

Supplementary Materials

Hotspot DNA Methyltransferase 3A (*DNMT3A*) and Isocitrate Dehydrogenase 1 and 2 (*IDH1/2*) Mutations in Acute Myeloid Leukemia and Their Relevance as Targets for Immunotherapy

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Table S1. HLA class I typing of EBV-B cell lines.

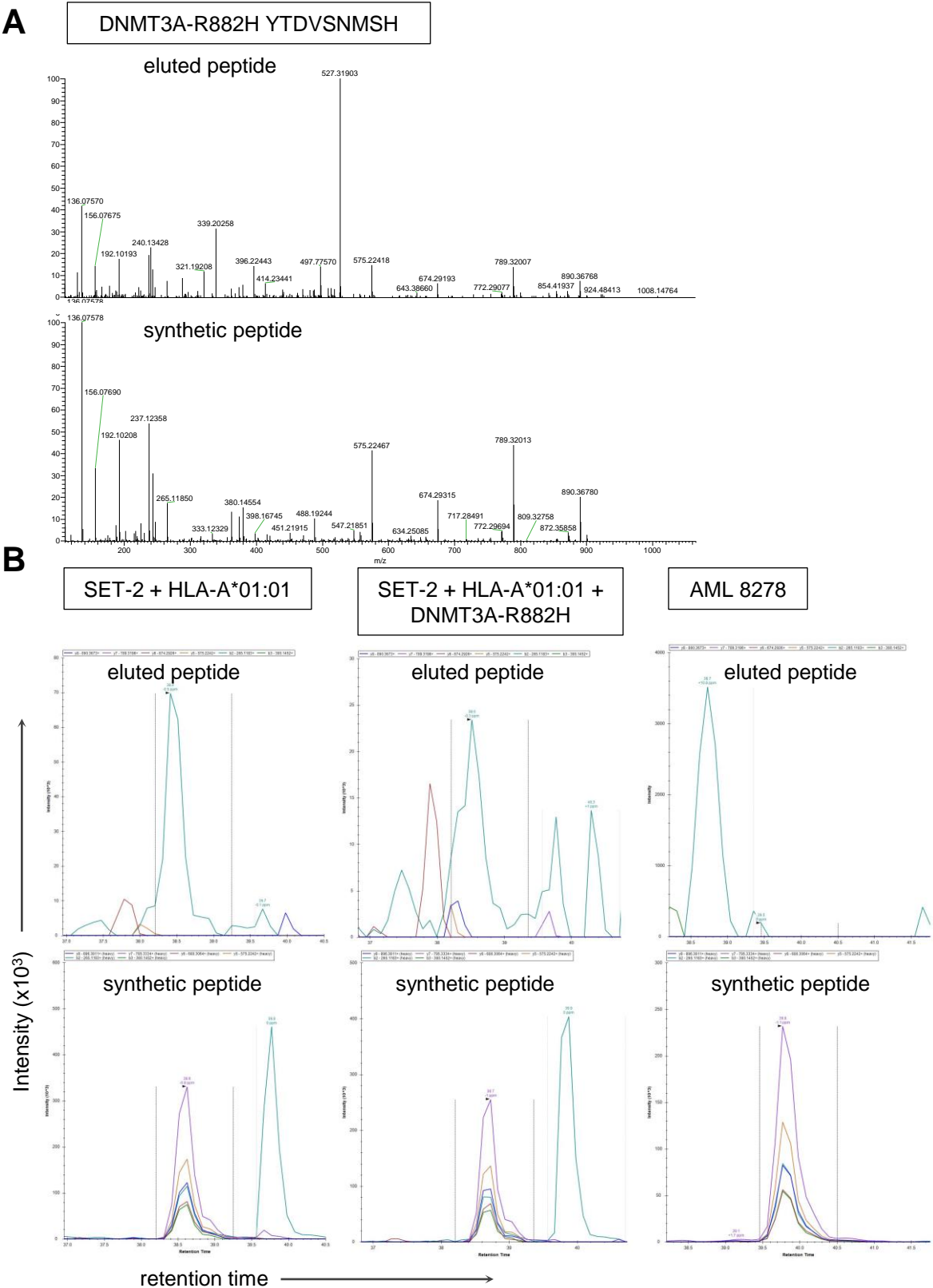
EBV-B cell line	HLA-A		HLA-B		HLA-C	
HG00337	A*03:01	A*03:01	B*07:02	B*07:02	C*07:02	C*07:02
HG00366	A*02:01	A*24:02	B*07:02	B*15:01	C*03:03	C*07:02
HG00265	A*01:01	A*01:01	B*08:01	B*08:01	C*07:01	C*07:01
HG00103	A*02:01	A*11:01	B*35:01	B*40:01	C*03:04	C*04:01
JYW5098	A*02:01	A*02:01	B*07:02	B*07:02	C*07:02	C*07:02

HLA class I immunopeptidomics was performed on five EBV-B cell lines transduced with a minigene construct coding for *DNMT3A*, *IDH1* and *IDH2* hotspot mutations. The EBV-B cell lines were selected for expression of common HLA class I alleles present in at least 20% of European populations, i.e. HLA-A*01:01, A*02:01, A*03:01, A*24:02, B*07:02, B*08:01, C*07:01, C*07:02 and C*03:03/03:04.

Table S2. Selected peptides, transitions and collision energy measurements during parallel reaction monitoring mass spectrometry.

Precursor m/z	Fragment m/z	Sequence	Fragment ion	Peptide variant	Collision energy (%)
528,787838	245,185969	SPNGTIQNIL	y2	light	17
528,787838	698,34678	SPNGTIQNIL	b7	light	17
528,787838	812,389707	SPNGTIQNIL	b8	light	17
528,787838	925,473771	SPNGTIQNIL	b9	light	17
528,787838	406,698492	SPNGTIQNIL	b8	light	17
528,787838	463,240524	SPNGTIQNIL	b9	light	17
528,787838	449,243066	SPNGTIQNIL	a9	light	17
531,794742	245,185969	SPNGTIQNIL	y2	heavy	17
531,794742	704,360589	SPNGTIQNIL	b7	heavy	17
531,794742	818,403516	SPNGTIQNIL	b8	heavy	17
531,794742	931,48758	SPNGTIQNIL	b9	heavy	17
531,794742	409,705396	SPNGTIQNIL	b8	heavy	17
531,794742	466,247428	SPNGTIQNIL	b9	heavy	17
531,794742	452,249971	SPNGTIQNIL	a9	heavy	17
527,218931	890,367257	YTDVSNMSH	y8	light	26
527,218931	789,319579	YTDVSNMSH	y7	light	26
527,218931	674,292636	YTDVSNMSH	y6	light	26
527,218931	575,224222	YTDVSNMSH	y5	light	26
527,218931	265,118283	YTDVSNMSH	b2	light	26
527,218931	380,145226	YTDVSNMSH	b3	light	26
530,225835	896,381066	YTDVSNMSH	y8	heavy	26
530,225835	795,333388	YTDVSNMSH	y7	heavy	26
530,225835	680,306445	YTDVSNMSH	y6	heavy	26
530,225835	575,224222	YTDVSNMSH	y5	heavy	26
530,225835	265,118283	YTDVSNMSH	b2	heavy	26
530,225835	380,145226	YTDVSNMSH	b3	heavy	26
535,216388	906,362172	YTDVSNM[+15.994915]SH	y8	light	26
535,216388	805,314494	YTDVSNM[+15.994915]SH	y7	light	26
535,216388	690,287551	YTDVSNM[+15.994915]SH	y6	light	26
535,216388	591,219137	YTDVSNM[+15.994915]SH	y5	light	26
535,216388	265,118283	YTDVSNM[+15.994915]SH	b2	light	26
535,216388	380,145226	YTDVSNM[+15.994915]SH	b3	light	26
538,223293	912,375981	YTDVSNM[+15.994915]SH	y8	heavy	26
538,223293	811,328303	YTDVSNM[+15.994915]SH	y7	heavy	26
538,223293	696,30136	YTDVSNM[+15.994915]SH	y6	heavy	26
538,223293	591,219137	YTDVSNM[+15.994915]SH	y5	heavy	26
538,223293	265,118283	YTDVSNM[+15.994915]SH	b2	heavy	26
538,223293	380,145226	YTDVSNM[+15.994915]SH	b3	heavy	26
627,276977	1090,48335	YTDVSNM[+15.994915]SHLA	y10	light	26
627,276977	989,435671	YTDVSNM[+15.994915]SHLA	y9	light	26
627,276977	874,408728	YTDVSNM[+15.994915]SHLA	y8	light	26
627,276977	775,340314	YTDVSNM[+15.994915]SHLA	y7	light	26
627,276977	265,118283	YTDVSNM[+15.994915]SHLA	b2	light	26
627,276977	380,145226	YTDVSNM[+15.994915]SHLA	b3	light	26
630,785559	1097,500514	YTDVSNM[+15.994915]SHLA	y10	heavy	26
630,785559	996,452835	YTDVSNM[+15.994915]SHLA	y9	heavy	26
630,785559	881,425892	YTDVSNM[+15.994915]SHLA	y8	heavy	26
630,785559	782,357478	YTDVSNM[+15.994915]SHLA	y7	heavy	26
630,785559	265,118283	YTDVSNM[+15.994915]SHLA	b2	heavy	26
630,785559	380,145226	YTDVSNM[+15.994915]SHLA	b3	heavy	26
619,27952	1074,488435	YTDVSNMSHLA	y10	light	26
619,27952	973,440756	YTDVSNMSHLA	y9	light	26
619,27952	858,413813	YTDVSNMSHLA	y8	light	26
619,27952	759,345399	YTDVSNMSHLA	y7	light	26
619,27952	265,118283	YTDVSNMSHLA	b2	light	26
619,27952	380,145226	YTDVSNMSHLA	b3	light	26
622,788102	1081,505599	YTDVSNMSHLA	y10	heavy	26
622,788102	980,45792	YTDVSNMSHLA	y9	heavy	26
622,788102	865,430977	YTDVSNMSHLA	y8	heavy	26
622,788102	766,362563	YTDVSNMSHLA	y7	heavy	26
622,788102	265,118283	YTDVSNMSHLA	b2	heavy	26
622,788102	380,145226	YTDVSNMSHLA	b3	heavy	26

Figure S1. DNMT3A^{R882H} 9mer YTDVSNMSH is not presented on AML.



(A) The HLA-A*01:01-binding DNMT3A^{R882H}-derived 9-mer YTDVSNMSH peptide was eluted from EBV-B cell line HG00265 transduced with the retroviral minigene construct. Peptide sequence was identified by MS/MS and validated by comparing eluted and synthetic peptide mass spectra. **(B)** Mass chromatograms of YTDVSNMSH by PRM-MS. HLA eluates from AML cell line SET-2 (DNMT3A^{R882H}) transduced with HLA-A*01:01, SET-2 transduced with HLA-A*01:01 and the full-length DNMT3A^{R882H} gene and AML sample 8278 (DNMT3A^{R882H}, HLA-A*01:01) were injected into the mass spectrometer and spiked with 20 fmol YTDVSNMSH (heavy amino acid bold and underlined) as a reference.

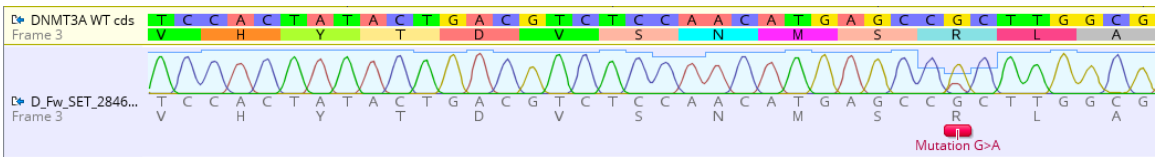
Table S3. Predicted HLA class I-binding DNMT3A, IDH1, and IDH2 neopeptides.

Gene	Mutation	Prevalence	Predicted peptide	HLA	NetMHCpan 4.1 predicted binding	Wildtype variant	NetMHCpan 4.1 predicted binding (%Rank)		
DNMT3A	c.2645G>A / R882H	9.6% ¹⁶	YTDVSNMSHLA	A*01:01	0.701	WB	YTDVSNMSRLA	1.089	WB
			YTDVSNMSHL	A*01:01	0.643	WB	YTDVSNMSRL	0.617	WB
			YTDVSNMSH	A*01:01	0.107	SB	YTDVSNMSR	0.438	SB
			VHYTDVSNMSH	A*01:01	1.389	WB	VHYTDVSNMSR	3.168	
			HYTDVSNMSH	A*01:01	0.808	WB	HYTDVSNMSR	1.885	WB
			MSHLARQRL	B*07:02	1.353	WB	MSRLARQRL	0.683	WB
			MSHLARQRL	B*08:01	1.569	WB	MSRLARQRL	1.164	WB
			SHLARQRLL	B*08:01	1.489	WB	SRLARQRLL	1.234	WB
			HLARQRLL	B*08:01	0.626	WB	RLARQRLL	2.895	
			YTDVSNMSHL	C*03:04	1.004	WB	YTDVSNMSRL	1.175	WB
			MSHLARQRL	C*03:04	0.381	SB	MSRLARQRL	1.525	WB
			SHLARQRLL	C*07:01	0.419	SB	SRLARQRLL	0.070	SB
			HYTDVSNMSHL	C*07:02	1.698	WB	HYTDVSNMSRL	2.423	
			SHLARQRLL	C*07:02	0.647	WB	SRLARQRLL	0.298	SB
	c.2644C>T / R882C	2.5% ¹⁶	YTDVSNMSCLA	A*01:01	1.061	WB	YTDVSNMSRLA	1.089	WB
			YTDVSNMSCL	A*01:01	0.787	WB	YTDVSNMSRL	0.617	WB
			YTDVSNMSC	A*01:01	0.793	WB	YTDVSNMSR	0.438	SB
			SCLARQRLL	B*08:01	1.685	WB	SRLARQRLL	1.234	WB
			YTDVSNMSCL	C*03:04	1.323	WB	YTDVSNMSRL	1.175	WB
IDH1	c.394C>T / R132C	3.6% ¹⁷	KPIIIGCHAY	B*07:02	1.390	WB	KPIIIGRHAY	0.762	WB
			KPIIIGCHA	B*07:02	1.526	WB	KPIIIGRHA	0.744	WB
	c.394C>G / R132G	0.6% ¹⁷	IIGGHAYGDQY	A*01:01	1.807	WB	IIGRHAYGDQY	2.528	
			KPIIIGGHAY	B*07:02	1.023	WB	KPIIIGRHAY	0.762	WB
	c.395G>A / R132H	0.4% ¹⁷	KPIIIGGHA	B*07:02	0.947	WB	KPIIIGRHA	0.744	WB
			IIGHHAYGDQY	A*01:01	1.997	WB	IIGRHAYGDQY	2.528	
			KPIIIGHHAY	B*07:02	1.054	WB	KPIIIGRHAY	0.762	WB
			KPIIIGHHA	B*07:02	0.669	WB	KPIIIGRHA	0.744	WB
			GHHAYGDQY	C*07:01	1.921	WB	GRHAYGDQY	0.302	SB
			GHHAYGDQY	C*07:02	1.082	WB	GRHAYGDQY	0.399	SB
	c.394C>A / R132S	0.4% ¹⁷	IIIGSHAYGDQY	A*01:01	1.594	WB	IIIGRHAYGDQY	3.413	
			IIGSHAYGDQY	A*01:01	1.044	WB	IIGRHAYGDQY	2.528	
			IGSHAYGDQY	A*01:01	0.686	WB	IGRHAYGDQY	4.466	
			PIIIGSHAY	A*01:01	1.950	WB	PIIIGRHAY	3.330	
			GSHAYGDQY	A*01:01	0.345	SB	GRHAYGDQY	6.370	
			KPIIIGSHAY	B*07:02	1.053	WB	KPIIIGRHAY	0.762	WB
KPIIIGSHA			B*07:02	0.652	WB	KPIIIGRHA	0.744	WB	
IDH2	c.419G>A / R140Q	5.4% ¹⁸	KSPNGTIQNI	B*07:02	0.811	WB	KSPNGTIRNI	0.838	WB
			SPNGTIQNIL	B*07:02	0.167	SB	SPNGTIRNIL	0.027	SB
			SPNGTIQNI	B*07:02	0.192	SB	SPNGTIRNI	0.146	SB
	c.515G>A / R172K	2.1% ¹⁸	ITIGKHAHGDQY	A*01:01	0.568	WB	ITIGRHAHGDQY	0.582	WB
			TIGKHAHGDQY	A*01:01	1.408	WB	TIGRHAHGDQY	1.461	WB
			VPGWTKPITIGK	A*03:01	0.983	WB	VPGWTKPITIGR	4.900	
			GWTKPITIGK	A*03:01	0.112	SB	GWTKPITIGR	1.139	WB
			WTKPITIGK	A*03:01	0.270	SB	WTKPITIGR	1.417	WB
			KPITIGKHAH	B*07:02	1.466	WB	KPITIGRHAH	1.108	WB
			KPITIGKHA	B*07:02	0.724	WB	KPITIGRHA	0.583	WB

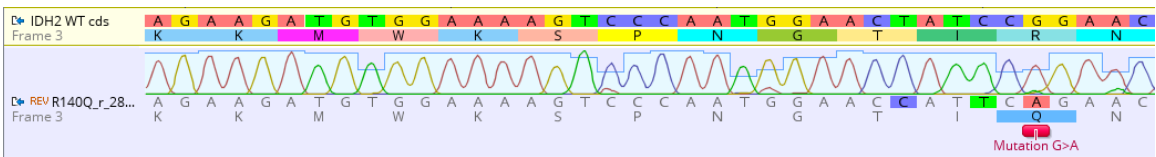
Peptide sequences of 21 amino acids with the mutated amino acid encoded by *DNMT3A*, *IDH1* or *IDH2* hotspot mutations at position 11 were searched for 9-, 10- and 11-mer neopeptides with predicted binding to common HLA class I alleles with population frequencies above 20% in European populations by NetMHCpan4.1. Indicated is the mutation prevalence according to Ley et al. (2010) [16], Schnittger et al. (2010) [17] and Koszarska et al. (2013) [18]. Peptides with predicted binding $\leq 0.5\%$ Rank are strong binders (SB), and peptides with predicted binding $0.5\% > \text{Rank} \leq 2.0\%$ are weak binders (WB). Predicted HLA class I binding is also shown for wildtype peptide variants.

Figure S2. *DNMT3A* and *IDH2* mutations in SET-2 and K562-R140Q.

A. SET-2

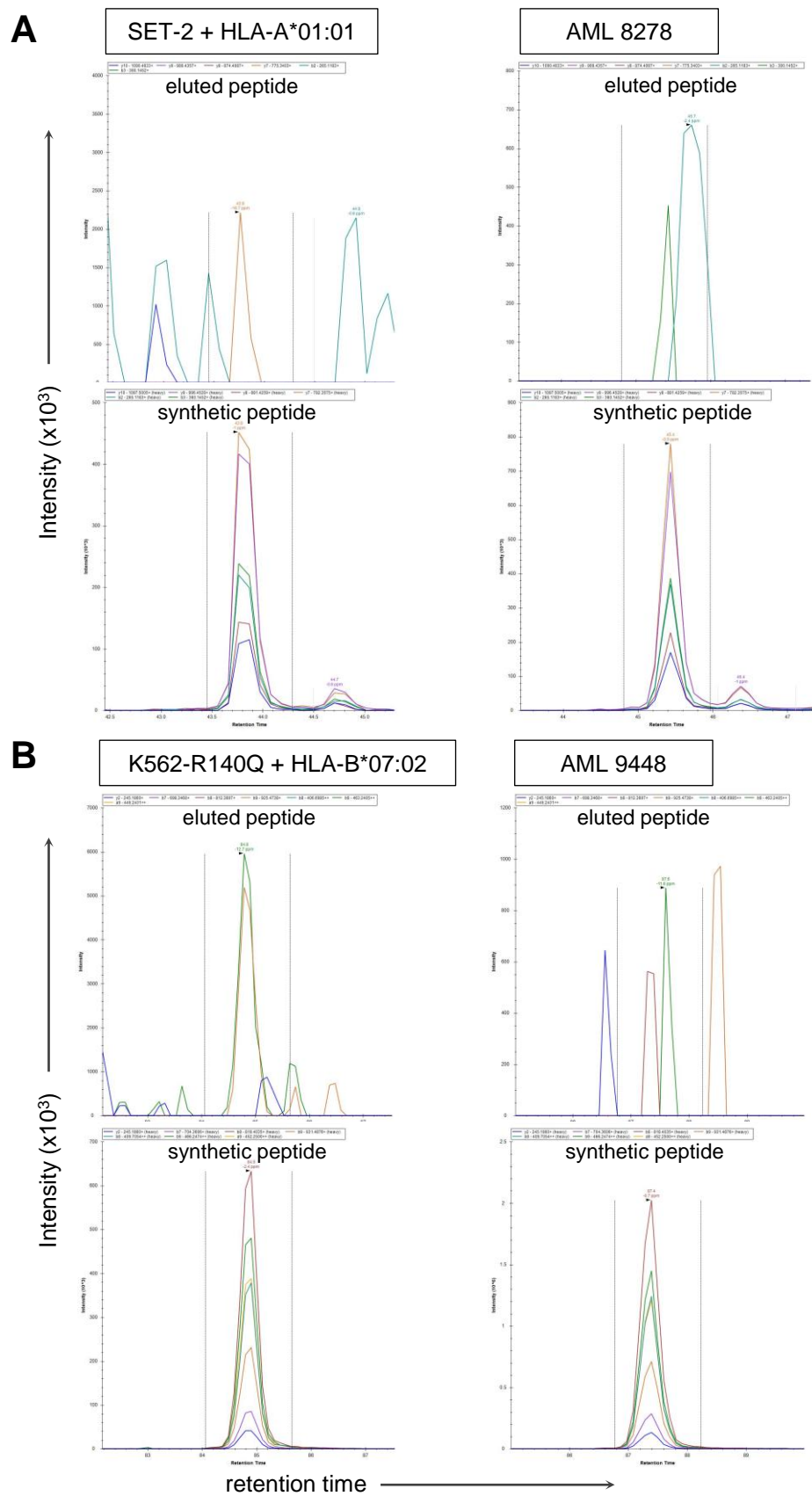


B. K562-R140Q



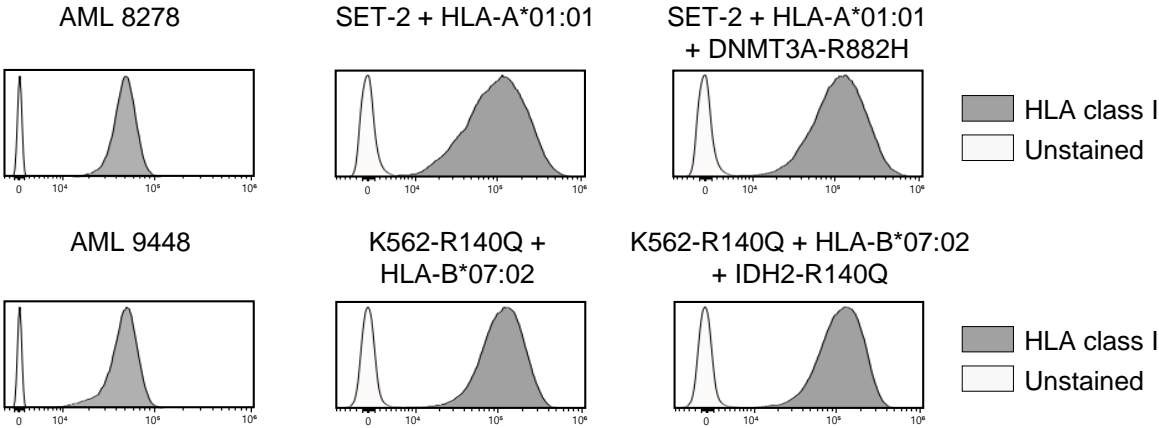
RNA was isolated and cDNA generated from AML cell line SET-2 (*DNMT3A*^{R882H}) and chronic myelogenous leukemia cell line K562-R140Q in which bi-allelic *IDH2*^{R140Q} mutations were introduced by CRISPR/Cas9 mediated genome editing [24]. Mutations were validated by targeted PCR and Sanger sequencing. **(A)** DNA sequence of the *DNMT3A* gene in SET-2 shows a heterozygous c.2645G>A (R882H) mutation. **(B)** DNA sequence of the *IDH2* gene in K562-R140Q shows a homozygous c.419G>A (R140Q) mutation.

Figure S3. Surface presentation of YTDVSNMSHLA and SPNGTIQNIL is not detected by PRM-MS on AML cells with *DNMT3A*^{R882H} or *IDH2*^{R140Q} mutations.



(A) Mass chromatograms showed no YTDVSNMSHLA in HLA-eluates of SET-2 (*DNMT3A*^{R882H}) transduced with HLA-A*01:01 and AML sample 8278 (*DNMT3A*^{R882H}, HLA-A*01:01). Heavy-isotope labeled YTDVSNMSHLA (heavy amino acid bold and underlined) was spiked-in as a reference. **(B)** Mass chromatograms showed no SPNGTIQNIL in HLA-eluates of K562-R140Q transduced with HLA-B*07:02 and AML sample 9448 (*IDH2*^{R140Q}, HLA-B*07:02). Heavy-isotope labeled SPNGTIQNIL was spiked-in as a reference.

Figure S4. HLA class I surface expression on AML samples and cell lines.



Mean fluorescence intensity after staining of patient AML samples and AML cell lines with a fluorescent antibody against HLA class I by flow cytometry.

Table S4. Isolation of DNMT3A^{R882H}-specific T-cell clones.

Donor	HLA-A		HLA-B		Reactivity against DNMT3A ^{R882H} peptide	Reactivity against transduced DNMT3A ^{R882H} gene
3	*01:01	*02:01	*07:02	*08:01	1	0
5	*01:01	*03:XX	*07:02	*08:01	1	0
8	*01:XX	*03:XX	*13:XX	*14:XX	3	1
10	*01:01	*03:01	*08:01	*35:01	0	
21	*01:01	*11:XX	*08:01	*56:01	0	
22	*01:01	*03:01	*08:01	*35:01	1	0
23	*01:01	*03:01	*08:01	*44:02	0	
24	*01:01	*03:01	*08:01	*13:02	0	
25	*01:01		*08:01		0	
30	*01:01	*03:01	*07:02	*08:01	0	
31	*01:01	*03:01	*35:01	*51:01	1	0
35	*01:01	*03:01	*07:02	*08:01	2	0
12	*01:XX	*03:XX	*07:XX	*18:XX	0	
16	*01:XX	*02:XX	*38:XX	*57:XX	0	
17-I ^C	*01:XX	*24:XX	*07:02	*15:17	7	2
17-II ^C	*01:XX	*24:XX	*07:02	*15:17	17	2
11	-	-	-	-	0	
13	-	-	-	-	0	
18	-	-	-	-	0	
19	-	-	-	-	0	
14	*11:01	*24:02	*35:01	*45:01	0	
15	*02:XX	*32:XX	*14:XX	*40:XX	0	
20	*03:XX	*24:XX	*14:01	*15:01	0	
32	*03:01	*25:01	*07:02	*18:01	0	
33	*02:01	*03:01	*07:02	*51:01	0	
34	*11:01	*68:01	*27:05	*57:01	0	
26					43	5

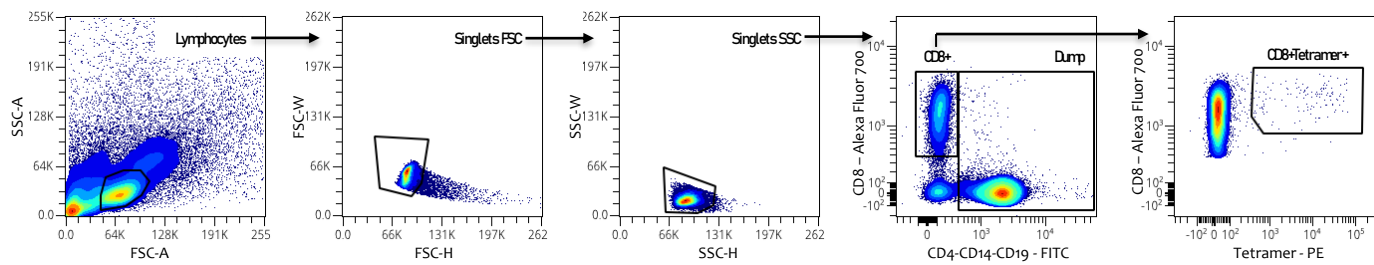
PBMC from 26 healthy individuals were screened for DNMT3A^{R882H}-specific CD8 T cells. Single cell sorted pHLA-tetramer-positive CD8 T-cell clones were isolated and screened for specific recognition of HLA-A*01:01 positive EBV-B cells loaded with YTDVSNMSHLA by IFN- γ ELISA. T-cell clones that were peptide-specific were tested for reactivity against K562 transduced with HLA-A*01:01 and the *DNMT3A^{R882H}* gene and HL-60 (HLA-A*01:01 positive) transduced with the *DNMT3A^{R882H}* gene. Five T cell clones from two healthy donors (donors 8 & 17) were reactive against the *DNMT3A^{R882H}* but not wildtype *DNMT3A* gene. Donor 17 has been screened twice (17-I^C & 17-II^C).

Table S5. Isolation of IDH2^{R140Q}-specific T-cell clones.

Donor	HLA-A		HLA-B		Reactivity against IDH2 ^{R140Q} peptide	Reactivity against the transduced IDH2 ^{R140Q} gene
1	*03:01	*26:01	*07:02	*27:05	1	1
2	*02:01	*03:01	*07:02	*51:01	0	
3	*01:01	*02:01	*07:02	*08:01	0	
4	*02:01	*03:01	*07:02	*44:02	0	
5	*01:01	*03:XX	*07:02	*08:01	1	1
6	*02:01	*03:01	*07:02	*51:01	1	0
7	*02:01	*03:01	*07:02	-	1	0
17-I ^C	*01:XX	*24:XX	*07:02	*15:17	0	
17-II ^C	*01:XX	*24:XX	*07:02	*15:17	0	
28	*03:01	*11:01	*07:02	*40:02	0	
30	*01:01	*03:01	*07:02	*08:01	1	0
32	*03:01	*25:01	*07:02	*18:01	0	
33	*02:01	*03:01	*07:02	*51:01	1	1
35	*01:01	*03:01	*07:02	*08:01	0	
12	*01:XX	*03:XX	*07:XX	*18:XX	0	
26	*02:XX	*11:XX	*07:XX	*15:XX	0	
11	-	-	-	-	0	
13	-	-	-	-	0	
18	-	-	-	-	0	
19	-	-	-	-	0	
9	*01:XX	*11:XX	*08:XX	*13:XX	0	
10	*01:01	*03:01	*08:01	*35:01	0	
14	*11:01	*24:02	*35:01	*45:01	0	
15	*02:XX	*32:XX	*14:XX	*40:XX	0	
16	*01:XX	*02:XX	*38:XX	*57:XX	0	
20	*03:XX	*24:XX	*14:01	*15:01	0	
27	*03:XX	*68/69:XX	*15:01	*27:XX	0	
29	*02:01	*03:01	*35:01	*57:01	0	
31	*01:01	*03:01	*35:01	*51:01	0	
34	*11:01	*68:01	*27:05	*57:01	0	
30					5	3

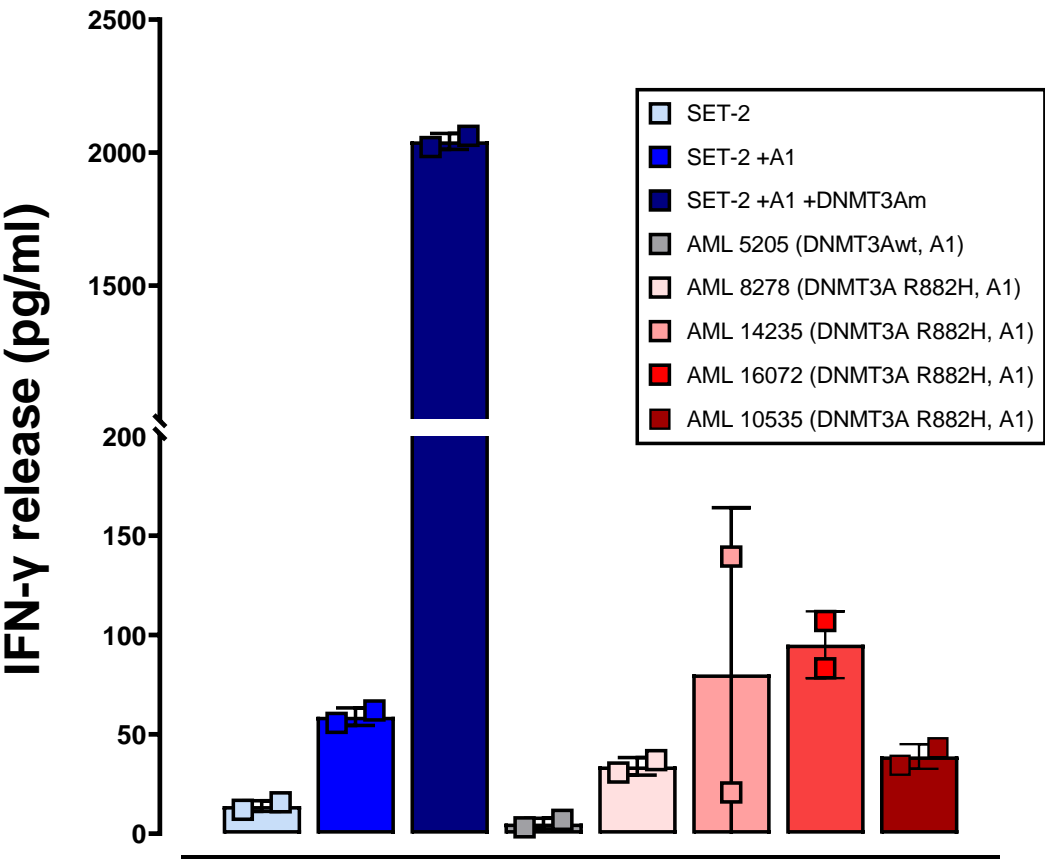
PBMC from 30 healthy individuals were screened for IDH2^{R140Q}-specific CD8 T cells. Single cell sorted pHLA-tetramer-positive CD8 T-cell clones were isolated and screened for specific recognition of HLA-B*07:02 positive EBV-B cells loaded with SPNGTIQNIL by IFN-γ ELISA. T-cell clones that were peptide-specific were tested for reactivity against K562 transduced with HLA-B*07:02 and the full-length IDH2^{R140Q} gene. Three T-cell clones were reactive against the IDH2^{R140Q} but not wildtype IDH2 gene.

Figure S5. Gating strategy for isolation of DNMT3A^{R882H} or IDH2^{R140Q}-specific CD8 T cells by flow cytometry.



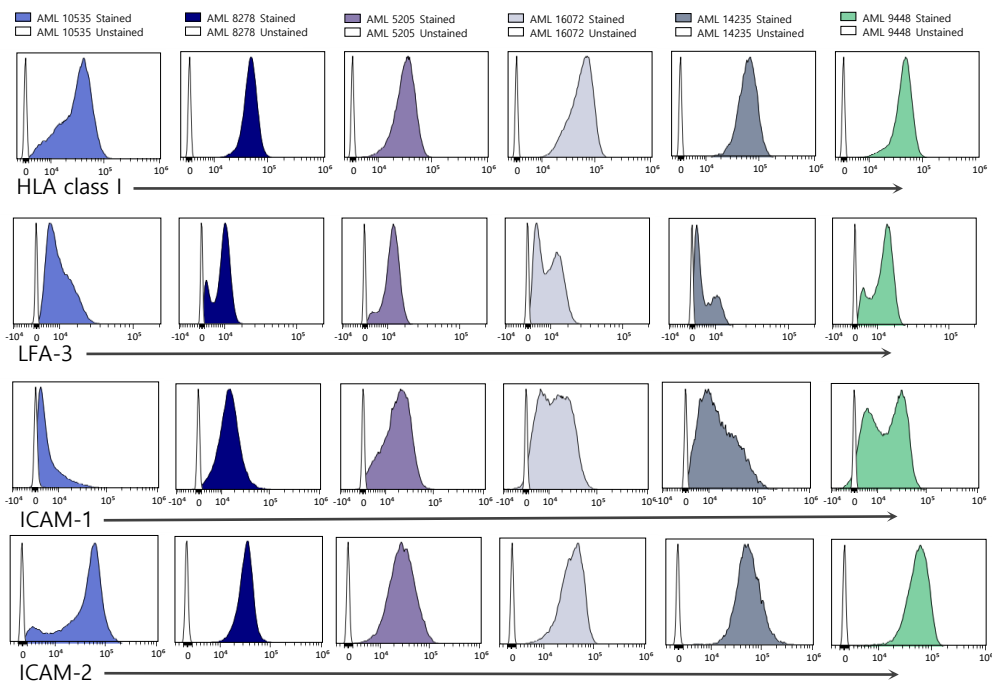
PBMC enriched for pHLA-tetramer-positive CD8 T cells were gated for single-cell lymphocytes that were negative for CD4, CD14 and CD19 (dump). pHLA-tetramer positive T cells within the CD8 gate were single-cell sorted by FACS.

Figure S6. Recognition of patient-derived AML samples by T-cell clone 8.6F10.



Reactivity of T-cell clone 8.6F10 was tested against AML cell line SET-2 (*DNMT3A^{R882H}*), SET-2 transduced with HLA-A*01:01 (SET-2 +A1), SET-2 transduced with HLA-A*01:01 and the full-length *DNMT3A^{R882H}* gene (SET-2 +A1 +DNMT3Am) and four HLA-A*01:01 and *DNMT3A^{R882H}* patient-derived AML samples 8278, 14235, 16072 and 10535. Patient-derived AML 5205 (HLA-A*01:01, wildtype *DNMT3A*) was included as negative control. IFN-γ release in duplicate wells of a single experiment are shown.

Figure S7. Surface expression of HLA class I and adhesion molecules on AML patient samples.



Mean Fluorescence Intensity (MFI) of patient AML samples stained with fluorescent antibodies against HLA class I, LFA-3 (Lymphocyte Function-Associated antigen 3), ICAM-1 (InterCellular Adhesion Molecule-1) or ICAM-2 (InterCellular Adhesion Molecule-2) by flow cytometry. Unstained samples are shown as controls.