium-based compounds		27.9	Covalent	n.d.	In vitro screening	[55]

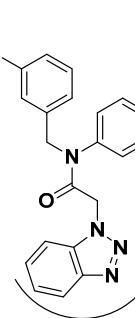
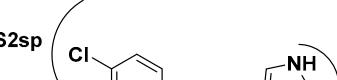
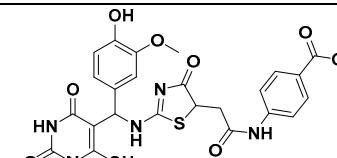
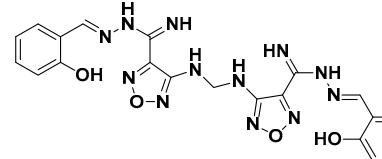
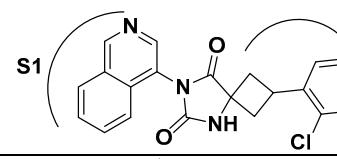
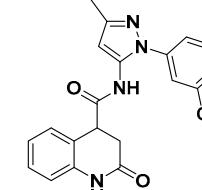
113	Selenium-based compounds		15.2	Covalent	n.d.	<i>In vitro</i> screening	[55]
114	Selenium-based compounds		27.4	Covalent	n.d.	<i>In vitro</i> screening	[55]
115	Selenium-based compounds		900	Covalent	11.2	Structure-based design	[56]
116	Selenium-based compounds		680	Covalent	11.7	Structure-based design	[56]
117	Selenium-based compounds		640	Covalent	18.2	Structure-based design	[56]
118	Selenium-based compounds		380	Covalent	2.0	Structure-based design	[56]
119	Selenium-based compounds		2800	Covalent	0.8	Structure-based design	[56]

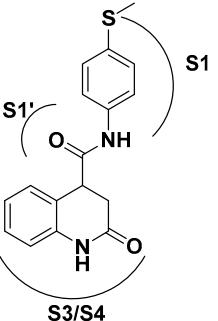
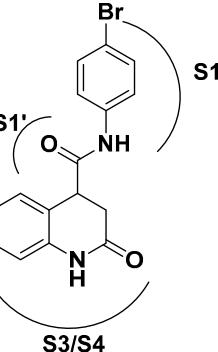
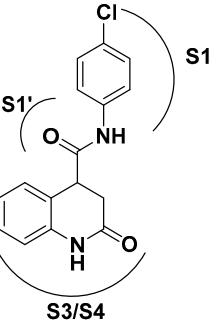
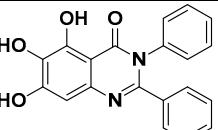
120 MR6-7-2	Selenium-based compounds		360	Covalent	4.5	Drug repurposing	[57]
121 MR6-18-4	Selenium-based compounds		340	Covalent	3.7	Drug repurposing	[57]
122 MR6-17-1	Selenium-based compounds		700	Covalent	n.a.	Drug repurposing	[57]
123 MR6-26-2	Selenium-based compounds		470	Covalent	3.2	Drug repurposing	[57]
124 MR6-31-2	Selenium-based compounds		820	Covalent	1.8	Drug repurposing	[57]
125 EBSULFUR	Sulfur-based compounds		490	Covalent	n.d.	In vitro screening	[58]
126	Sulfur-based compounds		110	Covalent	n.d.	In vitro screening	[58]

127	Selenium-based compounds		74	Covalent	n.d.	<i>In vitro</i> screening	[58]
128	Sulfur-based compounds		190	Covalent	n.d.	high-throughput screening	[59]
129	Sulfur-based compounds		160	Covalent	n.d.	Lead optimization	[59]
130	Sulfur-based compounds		180	Covalent	n.d.	Lead optimization	[59]
131	Sulfur-based compounds		140	Covalent	n.d.	Lead optimization	[59]
132	Sulfur-based compounds		150	Covalent	n.d.	Lead optimization	[59]

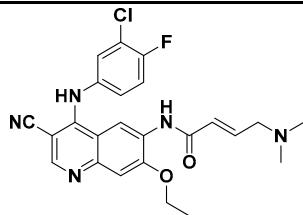
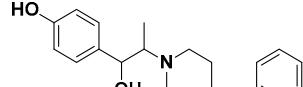
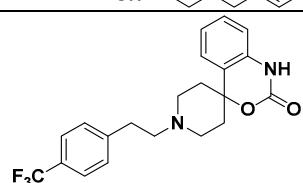
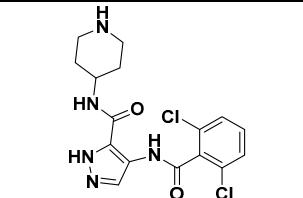
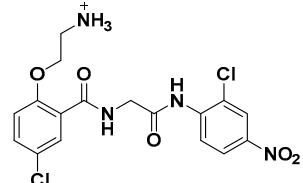
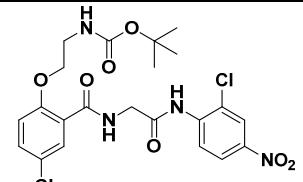
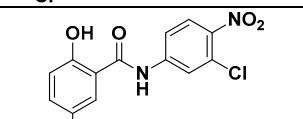
133	Sulfur-based compounds		150	Covalent	n.d.	Lead optimization	[59]
134	Sulfur-based compounds		120	Covalent	n.d.	Lead optimization	[59]
135	Sulfur-based compounds		116	Covalent	n.d.	Lead optimization	[59]
136	Sulfur-based compounds		165	Covalent	n.d.	Lead optimization	[59]
137	Sulfur-based compounds		165	Covalent	n.d.	Lead optimization	[59]

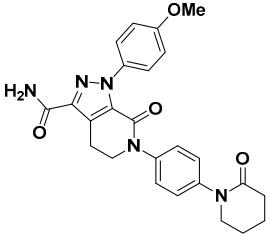
138	Miscellaneous		8600	Non Covalent	n.d.	Virtual and <i>in vitro</i> screening	[60]
139	Miscellaneous		96	Non Covalent	12.5	Lead optimization	[60]
140 S-217622	Miscellaneous		13	Non Covalent	0.5 – 0.29 ^e	Lead optimization	[60]
141 Z1244904919	Miscellaneous		730	Non Covalent	5	Virtual and <i>in vitro</i> screening	[61]
142 Z1759961356	Miscellaneous		690	Non Covalent	8.5	Virtual and <i>in vitro</i> screening	[61]

144 CCF981	Miscellaneous		S2sp 	68	Non Covalent	0.5	Lead optimization [62]
145	Miscellaneous			450	Non Covalent	0.77	Virtual and <i>in vitro</i> screening [63]
146	Miscellaneous			110	Non Covalent	0.11	Virtual and <i>in vitro</i> screening [63]
149	Miscellaneous			77	Non Covalent	0.11	Lead optimization [64]
150 ZINC00037365906 0	Miscellaneous			58000	Non Covalent	n.d.	Virtual screening [65]

156	Miscellaneous		1600	Non Covalent	n.d.	Virtual screening	[65]
157	Miscellaneous		2000	Non Covalent	n.d.	Virtual screening	[65]
158	Miscellaneous		5800	Non Covalent	n.d.	Virtual screening	[65]
160	Miscellaneous		83	Non Covalent	1.1	Structure-based design	[66]

161	Miscellaneous		100	Non Covalent	2.1	Structure-based design	[66]
162	Miscellaneous		4100	Non Covalent	n.d.	Warhead replacement	[67]
163 Neochinulin A	Miscellaneous		470	Non Covalent	n.d.	<i>in vitro</i> screening	[68]
164 Shikonin	Miscellaneous		1570	Non Covalent	n.d.	Virtual screening	[69]
165 C1	Miscellaneous		1500	Non Covalent	n.d.	Lead optimization	[70]
166 C2	Miscellaneous		1800	Non Covalent	n.d.	Lead optimization	[70]

167 Pelitinib	Miscellaneous		n.d.	Allosteric	1.25	X-ray screening Drug repurposing	[17]
168 Ifenprodil	Miscellaneous		n.d.	Allosteric	47	X-ray screening Drug repurposing	[17]
169 RS-102895	Miscellaneous		n.d.	Allosteric	19.8	X-ray screening Drug repurposing	[17]
170 AT7519	Miscellaneous		n.d.	Allosteric	25.2	X-ray screening Drug repurposing	[17]
171 JMX0286	Miscellaneous		4800	Allosteric	2.3	In vitro screening	[71]
172 JMX0301	Miscellaneous		4500	Allosteric	2.4	In vitro screening	[71]
173 JMX0941	Miscellaneous		3900	Allosteric	1.7	In vitro screening	[71]

174	Apixaban	Miscellaneous		9.7	Allosteric	1.8	Drug repurposing	[72]
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References

- [1] D. R. Owen *et al.*, ‘An oral SARS-CoV-2 M^{pro} inhibitor clinical candidate for the treatment of COVID-19’, *Science* (1979), vol. 374, no. 6575, pp. 1586–1593, Dec. 2021, doi: 10.1126/science.abl4784.
- [2] ‘WO2021250648A1 - Nitrile-containing antiviral compounds - Google Patents’. <https://patents.google.com/patent/WO2021250648A1/en> (accessed Mar. 05, 2023).
- [3] B. Bai *et al.*, ‘Peptidomimetic nitrile warheads as SARS-CoV-2 3CL protease inhibitors’, *RSC Med Chem*, vol. 12, no. 10, pp. 1722–1730, 2021, doi: 10.1039/D1MD00247C.
- [4] M. Zhu *et al.*, ‘Design, synthesis and biological evaluation of covalent peptidomimetic 3CL protease inhibitors containing nitrile moiety’, *Bioorg Med Chem*, vol. 87, p. 117316, May 2023, doi: 10.1016/J.BMC.2023.117316.
- [5] D. W. Kneller *et al.*, ‘Covalent narlaprevir- and boceprevir-derived hybrid inhibitors of SARS-CoV-2 main protease’, *Nat Commun*, vol. 13, no. 1, p. 2268, Apr. 2022, doi: 10.1038/s41467-022-29915-z.
- [6] L. Brewitz *et al.*, ‘Alkyne Derivatives of SARS-CoV-2 Main Protease Inhibitors Including Nirmatrelvir Inhibit by Reacting Covalently with the Nucleophilic Cysteine’, *J Med Chem*, vol. 66, no. 4, pp. 2663–2680, Feb. 2023, doi: 10.1021/acs.jmedchem.2c01627.
- [7] W. Dai *et al.*, ‘Structure-based design of antiviral drug candidates targeting the SARS-CoV-2 main protease’, *Science* (1979), vol. 368, no. 6497, pp. 1331–1335, Jun. 2020, doi: 10.1126/science.abb4489.
- [8] W. Dai *et al.*, ‘Design, Synthesis, and Biological Evaluation of Peptidomimetic Aldehydes as Broad-Spectrum Inhibitors against Enterovirus and SARS-CoV-2’, *J Med Chem*, vol. 65, no. 4, pp. 2794–2808, Feb. 2022, doi: 10.1021/acs.jmedchem.0c02258.
- [9] W. Vuong *et al.*, ‘Feline coronavirus drug inhibits the main protease of SARS-CoV-2 and blocks virus replication’, *Nat Commun*, vol. 11, no. 1, p. 4282, Aug. 2020, doi: 10.1038/s41467-020-18096-2.
- [10] W. Vuong *et al.*, ‘Improved SARS-CoV-2 Mpro inhibitors based on feline antiviral drug GC376: Structural enhancements, increased solubility, and micellar studies’, *Eur J Med Chem*, vol. 222, p. 113584, Oct. 2021, doi: 10.1016/j.ejmech.2021.113584.
- [11] M. D. Sacco *et al.*, ‘Structure and inhibition of the SARS-CoV-2 main protease reveal strategy for developing dual inhibitors against Mpro and cathepsin L’, *Sci Adv*, vol. 6, no. 50, Dec. 2020, doi: 10.1126/SCIAADV.ABE0751/SUPPL_FILE/ABE0751_SM.PDF.
- [12] H. Liu *et al.*, ‘Development of optimized drug-like small molecule inhibitors of the SARS-CoV-2 3CL protease for treatment of COVID-19’, *Nat Commun*, vol. 13, no. 1, p. 1891, Apr. 2022, doi: 10.1038/s41467-022-29413-2.
- [13] J. Qiao *et al.*, ‘SARS-CoV-2 M^{pro} inhibitors with antiviral activity in a transgenic mouse model’, *Science* (1979), vol. 371, no. 6536, pp. 1374–1378, Mar. 2021, doi: 10.1126/science.abf1611.

- [14] C. S. Dampalla *et al.*, 'Structure-Guided Design of Conformationally Constrained Cyclohexane Inhibitors of Severe Acute Respiratory Syndrome Coronavirus-2 3CL Protease', *J Med Chem*, vol. 64, no. 14, pp. 10047–10058, Jul. 2021, doi: 10.1021/acs.jmedchem.1c00319.
- [15] Z. Xia *et al.*, 'Rational Design of Hybrid SARS-CoV-2 Main Protease Inhibitors Guided by the Superimposed Cocrystal Structures with the Peptidomimetic Inhibitors GC-376, Telaprevir, and Boceprevir', *ACS Pharmacol Transl Sci*, vol. 4, no. 4, pp. 1408–1421, Aug. 2021, doi: 10.1021/acsphtsci.1c00099.
- [16] Y. R. Alugubelli *et al.*, 'A systematic exploration of boceprevir-based main protease inhibitors as SARS-CoV-2 antivirals', *Eur J Med Chem*, vol. 240, p. 114596, Oct. 2022, doi: 10.1016/j.ejmech.2022.114596.
- [17] S. Günther *et al.*, 'X-ray screening identifies active site and allosteric inhibitors of SARS-CoV-2 main protease', *Science* (1979), vol. 372, no. 6542, pp. 642–646, May 2021, doi: 10.1126/science.abf7945.
- [18] K. S. Yang *et al.*, 'A Quick Route to Multiple Highly Potent SARS-CoV-2 Main Protease Inhibitors**', *ChemMedChem*, vol. 16, no. 6, pp. 942–948, Mar. 2021, doi: 10.1002/cmdc.202000924.
- [19] E. Costanzi *et al.*, 'Structural and Biochemical Analysis of the Dual Inhibition of MG-132 against SARS-CoV-2 Main Protease (Mpro/3CLpro) and Human Cathepsin-L', *Int J Mol Sci*, vol. 22, no. 21, p. 11779, Oct. 2021, doi: 10.3390/ijms222111779.
- [20] Y. Ma *et al.*, 'A multi-pronged evaluation of aldehyde-based tripeptidyl main protease inhibitors as SARS-CoV-2 antivirals', *Eur J Med Chem*, vol. 240, p. 114570, Oct. 2022, doi: 10.1016/j.ejmech.2022.114570.
- [21] B. Bai *et al.*, 'Peptidomimetic α -Acylloxymethylketone Warheads with Six-Membered Lactam P1 Glutamine Mimic: SARS-CoV-2 3CL Protease Inhibition, Coronavirus Antiviral Activity, and *in Vitro* Biological Stability', *J Med Chem*, vol. 65, no. 4, pp. 2905–2925, Feb. 2022, doi: 10.1021/acs.jmedchem.1c00616.
- [22] S. Hattori *et al.*, 'A small molecule compound with an indole moiety inhibits the main protease of SARS-CoV-2 and blocks virus replication', *Nat Commun*, vol. 12, no. 1, p. 668, Jan. 2021, doi: 10.1038/s41467-021-20900-6.
- [23] N. Higashi-Kuwata *et al.*, 'Identification of SARS-CoV-2 Mpro inhibitors containing P1' 4-fluorobenzothiazole moiety highly active against SARS-CoV-2', *Nat Commun*, vol. 14, no. 1, p. 1076, Feb. 2023, doi: 10.1038/s41467-023-36729-0.
- [24] H. Yang *et al.*, 'Design, synthesis and biological evaluation of peptidomimetic benzothiazolyl ketones as 3CLpro inhibitors against SARS-CoV-2', *Eur J Med Chem*, vol. 257, p. 115512, Sep. 2023, doi: 10.1016/J.EJMECH.2023.115512.
- [25] W. Zhu *et al.*, 'Identification of SARS-CoV-2 3CL Protease Inhibitors by a Quantitative High-Throughput Screening', *ACS Pharmacol Transl Sci*, vol. 3, no. 5, pp. 1008–1016, Oct. 2020, doi: 10.1021/acsphtsci.0c00108.
- [26] A. Citarella *et al.*, 'Pseudo-Dipeptide Bearing α,α -Difluoromethyl Ketone Moiety as Electrophilic Warhead with Activity against Coronaviruses', *International Journal of Molecular Sciences* 2021, Vol. 22, Page 1398, vol. 22, no. 3, p. 1398, Jan. 2021, doi: 10.3390/IJMS22031398.
- [27] D. Shcherbakov *et al.*, 'Design and Evaluation of Bispidine-Based SARS-CoV-2 Main Protease Inhibitors', *ACS Med Chem Lett*, vol. 13, no. 1, pp. 140–147, Jan. 2022, doi: 10.1021/acsmedchemlett.1c00299.
- [28] J. K. Stille *et al.*, 'Design, synthesis and *in vitro* evaluation of novel SARS-CoV-2 3CLpro covalent inhibitors', *Eur J Med Chem*, vol. 229, p. 114046, Feb. 2022, doi: 10.1016/J.EJMECH.2021.114046.
- [29] N. Kitamura *et al.*, 'Expedited Approach toward the Rational Design of Noncovalent SARS-CoV-2 Main Protease Inhibitors', *J Med Chem*, vol. 65, no. 4, pp. 2848–2865, Feb. 2022, doi: 10.1021/ACS.JMEDCHEM.1C00509/SUPPL_FILE/JM1C00509_SI_002.CSV.
- [30] C. Ma *et al.*, 'Discovery of Di- and Trihaloacetamides as Covalent SARS-CoV-2 Main Protease Inhibitors with High Target Specificity', *J Am Chem Soc*, vol. 143, no. 49, pp. 20697–20709, Dec. 2021, doi: 10.1021/jacs.1c08060.

- [31] D. Yamane *et al.*, 'Selective covalent targeting of SARS-CoV-2 main protease by enantiopure chlorofluoroacetamide', *Chem Sci*, vol. 13, no. 10, pp. 3027–3034, 2022, doi: 10.1039/D1SC06596C.
- [32] M. Xiong, T. Nie, Q. Shao, M. Li, H. Su, and Y. Xu, 'In silico screening-based discovery of novel covalent inhibitors of the SARS-CoV-2 3CL protease', *Eur J Med Chem*, vol. 231, p. 114130, Mar. 2022, doi: 10.1016/j.ejmech.2022.114130.
- [33] A. Clyde *et al.*, 'High-Throughput Virtual Screening and Validation of a SARS-CoV-2 Main Protease Noncovalent Inhibitor', *J Chem Inf Model*, vol. 62, no. 1, pp. 116–128, Jan. 2022, doi: 10.1021/ACS.JCIM.1C00851/ASSET/IMAGES/LARGE/CI1C00851_0005.JPG.
- [34] S. Gao *et al.*, 'Discovery and Crystallographic Studies of Trisubstituted Piperazine Derivatives as Non-Covalent SARS-CoV-2 Main Protease Inhibitors with High Target Specificity and Low Toxicity', *J Med Chem*, vol. 65, no. 19, pp. 13343–13364, Oct. 2022, doi: 10.1021/ACS.JMEDCHEM.2C01146/SUPPL_FILE/JM2C01146_SI_002.CSV.
- [35] S. Gao *et al.*, 'Discovery and Crystallographic Studies of Nonpeptidic Piperazine Derivatives as Covalent SARS-CoV-2 Main Protease Inhibitors', *J Med Chem*, vol. 65, no. 24, pp. 16902–16917, Dec. 2022, doi: 10.1021/ACS.JMEDCHEM.2C01716/SUPPL_FILE/JM2C01716_SI_001.CSV.
- [36] P. Moon *et al.*, 'Discovery of Potent Pyrazoline-Based Covalent SARS-CoV-2 Main Protease Inhibitors', *bioRxiv*, p. 2022.03.05.483025, Mar. 2022, doi: 10.1101/2022.03.05.483025.
- [37] L. El Khoury *et al.*, 'Computationally driven discovery of SARS-CoV-2 Mpro inhibitors: from design to experimental validation', *Chem Sci*, vol. 13, no. 13, pp. 3674–3687, Mar. 2022, doi: 10.1039/D1SC05892D.
- [38] L. Zhang *et al.*, 'Crystal structure of SARS-CoV-2 main protease provides a basis for design of improved α -ketoamide inhibitors', *Science* (1979), vol. 368, no. 6489, pp. 409–412, Apr. 2020, doi: 10.1126/science.abb3405.
- [39] B. X. Quan *et al.*, 'An orally available Mpro inhibitor is effective against wild-type SARS-CoV-2 and variants including Omicron', *Nature Microbiology* 2022 7:5, vol. 7, no. 5, pp. 716–725, Apr. 2022, doi: 10.1038/s41564-022-01119-7.
- [40] X. Chen *et al.*, 'Inhibition mechanism and antiviral activity of an α -ketoamide based SARS-CoV-2 main protease inhibitor', *bioRxiv*, p. 2023.03.09.531862, Mar. 2023, doi: 10.1101/2023.03.09.531862.
- [41] C. Huang *et al.*, 'A new generation Mpro inhibitor with potent activity against SARS-CoV-2 Omicron variants', *Signal Transduction and Targeted Therapy* 2023 8:1, vol. 8, no. 1, pp. 1–13, Mar. 2023, doi: 10.1038/s41392-023-01392-w.
- [42] Z. Jin *et al.*, 'Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors', *Nature* 2020 582:7811, vol. 582, no. 7811, pp. 289–293, Apr. 2020, doi: 10.1038/s41586-020-2223-y.
- [43] S. Iketani *et al.*, 'Lead compounds for the development of SARS-CoV-2 3CL protease inhibitors', *Nature Communications* 2021 12:1, vol. 12, no. 1, pp. 1–7, Apr. 2021, doi: 10.1038/s41467-021-22362-2.
- [44] G. Amendola *et al.*, 'Lead Discovery of SARS-CoV-2 Main Protease Inhibitors through Covalent Docking-Based Virtual Screening', *J Chem Inf Model*, vol. 61, no. 4, pp. 2062–2073, Apr. 2021, doi: 10.1021/acs.jcim.1c00184.
- [45] S. Previti *et al.*, 'Structure-based lead optimization of peptide-based vinyl methyl ketones as SARS-CoV-2 main protease inhibitors', *Eur J Med Chem*, vol. 247, p. 115021, Feb. 2023, doi: 10.1016/j.ejmech.2022.115021.
- [46] S. Mondal *et al.*, 'Dual Inhibitors of Main Protease (M^{Pro}) and Cathepsin L as Potent Antivirals against SARS-CoV2', *J Am Chem Soc*, vol. 144, no. 46, pp. 21035–21045, Nov. 2022, doi: 10.1021/jacs.2c04626.
- [47] A. Citarella *et al.*, 'Synthesis of SARS-CoV-2 Mpro inhibitors bearing a cinnamic ester warhead with in vitro activity against human coronaviruses', *Org Biomol Chem*, 2023, doi: 10.1039/D3OB00381G.

- [48] D. Zaidman, P. Gehrtz, M. Filep, M. A. Walsh, F. Von Delft, and N. London Correspondence, 'An automatic pipeline for the design of irreversible derivatives identifies a potent SARS-CoV-2 M pro inhibitor', *Cell Chem Biol*, vol. 28, pp. 1795–1806.e5, 2021, doi: 10.1016/j.chembiol.2021.05.018.
- [49] M. Kuzikov *et al.*, 'Identification of Inhibitors of SARS-CoV-2 3CL-Pro Enzymatic Activity Using a Small Molecule in Vitro Repurposing Screen', *ACS Pharmacol Transl Sci*, vol. 4, no. 3, pp. 1096–1110, Jun. 2021, doi: 10.1021/ACSPTSCI.0C00216/ASSET/IMAGES/LARGE/PT0C00216_0007.jpeg.
- [50] H. Su *et al.*, 'Identification of pyrogallol as a warhead in design of covalent inhibitors for the SARS-CoV-2 3CL protease', *Nature Communications* 2021 12:1, vol. 12, no. 1, pp. 1–12, Jun. 2021, doi: 10.1038/s41467-021-23751-3.
- [51] S. I. Hattori *et al.*, 'GRL-0920, an indole chloropyridinyl ester, completely blocks SARS-CoV-2 infection', *mBio*, vol. 11, no. 4, pp. 1–16, Jul. 2020, doi: 10.1128/MBIO.01833-20/ASSET/11D12B15-E56A-4381-A5F9-AD6A79F97BE2/ASSETS/GRAFIC/MBIO.01833-20-F0005.jpeg.
- [52] A. K. Ghosh *et al.*, 'Indole Chloropyridinyl Ester-Derived SARS-CoV-2 3CLpro Inhibitors: Enzyme Inhibition, Antiviral Efficacy, Structure-Activity Relationship, and X-ray Structural Studies', *J Med Chem*, vol. 64, no. 19, pp. 14702–14714, Oct. 2021, doi: 10.1021/ACS.JMEDCHEM.1C01214/SUPPL_FILE/JM1C01214_SI_002.csv.
- [53] J. Breidenbach *et al.*, 'Targeting the Main Protease of SARS-CoV-2: From the Establishment of High Throughput Screening to the Design of Tailored Inhibitors', *Angewandte Chemie International Edition*, vol. 60, no. 18, pp. 10423–10429, Apr. 2021, doi: 10.1002/ANIE.202016961.
- [54] T. Pillaiyar *et al.*, 'Small-Molecule Thioesters as SARS-CoV-2 Main Protease Inhibitors: Enzyme Inhibition, Structure–Activity Relationships, Antiviral Activity, and X-ray Structure Determination', *J Med Chem*, vol. 65, no. 13, pp. 9376–9395, Jul. 2022, doi: 10.1021/acs.jmedchem.2c00636.
- [55] M. Zmudzinski *et al.*, 'Ebselen derivatives are very potent dual inhibitors of SARS-CoV-2 proteases - PLpro and Mpro in in vitro studies', *bioRxiv*, p. 2020.08.30.273979, Aug. 2020, doi: 10.1101/2020.08.30.273979.
- [56] S. Huff *et al.*, 'Discovery and Mechanism of SARS-CoV-2 Main Protease Inhibitors', *J Med Chem*, vol. 65, no. 4, pp. 2866–2879, Feb. 2022, doi: 10.1021/ACS.JMEDCHEM.1C00566/SUPPL_FILE/JM1C00566_SI_001.csv.
- [57] K. Amporndanai *et al.*, 'Inhibition mechanism of SARS-CoV-2 main protease by ebselen and its derivatives', *Nature Communications* 2021 12:1, vol. 12, no. 1, pp. 1–7, May 2021, doi: 10.1038/s41467-021-23313-7.
- [58] L. Y. Sun *et al.*, 'Ebsulfur and Ebselen as highly potent scaffolds for the development of potential SARS-CoV-2 antivirals', *Bioorg Chem*, vol. 112, p. 104889, Jul. 2021, doi: 10.1016/J.BIOORG.2021.104889.
- [59] W. Chen *et al.*, 'Discovery of highly potent SARS-CoV-2 Mpro inhibitors based on benzoisothiazolone scaffold', *Bioorg Med Chem Lett*, vol. 58, p. 128526, Feb. 2022, doi: 10.1016/j.bmcl.2022.128526.
- [60] Y. Unoh *et al.*, 'Discovery of S-217622, a Noncovalent Oral SARS-CoV-2 3CL Protease Inhibitor Clinical Candidate for Treating COVID-19', *J Med Chem*, vol. 65, no. 9, pp. 6499–6512, May 2022, doi: 10.1021/acs.jmedchem.2c00117.
- [61] J. Yang *et al.*, 'Structure-Based Discovery of Novel Nonpeptide Inhibitors Targeting SARS-CoV-2 M^{pro}', *J Chem Inf Model*, vol. 61, no. 8, pp. 3917–3926, Aug. 2021, doi: 10.1021/acs.jcim.1c00355.
- [62] S. H. Han *et al.*, 'Structure-Based Optimization of ML300-Derived, Noncovalent Inhibitors Targeting the Severe Acute Respiratory Syndrome Coronavirus 3CL Protease (SARS-CoV-2 3CLpro)', *J Med Chem*, vol. 65, no. 4, pp. 2880–2904, Feb. 2022, doi: 10.1021/ACS.JMEDCHEM.1C00598/SUPPL_FILE/JM1C00598_SI_002.csv.
- [63] S. A. Elseginy *et al.*, 'Promising anti-SARS-CoV-2 drugs by effective dual targeting against the viral and host proteases', *Bioorg Med Chem Lett*, vol. 43, p. 128099, Jul. 2021, doi: 10.1016/J.BMCL.2021.128099.

- [64] A. Luttens *et al.*, 'Ultralarge Virtual Screening Identifies SARS-CoV-2 Main Protease Inhibitors with Broad-Spectrum Activity against Coronaviruses', *J Am Chem Soc*, vol. 144, no. 7, pp. 2905–2920, Feb. 2022, doi: 10.1021/JACS.1C08402/ASSET/IMAGES/LARGE/JA1C08402_0004.jpeg.
- [65] G. G. Rossetti *et al.*, 'Non-covalent SARS-CoV-2 Mpro inhibitors developed from in silico screen hits', *Scientific Reports* 2022 12:1, vol. 12, no. 1, pp. 1–9, Feb. 2022, doi: 10.1038/s41598-022-06306-4.
- [66] K. Zhang *et al.*, 'Discovery of quinazolin-4-one-based non-covalent inhibitors targeting the severe acute respiratory syndrome coronavirus 2 main protease (SARS-CoV-2 Mpro)', *Eur J Med Chem*, vol. 257, p. 115487, Sep. 2023, doi: 10.1016/j.ejmech.2023.115487.
- [67] A. Citarella *et al.*, 'Discovery of a Novel Trifluoromethyl Diazirine Inhibitor of SARS-CoV-2 Mpro', *Molecules*, vol. 28, no. 2, Jan. 2023, doi: 10.3390/MOLECULES28020514.
- [68] H. A. Alhadrami *et al.*, 'Neoechinulin A as a Promising SARS-CoV-2 Mpro Inhibitor: In Vitro and In Silico Study Showing the Ability of Simulations in Discerning Active from Inactive Enzyme Inhibitors', *Mar Drugs*, vol. 20, no. 3, p. 163, Mar. 2022, doi: 10.3390/MD20030163/S1.
- [69] Y. Zhang *et al.*, 'Structure-Based Discovery and Structural Basis of a Novel Broad-Spectrum Natural Product against the Main Protease of Coronavirus', *J Virol*, vol. 96, no. 1, pp. 1253–1274, Jan. 2022, doi: 10.1128/JVI.01253-21/SUPPL_FILE/JVI.01253-21-S0001.PDF.
- [70] J. W. Zhang *et al.*, 'Discovery of 9,10-dihydrophenanthrene derivatives as SARS-CoV-2 3CLpro inhibitors for treating COVID-19', *Eur J Med Chem*, vol. 228, p. 114030, Jan. 2022, doi: 10.1016/J.EJMECH.2021.114030.
- [71] S. K. Samrat *et al.*, 'Allosteric inhibitors of the main protease of SARS-CoV-2', *Antiviral Res*, vol. 205, p. 105381, Sep. 2022, doi: 10.1016/j.antiviral.2022.105381.
- [72] O. A. Chaves *et al.*, 'Apixaban, an orally available anticoagulant, inhibits SARS-CoV-2 replication and its major protease in a non-competitive way', *J Mol Cell Biol*, vol. 14, no. 6, Aug. 2022, doi: 10.1093/jmcb/mjac039.