

Article

Efficient Stereo-Selective Fluorination on Vitamin D₃ Side-Chain Using Electrophilic Fluorination

 Fumihiro Kawagoe, Sayuri Mototani and Atsushi Kittaka * 

Faculty of Pharmaceutical Sciences, Teikyo University, 2-11-1 Kaga, Itabashi, Tokyo 173-8605, Japan; fkawagoe@pharm.teikyo-u.ac.jp (F.K.); 19dy10003vu@stu.teikyo-u.ac.jp (S.M.)

* Correspondence: akittaka@pharm.teikyo-u.ac.jp; Tel.: +81-3-3964-8109; Fax: +81-3-3964-8117

Abstract: Our research regarding side-chain fluorinated vitamin D₃ analogues has explored a series of efficient fluorination methods. In this study, a new electrophilic stereo-selective fluorination methodology at C24 and C22 positions of the vitamin D₃ side-chain was developed using *N*-fluorobenzenesulfonimide (NFSI) and CD-ring imides with an Evans chiral auxiliary (26,27,30).

Keywords: electrophilic fluorination; *N*-fluorobenzenesulfonimide (NFSI); side-chain fluorinated vitamin D₃; side-chain; Evans chiral auxiliary

1. Introduction

Regio- and stereo-selective fluorination of the vitamin D₃ side-chain is one of the efficient methods used to modulate biological activities, such as binding affinity to the vitamin D receptor (VDR), transactivation activity through the VDR, and metabolic stability against CYP24A1 [1]. We have reported the regio- and stereo-selective fluorination of the CD-ring side-chain starting from the Inhoffen–Lythgoe diol (1) and constructed C26,27-fluoro-CD-rings (2–4) [2,3], C24-fluoro-CD-rings (5–7) [4,5], C23-fluoro-CD-rings (8–10) [6,7], and C22-fluoro-CD-rings (11–13) [8] (Figure 1) to create a chemical library of side-chain fluorovitamin D₃ analogues using a convergent method (Scheme 1) [9–11].

1.1. Previous Studies of C24-Stereoselective Fluorination

On the vitamin D₃ side-chain, the C24 position is an important site for metabolic inactivation via CYP24A1 hydroxylation [12–14]. Therefore, structural modifications, especially fluorination and difluorination, of the C24 position have been actively investigated [15–25].



Citation: Kawagoe, F.; Mototani, S.; Kittaka, A. Efficient Stereo-Selective Fluorination on Vitamin D₃ Side-Chain Using Electrophilic Fluorination. *Biomolecules* **2024**, *14*, 37. <https://doi.org/10.3390/biom14010037>

Academic Editor: Thomas Müller

Received: 6 December 2023

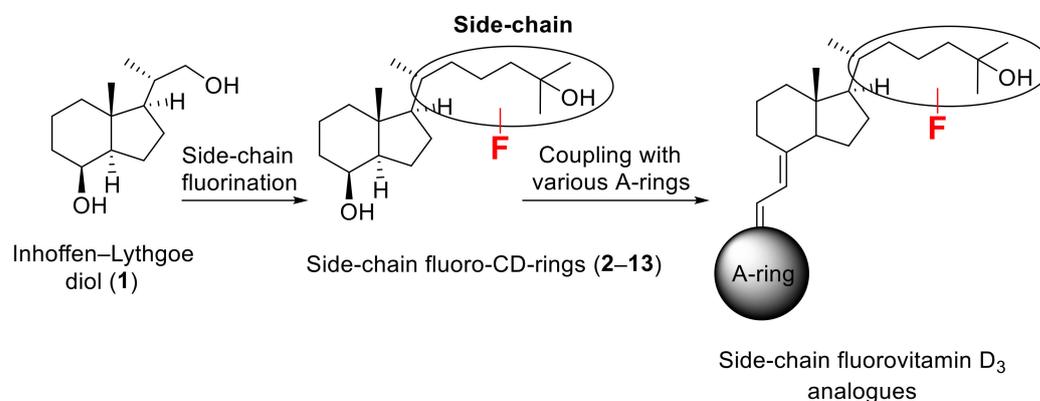
Revised: 21 December 2023

Accepted: 22 December 2023

Published: 26 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).



Scheme 1. Convergent synthesis of side-chain fluorinated vitamin D₃ analogues via side-chain fluoro-CD-ring fragments (2–13) starting from the Inhoffen–Lythgoe diol (1).

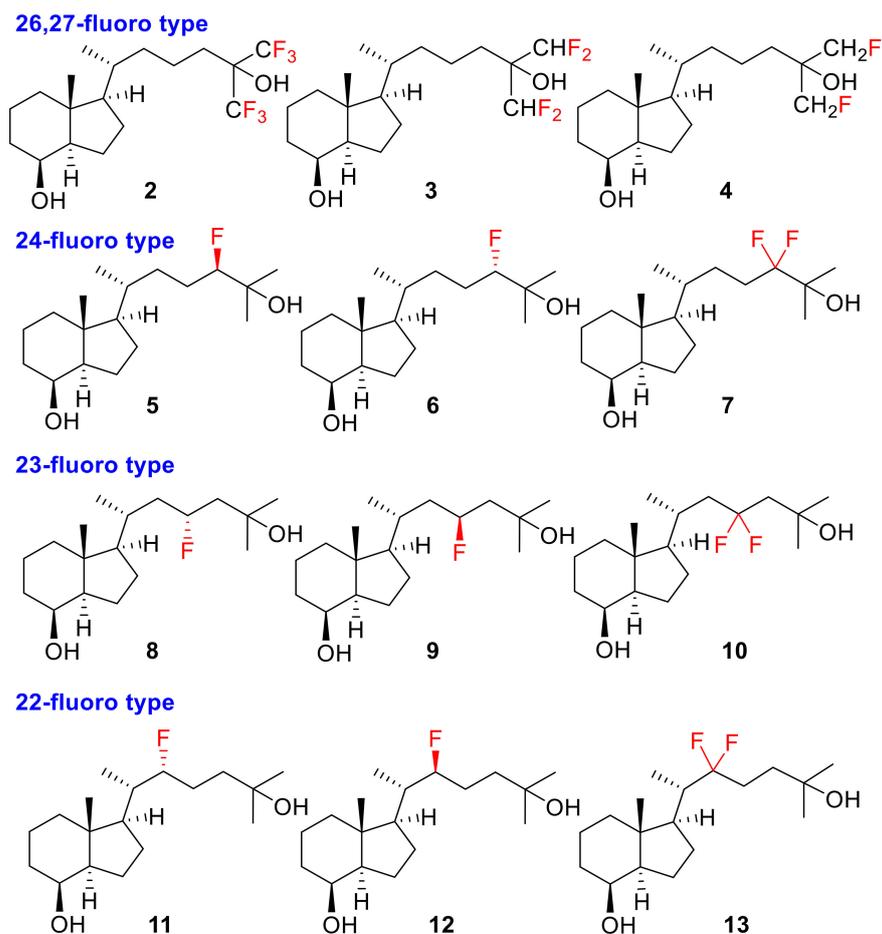
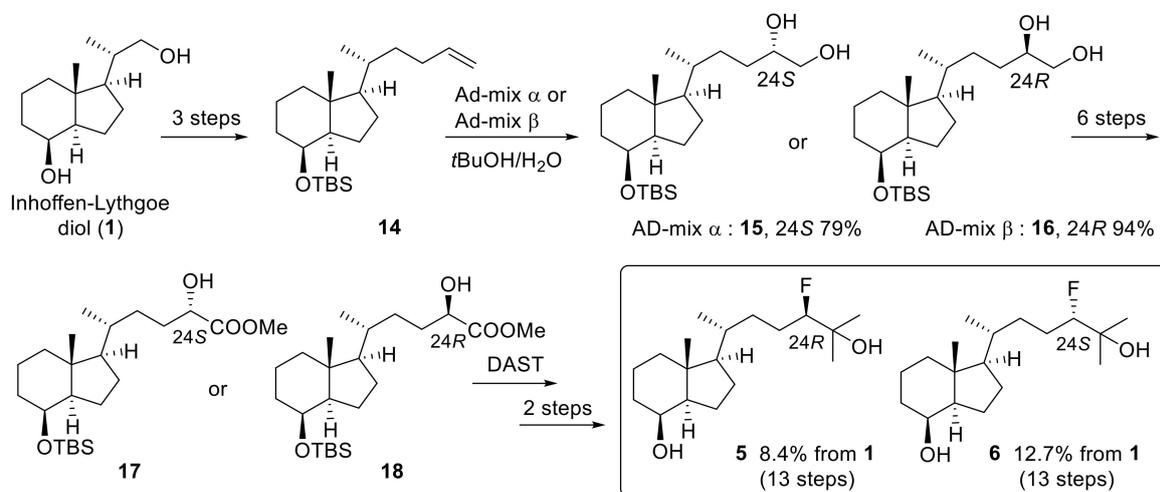


Figure 1. Structures of side-chain fluorinated CD-rings (2–13).

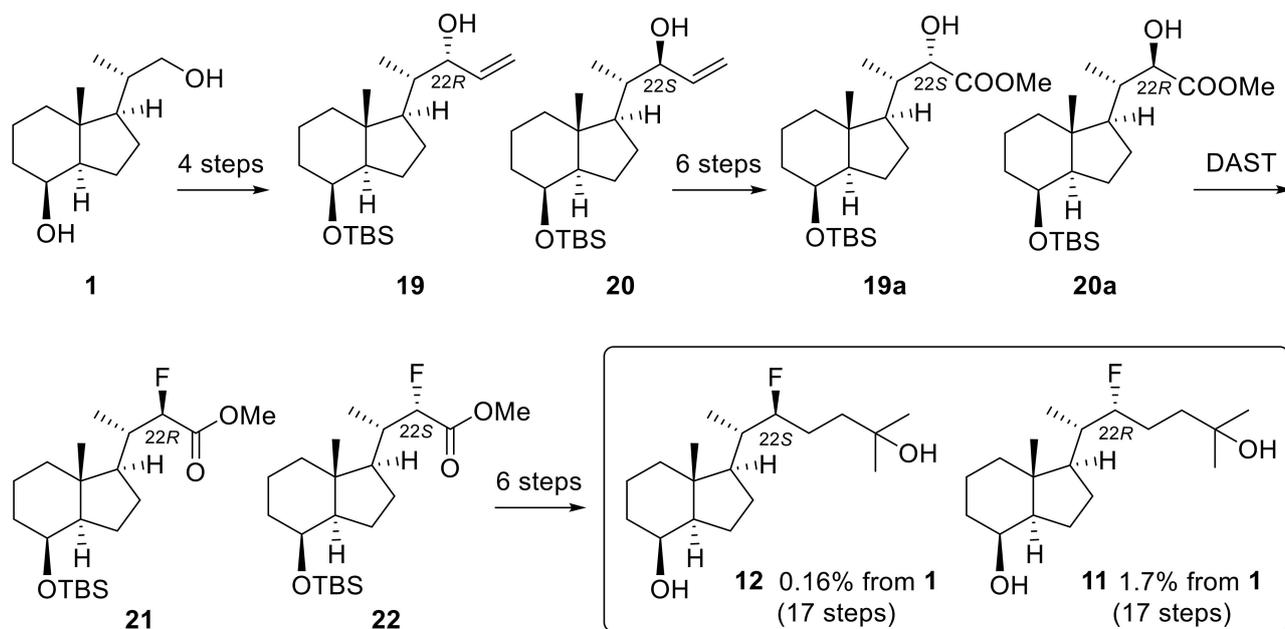
We previously reported the stereo-selective fluorination method at the C24 position of the CD-ring side-chain for stereo-selective synthesis of (24*R*)-fluoro-25-hydroxyvitamin D₃ and its 24*S* isomer [5]. The key step for introducing the C24-fluoro unit was achieved via osmium-catalyzed diastereoselective dihydroxylation followed by deoxy-fluorination reactions. However, multi-step synthesis was required (13 steps), and harmful osmium and excess amounts of the fluorinating reagent, DAST, were used in this synthetic route (Scheme 2).



Scheme 2. Synthetic route to C24-fluoro-CD-rings (5,6) using dihydroxylation/deoxyfluorination reactions.

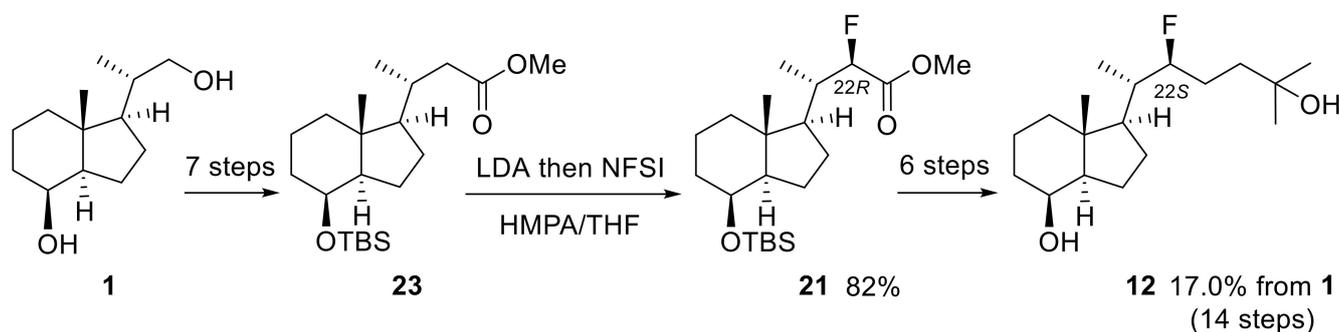
1.2. Previous Studies of C22-Stereoselective Fluorination

Until 2023, there was only one example of 22-fluorovitamin D synthesis, and the stereochemistry of C22 was unknown [26]. We recently reported the first stereo-selective fluorination at the C22 position of the CD-ring side-chain and synthesis of (22*R*)- and (22*S*)-fluoro-25-hydroxyvitamin D₃ [8]. However, multiple synthetic steps were required, including separation of the diastereomer mixture of C22-allyl alcohols (**19**,**20**) and the deoxyfluorination step using DAST, resulting in low overall yields (Scheme 3).



Scheme 3. Preparation of C22-fluoro-CD-rings (**11**,**12**) using DAST for the deoxyfluorination step.

In the report [8], we also developed an alternative C22-fluorination methodology using a cationic fluorination reaction between the methyl ester (**23**) and *N*-fluorobenzenesulfonylimide (NFSI) under basic conditions. However, this route yielded only the (22*R*)-fluoro product (**21**); efficient cationic fluorination of the (22*S*)-fluoro counterpart to yield the (22*R*)-CD-ring (**11**) has yet to be explored (Scheme 4).

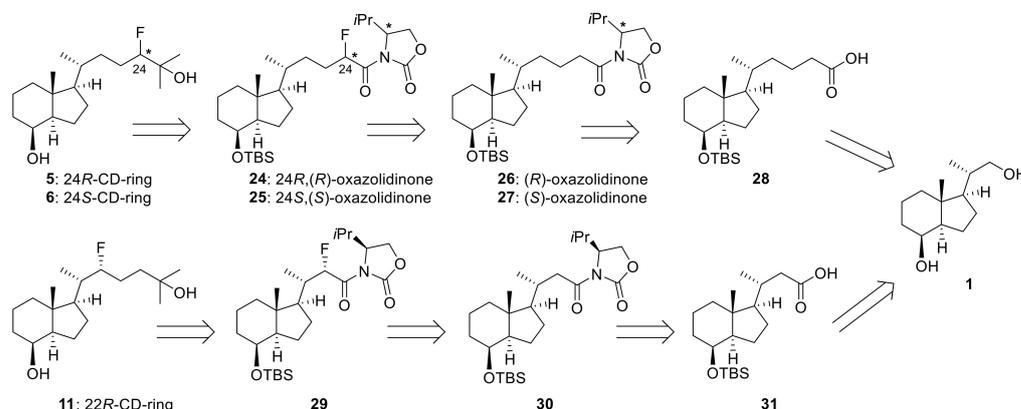


Scheme 4. Electrophilic fluorination at the C22-position of the CD-ring side-chain.

To overcome these drawbacks in order to introduce C24- and C22-monofluoro moieties, here we describe an alternative stereo-selective fluorination methodology at these positions via electrophilic fluorination using an Evans chiral auxiliary-based aldol reaction with NFSI [27–29].

The retrosynthetic paths are shown in Scheme 5. Evans oxazolidinone ligands could be introduced to the side-chain carboxylic acids (**28**,**31**), which were synthesized from the

Inhoffen–Lythgoe diol (**1**), and NFSI could be used as a cationic fluorination reagent in the presence of lithium hexamethyldisilazide (LHMDS).

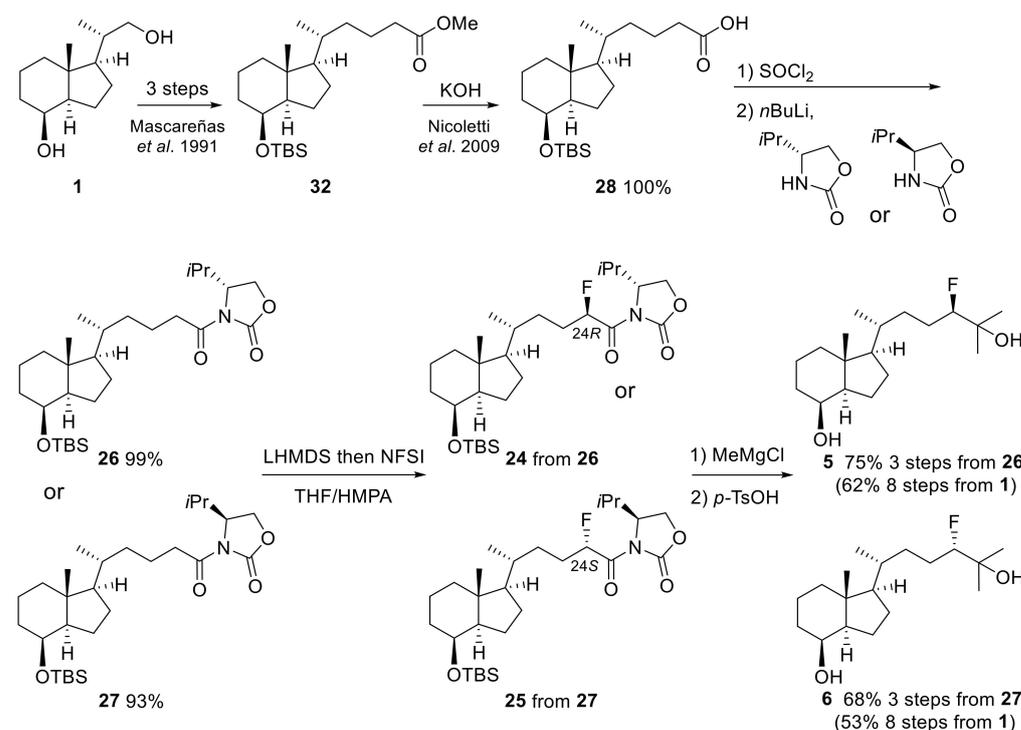


Scheme 5. Synthetic plans for the C24- and C22-fluoro-CD-rings (**5,6,11**). * means a chiral center.

2. Results and Discussion

2.1. Synthesis of 24-Fluoro-CD-Rings (**5,6**) via Electrophilic Fluorination

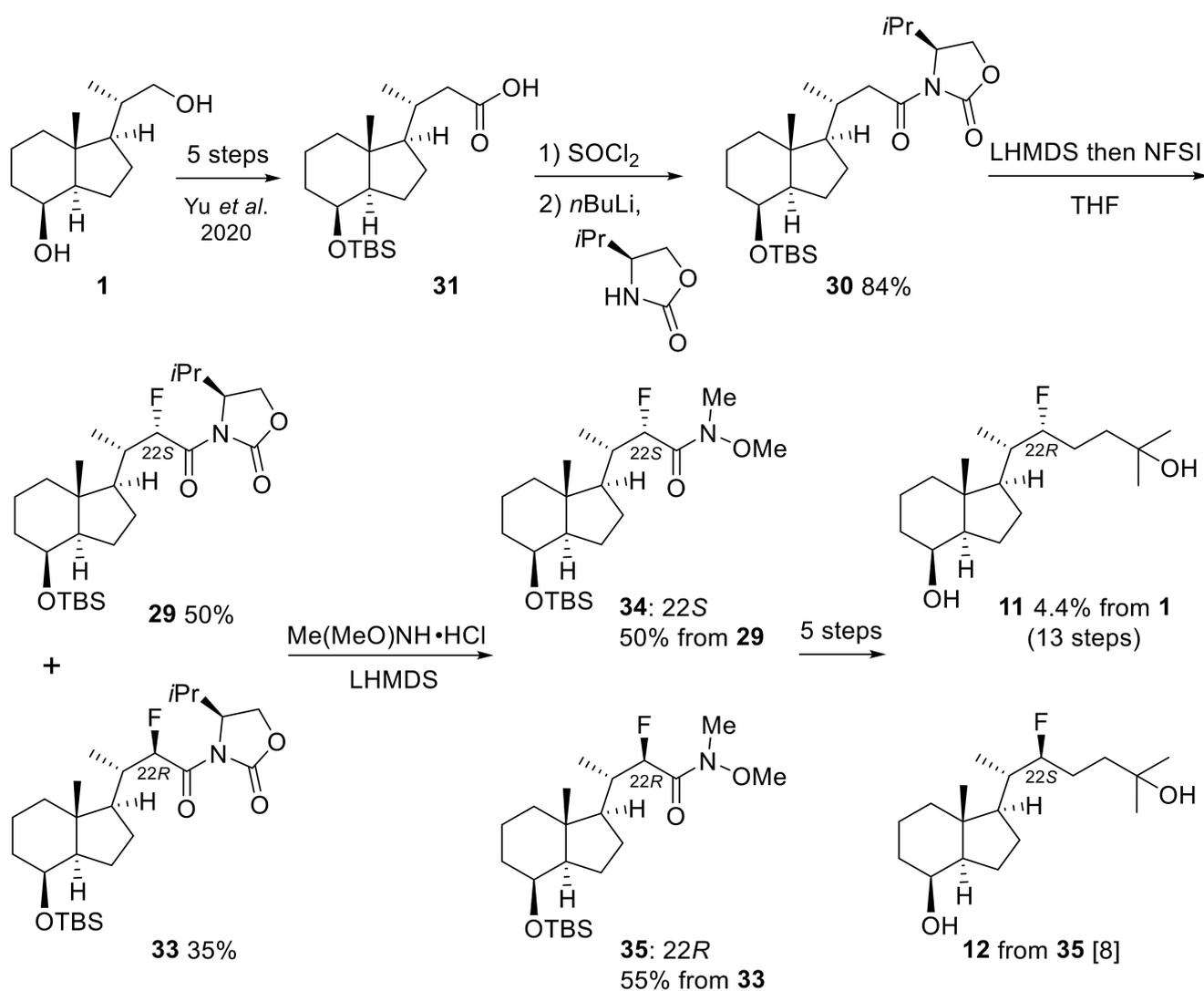
The synthetic route of the 24-fluoro-CD-rings (**5,6**) is illustrated in Scheme 6. We synthesized methyl ester (**32**) from the Inhoffen–Lythgoe diol (**1**) in three steps [30], and hydrolysis of **32** afforded the corresponding carboxylic acid (**28**) [31], which was subsequently converted to *N*-acyloxazolidinones (**26,27**) via acid chlorides. The C24-position was effectively fluorinated in a high diastereoselective manner to yield (24*R*)-product **24** from **26** and (24*S*)-product **25** from **27**, respectively. Displacement of the oxazolidinone ligand to 26,27-dimethyl units with the 25-hydroxy group using an excess amount of methyl magnesium chloride successfully proceeded with preservation of the stereochemistry at the C24-position, and removal of the silyl protective group under acidic conditions yielded the target (24*R*)-fluoro-CD-ring **5** (62% in 8 steps) and (24*S*)-fluoro-CD-ring **6** (53%).



Scheme 6. Improved synthesis of 24-fluoro-CD-rings (**5,6**) via electrophilic fluorination [30,31].

2.2. Synthesis of (22R)-Fluoro-CD-Ring (11) via Electrophilic Fluorination

The synthetic route to the (22R)-fluoro-CD-ring (11) started from carboxylic acid (31), which was readily available from the Inhoffen–Lythgoe diol in five steps [32]. This was converted to amide (30) followed by fluorination at the C22-position using NFSI in the presence of LHMDS to give the desired (22S)-fluoro-CD-ring (29) as a major product, along with (22R)-fluoro-CD-ring (33) at a ratio of 1.4:1. After separating 29 and 33 using silica gel column chromatography, they were converted to the corresponding Weinreb amides (34,35), which were the synthetic intermediates of C22-fluoro-CD-rings 11 and 12 [8], using methoxy(methyl)amine hydrochloride and LHMDS in moderate yields and with a rigid stereochemical integrity (Scheme 7).



Scheme 7. Improved synthesis of (22R)-fluoro-CD-ring (11) [32].

3. Conclusions

In conclusion, we developed an improved stereo-selective fluorination method at C24 and C22 positions to synthesize C24- and C22-fluoro-CD-rings utilizing an Evans chiral auxiliary for a cationic fluorination reaction as a key step. The side-chain carboxylic acids (28,31) were prepared from the Inhoffen–Lythgoe diol. These synthetic routes are more efficient than previous ones reported regarding both total yields and reaction steps.

Experimental Section

^1H and ^{13}C NMR spectra were recorded on JEOL AL-400 NMR (400 MHz) and ECP-600 NMR (600 MHz) spectrometers (Tokyo, Japan). ^1H NMR spectra were referenced using $(\text{CH}_3)_4\text{Si}$ (δ 0.00 ppm) or CHCl_3 (δ 7.26 ppm) as an internal standard. ^{13}C NMR spectra were referenced using deuterated solvent (δ 77.0 ppm for CDCl_3). IR spectra were recorded using a JASCO FT-IR-800 Fourier transform infrared spectrophotometer (Tokyo, Japan). High-resolution mass spectra were obtained using a SHIMADZU LCMS-IT-TOF mass spectrometer (Kyoto, Japan) using a positive electrospray ionization (ESI) method. Optical rotations were measured using a JASCO DIP-370 digital polarimeter (Tokyo, Japan). Column chromatography was performed on silica gel 60N (Kanto Chemical Co., Inc., 40–50 μm , Tokyo, Japan) or silica gel 60 (Merck, 0.040–0.063 mm, Tokyo, Japan). All experiments were performed under anhydrous conditions in an atmosphere of argon, unless otherwise stated. Supporting information regarding ^1H and ^{13}C NMR spectra of all new compounds (**26**, **27**, **29**, **30**, and **33**) is available via the link in Supplementary Materials.

(4R)-3-[(5R)-5-[(1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl]hexanoyl]-4-isopropylloxazolidin-2-one (**26**)

Carboxylic acid **28** [31] (403.6 mg, 1.05 mmol) was dissolved in thionyl chloride (3.0 mL, 4.97 g, 41.8 mmol) at room temperature, and the solution was refluxed for 1 h 30 min. The mixture was evaporated in vacuo, and the crude acyl chloride was used for the next reaction without further purification.

*n*BuLi (1.59 M in hexane, 5.3 mL, 8.40 mmol) was added to a precooled solution of (*R*)-4-isopropylloxazolidinone (1.09 g, 8.40 mmol) in THF (20 mL) at -78°C , and the mixture was stirred at the same temperature for 10 min. The above acyl chloride in THF (3 mL) was added to the solution and stirred at the same temperature for 10 min. After the reaction was quenched with H_2O and saturated aqueous NH_4Cl at room temperature, the mixture was extracted with EtOAc three times. The organic layer was dried over Na_2SO_4 , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 3:1) to obtain **26** (513.6 mg, 99%, in 2 steps) as a colorless oil.

26: $[\alpha]_{\text{D}}^{27} + 6.1$ (c 2.01, CHCl_3); IR (neat) 1786, 1704, 1383, 1252, 1208, 1085, 1023, 841 cm^{-1} ; ^1H NMR (600 MHz, CDCl_3) δ -0.02 (s, 3H), -0.01 (s, 3H), 0.86–0.91 (m, 21H), 1.02–1.12 (m, 3H), 1.18–1.25 (m, 2H), 1.29–1.44 (m, 5H), 1.47–1.56 (m, 2H), 1.64–1.66 (m, 1H), 1.69–1.83 (m, 3H), 1.92–1.95 (m, 1H), 2.34–2.39 (m, 1H), 2.80–2.95 (m, 2H), 3.97–3.99 (m, 1H), 4.18–4.27 (m, 2H), 4.42 (dt, $J = 3.0, 8.4$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ -5.2 , -4.8 , 13.7, 14.6, 17.6, 18.0, 18.0, 18.5, 21.0, 23.0, 25.8, 27.2, 28.4, 34.4, 35.1, 35.2, 35.9, 40.7, 42.1, 53.0, 56.5, 58.4, 63.3, 69.4, 154.0, 173.4; HRMS (ESI $^+$) calcd for $\text{C}_{28}\text{H}_{51}\text{NO}_4\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 516.3480, found 516.3489.

(4S)-3-[(5R)-5-[(1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl]hexanoyl]-4-isopropylloxazolidin-2-one (**27**)

Carboxylic acid **28** (606.2 mg, 1.58 mmol) was dissolved in thionyl chloride (3.4 mL, 5.6 g, 47.0 mmol) at room temperature, and the solution was refluxed for 1 h 30 min. The mixture was evaporated in vacuo, and the crude acyl chloride was used for the next reaction without further purification.

*n*BuLi (1.59 M in hexane, 5.9 mL, 9.42 mmol) was added to a precooled solution of (*S*)-4-isopropylloxazolidinone (1.24 g, 9.60 mmol) in THF (20 mL) at -78°C , and the mixture was stirred at the same temperature for 10 min. The above acyl chloride in THF (4 mL) was added to the solution and stirred at the same temperature for 20 min. After the reaction was quenched with H_2O and saturated aqueous NH_4Cl at room temperature, the mixture was extracted with EtOAc three times. The organic layer was dried over Na_2SO_4 , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 3:1) to obtain **27** (729.5 mg, 93%, in 2 steps) as a colorless oil.

27: $[\alpha]_{\text{D}}^{27} + 78.4$ (c 1.58, CHCl_3); IR (neat) 1786, 1704, 1387, 1255, 1208, 1089, 1027, 841 cm^{-1} ; ^1H NMR (600 MHz, CDCl_3) δ -0.02 (s, 3H), 0.00 (s, 3H), 0.86–0.92 (m, 21H), 1.00–1.11 (m, 3H), 1.19–1.25 (m, 2H), 1.29–1.44 (m, 5H), 1.49–1.57 (m, 2H), 1.61–1.83 (m, 4H),

1.92–1.95 (m, 1H), 2.33–2.40 (m, 1H), 2.76–2.99 (m, 2H), 3.97–3.99 (m, 1H), 4.18–4.27 (m, 2H), 4.42 (dt, $J = 3.3, 8.4$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ –5.2, –4.8, 13.7, 14.6, 17.6, 18.0, 18.0, 18.5, 21.0, 23.0, 25.8, 27.2, 28.4, 34.4, 35.1, 35.2, 36.0, 40.7, 42.1, 53.0, 56.5, 58.3, 63.3, 69.4, 154.1, 173.5; HRMS (ESI⁺) calcd for $\text{C}_{28}\text{H}_{51}\text{NO}_4\text{SiNa}$ $[\text{M}+\text{Na}]^+$ 516.3480, found 516.3495.

(1R,3aR,4S,7aR)-1-[(2R,5R)-5-Fluoro-6-hydroxy-6-methylheptan-2-yl]-7a-methyloctahydro-1H-inden-4-ol (5)

To a solution of **26** (168.9 mg, 0.34 mmol) in THF (2 mL) and hexamethylphosphoric triamide (HMPA) (200 μL) was added LHMDs (lithium hexamethyldisilazide) (359 μL , 1 M THF solution, 0.36 mmol) at -78°C , the mixture was stirred at the same temperature for 10 min, and *N*-fluorobenzenesulfonimide (NFSI) (117.8 mg, 0.37 mmol) was added. After being stirred at the same temperature for 40 min, the reaction mixture was quenched with H_2O and saturated aqueous NH_4Cl at -78°C . The mixture was extracted with EtOAc three times. The organic layer was washed with brine, dried over Na_2SO_4 , filtered, and concentrated. Crude **24** was used for the next reaction without further purification.

To a solution of the above crude **24** in THF (10 mL) was added MeMgCl (1.14 mL, 3.0 M THF solution, 3.42 mmol) at 0°C , and the mixture was stirred at 0°C for 10 min. After the reaction was quenched with H_2O , the mixture was extracted with EtOAc three times. The organic layer was washed with saturated aqueous NH_4Cl , dried over Na_2SO_4 , filtered, and concentrated. The crude residue was used for the next reaction without further purification.

To the above crude residue in MeOH (15 mL) was added *p*-toluenesulfonic acid monohydrate (479.9 mg, 2.52 mmol), and the mixture was stirred at room temperature for 63 h under air. After the reaction was quenched with H_2O and saturated aqueous NaHCO_3 at room temperature, the mixture was extracted with EtOAc three times. The organic layer was dried over Na_2SO_4 , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 2:1) to obtain **5** (77.5 mg, 75%, in 3 steps) as a white powder. The spectral data matched with those reported in the literature [5].

(1R,3aR,4S,7aR)-1-[(2R,5S)-5-Fluoro-6-hydroxy-6-methylheptan-2-yl]-7a-methyloctahydro-1H-inden-4-ol (6)

To a solution of **27** (111.0 mg, 0.23 mmol) in THF (2 mL) and hexamethylphosphoric triamide (HMPA) (200 μL) was added LHMDs (lithium hexamethyldisilazide) (236 μL , 1 M THF solution, 0.24 mmol) at -78°C , the mixture was stirred at the same temperature for 10 min, and *N*-fluorobenzenesulfonimide (NFSI) (76.4 mg, 0.24 mmol) was added. After being stirred at the same temperature for 40 min, the reaction mixture was quenched with H_2O and saturated aqueous NH_4Cl at -78°C . The mixture was extracted with EtOAc three times. The organic layer was washed with brine, dried over Na_2SO_4 , filtered, and concentrated. Crude **25** was used for the next reaction without further purification.

To a solution of the above crude **25** in THF (7 mL) was added MeMgCl (0.75 mL, 3.0 M THF solution, 2.25 mmol) at 0°C , and the mixture was stirred at 0°C for 10 min. After the reaction was quenched with H_2O , the mixture was extracted with EtOAc three times. The organic layer was washed with saturated aqueous NH_4Cl , dried over Na_2SO_4 , filtered, and concentrated. The crude residue was used for the next reaction without further purification.

To the above crude residue in MeOH (10 mL) was added *p*-toluenesulfonic acid monohydrate (558.3 mg, 2.94 mmol), and the mixture was stirred at room temperature for 18 h under air. After the reaction was quenched with H_2O and saturated aqueous NaHCO_3 at room temperature, the mixture was extracted with EtOAc three times. The organic layer was dried over Na_2SO_4 , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 2:1) to obtain **6** (45.9 mg, 68%, in 3 steps) as a white powder. The spectral data matched with those reported in the literature [5].

(4S)-3-[(3R)-3-[(1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl]butanoyl]-4-isopropylloxazolidin-2-one (30)

Carboxylic acid **31** [32] (613.8 mg, 1.73 mmol) was dissolved in thionyl chloride (2.5 mL, 4.12 g, 34.6 mmol) at room temperature, and the solution was refluxed for 1 h 30 min. The mixture was evaporated in vacuo, and the crude acyl chloride was used for the next reaction without further purification.

*n*BuLi (1.59 M in hexane, 3.3 mL, 5.19 mmol) was added to a precooled solution of (S)-4-isopropylloxazolidinone (673.3 mg, 5.21 mmol) in THF (6 mL) at $-78\text{ }^{\circ}\text{C}$, and the mixture was stirred at the same temperature for 15 min. The above acyl chloride in THF (6 mL) was added to the solution and stirred at the same temperature for 15 min. After the reaction was quenched with H₂O and saturated aqueous NH₄Cl at room temperature, the mixture was extracted with EtOAc three times. The organic layer was dried over Na₂SO₄, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **30** (679.3 mg, 84%, in 2 steps) as a colorless oil.

30: [α]_D²⁷ +67.9 (c 1.28, CHCl₃); IR (neat) 1778, 1700, 1471, 1383, 1259, 1212, 1104, 1019, 960, 849, 779 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ -0.02 (s, 3H), 0.00 (s, 3H), 0.86 (d, *J* = 6.6 Hz, 3H), 0.88 (s, 9H), 0.91 (d, *J* = 6.6 Hz, 3H), 0.94 (d, *J* = 6.6 Hz, 3H), 0.95 (s, 3H), 1.10–1.17 (m, 2H), 1.23–1.27 (m, 1H), 1.32–1.43 (m, 4H), 1.53–1.61 (m, 1H), 1.64–1.68 (m, 1H), 1.76–1.84 (m, 2H), 1.94 (dt, *J* = 3.0, 12.6 Hz, 1H), 2.03–2.10 (m, 1H), 2.32–2.40 (m, 1H), 2.58 (dd, *J* = 9.6, 16.2 Hz, 1H), 2.58 (dd, *J* = 3.6, 15.6 Hz, 1H), 3.99–4.00 (m, 1H), 4.19 (dd, *J* = 3.3, 9.3 Hz, 1H), 4.25 (t, *J* = 8.7 Hz, 1H), 4.45 (dt, *J* = 3.6, 8.4 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ -5.2, 4.8, 13.8, 14.5, 17.6, 18.0, 19.3, 23.0, 25.8, 27.0, 28.3, 32.6, 23.4, 40.6, 42.0, 42.3, 53.0, 56.9, 58.3, 63.1, 69.4, 154.0, 173.2; HRMS (ESI⁺) calcd for C₂₆H₄₇NO₄SiNa [M+Na]⁺ 488.3167, found 488.3183.

(4S)-3-[(2S,3S)-3-[(1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl]-2-fluorobutanoyl]-4-isopropylloxazolidin-2-one (29) and (4S)-3-[(2R,3S)-3-[(1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl]-2-fluorobutanoyl]-4-isopropylloxazolidin-2-one (33)

To a solution of **30** (207.6 mg, 0.45 mmol) in THF (3 mL) was added LHMDs (lithium hexamethyldisilazide) (472 μ L, 1 M THF solution, 0.472 mmol) at $-78\text{ }^{\circ}\text{C}$, the mixture was stirred at 0 $^{\circ}\text{C}$ for 1 h, and *N*-fluorobenzenesulfonimide (NFSI) (168.6 mg, 0.53 mmol) was added to the mixture at $-78\text{ }^{\circ}\text{C}$. After being stirred at the same temperature for 2 h, the reaction was quenched with H₂O and saturated aqueous NH₄Cl at $-78\text{ }^{\circ}\text{C}$. The mixture was extracted with EtOAc three times. The organic layer was washed with brine, dried over Na₂SO₄, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 2:1) to obtain **29** (108.4 mg, 50%) (less polar) and **33** (74.9 mg, 35%) (more polar), each as a colorless oil.

29: [α]_D²⁷ +83.1 (c 1.08, CHCl₃); IR (neat) 1786, 1718, 1384, 1256, 1201, 1025, 834 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ -0.02 (s, 3H), 0.00 (s, 3H), 0.88 (s, 9H), 0.89 (d, *J* = 7.2 Hz, 3H), 0.92 (s, 3H), 0.94 (d, *J* = 6.6 Hz, 3H), 1.14–1.39 (m, 9H), 1.52–1.67 (m, 3H), 1.74–1.94 (m, 3H), 2.08–2.18 (m, 1H), 2.51–2.56 (m, 1H), 3.98–4.00 (m, 1H), 4.29 (dd, *J* = 1.8, 9.0 Hz, 1H), 4.35 (t, *J* = 8.7 Hz, 1H), 4.38–4.40 (m, 1H), 5.74 (dd, *J* = 2.7, 47.4 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ -5.2, -4.8, 13.3, 14.4, 17.2 (d, *J* = 2.9 Hz), 17.5, 18.0, 18.0, 23.5, 25.8, 26.9, 28.0, 34.3, 40.1, 40.2, 40.3, 42.9, 51.6, 52.3, 59.1, 64.0, 69.3, 93.8 (d, *J* = 180.9 Hz), 153.5, 170.1 (d, *J* = 23.1 Hz); HRMS (ESI⁺) calcd for C₂₆H₄₆NO₄FSiNa [M+Na]⁺ 506.3072, found 506.3075.

33: [α]_D²⁷ +1.2 (c 0.57, CHCl₃); IR (neat) 1790, 1718, 1391, 1260, 1204, 1097, 1025, 978, 842, 774 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ -0.01 (s, 3H), 0.01 (s, 3H), 0.87 (d, *J* = 7.2 Hz, 3H), 0.88 (s, 9H), 0.92–0.95 (m, 9H), 1.18 (td, *J* = 2.4, 12.6 Hz, 1H), 1.32–1.43 (m, 4H), 1.49–1.68 (m, 4H), 1.76–1.93 (m, 4H), 2.03–2.17 (m, 1H), 2.30–2.37 (m, 1H), 4.00–4.02 (m, 1H), 4.26 (dd, *J* = 3.6, 9.0 Hz, 1H), 4.37 (t, *J* = 9.0 Hz, 1H), 4.57 (dt, *J* = 3.9, 9.0 Hz, 1H), 5.96 (dd, *J* = 1.5, 49.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ -5.2, -4.9, 12.4 (d, *J* = 4.4 Hz), 13.7, 14.3, 17.6, 17.7, 17.9, 22.7, 25.0, 25.8, 28.0, 34.2, 37.1, 37.2, 40.6, 42.0, 52.7, 52.9, 57.7, 64.0, 69.3,

91.9 (d, $J = 182.4$ Hz), 153.4, 169.7 (d, $J = 23.0$ Hz); HRMS (ESI⁺) calcd for C₂₆H₄₆NO₄FSiNa [M+Na]⁺ 506.3072, found 506.3091.

(2S,3S)-3-((1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl)-2-fluoro-N-methoxy-N-methylbutanamide (34)

To a solution of **29** (105.9 mg, 0.22 mmol) and Me(MeO)NH·HCl (48.8 mg, 0.50 mmol) in THF (10 mL) was added LHMDS (972 μL, 1 M in THF, 0.972 mmol) at 0 °C, and the mixture was stirred at the same temperature for 25 min. After the reaction was quenched with water and aqueous saturated NH₄Cl, the mixture was extracted with EtOAc three times. The organic layer was dried over Na₂SO₄, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **34** (45.4 mg, 50%) as a colorless oil. The spectral data matched with those reported in the literature [8].

(2R,3S)-3-((1R,3aR,4S,7aR)-4-[(*tert*-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl)-2-fluoro-N-methoxy-N-methylbutanamide (35)

To a solution of **33** (39.4 mg, 0.081 mmol) and Me(MeO)NH·HCl (17.2 mg, 0.18 mmol) in THF (5 mL) was added LHMDS (324 μL, 1 M in THF, 0.324 mmol) at 0 °C, and the mixture was stirred at the same temperature for 4 min. After the reaction was quenched with water and aqueous saturated NH₄Cl, the mixture was extracted with EtOAc three times. The organic layer was dried over Na₂SO₄, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **35** (18.7 mg, 55%) as a colorless oil. The spectral data matched with those reported in the literature [8].

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/biom14010037/s1>. File S1: ¹H and ¹³C NMR spectra of all new compounds **26**, **27**, **29**, **30**, and **33**.

Author Contributions: Conceptualization, F.K. and A.K.; investigation, F.K. and S.M.; original draft preparation, F.K.; writing—review and editing, A.K.; supervision, A.K.; funding acquisition, F.K. and A.K. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported in part by Grants-in-Aid from the Japan Society for the Promotion of Science (No. 22K14688 to F.K. and No. 23K06029 to A.K.).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Kawagoe, F.; Mototani, S.; Kittaka, A. Design and synthesis of fluoro analogues of vitamin D. *Int. J. Mol. Sci.* **2021**, *22*, 8191. [[CrossRef](#)] [[PubMed](#)]
2. Kawagoe, F.; Sugiyama, T.; Uesugi, M.; Kittaka, A. Recent developments for introducing a hexafluoroisopropanol unit into the vitamin D side chain. *J. Steroid Biochem. Mol. Biol.* **2018**, *177*, 250–254. [[CrossRef](#)] [[PubMed](#)]
3. Kawagoe, F. Synthesis of new 26,26,27,27-tetrafluoro and 26,27-difluoro-25-hydroxyvitamin D₃: The effects of the terminal fluorine atoms on affinity to VDR and CYP24A1-dependent metabolism. *Chem. Pharm. Bull.* **2023**, *71*, 717–723. [[CrossRef](#)] [[PubMed](#)]
4. Kawagoe, F.; Mototani, S.; Yasuda, K.; Nagasawa, K.; Uesugi, M.; Sakaki, T.; Kittaka, A. Introduction of fluorine atoms to vitamin D₃ side-chain and synthesis of 24,24-difluoro-25-hydroxyvitamin D₃. *J. Steroid Biochem. Mol. Biol.* **2019**, *195*, 105477. [[CrossRef](#)] [[PubMed](#)]
5. Kawagoe, F.; Mototani, S.; Yasuda, K.; Mano, H.; Sakaki, T.; Kittaka, A. Stereoselective synthesis of 24-fluoro-25-hydroxyvitamin D₃ analogues and their stability to hCYP24A1-dependent catabolism. *Int. J. Mol. Sci.* **2021**, *22*, 11863. [[CrossRef](#)]
6. Mototani, S.; Kawagoe, F.; Yasuda, K.; Mano, H.; Sakaki, T.; Kittaka, A. The first convergent synthesis of 23,23-difluoro-25-hydroxyvitamin D₃ and its 24-hydroxy derivatives: Preliminary assessment of biological activities. *Molecules* **2022**, *27*, 5352. [[CrossRef](#)]

7. Kawagoe, F.; Yasuda, K.; Mototani, S.; Sugiyama, T.; Uesugi, M.; Sakaki, T.; Kittaka, A. Synthesis and CYP24A1-dependent metabolism of 23-fluorinated vitamin D₃ analogues. *ACS Omega* **2019**, *4*, 11332–11337. [[CrossRef](#)]
8. Kawagoe, F.; Mototani, S.; Yasuda, K.; Takeuchi, A.; Mano, H.; Kakuda, S.; Saitoh, H.; Sakaki, T.; Kittaka, A. Synthesis of (22R)-, (22S)-22-fluoro-, and 22,22-difluoro-25-hydroxyvitamin D₃ and effects of side-chain fluorination on biological activity and CYP24A1-dependent metabolism. *J. Org. Chem.* **2023**, *88*, 12394–12408. [[CrossRef](#)]
9. Nagata, A.; Akagi, Y.; Asano, L.; Kotake, K.; Kawagoe, F.; Mendoza, A.; Masoud, S.S.; Usuda, K.; Yasui, K.; Takemoto, Y.; et al. Synthetic chemical probes that dissect vitamin D activities. *ACS Chem. Biol.* **2019**, *14*, 2851–2858. [[CrossRef](#)]
10. Kawagoe, F.; Mendoza, A.; Hayata, Y.; Asano, L.; Kotake, K.; Mototani, S.; Kawamura, S.; Kurosaki, S.; Akagi, Y.; Takemoto, Y.; et al. Discovery of a vitamin D receptor-silent vitamin D derivative that impairs sterol regulatory element-binding protein in vivo. *J. Med. Chem.* **2021**, *64*, 5689–5709. [[CrossRef](#)]
11. Kawagoe, F.; Mototani, S.; Mendoza, A.; Takemoto, Y.; Uesugi, M.; Kittaka, A. Structure-activity relationship studies on vitamin D-based selective SREBP/SCAP inhibitor KK-052. *RSC Med. Chem.* **2023**, *14*, 2030–2034. [[CrossRef](#)]
12. Sakaki, T.; Sawada, N.; Komai, K.; Shiozawa, S.; Yamada, S.; Yamamoto, K.; Ohyama, Y.; Inouye, K. Dual metabolic pathway of 25-hydroxyvitamin D₃ catalyzed by human CYP24. *Eur. J. Biochem.* **2000**, *267*, 6158–6165. [[CrossRef](#)] [[PubMed](#)]
13. Sakaki, T.; Kagawa, N.; Yamamoto, K.; Inouye, K. Metabolism of vitamin D₃ by cytochromes P450. *Front. Biosci.* **2005**, *10*, 119–134.
14. Yasuda, K.; Nishikawa, M.; Okamoto, K.; Horibe, K.; Mano, H.; Yamaguchi, M.; Okon, R.; Nakagawa, K.; Tsugawa, N.; Okano, T.; et al. Elucidation of metabolic pathways of 25-hydroxyvitamin D₃ mediated by Cyp24A1 and Cyp3A using *Cyp24a1* knockout rats generated by CRISPR/Cas9 system. *J. Biol. Chem.* **2021**, *296*, 100668. [[CrossRef](#)] [[PubMed](#)]
15. Kobayashi, Y.; Taguchi, T.; Terada, T.; Oshida, J.-I.; Morisaki, M.; Ikekawa, N. Synthesis of 24,24-difluoro- and 24 ξ -fluoro-25-hydroxyvitamin D₃. *Tetrahedron Lett.* **1979**, *20*, 2023–2026. [[CrossRef](#)]
16. Shiuey, S.-J.; Partridge, J.J.; Chadha, N.K.; Boris, A.; Uskoković, M.R. Stereospecific synthesis of 1 α ,25-dihydroxy-24R-fluorocholecalciferol (Ro23-0233). In *Vitamin D, Chemical, Biochemical and Clinical Update*; Walter de Gruyter: Berlin, Germany, 1985; pp. 765–766.
17. Shiuey, S.-J.; Partridge, J.J.; Uskoković, M.R. Triply convergent synthesis of 1 α ,25-dihydroxy-24(R)-fluorocholecalciferol. *J. Org. Chem.* **1988**, *53*, 1040–1046. [[CrossRef](#)]
18. Yamada, S.; Ohmori, M.; Takayama, H. Synthesis of 24,24-difluoro-25-hydroxyvitamin D₃. *Tetrahedron Lett.* **1979**, *20*, 1859–1862. [[CrossRef](#)]
19. Gill, H.S.; Londowski, J.M.; Corradino, R.A.; Zinsmeister, A.R.; Kumar, R. Synthesis and biological activity of novel vitamin D analogues: 24,24-difluoro-25-hydroxy-26,27-dimethylvitamin D₃ and 24,24-difluoro-1 α ,25-dihydroxy-26,27-dimethylvitamin D₃. *J. Med. Chem.* **1990**, *33*, 480–490. [[CrossRef](#)]
20. Konno, K.; Ojima, K.; Hayashi, T.; Takayama, H. An alternative and efficient synthesis of 24,24-difluoro-1 α ,25-dihydroxyvitamin D₃. *Chem. Pharm. Bull.* **1992**, *40*, 1120–1124. [[CrossRef](#)]
21. Ando, K.; Kondo, F.; Koike, F.; Takayama, H. An improved synthesis of 24,24-difluoro-1 α ,25-dihydroxyvitamin D₃ from vitamin D₂. *Chem. Pharm. Bull.* **1992**, *40*, 1662–1664. [[CrossRef](#)]
22. Kondo, F.; Maki, S.; Konno, K.; Takayama, H. The first synthesis of 24,24-difluoro-1 α -hydroxyvitamin D₃ by means of radical deoxygenation of alcohols. *Chem. Pharm. Bull.* **1996**, *44*, 62–66. [[CrossRef](#)]
23. Iwasaki, H.; Hosotani, R.; Miyamoto, Y.; Nakano, Y.; Yamamoto, K.; Yamada, S.; Shinki, T.; Suda, T.; Yamaguchi, K.; Konno, K.; et al. Stereoselective synthesis and structural establishment of (25S)-24,24-difluoro-1 α ,25,26-trihydroxyvitamin D₃, a major metabolite of 24,24-difluoro-1 α ,25-dihydroxyvitamin D₃. *Tetrahedron* **1998**, *54*, 14705–14724. [[CrossRef](#)]
24. Flores, A.; Massarelli, I.; Thoden, J.B.; Plum, L.A.; DeLuca, H.F. A methylene group on C-2 of 24,24-difluoro-19-nor-1 α ,25-dihydroxyvitamin D₃ markedly increases bone calcium mobilization in vivo. *J. Med. Chem.* **2015**, *58*, 9731–9741. [[CrossRef](#)] [[PubMed](#)]
25. Tanaka, Y.; DeLuca, H.F.; Schnoes, H.K.; Ikekawa, N.; Kobayashi, Y. 24,24-difluoro-1,25-dihydroxyvitamin D₃: In vitro production, isolation, and biological activity. *Arch. Biochem. Biophys.* **1980**, *199*, 473–478. [[CrossRef](#)] [[PubMed](#)]
26. Gill, H.S.; Londowski, J.M.; Corradino, R.A.; Kumar, R. The synthesis and biological activity of 22-fluorovitamin D₃: A new vitamin D analog. *Steroids* **1986**, *48*, 93–108.
27. Davis, F.A.; Kasu, P.V.N. Asymmetric synthesis of α -fluoro ketones using α -fluoro oxazolidinone carboximides. *Tetrahedron Lett.* **1998**, *39*, 6135–6138. [[CrossRef](#)]
28. Less, S.L.; Handa, S.; Millburn, K.; Leadlay, P.F.; Dutton, C.J.; Staunton, J. Biosynthesis of Tetronasin: Part 6. Preparation of structural analogues of the diketide and triketide biosynthetic precursors to Tetronasin. *Tetrahedron Lett.* **1996**, *37*, 3515–3518. [[CrossRef](#)]
29. Brunet, V.A.; O'Hagan, D.; Slawin, A.M.Z. Titanium mediated asymmetric aldol reaction with α -fluoropropionimide enolates. *J. Fluor. Chem.* **2007**, *128*, 1271–1279. [[CrossRef](#)]
30. Mascareñas, J.L.; Pérez-Sestelo, J.; Castedo, L.; Mouriño, A. A short, flexible route to vitamin D metabolites and their side chain analogues. *Tetrahedron Lett.* **1991**, *32*, 2813–2816. [[CrossRef](#)]
31. Nicoletti, D.; Mouriño, A.; Torneiro, M.S. Synthesis of 25-hydroxyvitamin D₃ and 26,26,26,27,27,27-hexadeutero-25-hydroxyvitamin D₃ on solid support. *J. Org. Chem.* **2009**, *74*, 4782–4786. [[CrossRef](#)]
32. Yu, O.B.; Mutchie, T.R.; Di Milo, E.S.; Arnold, L.A. Synthesis and biological evaluation of calcioic acid. *Steroids* **2020**, *154*, 108536. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.