



Potential Immunohistochemical Biomarkers for Grading Oral Dysplasia: A Literature Review

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Abstract: Oral cancer is a prevalent global health issue, with significant morbidity and mortality rates. Despite available preventive measures, it remains one of the most common cancers, emphasising the need for improved diagnostic and prognostic tools. This review focuses on oral potentially malignant disorders (OPMDs), precursors to oral cancer, specifically emphasising oral epithelial dysplasia (OED). The World Health Organisation (WHO) provides a three-tier grading system for OED, and recent updates have expanded the criteria to enhance diagnostic precision. In the prognostic evaluation of OED, histological grading is presently regarded as the gold standard; however, its subjectivity and unreliability in anticipating malignant transformation or recurrence pose notable limitations. The primary objective is to investigate whether specific immunohistochemical biomarkers can enhance OED grading assessment according to the WHO classification. Biomarkers exhibit significant potential for comprehensive cancer risk evaluation, early detection, diagnosis, prognosis, and treatment optimisation. Technological advancements, including sequencing and nanotechnology, have expanded detection capabilities. Some analysed biomarkers are most frequently chosen, such as p53, Ki-67, cadherins/catenins, and other proteins used to differentiate OED grades. However, further research is needed to confirm these findings and discover new potential biomarkers for precise dysplasia grading and minimally invasive assessment of the risk of malignant transformation.

Keywords: oral dysplasia; oral epithelial dysplasia; immunohistochemistry; histological grading; immunoexpression

1. Introduction

Oral cancer is becoming more and more frequent worldwide [1]. Despite the widely available prevention, it is one of the most common cancers in the world, with 476,125 new cases and 225,900 deaths in 2020 [2]. Among the causes of carcinogenesis in the oral cavity, tobacco smoking or chewing, alcohol consumption, occupational exposure, risky sexual behaviour, genetic factors, and environmental pollution are widely mentioned [3]. Smoking is the most prominent risk factor for oral cancer due to the carcinogenic chemicals in cigarette smoke, including nitrosamines, benzopyrenes, and aromatic amines [4]. The risk of oral cancer is three times higher in smokers compared to non-smokers. In addition, the combination of cigarette smoking and frequent heavy alcohol consumption increases the risk of developing cancer by several times [5,6].



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Neoplastic lesions are often preceded by oral potentially malignant disorders (OP-MDs) [7]. The World Health Organisation's (WHO) classification of head and neck cancers defines OPMDs as "clinical symptoms carrying the risk of developing oral cancer, whether clinically definable precursor lesions or clinically normal mucosa" [8]. This group includes lesions such as leukoplakia, oral lichen planus, and oral lichenoid lesions [9]. Until recently, oral epithelial dysplasia (OED), proliferative verrucous leukoplakia, submucous fibrosis, and HPV-associated dysplasia were classified as OPMDs [10]. The histological presence of OED is currently the strongest predictor of malignant transformation in OPMDs [11]. According to the WHO classification, OED is characterised as "a spectrum of architectural and cytological epithelial changes resulting from the accumulation of genetic alterations, usually arising in a range of OPMD and indicating a risk of malignant transformation to OSCC" [9]. These structural changes reflect the loss of normal maturation and stratified epithelium [12].

Therefore, a biopsy is conventionally performed to assess precancerous changes (dysplasia) in the tissue and obtain a histopathological diagnosis of a potentially malignant disease. The terminology of dysplasia was re-adopted by the WHO in the Classification of Tumours of the Oral Cavity and Oropharynx in 2005. However, instead of using the term dysplasia, some authors suggest employing the term squamous intraepithelial neoplasia (SIN) or variations such as oral intraepithelial neoplasia (OIN) [13], which are modifications of cervical pre-malignant lesions [14]. This change in terminology to OIN aims to avoid confusion with the WHO's term of CIS (carcinoma in situ) and to emphasise the characteristics of OSCC that differ from those of SCC of the uterine cervix [15]. The WHO refrained from endorsing this suggestion. The decision against adopting the SIN terminology stemmed from its perceived inadequacy in clarifying the situation in a manner significant enough to replace the globally utilised concept of dysplasia [16]. Furthermore, it was not demonstrated at that time that many OPMDs lead to cancer [17,18].

To assess the extent of dysplasia, a set of grading criteria was implemented to categorise the progression of the lesion. According to the WHO three-tier OED classification, dysplasia is classified as mild, moderate, or severe, considering both architectural features (tissue changes) and cytological alterations (changes in individual cells/cytological pattern) [14]. In the most recent WHO classification as of 2022, the OED grading criteria were expanded to encompass additional architectural and cytologic features, as detailed in Table 1 [19]. This expansion aims to enhance the diagnostic precision of dysplasia, emphasising that architectural features alone may indicate the presence of dysplasia. Despite the inherent challenges in dysplasia grading, the WHO maintains a three-tiered grading system [10,20].

Architectural Features	Cytological Features
Irregular epithelial stratification	Abnormal variation in nuclear size
Loss of polarity of basal cells	Abnormal variation in nuclear shape
Drop-shaped rete ridges	Abnormal variation in cell size
Mitoses high in epithelium	Abnormal variation in cell shape
Generalised premature keratinisation	Increased N/C ratio
Keratin pearls within rete ridges	Atypical mitotic figures
Loss of epithelial cell cohesion	Increased number and size of nucleoli
Altered keratin pattern for oral sub-site	Hyperchromasia
Verrucous or papillary architecture	Increased number of mitotic figures
Extension of changes along minor gland ducts	Single-cell keratinisation
Sharply defined margin of changes	Apoptotic mitoses
Multiple different patterns of dysplasia	Increased nuclear size
Multifocal or skip lesions	
Expanded proliferative compartment	
Basal cell clustering/nesting	

Table 1. The WHO diagnostic criteria for oral epithelial dysplasia—update 2022—according to Muller and Tilakaratne [10].

Moreover, the binary classification system would be an alternative approach to the WHO classification. This system categorises OED into low- and high-risk dysplasia, utilising a quantitative threshold of dysplastic pathological features and aiming to enhance reliability [19]. Furthermore, the binary system offers promising results in predicting malignant transformations, overcoming "opt-out" judgments associated with the fourscale or five-scale grading system [21,22]. While this may facilitate disease categorisation and reduce observer variability, the clinical prognostic value remains largely untested and widespread acceptance of this system necessitates additional international validation before it can be fully endorsed [9,21–23].

In OED prognostication, histological grading is found as the current gold standard but is subjective and unreliable to predict malignant transformation or recurrence [24]. Therefore, we aimed to answer whether alterations in the expression of specific immunohistochemical biomarkers could help to facilitate OED grading assessment according to the WHO classification. For this purpose, we prepared a literature review covering original articles published between 2017 and 2022 and indexed in databases, such as PubMed, Scopus, and the Web of Science.

2. Discussion of Potential Immunohistochemical Biomarkers in Grading of Oral Epithelial Dysplasia

Researchers are still conducting studies that will establish a biomarker that can unambiguously diagnose and differentiate between the stages of dysplasia. Biomarkers can be genes, proteins, or other substances whose levels or presence are tested to detect cell changes [25]. Not all cells affected by carcinogenesis are the same, as they may present gene changes or differences in the levels of given metabolites and proteins [26]. Detection technologies have developed significantly in recent decades, including sequencing, nanotechnology, or methods determining circulating tumour DNA/RNA or exosomes [27]. The clinical applications of biomarkers are broad. They can be used as tools for cancer risk assessment, screening and early cancer detection, accurate diagnosis, prognosing patients' condition, and predicting responses to treatment [28]. Also, they help in the optimisation of the treatment process. This is essential for targeted therapy, as it is only effective in patients with specific cancer genetic mutations, and biomarkers are used to identify these subgroups [29]. Further research is required to overcome the scientific challenges of developing new biomarkers with greater sensitivity, specificity, and positive predictive value.

Interestingly, many biomarkers are emerging in studies regarding the differentiation of dysplasia grades. De Vicente et al. [30] observed an association between NANOG (a key regulator of pluripotency and self-renewal in embryonic and adult stem cells) and the grade of dysplasia. It was noted that expression of NANOG increased with the grade of dysplasia. The importance of NANOG was also confirmed in the study by Grubelnik et al. [31], which stated that this marker can be used to differentiate dysplasia grades. NANOG protein detection has a diagnostic potential for oral high-grade dysplasia, distinguishing it from low-grade dysplasia and non-neoplastic reactive lesions.

The study by Wang et al. [32] showed that significantly increased Orai1 and STIM1 protein levels were noted in OPMD with mild, moderate, and severe OED in comparison with normal oral mucosa. Orai1 is calcium release-activated calcium modulator 1. This protein is a membrane calcium channel subunit activated by the calcium sensor STIM1 when calcium stores are depleted. Disruption of normal intracellular Ca²⁺ is reported to be associated with the formation of cancer in some studies [33].

Given the different mechanisms of dysplasia development, the sophistication of the malignant transformation processes in the cells, and the individual changes in each person subjected to different environmental factors, it is very difficult to isolate a single comprehensive biomarker. For this literature review, the most commonly mentioned biomarkers were proteins, such as p53, Ki-67, cadherins/catenins, and others.

2.1. Biomarkers Related to Cell Division and Proliferation

The cell cycle is regulated by the activity of various cyclins and cyclin-dependent kinases (Cdks). Cyclins form a complex with Cdks, and complex formation results in the activation of the Cdk active site. Cyclins without Cdk activation have no enzymatic activity but have binding sites for some substrates [34]. Cyclins are some of the most important cell cycle regulatory proteins and are linked to a specific phase of the cycle [35]. Both cyclins and their associated proteins are currently the subject of intense research, as perturbations of their expression and regulation can lead to tumorigenesis [36]. The majority of findings have reported on the overexpression of cyclins D and E in the development of many types of cancer [37].

In many studies, p63 and CD31 are the primarily examined markers. The p63 protein in normal cells is found in the basal layer of squamous epithelium [38]. Bavle et al. [39] found that p63 expression rose with increased severity of dysplasia and increased expression in suprabasal cells. The studies showed that p63 is required to maintain cell proliferation. It was observed that as the severity of dysplasia rose, the proliferation rate increased; however, cell differentiation was jeopardised [40]. As the disease progressed, the number of blood vessels increased and angiogenesis occurred. This is one of the factors that plays an important role in tumour growth and metastasis, providing nutrition to the developing tumour [41]. CD31 protein is a marker of angiogenesis, so it was used to detect vascular changes near the epithelium. The correlation of p63 with CD31 added value to the categorisation of leukoplakic lesions in the cases of low and moderate dysplasia [39].

Patel et al. [42] assessed p63 expression in different grades of dysplasia and Cyclin D1 expression. Cyclin D1 is classified as a proto-oncogene. *P63* expression showed no statistically significant differences in different grades of dysplasia, and cyclin D1 showed only statistically significant differences between severe and mild grades of dysplasia. Gupta et al. [43] used VEGF and CD34 as dysplasia markers. The study evaluated the percentage of VEGF immunoreactivity, the intensity of VEGF staining, and CD34 immunostaining. The expression of VEGF and CD34 increased significantly during the transition from normal oral mucosa to severe OED.

CD44—cluster of differentiation 44—is a transmembrane glycoprotein [44]. Venkat Naga et al. [45] used a cluster of differentiation 44 (CD44) antibody to assess the correlation between this marker and oral dysplasia grading. The authors compared four groups: control tissue, mild epithelial dysplasia, moderate epithelial dysplasia, and severe epithelial dysplasia. A comparison of the groups showed statistically significant results. It suggested that CD44 may be a useful marker for diagnosing dysplastic lesions.

Interestingly, Aravind et al. [46] evaluated the osteopontin (OPN) expression in premalignant and malignant lesions. The authors observed a progressive increase in OPN expression, which was seen with increasing grades of dysplasia. Osteopontin seemed to be a promising biomarker in predicting the malignant potential of a premalignant lesion. Osteopontin, a phosphorylated sialoprotein, is a component of the mineralised extracellular matrices of bones and teeth [47] that has many functions in inflammation, immune responses, wound healing, cell adhesion, and cell migration through interactions with integrins and CD44 variants [48].

P53, also known as *TP53*, is a gene that encodes a protein that regulates the cell cycle and, therefore, acts as a tumour suppressor, regulating cell division by stopping cells from growing and proliferating too rapidly or in an uncontrolled manner [49]. As presented in Figure 1, p53 plays a critical role in the regulation of the DNA damage response. Under normal conditions, p53 is expressed at an extremely low level. The regulation of p53 activity is caused by the MDM2 protein, which contributes to the proteasomal degradation of this suppressor [50]. When DNA damage or energetic stress occurs in a cell, p53 expression is induced, causing the cell cycle to stop. This is a chance for the repairment processes, or the cells will develop apoptosis. The most important purpose of this protein is to eliminate cancer-prone cells from the replication pool [51]. When DNA damage, mitotic impairment, and oxidative stress are excessive, the p53 protein can be mutated to wild-type p53 protein



(wtp53), which is inactivated under physiological conditions [52]. Mutations in the *P53* gene and the functions of wtp53 expression have been linked to various human cancers [49].

Figure 1. The mechanism of p53 regulation in DNA damage response.

Researchers demonstrated p53's role in differentiating grades of dysplasia. Pandya et al. [53] showed that the difference in expression was statistically significant between mild and severe dysplasia. The difference in TP53 expression between mild and severe dysplasia was statistically significant, according to Patil et al. [54]. The expression also increased with the increasing grades of epithelial dysplasia. Deregulation of this oncosuppressive protein may be important for the liability of the lesions to carcinogenesis. In the study by Sawada et al. [55], the higher the grade of dysplasia, the more frequently a TP53 mutation was observed. Imaizumi et al. [56] assessed p53 expression by immunofluorescence as a biomarker to differentiate between oral squamous epithelial lesions. The study consisted of 129 archival oral biopsy samples, including 18 benign squamous lesions, 37 low-grade dysplasias, 22 high-grade dysplasias, and 52 OSCCs. The authors found that the expression of p53 can be a valuable biomarker that helps to estimate the grade of oral epithelial dysplasia.

 Δ Np63 is in the p53 family and is a p63 isoform, guiding the maturation of these stem cells through the regulation of their self-renewal and terminal differentiation. Yesassociated protein (YAP) is an oncoprotein in the cytoplasm in an inactive form [57]. YAP moves to the cell nucleus and activates the transcription of genes responsible for cell division and apoptosis [58]. Ono et al. [59] assessed the correlation between the expression of Δ Np63 and YAP and the grade of oral dysplasia. The authors found that in oral dysplasia, the expression of YAP and Δ Np63 was higher in high-grade than in low-grade disease. YAP and Δ Np63 expression correlated with grades of oral dysplasia.

The Ki-67 protein is widely used as a marker of human cancer cell proliferation [60]. Ki-67 plays a role in interphase and mitotic cells, and its distribution changes during the cell cycle. These localisations are associated with distinct functions [61]. Increased tumour cell proliferation is considered a significant natural factor in cancer detection. Ki-67 plays a significant role in cancer formation due to its positive association with tumour proliferation and invasion [62]. Ki-67 is the most suitable biological marker of mitotic activity due to its expression in the nucleus in a specific cell cycle period [63].

Mutations of P53 and high levels of Ki-67 protein are frequently observed in various types of human cancer. Ki-67 shows a stronger association with poor tumour differentiation and negatively affects patients' survival in advanced stages [64]. Both P53 mutational status/type and high Ki-67 can also significantly impact overall survival [65]. The expression of p53 and Ki-67 increases as normal oral mucosa becomes dysplastic and undergoes malignant transformation [66]. Co-expression of p53 and Ki-67 is related to larger tumours and

metastasis to lymph nodes; thus, this observation suggests that it can be used to identify high-risk lesions [67].

In their study, Kamala et al. [68] observed an increase in Ki-67 expression with the severity of dysplasia. The Ki-67 antigen can be used as a marker for histological evaluations of OED. According to Dash et al. [69], as the severity of OED increases, the number of cells showing positive Ki-67 expression also increases. This is also confirmed by Mondal et al. [70], who found that the differences in Ki-67 expression were statistically significant between normal mucosa and mild dysplasia, as well as between mild, moderate, and severe dysplasia. Ki-67 not only detects the hyperactive cells in OED, but its expression of Ki-67 can also be comparable to the clinical course or prognostication of a disease.

According to the study by Takkem et al. [71], Ki-67 expression was restricted to the basal layers of normal oral epithelium, while Ki-67-positive cells in OED were localised in the basal, suprabasal, and squamous layers; Ki-67 expression was increased in patients at high case risk. Ki-67-positive cells in well-differentiated OSCC were mainly located at the periphery of tumour nests; in moderately differentiated OSCC, they were located both at the periphery and in part of the centre of tumour nests, while they were scattered in the most poorly differentiated lesions. The study by Kamala et al. [68] aimed to determine the degree and pattern of expression of aberrant Ki67 in OSMF. The study confirmed a statistically significant correlation between the expression of Ki-67 with the clinical and histological grading of OSMF and the histological grading of OSCC.

Moreover, Swain et al. [72] examined Ki-67 with MCM2 expression in OED, OSCC, and normal mucosa. The study confirmed that the expression of these proteins increased progressively. The expression profile of MCM 2 and Ki-67 was increased with the increasing grades of epithelial dysplasia. In their studies, Gadbail et al. [73,74] used Ki-67, CD105, and α -SMA antigen to differentiate the OED grades. The expressions of Ki-67, CD105, and α -SMA markers complement the binary grading system of OED. Ki-67 showed significant increases from normal oral mucosa to low-grade and high-grade epithelial dysplasia.

Additionally, Suwasini et al. [75] found a statistically significant association between p53 and Ki-67. The results highlighted the potential use of the p53 protein and the Ki-67 antigen as significant molecular markers for early PMD detection and OSCC risk. This observation was also confirmed by Leung et al. [76]—Ki-67 and p53 were significantly increased with higher histological grades of OD. These observations showed the role of DNA-replicative stress in higher grades of dysplasia and transformation from OD to OSCC.

Monteiro et al. [77] analysed the immunoexpression of BubR1, Mad2, Bub3, Spindly, and Ki-67 proteins in 64 oral biopsies. Spindly is a protein that targets dynein/dynactin to kinetochores in mitosis. The authors observed that the expression of Spindly was significantly correlated with a high Ki-67 score and the grade of dysplasia. This observation confirmed that the expression of Ki-67 protein is associated with an increased risk for malignant transformation.

Stathmin is a member of a family of proteins that plays important roles in regulating the microtubule cytoskeleton [78]. This protein regulates microtubule dynamics by promoting the depolymerisation of microtubules and/or preventing the polymerisation of tubulin heterodimers [79]. Vadla et al. [80] evaluated the role of stathmin in OSCC and oral dysplasia and the correlation of stathmin expression with dysplasia grading. The study presented a statistically significant correlation between increased grades of oral dysplasia and expression levels of stathmin. This study confirmed the positive role of stathmin in disease progression and suggested that stathmin could be an early diagnostic biomarker for oral dysplasia.

2.2. Biomarkers Related to Epithelial–Mesenchymal Transition (EMT)

Epithelial stem cells maintain tissues throughout adult life and are controlled by epithelial–mesenchymal interactions to balance cell production and loss. A defining characteristic of an epithelium is the close contact that these cells have with the underlying mesenchyme [81]. Polarised epithelial cells normally interact with the basement membrane,

causing several biochemical changes that enable them to adopt a mesenchymal cell phenotype, including enhanced migratory capacity, invasiveness, elevated resistance to apoptosis, and greatly increased production of ECM components. This biological process is called an epithelial–mesenchymal transition (EMT) [82]. This transformation can occur in physiological processes during embryogenesis, organ development, and tissue regeneration, as well as in tumorigenesis and cancer progression, including tumour cell invasion and metastasis [83].

Mesenchymal stem cells are stromal cells capable of self-renewal and multilineage differentiation. They show a greater ability to infiltrate the capillaries at the site of the primary tumour lesions [84,85]. This mechanism is a critical mechanism for the acquisition of the malignant phenotype in neoplastic epithelial processes. This subtype accompanies the formation of distant metastases, where, in secondary foci, cells change their phenotype through a reverse mesenchymal–epithelial transition (MET) [86,87].

The role of EMT in OSCC is to transform normal epithelial cells into malignant mesenchymal cells by losing intercellular adhesion, causing metastatic progression and infiltration [88]. In the epithelial stage, tumour cells are cubic and adherent to each other. Also, in this stage, tumour cells show positive E-cadherin expression and negative vimentin expression. In the mesenchymal stage, the tumour cells show higher vimentin expression, but the expression of E-cadherin is repressed. The tumour cells are fibroblast-like and lose their cell–cell junctions [89].

The hallmark of EMT is the upregulation of N-cadherin followed by the downregulation of E-cadherin, and this process is regulated by a complex network of signalling pathways and transcription factors. The breakdown of cell–cell connections is caused by a change in cadherin expression (E-cadherin replaced by N-cadherin). Then, cells lose their apical–basal polarity, which is converted into a front–rear polarity. The downregulation of E-cadherin is often found in malignant epithelial cancers. N-cadherin indicates ongoing EMT and its expression has been correlated with the development of various types of carcinoma [90].

Also, MMPs can induce EMT, contributing directly to cell migration and invasion by degrading specific substrates and implicating many steps of carcinogenesis, including primary tumour growth, angiogenesis, basal membrane and stroma invasion, and metastatic progression [91]. Structural and functional support to the cell is provided by vimentin-filamentous protein. In the early stages of cancer, vimentin is at a very low level. Its concentration increases when the tumour invades the surrounding areas [92].

Remodelling the cytoskeleton results in altered cell morphology and increased motility. EMT is dictated by a series of changes in the expression levels of proteins regulated by the activity of proteins responsible for intercellular interactions (Figure 2). Markers of EMT are proteins specific to the epithelial phenotype, e.g., E-cadherin, mucin-1, cytokeratins, occludin, or desmoplakin, whose activity is reduced. As a result of EMT, the levels of N-cadherin, vimentin, fibronectin, or vitronectin are increased [93]. The expression of the EMT-associated protein markers can be used by pathologists as specific indicators of risk of malignancy processes. Moreover, the ability to adapt to different environmental conditions or in the presence of chemotherapeutics is the main characteristic of malignant tumours and is closely linked to EMT. This relationship can be helpful for oncological therapeutic strategies [94]. Understanding EMT and MET may help to identify specific markers to distinguish normal stem cells from cancer stem cells in the future [86].

One of these biomarkers are cytokeratins (CKs). CKs are keratin proteins located in the intracytoplasmic cytoskeleton of epithelial tissue. They are an important component of the intermediate filaments that help cells resist mechanical loads [95]. Batool et al. [96] found a strong correlation between the intensity of CK5\6 staining and the different stages of dysplasia. Additionally, this marker allows for the differentiation of healthy mucosa from dysplastic mucosa. A gradual increase in staining intensity for CK5\6 was observed with increasing grades of dysplasia. They found a highly significant association with CK5\6 immunopositivity and transforming normal mucosa into various grades of oral dysplastic

lesions. Also, CK19 belongs to a family of keratins. CK19 is an odontogenic epithelial marker reported to exhibit increased expression in various cancers, including OSCC [97]. Rajeswari et al. [98] noticed an increased expression of CK19 in severe dysplasia, but in mild and moderate dysplasia, CK19 expression was lower than the normal mucosa. This study showed that CK19 cannot be a marker to assess the grading of dysplasia.



Figure 2. The process of epithelial-mesenchymal transition (EMT).

 β -Catenin regulates cell adhesion and migration as an intercellular junction-forming element in complex with E-cadherin [99]. Intercellular junctions determine their polarity and enable tissue integrity, growth, and maturation [100]. They enable interaction and signal transmission between neighbouring cells and between neighbouring cells and the extracellular matrix. Weakening intracellular junctions can lead to the disruption of cell cycle control, resulting in the separation of individual cells from the primary tumour, thus creating the conditions for tumour metastasis [101]. E-cadherin is produced on the surface of the epithelial cells of many organs. It is responsible for the integrity of the mucosal tissue, the first line of defence against environmental toxic molecules [102].

The study by Chowdhury et al. [103] confirmed the role of β -catenin in differentiating the respective grades of dysplasia. The concentration of β -catenin increased in the individual grades of dysplasia. In the study by Prgomet et al. [104], there were statistically significantly higher expressions of β -catenin in dysplasia compared with normal-appearing oral mucosa. Still, the authors did not compare the results in different grades of oral dysplasia. Decreased E-cadherin and increased VEGF expression could be involved in the tissue growth and transformation of OPMDs, correlating with their different histological grades in numerous studies. This was confirmed in the study by Sharada et al. [105], as these association markers can be used to predict the potential risk of malignant transformation in OED. In their research, Sharma et al. [106] also evaluated the importance of E-cadherin in differentiating the dysplasia grade. E-cadherin expression decreased significantly with increasing dysplasia grade.

Similarly, Puneeta et al. [107] assessed the expression of vimentin and E-cadherin in different grades of OED and OSCC. In the OED group, a progressive involvement of all layers was observed, with 5% of mild OED, 10% of moderate OED and 70% of severe OED showing expression of E-cadherin up to the superficial layers, which was statistically significant. Vimentin expression was low in mild OED, while high expression

was more prevalent in moderate and severe OED. This finding was statistically significant. Furthermore, the study by Miguel et al. [108] aimed to investigate the immunoexpression of matrix metalloproteinase 9, tissue inhibitor of metalloproteinase 1, and vimentin. The authors confirmed the role of the epithelial expression of vimentin in the malignant process. Also, they found that smokers had a higher epithelial expression of MMP-9 and vimentin.

As mentioned earlier, N-cadherin is upregulated while E-cadherin is downregulated during EMT in carcinogenesis. This process is associated with enhanced migratory and invasive traits, which causes an inferior patient survival rate [90]. Chandolia et al. [109] assessed N-cadherin expression in 100 cases (epithelium with normal oral mucosa, OED lesions, and OSCC). The differences were statistically significant, and the study showed that N-cadherin expression was more evident than in OED, followed by the normal oral epithelium.

Importantly, the Wnt pathway stabilises the ß-catenin protein and interferes in the ß-catenin and E-cadherin complex. The Wnt pathway is involved in the dysplastic changes that downregulate E-cadherin by TWIST (Figure 2). TWIST binds to E-cadherin and suppresses the transcription of E-cadherin [110]. Qahtani et al. [111] examined the expression of the TWIST protein. The authors found significant differences between severe dysplasia and other grades of oral dysplasia. The study confirmed that the cadherin–catenin complex and the proteins involved in their regulation play a role in carcinogenesis.

Podoplanin (PDPN) is a small cell-surface mucin-like glycoprotein [112]. Podoplanin expression is upregulated in different cell types, including fibroblasts, macrophages, T helper cells, and epithelial cells, during inflammation and cancer, where it plays important roles [113]. Podoplanin interacts with other proteins in the same or neighbouring cells. The binding of podoplanin to ligands leads to the modulation of signalling pathways, which regulate proliferation, contractility, migration, epithelial-mesenchymal transition, and the remodelling of the extracellular matrix [114]. Karunagaran et al. [115] showed a significant association between dysplasia and podoplanin expression, with increasing dysplasia grade corresponding with podoplanin expression. Podoplanin seemed to have an increased expression as the dysplasia grade increased, suggesting its role in the progression of the disease toward malignancy. Lunawat et al. [116] investigated podoplanin immunoexpression in lymphatic vessels of OED. Podoplanin expression significantly increased with higher grades of dysplasia. This observation might help to diagnose the wider progression of dysplastic lesions to carcinoma. The study by Monteiro et al. [117] aimed to evaluate the expression of biomarkers CD44v6, CD147, EGFR, p53, p63, p73, p16, and podoplanin in oral leukoplakia. In a multivariate analysis, the authors observed a significant increase in high expression from normal tissue to low-grade dysplasia and high-grade dysplasia cases in CD44v6, p53, p73, and podoplanin. In conclusion, podoplanin expression could be a useful predictive marker in malignant transformation. Similarly, Abidullah et al. [118] found that the staining of MUC4 increased from mild to moderate to severe dysplasia. Mucin MUC4 is membrane-associated and plays a protective role [119]. Therefore, MUC4 can be a marker for the diagnosis of OED.

2.3. Biomarkers Related to Cell Death Regulation

Other altered proteins are the members of the Bcl-2 family. These proteins are considered as the principal players in the cascade of events that activate or inhibit apoptosis [120]. In this family, there are, for example, Bcl-XL, Bcl-2, and Bax. Bcl-2 acts as a checkpoint upstream of caspases and mitochondrial dysfunction [121]. Also, Bcl-2 can rescue maturation at several points of lymphocyte development. The Bcl-2 proto-oncogene was discovered at the chromosomal breakpoint of t (14;18) found in a human follicular lymphoma [122]. Pathak et al. [123] observed that the level of Bcl-2 increased with the grade of dysplasia. However, Bcl-2 expression was decreased in OSCC. Pallavi et al. [124] assessed the expression of Bcl-2 and c-Myc in OED and OSCC. Similarly, the authors noticed that Bcl-2 increased with grades of dysplasia. Bcl-2 proteins could positively affect lesion progression from premalignancy to malignancy.

Also, the PD-1/PD-L1 pathway can be a potential marker for oral dysplasia. Programmed Cell Death Protein 1 (PD-1) inhibits immune responses and modulates T-cell activity [125]. Kujan et al. [126] investigated the role of the PD-1/PD-L1 pathway in the development of dysplasia and OSCC. The study found that the PD-1/PD-L1 pathway can be associated with the development of OSCC and the grade of dysplasia. Programmed cell death 4 (PDCD4) functions as a tumour suppressor and an inhibitor of protein translation [127]. PDCD4 expression was observed in normal oral mucosa, OED, and OSCC. Desai and Kale [128] showed that the maximum expression was observed in normal oral mucosa, which reduced significantly in OED and OSCC.

Heat shock protein 27 (HSP27) belongs to the small-molecular-weight heat shock protein family and has a molecular weight of approximately 27 KDa [129]. This protein protects other proteins from damage due to environmental factors such as heat, toxins, free radicals, and ischaemia [130]. Karri et al. [131] found that a low expression of HSP27 could be an early molecular indicator of initial dysplastic changes in normal mucosa. Conversely, the overexpression of HSP27 could be a prognostic value of malignant transformation from oral dysplasia to oral squamous cell carcinoma. Cornulin (known as C1 Orf10, or squamous epithelial heat shock protein 53) is a member of the heat shock protein 70 (HSP70) family [132]. Cornulin plays an important role in the differentiation of the epidermis. The expression of cornulin causes cell cycle arrest at G1, and its downregulation plays a role in oral carcinogenesis [133]. Santosh et al. [134] found that cornulin expression decreased in oral dysplasia compared with normal oral mucosa and was absent in OSCC.

2.4. Biomarkers Related to Cellular Metabolism

A major component of the cellular response to oxygen deprivation is the transcription factor HIF-1 (hypoxia-inducible factor-1). HIF-1 consists of an HIF-1 beta unit and one of three units of HIF-1alpha, HIF-2alpha, or HIF-3alpha [135]. Patel et al. [136] assessed the expression of HIF-1alpha in OED and compared the expression between grades. The authors noticed that the expression of HIF-alpha statistically significantly increased as grades of oral dysplasia were higher. Also, HIF-alpha could be a marker of risk of malignant transformation.

Inducible nitric oxide synthase (iNOS) is an enzyme in oxygen and nitrogen metabolite metabolism [137]. Using immunohistochemical methods, Singh et al. [138] compared iNOS expression between oral leukoplakia and OSCC. The authors found that the expression of iNOS rose with the progressing clinical stages of oral leukoplakia and OSCC. Therefore, iNOS might be a diagnostic marker in oral leukoplakia and a prognostication marker of OSCC. Another enzyme, cyclooxygenase (COX or prostaglandin–endoperoxide synthase), is required to change arachidonic acid to prostaglandins [139]. Sharada et al. [140] examined the expression of COX-2 and type IV collagen in OED. The study found that its expression increased significantly as the grade of dysplasia was higher. This marker could be applied to assess the malignant potential.

2.5. Biomarkers Related to Extracellular Signalling Pathways

Paxillin is a 68 kDa, phosphotyrosine-containing protein that may play a role in several signalling pathways [141]. The study by Alam et al. [142] presented a statistically significant correlation between increased grades of oral dysplasia and expression of paxillin. Paxillin may play an important role in the pathogenesis of oral dysplasia and OSCC.

EGFR is a 170 kDa transmembrane glycoprotein receptor [143]. EGFR regulates cell growth, differentiation, and gene expression [144]. Fakurnejad et al. [145] demonstrated that an anti-EGFR agent could successfully discriminate high-grade dysplastic lesions from low-grade dysplasia. Melanoma inhibitory activity (MIA) and MIA2 are other receptors participating in tumour growth and invasion. Kawai et al. [146] evaluated MIA and MIA2 as expressed in the oral mucosa within early neoplastic lesions and suggested that MIA and MIA2 are useful novel immunohistochemical markers for discriminating between normal tissue and OED.

Laminins are another family of structural proteins. Laminins participate in organising the complex interactions of the basement membranes. Laminin-1 is in the Reichert membrane (extraembryonic basement membrane) [147]. A study by Vageli et al. [148] assessed laminin immunostaining in biopsies as a useful biomarker of actinic cheilitis and differential diagnosis between actinic cheilitis and lip cancer. This marker can differentiate between low- and high-grade dysplasia. This study can provide new insight into the mechanism of progression of actinic cheilitis into lip cancer. Also, Nguyen et al. [149] evaluated the immunoexpression of LAMC2. The expression of LAMC2 was significantly associated with the grade of dysplasia. LAMC2 may be a predictive marker for the malignant progression of leukoplakia.

In the study by Debta et al. [150], GLUT-1 also appeared as a marker for differentiating dysplasia severity. A statistically significant increasing level of GLUT-1 corresponded to more advanced grades of dysplasia and was consistent with the WHO system. GLUT-1 expression was significantly increased from normal to mild, moderate, and severe dysplasia. The expression of the GLUT-1 marker complemented the WHO grading system of OED. Also, Patlolla et al. [151] confirmed a significant correlation between the location of GLUT-1 within the cell and the grade of dysplasia.

Moreover, Udompatanakorn and Taebunpakul [152] assessed the pattern of expression of METTL3 in OED. METTL3 is an enzyme involved in the post-transcriptional methylation of internal adenosine residues [153]. The authors observed that the expression of METTL3 increased in oral dysplasia and OSCC. METTL3 expression might be a marker for the progression of oral dysplasia and transformation to OSCC.

Another marker is the minichromosome maintenance protein (MCM-2), which is a key component of the pre-replication complex. This protein may be involved in the formation of replication forks and in the migration of other proteins during DNA replication [154]. The study by Zakaria et al. [155] aimed to assess MCM-2 activity in oral epithelial dysplastic lesions. The MCM-2 immunostaining showed a statistically significant increase from mild to severe dysplasia, and the highest value was in invasive squamous cell carcinoma. MCM-2 activity is associated with the grade of dysplasia. This observation suggests that MCM-2 may be a potential biomarker for early squamous cell carcinoma.

2.6. Limitations and Challenges

The limitations of this review include the heterogeneity of the study designs in terms of clinical and histopathologic diagnoses, as well as laboratory methods determining markers of oral dysplasia. The included studies focused on a wide range of phenomena detected using immunochemical methods. However, changes in EMT markers (i.e., cadherin, vimentin, etc.) and p53 or Ki-67 were most frequently described. These molecules are known to affect the cell cycle, proliferation, and differentiation of cells, including cancer cells. Laboratory testing is important in assessing the levels of markers, but the results may be influenced by the quality of the collected specimens and the storage time.

It should be noted that the pathologists manually assigned the different degrees of dysplasia. The quality of the collected samples, the experience of the researchers, and the type of classification can impact the findings. The complexity of cell tumorigenesis and the number of pathways involved in this process (considering the relationships between different pathways) creates a problem in identifying a single universal marker for OED grading. Therefore, developing immunohistochemical marker panels with high sensitivity and specificity to detect early stages of oral dysplasia should be considered in the future.

To summarise this review, we include Table 2, presenting the main potential immunohistochemical markers for oral dysplasia. In the Supplementary Materials, we attach Table S1, reporting all potential immunohistochemical markers with methodological descriptions of the tested samples.

Relation of Biomarkers	Examples of Biomarkers for Grading of Oral Dysplasia
Cell division and proliferation	p53 [53–55,75,76,117], Ki-67 [56,68–76], CD105 [73,74], p63 [39,42,117], CD31 [39], CD34 [43], cycD1 [42], VEGF [43], YAP, Np63 [59], stathmin [80], CDKN1A [53]
Epithelial-mesenchymal transition	CK5\6 [96], CK19 [98], β-catenin [103,104], N-cadherin [109], E-cadherin [104–107], TWIST [111], VIM [107,108], PDPN [115,117], MMP-9 [108]
Cell death regulation	Bcl-2 [123,124], PDCD4 [128], HSP27 [131], cornulin [134]
Cellular metabolism	HIF-1a [136], iNOS [138], COX-2 [140]
	Paxillin [142], EGFR [145], MIA, MIA2 [146], laminin [148], LAMC2 [149],
Extracellular signalling pathways	GLUT-1 [150,151], METTL3 [152], MCM2 [155], 8-OHdG, Ref-1, XRCC-1 [156], Orai1,
	STIM1 [32], NANOG [30,31]

Table 2. Summary of main potential immunohistochemical biomarkers for assessing grading of oral dysplasia.

Legend: 8-OHdG, 8-hydroxy-2-deoxyguanosine; Bcl-2, B-cell lymphoma 2; CD, cluster differentiation; CDKN1A, cyclin-dependent kinase inhibitor 1A; CK, cytokeratin; COX-2, cyclooxygenase 2; CycD1, cyclin D1; EGFR, epidermal growth factor receptor; GLUT-1, glucose transporter-1; HIF, hypoxia-inducible factor; HSP, heat shock protein; iNOS, inducible nitric oxide synthase; LAMC2, laminin subunit gamma 2; MCM2, minichromosome maintenance complex component 2; METTL3, methyltransferase-like 3; MMP-9, matrix metalloproteinase 9; MIA, melanoma inhibitory activity; PDCD4, programmed cell death 4; PDPN, podoplanin; Ref-1, Redox factor-1; STIM1, stromal interaction molecule 1; VEGF, vascular endothelial growth factor; VIM, vimentin; XRCC-1, X-ray Repair Cross Complementing-1; YAP, Yes-associated protein.

3. Conclusions

According to our review, there are many various immunohistochemical biomarkers for dysplasia grading. The researchers most commonly used p53 protein, Ki-67 protein, cadherins/catenins, and other proteins as markers to differentiate grades of oral epithelial dysplasia. However, further research is desirable to confirm these outcomes and detect new potential biomarkers to properly establish the dysplasia grade and the risk of malignant transformation in a minimally invasive way.

Supplementary Materials: The following supporting information can be downloaded at https://www.mdpi.com/article/10.3390/biomedicines12030577/s1: Table S1. Detailed summary of potential immunohistochemical biomarkers for assessing grading of oral dysplasia with a brief description of the sample characteristics and the study setting.

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References

- 1. Gupta, N.; Gupta, R.; Acharya, A.K.; Patthi, B.; Goud, V.; Reddy, S.; Garg, A.; Singla, A. Changing Trends in Oral Cancer—A Global Scenario. *Nepal J. Epidemiol.* **2016**, *6*, 613–619. [CrossRef]
- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* 2021, 71, 209–249. [CrossRef]
- 3. Nijakowski, K.; Gruszczyński, D.; Kopała, D.; Surdacka, A. Salivary Metabolomics for Oral Squamous Cell Carcinoma Diagnosis: A Systematic Review. *Metabolites* **2022**, *12*, 294. [CrossRef]
- 4. Centers for Disease Control and Prevention (US); National Center for Chronic Disease Prevention and Health Promotion (US); Office on Smoking and Health (US). *How Tobacco Smoke Causes Disease: The Biology and Behavioral Basis for Smoking-Attributable Disease: A Report of the Surgeon General*; Centers for Disease Control and Prevention (US): Atlanta, GA, USA, 2010.
- 5. Yang, Y.; Zhou, M.; Zeng, X.; Wang, C. The Burden of Oral Cancer in China, 1990–2017: An Analysis for the Global Burden of Disease, Injuries, and Risk Factors Study 2017. *BMC Oral Health* **2021**, *21*, 44. [CrossRef]
- Edirisinghe, S.T.; Weerasekera, M.; De Silva, D.K.; Liyanage, I.; Niluka, M.; Madushika, K.; Deegodagamage, S.; Wijesundara, C.; Rich, A.M.; De Silva, H.; et al. The Risk of Oral Cancer among Different Categories of Exposure to Tobacco Smoking in Sri Lanka. *Asian Pac. J. Cancer Prev. APJCP* 2022, 23, 2929–2935. [CrossRef]

- 7. Bouaoud, J.; Bossi, P.; Elkabets, M.; Schmitz, S.; van Kempen, L.C.; Martinez, P.; Jagadeeshan, S.; Breuskin, I.; Puppels, G.J.; Hoffmann, C.; et al. Unmet Needs and Perspectives in Oral Cancer Prevention. *Cancers* **2022**, *14*, 1815. [CrossRef]
- 8. Gupta, S.; Jawanda, M.K.; Madhushankari, G. Current Challenges and the Diagnostic Pitfalls in the Grading of Epithelial Dysplasia in Oral Potentially Malignant Disorders: A Review. J. Oral Biol. Craniofacial Res. 2020, 10, 788–799. [CrossRef]
- Rich, A.M.; Hussaini, H.M.; Nizar, M.A.M.; Gavidi, R.O.; Tauati-Williams, E.; Yakin, M.; Seo, B. Diagnosis of Oral Potentially Malignant Disorders: Overview and Experience in Oceania. *Front. Oral Health* 2023, 4, 1122497. [CrossRef]
- 10. Muller, S.; Tilakaratne, W.M. Update from the 5th Edition of the World Health Organization Classification of Head and Neck Tumors: Tumours of the Oral Cavity and Mobile Tongue. *Head Neck Pathol.* **2022**, *16*, 54–62. [CrossRef]
- Bernard, C.; Zhang, J.Z.; Klieb, H.; Blanas, N.; Xu, W.; Magalhaes, M. Clinical Outcomes of Oral Epithelial Dysplasia Managed by Observation versus Excision. *Head Neck* 2023, 45, 3096–3106. [CrossRef]
- 12. Mukherjee, A.; Ferrao, T.; Spadigam, A.E.; Dhupar, A.; Mukherjee, A.; Ferrao, T.; Spadigam, A.E.; Dhupar, A. Oral Epithelial Dysplasia in Tobacco Non-Habitués: A Case Report and Review of Literature. *Cureus* **2023**, *15*, 47362. [CrossRef]
- Küffer, R.; Lombardi, T. Premalignant Lesions of the Oral Mucosa. A Discussion about the Place of Oral Intraepithelial Neoplasia (OIN). Oral Oncol. 2002, 38, 125–130. [CrossRef]
- 14. Ranganathan, K.; Kavitha, L. Oral Epithelial Dysplasia: Classifications and Clinical Relevance in Risk Assessment of Oral Potentially Malignant Disorders. *J. Oral Maxillofac. Pathol. JOMFP* **2019**, *23*, 19–27. [CrossRef]
- 15. Izumo, T. Oral Premalignant Lesions: From the Pathological Viewpoint. Int. J. Clin. Oncol. 2011, 16, 15–26. [CrossRef]
- Warnakulasuriya, S.; Reibel, J.; Bouquot, J.; Dabelsteen, E. Oral Epithelial Dysplasia Classification Systems: Predictive Value, Utility, Weaknesses and Scope for Improvement. J. Oral Pathol. Med. Off. Publ. Int. Assoc. Oral Pathol. Am. Acad. Oral Pathol. 2008, 37, 127–133. [CrossRef]
- 17. Warnakulasuriya, S.; Johnson, N.W.; van der Waal, I. Nomenclature and Classification of Potentially Malignant Disorders of the Oral Mucosa. J. Oral Pathol. Med. Off. Publ. Int. Assoc. Oral Pathol. Am. Acad. Oral Pathol. 2007, 36, 575–580. [CrossRef]
- 18. Napier, S.S.; Speight, P.M. Natural History of Potentially Malignant Oral Lesions and Conditions: An Overview of the Literature. J. Oral Pathol. Med. Off. Publ. Int. Assoc. Oral Pathol. Am. Acad. Oral Pathol. 2008, 37, 1–10. [CrossRef]
- Hankinson, P.; Mahmood, H.; Walsh, H.; Speight, P.M.; Khurram, S.A. Demystifying Oral Epithelial Dysplasia: A Histological Guide. *Pathology* 2023, 56, 11–23. [CrossRef]
- 20. WHO Classification of Tumours Editorial Board. *Head and Neck Tumours*, 5th ed.; WHO Classification of Tumours Series; International Agency for Research on Cancer: Lyon, France, 2022.
- Kujan, O.; Khattab, A.; Oliver, R.J.; Roberts, S.A.; Thakker, N.; Sloan, P. Why Oral Histopathology Suffers Inter-Observer Variability on Grading Oral Epithelial Dysplasia: An Attempt to Understand the Sources of Variation. *Oral Oncol.* 2007, 43, 224–231. [CrossRef]
- 22. Krishnan, L.; Karpagaselvi, K.; Kumarswamy, J.; Sudheendra, U.S.; Santosh, K.V.; Patil, A. Inter- and Intra-Observer Variability in Three Grading Systems for Oral Epithelial Dysplasia. J. Oral Maxillofac. Pathol. JOMFP 2016, 20, 261–268. [CrossRef]
- 23. Odell, E.; Kujan, O.; Warnakulasuriya, S.; Sloan, P. Oral Epithelial Dysplasia: Recognition, Grading and Clinical Significance. *Oral Dis.* **2021**, *27*, 1947–1976. [CrossRef]
- Mahmood, H.; Bradburn, M.; Rajpoot, N.; Islam, N.M.; Kujan, O.; Khurram, S.A. Prediction of Malignant Transformation and Recurrence of Oral Epithelial Dysplasia Using Architectural and Cytological Feature Specific Prognostic Models. *Mod. Pathol.* 2022, 35, 1151–1159. [CrossRef]
- 25. Strimbu, K.; Tavel, J.A. What Are Biomarkers? Curr. Opin. HIV AIDS 2010, 5, 463–466. [CrossRef]
- 26. Pavlova, N.N.; Thompson, C.B. The Emerging Hallmarks of Cancer Metabolism. Cell Metab. 2016, 23, 27–47. [CrossRef]
- 27. Hsu, C.-C.; Wu, Y. Recent Advances in Nanotechnology-Enabled Biosensors for Detection of Exosomes as New Cancer Liquid Biopsy. *Exp. Biol. Med.* **2022**, 247, 2152–2172. [CrossRef]
- Kourou, K.; Exarchos, T.P.; Exarchos, K.P.; Karamouzis, M.V.; Fotiadis, D.I. Machine Learning Applications in Cancer Prognosis and Prediction. Comput. Struct. Biotechnol. J. 2015, 13, 8–17. [CrossRef]
- Kamel, H.F.M.; Al-Amodi, H.S.A.B. Exploitation of Gene Expression and Cancer Biomarkers in Paving the Path to Era of Personalized Medicine. *Genom. Proteom. Bioinform.* 2017, 15, 220–235. [CrossRef]
- de Vicente, J.C.; Rodríguez-Santamarta, T.; Rodrigo, J.P.; Allonca, E.; Vallina, A.; Singhania, A.; Donate-Pérez Del Molino, P.; García-Pedrero, J.M. The Emerging Role of NANOG as an Early Cancer Risk Biomarker in Patients with Oral Potentially Malignant Disorders. *J. Clin. Med.* 2019, *8*, 1376. [CrossRef]
- Grubelnik, G.; Boštjančič, E.; Aničin, A.; Dovšak, T.; Zidar, N. MicroRNAs and Long Non-Coding RNAs as Regulators of NANOG Expression in the Development of Oral Squamous Cell Carcinoma. *Front. Oncol.* 2020, 10, 579053. [CrossRef]
- 32. Wang, Y.-Y.; Wang, W.-C.; Su, C.-W.; Hsu, C.-W.; Yuan, S.-S.; Chen, Y.-K. Expression of Orai1 and STIM1 in Human Oral Squamous Cell Carcinogenesis. J. Dent. Sci. 2022, 17, 78–88. [CrossRef]
- 33. Lunz, V.; Romanin, C.; Frischauf, I. STIM1 Activation of Orai1. Cell Calcium 2019, 77, 29–38. [CrossRef]
- Wolgemuth, D.J. Function of Cyclins in Regulating the Mitotic and Meiotic Cell Cycles in Male Germ Cells. *Cell Cycle Georget*. 2008, 7, 3509–3513. [CrossRef]
- 35. Wang, Z. Cell Cycle Progression and Synchronization: An Overview. Methods Mol. Biol. 2022, 2579, 3–23. [CrossRef]
- 36. Loyer, P.; Trembley, J.H. Roles of CDK/Cyclin Complexes in Transcription and Pre-mRNA Splicing: Cyclins L and CDK11 at the Cross-Roads of Cell Cycle and Regulation of Gene Expression. *Semin. Cell Dev. Biol.* **2020**, *107*, 36–45. [CrossRef]

- Zhang, W.; Liu, Y.; Jang, H.; Nussinov, R. Cell Cycle Progression Mechanisms: Slower Cyclin-D/CDK4 Activation and Faster Cyclin-E/CDK2. *BioRxiv Prepr. Serv. Biol.* 2023. [CrossRef]
- Steurer, S.; Riemann, C.; Büscheck, F.; Luebke, A.M.; Kluth, M.; Hube-Magg, C.; Hinsch, A.; Höflmayer, D.; Weidemann, S.; Fraune, C.; et al. P63 Expression in Human Tumors and Normal Tissues: A Tissue Microarray Study on 10,200 Tumors. *Biomark. Res.* 2021, 9,7. [CrossRef]
- 39. Bavle, R.M.; Paremala, K.; Venugopal, R.; Rudramuni, A.S.; Khan, N.; Hosthor, S.S. Grading of Oral Leukoplakia: Can It Be Improvised Using Immunohistochemical Markers P63 and CD31. *Contemp. Clin. Dent.* **2021**, *12*, 37–43. [CrossRef]
- Truong, A.B.; Kretz, M.; Ridky, T.W.; Kimmel, R.; Khavari, P.A. P63 Regulates Proliferation and Differentiation of Developmentally Mature Keratinocytes. *Genes Dev.* 2006, 20, 3185–3197. [CrossRef]
- 41. Bergholz, J.; Xiao, Z.-X. Role of P63 in Development, Tumorigenesis and Cancer Progression. *Cancer Microenviron. Off. J. Int. Cancer Microenviron. Soc.* 2012, *5*, 311–322. [CrossRef]
- 42. Patel, S.B.; Manjunatha, B.S.; Shah, V.; Soni, N.; Sutariya, R. Immunohistochemical Evaluation of P63 and Cyclin D1 in Oral Squamous Cell Carcinoma and Leukoplakia. *J. Korean Assoc. Oral Maxillofac. Surg.* **2017**, *43*, 324–330. [CrossRef]
- 43. Gupta, S.; Gupta, V.; Tyagi, N.; Vij, R.; Vij, H.; Sharma, E. Analysis of Role of Angiogenesis in Epithelial Dysplasia: An Immunohistochemical Study. J. Clin. Diagn. Res. 2017, 11, EC29–EC34. [CrossRef]
- 44. Tawara, M.; Suzuki, H.; Goto, N.; Tanaka, T.; Kaneko, M.K.; Kato, Y. A Novel Anti-CD44 Variant 9 Monoclonal Antibody C44Mab-1 Was Developed for Immunohistochemical Analyses against Colorectal Cancers. *Curr. Issues Mol. Biol.* **2023**, 45, 3658–3673. [CrossRef]
- Venkat Naga, S.K.S.; Shekar, P.C.; Kattappagari, K.K.; Prakash Chandra, K.L.; Reddy, G.S.; Ramana Reddy, B.V. Expression of Cluster Differentiation-44 Stem Cell Marker in Grades of Oral Epithelial Dysplasia: A Preliminary Study. J. Oral Maxillofac. Pathol. JOMFP 2019, 23, 203–207. [CrossRef]
- 46. Aravind, T.; Janardhanan, M.; Rakesh, S.; Savithri, V.; Unnikrishnan, U.G. Immunolocalization of Osteopontin in Dysplasias and Squamous Cell Carcinomas Arising from Oral Epithelium. *J. Oral Maxillofac. Pathol. JOMFP* **2017**, *21*, 18–23. [CrossRef]
- Sodek, J.; Ganss, B.; McKee, M.D. Osteopontin. Crit. Rev. Oral Biol. Med. Off. Publ. Am. Assoc. Oral Biol. 2000, 11, 279–303. [CrossRef]
- 48. Mrochem, J.; Bartnik, W. Osteopontin—A New Marker in Neoplastic Diseases. Contemp. Oncol. Onkol. 2008, 12, 349–353.
- 49. Ozaki, T.; Nakagawara, A. Role of P53 in Cell Death and Human Cancers. Cancers 2011, 3, 994–1013. [CrossRef]
- 50. Williams, A.B.; Schumacher, B. P53 in the DNA-Damage-Repair Process. *Cold Spring Harb. Perspect. Med.* **2016**, *6*, a026070. [CrossRef]
- 51. Babamohamadi, M.; Babaei, E.; Ahmed Salih, B.; Babamohammadi, M.; Jalal Azeez, H.; Othman, G. Recent Findings on the Role of Wild-Type and Mutant P53 in Cancer Development and Therapy. *Front. Mol. Biosci.* **2022**, *9*, 903075. [CrossRef]
- 52. Borrero, L.J.H.; El-Deiry, W.S. Tumor Suppressor P53: Biology, Signaling Pathways, and Therapeutic Targeting. *Biochim. Biophys. Acta Rev. Cancer* **2021**, *1876*, 188556. [CrossRef]
- Pandya, J.A.; Boaz, K.; Natarajan, S.; Manaktala, N.; Nandita, K.P.; Lewis, A.J. A Correlation of Immunohistochemical Expression of TP53 and CDKN1A in Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma. *J. Cancer Res. Ther.* 2018, 14, 666–670. [CrossRef]
- 54. Patil, S.; Gawande, M.; Chaudhari, M.; Sharma, P.; Hande, A.; Sonone, A. Prognostic Significance of P53 Expression in Various Grades of Epithelial Dysplasia. *J. Datta Meghe Inst. Med. Sci. Univ.* **2022**, *17*, 306–310. [CrossRef]
- 55. Sawada, K.; Momose, S.; Kawano, R.; Kohda, M.; Irié, T.; Mishima, K.; Kaneko, T.; Horie, N.; Okazaki, Y.; Higashi, M.; et al. Immunohistochemical Staining Patterns of P53 Predict the Mutational Status of TP53 in Oral Epithelial Dysplasia. *Mod. Pathol. Off. J. US Can. Acad. Pathol. Inc.* 2022, 35, 177–185. [CrossRef]
- Imaizumi, T.; Matsuda, K.; Tanaka, K.; Kondo, H.; Ueki, N.; Kurohama, H.; Otsubo, C.; Matsuoka, Y.; Akazawa, Y.; Miura, S.; et al. Detection of Endogenous DNA Double-Strand Breaks in Oral Squamous Epithelial Lesions by P53-Binding Protein 1. *Anticancer Res.* 2021, 41, 4771–4779. [CrossRef]
- 57. Napoli, M.; Wu, S.J.; Gore, B.L.; Abbas, H.A.; Lee, K.; Checker, R.; Dhar, S.; Rajapakshe, K.; Tan, A.C.; Lee, M.G.; et al. ΔNp63 Regulates a Common Landscape of Enhancer Associated Genes in Non-Small Cell Lung Cancer. *Nat. Commun.* 2022, 13, 614. [CrossRef]
- 58. Abylkassov, R.; Xie, Y. Role of Yes-Associated Protein in Cancer: An Update. Oncol. Lett. 2016, 12, 2277–2282. [CrossRef]
- 59. Ono, S.; Nakano, K.; Takabatake, K.; Kawai, H.; Nagatsuka, H. Immunohistochemistry of YAP and dNp63 and Survival Analysis of Patients Bearing Precancerous Lesion and Oral Squamous Cell Carcinoma. *Int. J. Med. Sci.* **2019**, *16*, 766–773. [CrossRef]
- 60. Sun, X.; Kaufman, P.D. Ki-67: More than a Proliferation Marker. *Chromosoma* **2018**, *127*, 175–186. [CrossRef]
- 61. Booth, D.G.; Takagi, M.; Sanchez-Pulido, L.; Petfalski, E.; Vargiu, G.; Samejima, K.; Imamoto, N.; Ponting, C.P.; Tollervey, D.; Earnshaw, W.C.; et al. Ki-67 Is a PP1-Interacting Protein That Organises the Mitotic Chromosome Periphery. *eLife* **2014**, *3*, e01641. [CrossRef]
- 62. Liang, Y.; Ma, C.; Li, F.; Nie, G.; Zhang, H. The Role of Contactin 1 in Cancers: What We Know So Far. *Front. Oncol.* 2020, 10, 574208. [CrossRef]
- Iqbal, A.; Tamgadge, S.; Tamgadge, A.; Pereira, T.; Kumar, S.; Acharya, S.; Jadhav, A. Evaluation of Ki-67 Expression in Oral Submucous Fibrosis and Its Correlation with Clinical and Histopathological Features. J. Microsc. Ultrastruct. 2019, 8, 20–24. [CrossRef]

- 64. Kim, C.-H.; Lee, H.S.; Park, J.-H.; Choi, J.-H.; Jang, S.-H.; Park, Y.-B.; Lee, M.G.; Hyun, I.G.; Kim, K.I.; Kim, H.S.; et al. Prognostic Role of P53 and Ki-67 Immunohistochemical Expression in Patients with Surgically Resected Lung Adenocarcinoma: A Retrospective Study. J. Thorac. Dis. 2015, 7, 822–833. [CrossRef]
- 65. Lalkota, B.P.; Srinivasa, B.J.; Swamy, M.V.; Hazarika, D.; Jeet, B.M.; Jyothi, K.; Ghosh, M.; Sayeed, S.M.; Nasiruddin, M.; Naik, R. The Role of P53 and Ki67 in Predicting Clinical Outcome in Breast Cancer Patients. J. Cancer Res. Ther. **2023**, *19*, 208. [CrossRef]
- 66. Humayun, S.; Prasad, V.R. Expression of P53 Protein and Ki-67 Antigen in Oral Premalignant Lesions and Oral Squamous Cell Carcinomas: An Immunohistochemical Study. *Natl. J. Maxillofac. Surg.* **2011**, *2*, 38–46. [CrossRef]
- 67. Kumar, P.; Kane, S.; Rathod, G.P. Coexpression of P53 and Ki 67 and Lack of C-erbB2 Expression in Oral Leukoplakias in India. *Braz. Oral Res.* 2012, *26*, 228–234. [CrossRef]
- 68. Kamala, K.A.; Kanetkar, S.R.; Datkhile, K.D.; Sankethguddad, S. Expression of Ki67 Biomarker in Oral Submucous Fibrosis with Clinico-Pathological Correlations: A Prospective Study. *Asian Pac. J. Cancer Prev. APJCP* **2022**, *23*, 253–259. [CrossRef]
- 69. Dash, K.C.; Mahapatra, N.; Bhuyan, L.; Panda, A.; Behura, S.S.; Mishra, P. An Immunohistochemical Study Showing Ki-67 as an Analytical Marker in Oral Malignant and Premalignant Lesions. *J. Pharm. Bioallied Sci.* **2020**, *12*, S274–S278. [CrossRef]
- Mondal, K.; Mandal, R.; Sarkar, B.C. Importance of Ki-67 Labeling in Oral Leukoplakia with Features of Dysplasia and Carcinomatous Transformation: An Observational Study over 4 Years. S. A. J. Cancer 2020, 9, 99–104. [CrossRef]
- Takkem, A.; Barakat, C.; Zakaraia, S.; Zaid, K.; Najmeh, J.; Ayoub, M.; Seirawan, M.Y. Ki-67 Prognostic Value in Different Histological Grades of Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma. *Asian Pac. J. Cancer Prev. APJCP* 2018, 19, 3279–3286. [CrossRef]
- Swain, S.; Nishat, R.; Ramachandran, S.; Raghuvanshi, M.; Behura, S.S.; Kumar, H. Comparative Evaluation of Immunohistochemical Expression of MCM2 and Ki67 in Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma. *J. Cancer Res. Ther.* 2022, 18, 997–1002. [CrossRef]
- 73. Gadbail, A.R.; Chaudhary, M.; Sarode, S.C.; Gondivkar, S.; Tekade, S.A.; Zade, P.; Hande, A.; Sarode, G.S.; Patil, S. Ki67, CD105, and α-SMA Expression Supports the Transformation Relevant Dysplastic Features in the Atrophic Epithelium of Oral Submucous Fibrosis. *PLoS ONE* 2018, *13*, e0200171. [CrossRef]
- 74. Gadbail, A.R.; Chaudhary, M.S.; Sarode, S.C.; Gawande, M.; Korde, S.; Tekade, S.A.; Gondivkar, S.; Hande, A.; Maladhari, R. Ki67, CD105, and α-SMA Expressions Better Relate the Binary Oral Epithelial Dysplasia Grading System of World Health Organization. J. Oral Pathol. Med. Off. Publ. Int. Assoc. Oral Pathol. Am. Acad. Oral Pathol. 2017, 46, 921–927. [CrossRef]
- Suwasini, S.; Chatterjee, K.; Purkait, S.K.; Samaddar, D.; Chatterjee, A.; Kumar, M. Expression of P53 Protein and Ki-67 Antigen in Oral Leukoplakia with Different Histopathological Grades of Epithelial Dysplasia. *J. Int. Soc. Prev. Community Dent.* 2018, *8*, 513–522. [CrossRef]
- Leung, E.Y.; McMahon, J.D.; McLellan, D.R.; Syyed, N.; McCarthy, C.E.; Nixon, C.; Orange, C.; Brock, C.; Hunter, K.D.; Adams, P.D. DNA Damage Marker Phosphorylated Histone H2AX Is a Potential Predictive Marker for Progression of Epithelial Dysplasia of the Oral Cavity. *Histopathology* 2017, *71*, 522–528. [CrossRef]
- Monteiro, L.; Silva, P.; Delgado, L.; Amaral, B.; Garcês, F.; Salazar, F.; Pacheco, J.-J.; Lopes, C.; Bousbaa, H.; Warnakulasuriya, S. Expression of Spindle Assembly Checkpoint Proteins BubR1 and Mad2 Expression as Potential Biomarkers of Malignant Transformation of Oral Leukoplakia: An Observational Cohort Study. *Med. Oral Patol. Oral Cirugia Bucal* 2021, 26, e719–e728. [CrossRef]
- 78. Rubin, C.I.; Atweh, G.F. The Role of Stathmin in the Regulation of the Cell Cycle. J. Cell. Biochem. 2004, 93, 242–250. [CrossRef]
- 79. Feng, S.; Song, Y.; Shen, M.; Xie, S.; Li, W.; Lu, Y.; Yang, Y.; Ou, G.; Zhou, J.; Wang, F.; et al. Microtubule-Binding Protein FOR20 Promotes Microtubule Depolymerization and Cell Migration. *Cell Discov.* **2017**, *3*, 17032. [CrossRef]
- Vadla, P.; Deepthi, G.; Kumar, C.A.; Bashamalla, R.; Syeda, N.; Naramala, S. Immunohistochemical Expression of Stathmin in Oral Dysplasia: An Original Study with an Insight of Its Action on Microtubules. J. Oral Maxillofac. Pathol. JOMFP 2021, 25, 247–252. [CrossRef]
- 81. Blanpain, C.; Horsley, V.; Fuchs, E. Epithelial Stem Cells: Turning over New Leaves. Cell 2007, 128, 445–458. [CrossRef]
- 82. Kalluri, R.; Weinberg, R.A. The Basics of Epithelial-Mesenchymal Transition. J. Clin. Investig. 2009, 119, 1420–1428. [CrossRef]
- Dongre, A.; Weinberg, R.A. New Insights into the Mechanisms of Epithelial-Mesenchymal Transition and Implications for Cancer. Nat. Rev. Mol. Cell Biol. 2019, 20, 69–84. [CrossRef]
- 84. Ding, D.-C.; Shyu, W.-C.; Lin, S.-Z. Mesenchymal Stem Cells. Cell Transplant. 2011, 20, 5–14. [CrossRef]
- 85. Huang, Y.; Hong, W.; Wei, X. The Molecular Mechanisms and Therapeutic Strategies of EMT in Tumor Progression and Metastasis. *J. Hematol. Oncol.* **2022**, *15*, 129. [CrossRef]
- Chen, T.; You, Y.; Jiang, H.; Wang, Z.Z. Epithelial-Mesenchymal Transition (EMT): A Biological Process in the Development, Stem Cell Differentiation, and Tumorigenesis. J. Cell. Physiol. 2017, 232, 3261–3272. [CrossRef]
- Toriumi, K.; Berto, S.; Koike, S.; Usui, N.; Dan, T.; Suzuki, K.; Miyashita, M.; Horiuchi, Y.; Yoshikawa, A.; Asakura, M.; et al. Combined Glyoxalase 1 Dysfunction and Vitamin B6 Deficiency in a Schizophrenia Model System Causes Mitochondrial Dysfunction in the Prefrontal Cortex. *Redox Biol.* 2021, 45, 102057. [CrossRef]
- Krisanaprakornkit, S.; Iamaroon, A. Epithelial-Mesenchymal Transition in Oral Squamous Cell Carcinoma. ISRN Oncol. 2012, 2012, 681469. [CrossRef]
- 89. Ling, Z.; Cheng, B.; Tao, X. Epithelial-to-Mesenchymal Transition in Oral Squamous Cell Carcinoma: Challenges and Opportunities. *Int. J. Cancer* 2021, *148*, 1548–1561. [CrossRef]

- Loh, C.-Y.; Chai, J.Y.; Tang, T.F.; Wong, W.F.; Sethi, G.; Shanmugam, M.K.; Chong, P.P.; Looi, C.Y. The E-Cadherin and N-Cadherin Switch in Epithelial-to-Mesenchymal Transition: Signaling, Therapeutic Implications, and Challenges. *Cells* 2019, *8*, 1118. [CrossRef]
- 91. Radisky, E.S.; Radisky, D.C. Matrix Metalloproteinase-Induced Epithelial-Mesenchymal Transition in Breast Cancer. J. Mammary Gland Biol. Neoplasia 2010, 15, 201–212. [CrossRef]
- 92. Usman, S.; Waseem, N.H.; Nguyen, T.K.N.; Mohsin, S.; Jamal, A.; Teh, M.-T.; Waseem, A. Vimentin Is at the Heart of Epithelial Mesenchymal Transition (EMT) Mediated Metastasis. *Cancers* **2021**, *13*, 4985. [CrossRef]
- Lamouille, S.; Xu, J.; Derynck, R. Molecular Mechanisms of Epithelial-Mesenchymal Transition. *Nat. Rev. Mol. Cell Biol.* 2014, 15, 178–196. [CrossRef]
- 94. Fedele, M.; Sgarra, R.; Battista, S.; Cerchia, L.; Manfioletti, G. The Epithelial–Mesenchymal Transition at the Crossroads between Metabolism and Tumor Progression. *Int. J. Mol. Sci.* 2022, 23, 800. [CrossRef]
- 95. Herrmann, H.; Bär, H.; Kreplak, L.; Strelkov, S.V.; Aebi, U. Intermediate Filaments: From Cell Architecture to Nanomechanics. *Nat. Rev. Mol. Cell Biol.* **2007**, *8*, 562–573. [CrossRef]
- Batool, S.; Fahim, A.; Qureshi, A.; Jabeen, H.; Ali, S.N.; Khoso, M.Y. Role Of Alteration Of Ck5\6 Profile In Dysplastic Progression Of Oral Mucosa In Tobacco Users. J. Ayub Med. Coll. Abbottabad JAMC 2020, 32, 527–530.
- 97. Lindberg, K.; Rheinwald, J.G. Suprabasal 40 Kd Keratin (K19) Expression as an Immunohistologic Marker of Premalignancy in Oral Epithelium. *Am. J. Pathol.* **1989**, 134, 89–98.
- Rajeswari, P.; Janardhanan, M.; Suresh, R.; Savithri, V.; Aravind, T.; Raveendran, G.C. Expression of CK 19 as a Biomarker in Early Detection of Oral Squamous Cell Carcinoma. J. Oral Maxillofac. Pathol. JOMFP 2020, 24, 523–529. [CrossRef]
- 99. Tian, X.; Liu, Z.; Niu, B.; Zhang, J.; Tan, T.K.; Lee, S.R.; Zhao, Y.; Harris, D.C.H.; Zheng, G. E-Cadherin/β-Catenin Complex and the Epithelial Barrier. *J. Biomed. Biotechnol.* **2011**, 2011, 567305. [CrossRef]
- Gumbiner, B.M. Regulation of Cadherin-Mediated Adhesion in Morphogenesis. Nat. Rev. Mol. Cell Biol. 2005, 6, 622–634.
 [CrossRef]
- 101. Hulpiau, P.; van Roy, F. Molecular Evolution of the Cadherin Superfamily. Int. J. Biochem. Cell Biol. 2009, 41, 349–369. [CrossRef]
- Yuksel, H.; Ocalan, M.; Yilmaz, O. E-Cadherin: An Important Functional Molecule at Respiratory Barrier Between Defence and Dysfunction. *Front. Physiol.* 2021, 12, 720227. [CrossRef]
- 103. Chowdhury, P.; Nagamalini, B.R.; Singh, J.; Ashwini, B.K.; Sharada; Swaminathan, U. Expression of β-Catenin in Oral Leukoplakia and Oral Submucous Fibrosis: An Immunohistochemical Study. J. Oral Maxillofac. Pathol. JOMFP 2021, 25, 124–130. [CrossRef]
- 104. Prgomet, Z.; Andersson, T.; Lindberg, P. Higher Expression of WNT5A Protein in Oral Squamous Cell Carcinoma Compared with Dysplasia and Oral Mucosa with a Normal Appearance. *Eur. J. Oral Sci.* **2017**, *125*, 237–246. [CrossRef]
- 105. Sharada, P.; Swaminathan, U.; Nagamalini, B.R.; Kumar, K.V.; Ashwini, B.K.; Lavanya, V. Coalition of E-Cadherin and Vascular Endothelial Growth Factor Expression in Predicting Malignant Transformation in Common Oral Potentially Malignant Disorders. J. Oral Maxillofac. Pathol. JOMFP 2018, 22, 40–47. [CrossRef]
- 106. Sharma, J.; Bhargava, M.; Aggarwal, S.; Aggarwal, A.; Varshney, A.; Chopra, D. Immunohistochemical Evaluation of E-Cadherin in Oral Epithelial Dysplasia and Squamous Cell Carcinoma. *Indian J. Pathol. Microbiol.* **2022**, *65*, 755–760. [CrossRef]
- 107. Puneeta, N.; Santosh, T.; Mishra, I.; Gaikwad, P.; Sahu, A. Evaluation of E-Cadherin and Vimentin Expression for Different Grades of Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma—An Immunohistochemical Study. J. Oral Maxillofac. Pathol. JOMFP 2022, 26, 285–286. [CrossRef]
- 108. Miguel, A.F.P.; Embaló, B.; Alves Dias, H.B.; Rivero, E.R.C. Immunohistochemical Expression of MMP-9, TIMP-1, and Vimentin and Its Correlation With Inflammatory Reaction and Clinical Parameters in Oral Epithelial Dysplasia. *Appl. Immunohistochem. Mol. Morphol. AIMM* 2021, 29, 382–389. [CrossRef]
- Chandolia, B.; Rajliwal, J.P.; Bajpai, M.; Arora, M. Prognostic Potential of N-Cadherin in Oral Squamous Cell Carcinoma via Immunohistochemical Methods. J. Coll. Physicians Surg.-Pak. JCPSP 2017, 27, 475–478.
- 110. Nelson, W.J.; Nusse, R. Convergence of Wnt, β-Catenin, and Cadherin Pathways. Science 2004, 303, 1483–1487. [CrossRef]
- 111. Qahtani, M.S.; El-Deeb, A.M.; Metwaly, H.A.M. Evaluation of Immunohistochemical Expression of TWIST in Oral Epithelial Dysplasia and Squamous Cell Carcinoma. *Int. J. Health Sci.* 2020, *14*, 33–39.
- 112. Quintanilla, M.; Montero-Montero, L.; Renart, J.; Martín-Villar, E. Podoplanin in Inflammation and Cancer. *Int. J. Mol. Sci.* 2019, 20, 707. [CrossRef]
- 113. Zhang, J.; Du, Y.; Wei, Z.; Tai, B.; Jiang, H.; Du, M. The Prevalence and Risk Indicators of Tooth Wear in 12- and 15-Year-Old Adolescents in Central China. *BMC Oral Health* **2015**, *15*, 120. [CrossRef]
- 114. Ward, L.S.C.; Sheriff, L.; Marshall, J.L.; Manning, J.E.; Brill, A.; Nash, G.B.; McGettrick, H.M. Podoplanin Regulates the Migration of Mesenchymal Stromal Cells and Their Interaction with Platelets. *J. Cell Sci.* **2019**, 132, jcs222067. [CrossRef]
- Karunagaran, M.; Murali, P.; Palaniappan, V.; Sivapathasundharam, B. Expression and Distribution Pattern of Podoplanin in Oral Submucous Fibrosis with Varying Degrees of Dysplasia—An Immunohistochemical Study. J. Histotechnol. 2019, 42, 80–86. [CrossRef]
- 116. Lunawat, S.D.; Prakash, N.; Pradeep, G.L.; Chaware, S.J.; Chaudhari, N.R.; Salunkhe, V.P. Assessment of Podoplanin Lymphatic Vessel Density in Oral Epithelial Dysplasia. *J. Oral Maxillofac. Pathol. JOMFP* **2021**, *25*, 548. [CrossRef]

- 117. Monteiro, L.; do Amaral, B.; Delgado, L.; Garcês, F.; Salazar, F.; Pacheco, J.J.; Lopes, C.; Warnakulasuriya, S. Podoplanin Expression Independently and Jointly with Oral Epithelial Dysplasia Grade Acts as a Potential Biomarker of Malignant Transformation in Oral Leukoplakia. *Biomolecules* 2022, 12, 606. [CrossRef]
- 118. Abidullah, M.; Nahar, P.; Ahmed, S.A. Expression of MUC4 in Oral Dysplastic Epithelium. *Int. J. Pharm. Investig.* **2019**, *9*, 85–88. [CrossRef]
- 119. Carraway, K.L.; Theodoropoulos, G.; Kozloski, G.A.; Carothers Carraway, C.A. Muc4/MUC4 Functions and Regulation in Cancer. *Future Oncol.* 2009, *5*, 1631–1640. [CrossRef]
- 120. Hardwick, J.M.; Soane, L. Multiple Functions of BCL-2 Family Proteins. *Cold Spring Harb. Perspect. Biol.* 2013, 5, a008722. [CrossRef]
- 121. Chao, D.T.; Korsmeyer, S.J. BCL-2 Family: Regulators of Cell Death. Annu. Rev. Immunol. 1998, 16, 395–419. [CrossRef]
- 122. Hua, C.; Zorn, S.; Jensen, J.P.; Coupland, R.W.; Ko, H.S.; Wright, J.J.; Bakhshi, A. Consequences of the t(14;18) Chromosomal Translocation in Follicular Lymphoma: Deregulated Expression of a Chimeric and Mutated BCL-2 Gene. *Oncogene Res.* **1988**, *2*, 263–275.
- 123. Pathak, A.; Shetty, D.C.; Dhanapal, R.; Kaur, G. To Analyse the Mitotic and Keratinisation Correlation with Bcl-2 Expression in Varying Grades of Oral Epithelial Dysplasia and Squamous Cell Carcinoma. J. Oral Maxillofac. Pathol. JOMFP 2022, 26, 316–321. [CrossRef]
- 124. Pallavi, N.; Nalabolu, G.R.K.; Hiremath, S.K.S. Bcl-2 and c-Myc Expression in Oral Dysplasia and Oral Squamous Cell Carcinoma: An Immunohistochemical Study to Assess Tumor Progression. J. Oral Maxillofac. Pathol. JOMFP **2018**, 22, 325–331. [CrossRef]
- 125. Han, Y.; Liu, D.; Li, L. PD-1/PD-L1 Pathway: Current Researches in Cancer. Am. J. Cancer Res. 2020, 10, 727–742.
- 126. Kujan, O.; Agag, M.; Smaga, M.; Vaishnaw, Y.; Idrees, M.; Shearston, K.; Farah, C.S. PD-1/PD-L1, Treg-Related Proteins, and Tumour-Infiltrating Lymphocytes Are Associated with the Development of Oral Squamous Cell Carcinoma. *Pathology* 2022, 54, 409–416. [CrossRef]
- 127. Wang, Q.; Yang, H.-S. The Role of Pdcd4 in Tumor Suppression and Protein Translation. Biol. Cell 2018, 110, 169–177. [CrossRef]
- 128. Desai, K.M.; Kale, A.D. Immunoexpression of Programmed Cell Death 4 Protein in Normal Oral Mucosa, Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma. J. Oral Maxillofac. Pathol. JOMFP 2017, 21, 462. [CrossRef]
- 129. Ferns, G.; Shams, S.; Shafi, S. Heat Shock Protein 27: Its Potential Role in Vascular Disease. *Int. J. Exp. Pathol.* 2006, 87, 253–274. [CrossRef]
- 130. Vidyasagar, A.; Wilson, N.A.; Djamali, A. Heat Shock Protein 27 (HSP27): Biomarker of Disease and Therapeutic Target. *Fibrogenesis Tissue Repair* **2012**, *5*, 7. [CrossRef]
- 131. Karri, R.L.; Subramanyam, R.V.; Venigella, A.; Babburi, S.; Pinisetti, S.; Rudraraju, A. Differential Expression of Heat Shock Protein 27 in Oral Epithelial Dysplasias and Squamous Cell Carcinoma. *J. Microsc. Ultrastruct.* **2020**, *8*, 62–68. [CrossRef]
- Yagui-Beltran, A.; Craig, A.L.; Lawrie, L.; Thompson, D.; Pospisilova, S.; Johnston, D.; Kernohan, N.; Hopwood, D.; Dillon, J.F.; Hupp, T.R. The Human Oesophageal Squamous Epithelium Exhibits a Novel Type of Heat Shock Protein Response. *Eur. J. Biochem.* 2001, 268, 5343–5355. [CrossRef]
- 133. Chen, K.; Li, Y.; Dai, Y.; Li, J.; Qin, Y.; Zhu, Y.; Zeng, T.; Ban, X.; Fu, L.; Guan, X.-Y. Characterization of Tumor Suppressive Function of Cornulin in Esophageal Squamous Cell Carcinoma. *PLoS ONE* **2013**, *8*, e68838. [CrossRef]
- 134. Santosh, N.; McNamara, K.K.; Beck, F.M.; Kalmar, J.R. Expression of Cornulin in Oral Premalignant Lesions. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol.* 2019, 127, 526–534. [CrossRef]
- 135. Lee, J.-W.; Bae, S.-H.; Jeong, J.-W.; Kim, S.-H.; Kim, K.-W. Hypoxia-Inducible Factor (HIF-1)Alpha: Its Protein Stability and Biological Functions. *Exp. Mol. Med.* **2004**, *36*, 1–12. [CrossRef]
- 136. Patel, N.R.; Jain, L.; Mahajan, A.M.; Hiray, P.V.; Shinde, S.S.; Patel, P.A. An Immunohistochemical Study of HIF-1 Alpha in Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma. *Indian J. Otolaryngol. Head Neck Surg. Off. Publ. Assoc. Otolaryngol. India* 2019, 71, 435–441. [CrossRef]
- Kleinert, H.; Forstermann, U. Inducible Nitric Oxide Synthase. In *xPharm: The Comprehensive Pharmacology Reference*; Enna, S.J., Bylund, D.B., Eds.; Elsevier: New York, NY, USA, 2007; pp. 1–12, ISBN 978-0-08-055232-3.
- 138. Singh, D.N.; Srivastava, K.C.; Potsangbam, A.D.; Shrivastava, D.; Nandini, D.B.; Singh, W.T.; Singh, K.S. A Case-Control Study Comparing and Correlating iNOS Expression among Various Clinicopathological Variants of Oral Leukoplakia and Oral Squamous Cell Carcinoma: A Immunohistochemistry Study. J. Pharm. Bioallied Sci. 2020, 12, S324–S331. [CrossRef]
- 139. Turini, M.E.; DuBois, R.N. Cyclooxygenase-2: A Therapeutic Target. Annu. Rev. Med. 2002, 53, 35–57. [CrossRef]
- 140. Sharada, P.; Swaminathan, U.; Nagamalini, B.; Vinod Kumar, K.; Ashwini, B. Histoscore and Discontinuity Score—A Novel Scoring System to Evaluate Immunohistochemical Expression of COX-2 and Type IV Collagen in Oral Potentially Malignant Disorders and Oral Squamous Cell Carcinoma. *J. Orofac. Sci.* **2021**, *13*, 96–104. [CrossRef]
- 141. Schaller, M.D. Paxillin: A Focal Adhesion-Associated Adaptor Protein. Oncogene 2001, 20, 6459–6472. [CrossRef]
- 142. Alam, S.; Astekar, M.S.; Sapra, G.; Agarwal, A.; Agarwal, A.M.; Vishnu Rao, S.G. Immunohistochemical Expression of Paxillin in Potentially Malignant Disorders and Squamous Cell Carcinoma Patients. J. Oral Maxillofac. Pathol. JOMFP 2022, 26, 322–329. [CrossRef]
- 143. Soonthornthum, T.; Arias-Pulido, H.; Joste, N.; Lomo, L.; Muller, C.; Rutledge, T.; Verschraegen, C. Epidermal Growth Factor Receptor as a Biomarker for Cervical Cancer. *Ann. Oncol.* **2011**, *22*, 2166–2178. [CrossRef]

- 144. Kim, J.W.; Kim, Y.T.; Kim, D.K.; Song, C.H.; Lee, J.W. Expression of Epidermal Growth Factor Receptor in Carcinoma of the Cervix. *Gynecol. Oncol.* **1996**, *60*, 283–287. [CrossRef]
- 145. Fakurnejad, S.; van Keulen, S.; Nishio, N.; Engelen, M.; van den Berg, N.S.; Lu, G.; Birkeland, A.; Baik, F.; Colevas, A.D.; Rosenthal, E.L.; et al. Fluorescence Molecular Imaging for Identification of High-Grade Dysplasia in Patients with Head and Neck Cancer. Oral Oncol. 2019, 97, 50–55. [CrossRef]
- 146. Kawai, R.; Sugita, Y.; Suzumura, T.; Hattori, T.; Yoshida, W.; Kubo, K.; Maeda, H. Melanoma Inhibitory Activity and Melanoma Inhibitory Activity 2 as Novel Immunohistochemical Markers of Oral Epithelial Dysplasia. *J. Clin. Med.* **2021**, *10*, 3661. [CrossRef]
- 147. Ekblom, P.; Lonai, P.; Talts, J.F. Expression and Biological Role of Laminin-1. *Matrix Biol. J. Int. Soc. Matrix Biol.* 2003, 22, 35–47. [CrossRef]
- 148. Vageli, D.; Doukas, P.G.; Zacharouli, K.; Kakanis, V.; Strataki, M.; Zioga, A.; Skoulakis, C.; Koukoulis, G.; Ioannou, M. Laminin Immunostaining in Biopsies as a Useful Biomarker of Early Invasion in Actinic Cheilitis and Differential Diagnosis Between Actinic Cheilitis and Lip Cancer: New Insights. *Head Neck Pathol.* **2022**, *17*, 331–338. [CrossRef]
- Nguyen, C.T.K.; Okamura, T.; Morita, K.-I.; Yamaguchi, S.; Harada, H.; Miki, Y.; Izumo, T.; Kayamori, K.; Yamaguchi, A.; Sakamoto, K. LAMC2 Is a Predictive Marker for the Malignant Progression of Leukoplakia. J. Oral Pathol. Med. Off. Publ. Int. Assoc. Oral Pathol. Am. Acad. Oral Pathol. 2017, 46, 223–231. [CrossRef]
- 150. Debta, P.; Sarode, G.; Siddhartha, S.; Sarode, S.; Debta, F.M.; Swain, S.K.; Sahu, M.C.; Patro, S.; Patil, S. GLUT-1 Expression: An Aid in Complementing the WHO Oral Epithelial Dysplasia Grading System. *J. Contemp. Dent. Pract.* **2020**, *21*, 951–955.
- Patlolla, P.; Shyam, N.D.V.; Kumar, G.K.; Narayen, V.; Konda, P.; Mudududla, P. Evaluation of Glucose Transporter-1 Expression in Oral Epithelial Dysplasia and Oral Squamous Cell Carcinoma: An Immunohistochemical Study. J. Oral Maxillofac. Pathol. JOMFP 2020, 24, 578. [CrossRef]
- 152. Udompatanakorn, C.; Taebunpakul, P. The Expression of Methyltransferase-Like 3 in Oral Precancerous Lesions and Oral Squamous Cell Carcinoma. *Eur. J. Dent.* **2022**, *17*, 349–356. [CrossRef]
- 153. Singh, D.; Nishi, K.; Khambata, K.; Balasinor, N.H. Introduction to Epigenetics: Basic Concepts and Advancements in the Field. In *Epigenetics and Reproductive Health*; Tollefsbol, T., Ed.; Translational Epigenetics; Academic Press: Cambridge, MA, USA, 2020; Volume 21, pp. xxv–xliv.
- 154. Takisawa, H.; Mimura, S.; Kubota, Y. Eukaryotic DNA Replication: From Pre-Replication Complex to Initiation Complex. *Curr. Opin. Cell Biol.* **2000**, *12*, 690–696. [CrossRef]
- 155. Zakaria, S.H.; Farag, H.A.; Khater, D.S. Immunohistochemical Expression of MCM-2 in Oral Epithelial Dysplasias. *Appl. Immunohistochem. Mol. Morphol. AIMM* 2018, 26, 509–513. [CrossRef]
- 156. da Silva Barros, C.C.; de Almeida Freitas, R.; da Costa Miguel, M.C.; da Silveira, É.J.D. DNA Damage through Oxidative Stress Is an Important Event in Oral Leukoplakia. *Arch. Oral Biol.* **2022**, *135*, 105359. [CrossRef]

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