

Review

Antidiabetic Medicinal Plants Used in the Eastern Cape Province of South Africa: An Updated Review

Idowu Jonas Sagbo * and Ahmed A. Hussein * 

Chemistry Department, Cape Peninsula University of Technology, Symphony Road, Bellville Campus, Bellville 7535, South Africa

* Correspondence: sagboi@cput.ac.za (I.J.S.); mohammedam@cput.ac.za (A.A.H.)

Abstract: Oral antidiabetic drugs are usually costly and are associated with several adverse side effects. This has led to the use of medicinal plants that are considered to have multiple therapeutic targets and are readily accessible. In the Eastern Cape province of South Africa, the number of people using medicinal plants for the management of diabetes has been climbing steadily over the past two decades due to their cultural acceptability, accessibility, affordability, efficacy, and safety claims. In this study, a review of antidiabetic medicinal plants used in the Eastern Cape province of South Africa was conducted. A comprehensive literature survey was thoroughly reviewed using several scientific databases, ethnobotanical books, theses and dissertations. About forty-eight (48) plant species were identified as being used to treat diabetes by the people of Eastern Cape province. Among the plant species, only eight (8) species have not been scientifically evaluated for their antidiabetic activities and twenty antidiabetic compounds were isolated from these medicinal plants. This review has confirmed the use and potential of the antidiabetic medicinal plants in the Eastern Cape province and identified several promising species for further scientific investigation.

Keywords: antidiabetic drugs; medicinal plants; diabetes mellitus; eastern cape; hyperglycaemia; hypoglycaemia



Citation: Sagbo, I.J.; Hussein, A.A. Antidiabetic Medicinal Plants Used in the Eastern Cape Province of South Africa: An Updated Review. *Processes* **2022**, *10*, 1817. <https://doi.org/10.3390/pr10091817>

Academic Editor: Adriana Trifan

Received: 16 August 2022

Accepted: 3 September 2022

Published: 9 September 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Diabetes mellitus (DM), generally known as diabetes, is a non-communicable metabolic disease described by an abnormal increase in blood sugar (glucose) levels due to complete or relative lack of insulin secretion, with concomitant modifications in the metabolism of lipids, proteins and carbohydrates [1,2]. The chronic hyperglycaemic status in diabetics yields an increased risk of complications due to the long-term damage and dysfunction of several organs such as the eyes, kidneys, heart, and nerves. Symptoms of clear hyperglycaemia include weight loss, blurred vision, polyuria (excessive passage of urine), polydipsia (excessive thirst), and susceptibility to infections, whereas long-term complications may include retinopathy (damage to the retina of the eyes) with possible loss of vision, nephropathy (deterioration of kidney function), and cardiovascular complications such as high blood pressure (hypertension).

Some specific cases of diabetes have been documented, but the vast majority of cases fall into two broad classes, namely, Type 1 diabetes and Type II diabetes. Type 1 diabetes or insulin-dependent diabetes is caused by a relative or absolute deficiency of insulin secretion, commonly due to cell-mediated autoimmune destruction of beta cells of the pancreas, which may have a genetic predisposition [3]. Type 1 diabetes accounts for 5–10% of patients with diabetes. On the other hand, Type 2 diabetes or non-insulin dependent diabetes, or adult-onset diabetes accounts for about 90–95% of diabetic patients and is linked with reduced insulin sensitivity, also known as insulin resistance and/or impaired insulin secretion. The pathogenesis of type 2 diabetes is more flexible, though the autoimmune destruction of beta cells does not occur [4].

1.1. Prevalence of Diabetes

Over the past two decades, the number of people diagnosed with diabetes has reached an unprecedented high and a further increase is expected. The International diabetes federation (IDF) reported that there are 463 million (20–79 years) people globally suffering from this life-threatening disease and this figure is anticipated to increase to 700 million by 2040 [5]. The World Health Organization (WHO) also indicated that if the existing trend lingers, diabetes will be the second highest killer by 2040 unless robust and rigorous actions are made by individuals, communities and governments [6]. These are part of an awareness campaign on the burden of diabetes and the urgency to intensify prevention and control activities. Globally, China has the highest number of people living with diabetes, followed by India and the United States [5]. Asia and Africa have been identified as areas with high diabetic populations which could increase beyond projected levels if urgent attention is not given [5]. In South Africa, there were about 4.5 million cases of diabetes in 2019, with occurrences in the same period placed at 12.7% among adults 20–79 years old [5]. Inadequate promising therapy to cure diabetes could be accountable for the predicted figure in this part of the world.

1.2. Oral Antidiabetic Drugs and Their Limitations in the Treatment of Diabetes Mellitus

Currently, existing oral hypoglycaemic drugs are categorized into different groups according to their mechanisms of action (Figure 1). The list includes, but is not limited to sulfonylureas, meglitinides, biguanides, thiazolidinediones, alpha-glucosidase inhibitors and dipeptidyl peptidase 4 (DPP-IV) inhibitors.

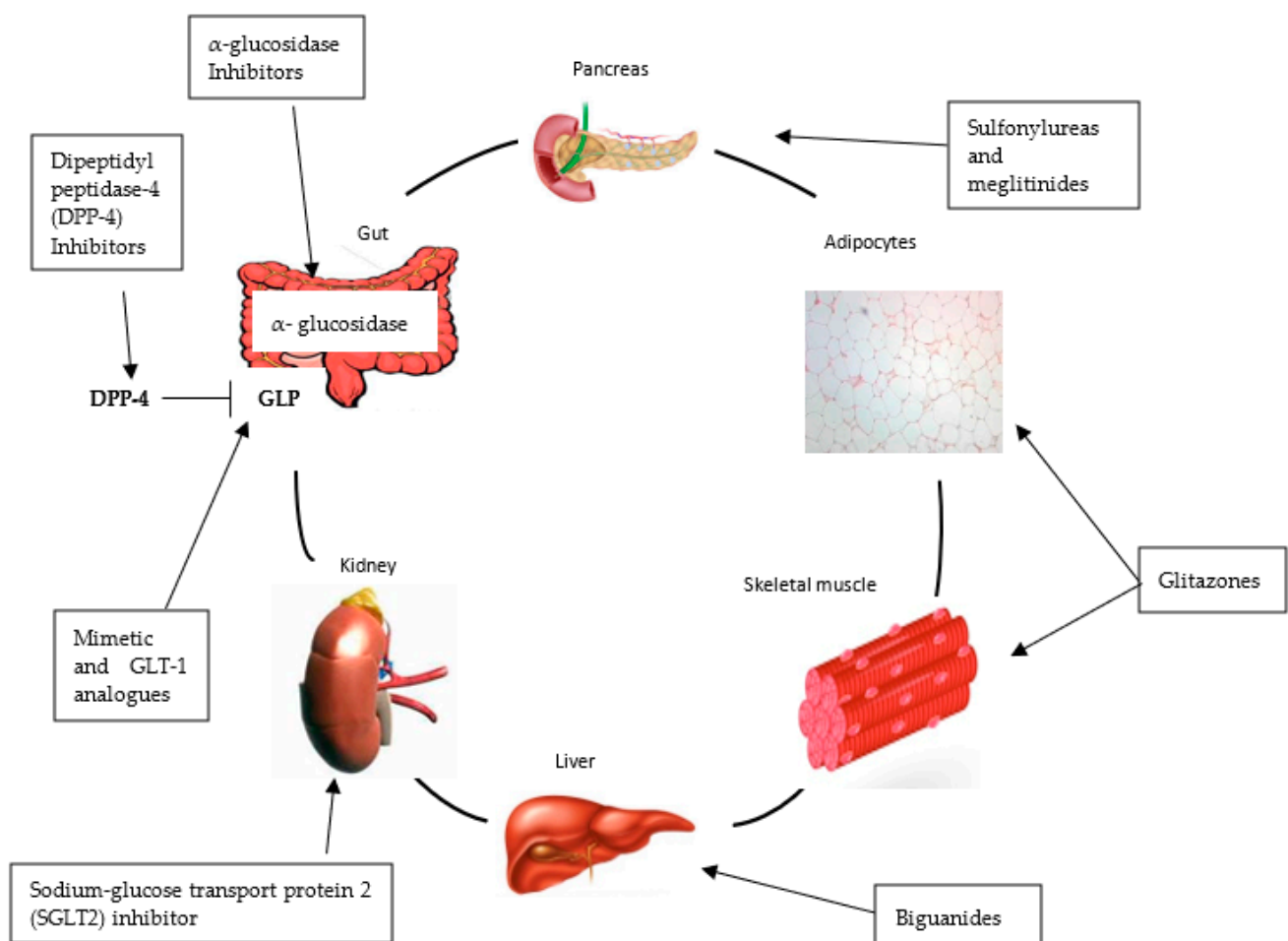


Figure 1. Target tissue of current antidiabetic drugs.

The sulfonylureas act by stimulating insulin release from the pancreatic beta cells [7]. They bind to the SUR-1 (sulfonylurea receptor-1), expressed on the pancreatic beta cell membranes, thereby inhibiting the efflux of potassium ions through the channels that cause depolarization [7]. This depolarization causes the opening of voltage-gated calcium channels, leading to the increased influx of calcium, and this rise in intracellular calcium stimulates insulin release [8]. However, it has been proven that these antidiabetic drugs do not reduce the long-term complication of diabetes and may also enhance appetite hereafter, resulting in weight gain [9]. Examples of drugs in this class are tolbutamide, tolazamide, acetohexamide chlorpropamide, glyburide, glipizide, glibenclamide and glimepiride.

Meglitinides are another class of oral antidiabetic drugs. These drugs reduce blood sugar levels, thereby increasing insulin release from the beta-cell pancreatic [10]. This is achieved by modulating beta cells to secrete insulin by controlling the efflux of potassium through potassium channels. Meglitinides do not have a direct effect on insulin exocytosis as it is in sulfonylureas [11]. This class of antidiabetic drug is taken by diabetic patients shortly before a meal to increase the insulin response to each meal [12]. If a meal is skipped the medication is also skipped. The reported side effects associated with meglitinides include weight gain and hypoglycaemia [13]. Typical examples of the antidiabetic drugs in this class are repaglinide and nateglinide.

The biguanides, another class of oral antidiabetic drugs, work by enhancing insulin sensitivity in peripheral tissues through the alteration of post-receptor signalling in the insulin signalling pathway. Their effects on hepatic tissue result in reduced hepatic glucose production through a reduction in gluconeogenesis and glycogenolysis [14,15]. The best example of this class is metformin. However, metformin has been described to have certain adverse side effects such as heart failure, hepatic impairment, gastrointestinal disturbances and renal impairment [7,16].

The thiazolidinediones are another class of oral antidiabetic drugs that mediate their function by binding to the PPAR γ (peroxisome proliferator-activated receptor gamma) mainly expressed in adipocytes. Binding to PPAR γ stimulates interaction with the retinoid X receptor, which hetero-dimerizes and activates genes that play a significant role in lipid and carbohydrate metabolism [17]. They help to improve muscle and fat sensitivity to insulin and, to a lesser extent, reduce hepatic glucose production. However, thiazolidinediones have been described to be linked with the pathophysiology of fluid-retention and weight increase. The complete effects of fluid retention include oedema, heart failure, liver toxicity and anaemia [18]. Examples of drugs in this class include troglitazone, rosiglitazone and pioglitazone.

The alpha-glucosidase inhibitor class of oral antidiabetic drugs is occasionally called “starch blockers”. They block the action of intestinal enzymes that break down carbohydrates in the small intestine, thus slowing down the absorption of ingested carbohydrates and decreasing post-prandial hyperglycaemia in diabetic patients [19]. However, one of the biggest drawbacks of these oral antidiabetic drugs is their side effects. The prominent side effects include nausea, flatulence, diarrhoea, bloating and abdominal pains [11]. Acarbose and miglitol are examples of this type of oral antidiabetic drug.

The dipeptidyl peptidase 4 (DPP-IV) inhibitors are a new class of oral antidiabetic drugs. They attenuate incretin degradation, thus increasing the half-lives of incretin and enhancing the stimulation of pancreatic insulin secretion and beta cell growth [20]. Incretin hormones (glucose-dependent insulintropic peptide (GIP) and glucagon-like peptide (GLP-1)) contribute meaningfully to glucose-dependent insulin secretion by increasing beta cell mass and decreasing glucagon secretion. These gut hormones (GIP and GLP-1) are highly sensitive to degradation by DPP-IV, a serine protease that cleaves polypeptides containing proline and alanine residues at the penultimate N-terminal position and thus decreases the effectiveness of the hormones. Adverse side effects of DPP-IV inhibitors include headache, runny nose, diarrhea, nausea, stomach pain, and sore throat. The best examples of this class are alogliptin, linagliptin, sitagliptin and saxagliptin.

1.3. Plants as an Alternative Source of Antidiabetic Agents

The use of herbs to treat diabetes mellitus has been highly acceptable as part of medical intervention. This is due to their efficacy, probable fewer side effects and lesser costs [21]. In developing countries, medicinal plants are used to treat diabetes to overwhelm the problem of the cost of western medicines to the populace [22]. These medicinal plants have been indicated to contain several active chemical constituents that are accountable to treat diabetes [22]. Plants have always served as a good source of drugs with some of the existing antidiabetic drugs being acquired directly or indirectly from them [23–25]. For example, metformin, the favoured first-line oral antidiabetic drug, emanates from a derivate of French lilac (*Galega officinalis*) [26].

There have been several review reports on plants with probable antidiabetic activity from different parts of the world in the literature [27–29]. In South Africa, several ethnobotanical surveys have shown numerous plants used traditionally to treat diabetes [30–32]. Some plants with antidiabetic effects from South Africa are available in the form of herbal supplements, possessing the NAPPi code, a unique coding identifier for medicine, surgical products and medical procedures. For example, Probetix (Nappi code: 711050-001) is an herbal supplement made from the *Sutherlandia frutescens* extract (main active component: pinitol). This supplement has been reported to cause a reversal of insulin resistance and reduce intestinal glucose uptake [33]. Manna DFM43 (Nappi code: 705846-001) developed from the pods of *Prosopis glandulosa* (main active component: galactomannan) is known to slow down glucose absorption and also reduce the glycaemic index of foods. Despite the enormous use of these plants in the treatment of life-threatening diseases such as diabetes, there are still several plants that remain to be recorded.

For many years, people from the Eastern Cape province, one of the poorest provinces in South Africa, combined with the highest provincial unemployment rate (55%), relied heavily on the use of medicinal plants to treat diabetes. This is mainly due to their cultural acceptability, accessibility, inexpensiveness, efficacy, and safety claims [34,35]. The Xhosas are the major inhabitants of the Eastern Cape province. Despite the western influence, the Xhosa people still believe in the effectiveness of medicinal plants and prefer to use these plants currently. This review aims to highlight the antidiabetic medicinal plants used in the Eastern Cape province of South Africa with the view of preventing the loss of vital traditional knowledge of plants used to treat diabetes. This review is expected to identify the existing knowledge gaps and serves as an important baseline for future research on scientifically underexploited plant species.

2. Methods

A comprehensive literature survey was thoroughly conducted from January 2021 to August 2022. A report about the antidiabetic medicinal plants used traditionally in the management of diabetes in the Eastern Cape province was thoroughly retrieved from various scientific databases such as Google Scholar, Science Direct, PubMed, Medline, Scopus and Web of Science. In addition, theses, dissertations, and ethnobotanical books were also retrieved from various university libraries. The keywords and terms used to obtain relevant articles or information were the “scientific name of the plants”, “antidiabetic”, “hypoglycaemia”, mode of action, “diabetes” and “ethnopharmacology”.

3. Results and Discussion

3.1. Plants Used in the Eastern Cape Province with Antidiabetic Potentials

Forty-eight plant species were identified as being used to treat diabetes by the people of Eastern Cape province (Table 1), although some of these plants have been scientifically examined for their antidiabetic activity, but eight of these plant species are yet to be scientifically investigated (Table 2). For the purpose of this review, a comprehensive description of the traditional usage, antidiabetic mechanism of action and the active molecules (Figure 2) of some of the plants used in the Eastern Cape province for the treatment of diabetes are as follows:

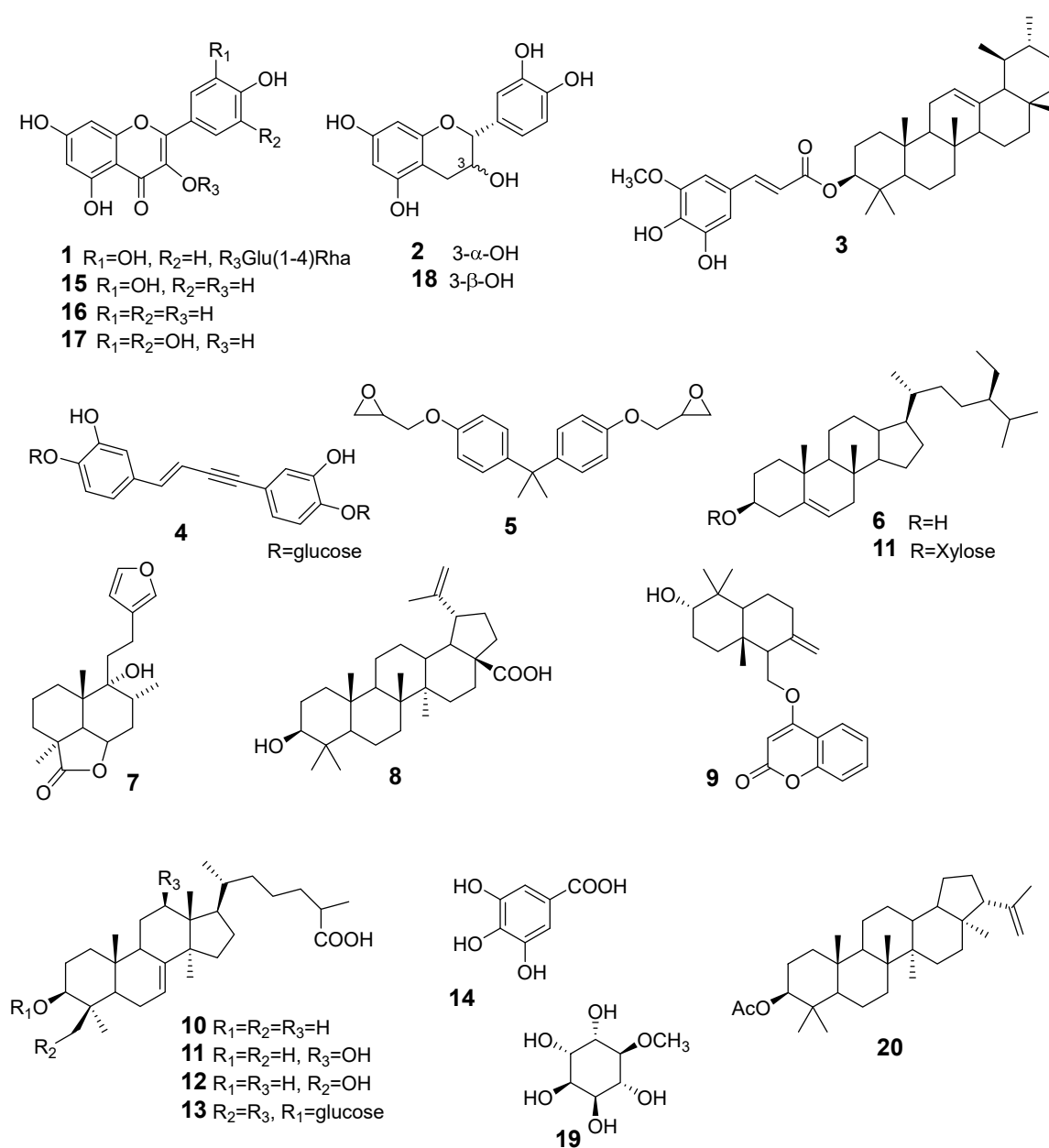


Figure 2. Antidiabetic compounds isolated from Eastern Cape antidiabetic medicinal plants. The numbers 1–20 correspond to the compounds reported in Table 2.

3.2. *Albuca setosa* Jacq.

Albuca setosa (Figure 3) (Hyacinthaceae family) is a plant that has narrow leaves that become broader at the base. It is a perennial, very hardy, evergreen plant that grows from 0.15 m to 0.5 m [36]. The plant is dispersed in most parts of South African provinces, except for the extreme South-Western Cape province. Some of these provinces include the Eastern Cape, Limpopo, Free State Northern Cape, KwaZulu-Natal, Gauteng and Mpumalanga. Traditionally, the fresh corms of the plant are crushed, boiled and taken orally to treat diabetes [37]. Additionally, the plant is used for the treatment of wounds, articulation complications, rheumatoid arthritis, digestive disorder and venereal diseases in human beings [38]. The aqueous extract of *A. setosa* corms showed high glucose utilization in cultured L6 muscle and 3T3-L1 cells [37]. The extract also exhibited weak inhibition against alpha-amylase activity but strongly inhibit alpha glucosidase with IC_{50} value of 7.725 mg/mL [37]. This antidiabetic activity of *A. setosa* has been attributed to its main bioactive compounds such as phenols and flavonoids [37].

3.3. *Artemisia afra* Jacq. ex Willd.

Artemisia afra is one of the most frequently used medicinal plants in South Africa. The plant belongs to the Asteraceae family. It grows in bushy, slightly untidy clumps, commonly with high stems up to 2 m in height, but occasionally as low as 0.6 m [39]. The plant is widespread in all South African provinces, except for the Northern Cape province. It is also found in Swaziland, Lesotho, and northwards into tropical Africa [31]. The leaves or roots of *A. afra* are used mostly as an infusion and then taken orally by the people of the Eastern Cape to treat diabetes [31]. Studies investigated by Afolayan and Sumonu [40] revealed that *A. afra* exhibited a strong ability to reverse diabetic oxidative stress in streptozotocin-induced diabetic rats. One report also indicated that *A. afra* extract showed a significant reduction in blood glucose levels with the greatest reduction seen in the 200 mg/kg concentration, which is almost the same effect as the standard positive control, glibenclamide, used in the study [41]. It has also been reported that acetone extract of *A. afra* showed weak inhibition of alpha-glucosidase [41]. In a separate study, the active antidiabetic molecules reported from *A. afra* are polyphenols, flavonoids, sterols, alkaloids and terpenoids [42]. Literature surveys revealed no reported antidiabetic isolated compound from this plant.

3.4. *Brachylaena discolor* DC.

Brachylaena discolor (Asteraceae family) is an evergreen shrub with a height of 4 to 10 m. It is found in coastal forests, bushes and on the margins of the evergreen forest in the Eastern Cape province of South Africa to Mozambique. The leaf infusion of the plant is used to treat diabetes in the Eastern Cape province [31]. In addition, the infusion is also used as a tonic to treat intestinal parasites and chest pain. A literature report revealed that the aqueous extract of *B. discolor* stimulated glucose uptake in 3T3-L1 and C2C12 muscle cells [43]. Mellem et al. [44] also reported that aqueous extract of *B. discolor* showed a strong inhibition against alpha-glucosidase. In the vivo studies, the methanol extract of *B. discolor* exhibited a marked decline in the blood glucose level of a diabetic rat at the tested concentration [45]. The literature survey showed no report on the antidiabetic activity of isolated compounds. Therefore, additional studies are required to elucidate the antidiabetic molecules present in the plant.

3.5. *Bulbine frutescens* (L.) Willd.

Bulbine frutescens is a common, waterwise garden plant that belongs to the Xanthorrhoeaceae family. It is a fast-growing plant with linear green leaves in opposite rows and grasping the stems at the base. The plant occurs extensively throughout parts of the Western Cape, Eastern Cape and Northern Cape provinces of South Africa. The root infusion of the plant is taken orally to treat diabetes by the people of the Eastern Cape [34]. An infusion of fresh leaves in a cup of boiling water is taken for coughs, arthritis and colds [46]. A report by van Huyssteen et al. [47] showed that whole plant aqueous extract of *B. frutescens* increased glucose uptake in Chang liver cells. This effect was greater than the standard positive control, metformin, used in the study. Nevertheless, there are a lack of scientific data on the antidiabetic in vivo studies of *B. frutescens* in the literature. No reports on antidiabetic isolated compounds from *B. frutescens* have yet been carried out.

3.6. *Catha edulis* (Vahl) Forrsk. ex Endl.

Catha edulis belongs to the Celastraceae family, normally known as the spike thorn family. The plant is very attractive and is found in woodlands and on rocky outcrops. It is distributed in the Eastern Cape province, mostly from the mist belt, moving inland. The fresh leaves of the plant are chewed to treat diabetes [48]. The in vitro studies revealed that *C. edulis* dichloromethane/methanol (1:1) extract stimulated glucose uptake in C2C12 and 3T3-L1 fat cells. In animal studies, the aqueous extract of the plant exhibited significant hypoglycaemic and weight reduction effects in normal streptozotocin-induced diabetic rats [48]. Piero et al. [49] also reported that the aqueous extract of *C. edulis* effectively lowered blood

glucose levels to normal in alloxan-induced diabetic rats compared to insulin used in the study. This antidiabetic effect of the plant has been ascribed to the presence of chemical components, but the elucidation or isolation of the antidiabetic molecules present in the plant is still required.

3.7. *Conyza scabrida* DC.

Conyza scabrida is a plant that consists of fresh or dried leaves and small stems. The plant belongs the Asteraceae family and is found in streamside and forest margins of the Western and Eastern Cape provinces of South Africa. The fresh leaves of the plant are taken orally as an infusion to treat diabetes [50]. The plant is also used as an external application to treat sores and inflammation [51]. The literature survey revealed no scientific investigation on its antidiabetic properties.

3.8. *Dianthus thunbergii* S.S. Hooper

Dianthus thunbergii is referred to as “wild pink”, due to the color of its flowers, belonging to the genus *Dianthus*, family Caryophyllaceae. The plant is 30 cm high, and its flowers are pale pink with bracts approximately 4 cm long with fine grey-blue leaves at the base. It is found in South Africa and occurs often in the Eastern part of the country [52]. The extract from the freshly crushed roots of the plant is reportedly used against diabetes [30]. The ethanol and aqueous root extract of *D. thunbergii* have been reported to exhibit moderate glucose uptake in L6 muscle cells in vitro [53], but the scientific efficacy of the plant in in vivo studies has not been validated. Additionally, isolated antidiabetic compounds from *D. thunbergia* are yet to be reported in the literature.

3.9. *Euclea undulata* Thunb.

Euclea undulata is a dense, erect, grassy, perennial dioecious shrub belonging to the family Ebenaceae. It is one of the most common small trees across the enormous subtropical and central interior regions of Southern Africa. Its flowers are very small and pale in ancillary racemes up to 2 cm long. The plant is widespread in Southern African countries. In South Africa, the plant occurs extensively on rocky slopes throughout all provinces [31]. Traditionally, an infusion from the ground root bark of the plant is drunk as a tea to treat diabetes by Eastern Cape people [54]. The root bark acetone extract of the plant has been described to increase glucose uptake in C2C12, Chang liver and 3T3-L1 cells exhibited a strong effect against alpha-glucosidase [7]. It has been reported that the root bark acetone extract of *E. undulata* also decreased fasting blood glucose levels, elevated cholesterol and triglyceride levels to near normal in streptozotocin–nicotinamide-induced type 2 diabetic rats without any weight gain at a dose of 100 mg/kg body weight [55]. Several antidiabetic compounds isolated from this plant have also been reported for their antidiabetic activity [56]. Epicatechin isolated from *E. undulata* was reported to reduce blood glucose levels in C2C12 cells [56]. Another compound, α -amyrin-3O- β -(5-hydroxy) ferulic acid isolated from *E. undulata* showed strong inhibition against alpha-glucosidase activity with an IC₅₀ value of 4.79 [56].

3.10. *Hypoxis colchicifolia* Bak.

Hypoxis colchicifolia (Hypoxidaceae family) is the second most vital *Hypoxis* medicinal species with marketable value in South Africa. It grows in grasslands between 25 and 60 cm in height. The plant possesses large underground tubers which enable it to subsist the constant grass fires common to this vegetation type. *H. colchicifolia* is found on sandy or poor soil in grassland across the provinces of Eastern Cape and KwaZulu-Natal [57]. In traditional medicine, the fresh corms of the plant are milled, boiled in water and then taken orally to treat diabetes. The in vitro antidiabetic studies conducted by Cumbe [58] revealed that the methanol extract of the plant exhibited moderate glucose utilization in C2C12 muscle cells and Chang liver cells, but there is no scientific studies supporting this claim in the animal model. Active antidiabetic molecules isolated from *H. colchicifolia* include

hypoxoside and bisphenol A diglycidyl ether [58]. It should be noted that bisphenol A diglycidyl ether is an environmental impurity. This antidiabetic activity of *H. colchicifolia* has been attributed to its main bioactive compounds.

3.11. *Leonotis leonurus* (L.) R.Br.

Leonotis leonurus (Lamiaceae family) is referred to as a lion's tail. It is a fast-growing, soft-woody, strong shrub that grows up to 2–3 m high and 1.5 m wide. It is very common and widespread across all South African provinces, most especially the Eastern Cape, where it grows between rocks and in grassland [59]. A decoction from the whole part of the plant is taken orally for the treatment of diabetes by the people of the Eastern Cape [30]. Very little has been done to establish the in vitro antidiabetic properties of this plant. The aqueous extract of *L. leonurus* has been reported to exhibit hypoglycaemic effects in streptozotocin-induced diabetic rats, thereby decreasing blood glucose levels as well as low-density lipoprotein [60]. Another study conducted by Odei-Addo et al. [61] revealed that the extract Nanostructured lipid carriers (NLCs) formulation improved glucose uptake in liver cells. The antidiabetic activity of this plant has been ascribed to various polyphenolics, diterpenoids, and flavonoids present in the extract [62]. Marrubin, an active antidiabetic compound, has been isolated from the organic extracts of *L. leonurus* [63].

3.12. *Momordica foetida* Schumach.

Momordica foetida (Cucurbitaceae family) is an herbaceous, climbing, perennial herb producing annual stems up to 4.5 m. The plant has dark green flecks when young and woody when old. It is mostly found in the Eastern Cape province of South Africa. Traditionally, the whole plant is apparently used to treat diabetes in the Eastern Cape [30]. In addition, the juice of crushed leaves from the plant is used to relieve intestinal disorders [30]. Van der venter et al. [43] reported the in vitro antidiabetic activity of *M. foetida* dichloromethane/methanol (1:1) extract, thereby showing an increase in glucose uptake in C2C12 muscle cells. Akinwumi [64] also reported that the effects of the methanol extract of *M. foetida* had low inhibitory activity against alpha-amylase and alpha-glucosidase activities. Not much has been done to validate the effect of the plant in animal model studies. However, foetidin, an isolated compound from *M. foetida*, has been reported to decrease blood glucose levels in fasting and alloxan-treated rats [65].

3.13. *Psidium guajava* L.

Psidium guajava (Myrtaceae family) is a well-known tropical tree rich in fruit. It is a perennial shrub-like tree that ranges in height from 6 to 25 ft [31]. The plant has a wide-ranging dispersed network of branches. The leaves of the plant are extensive and green in colour, with noticeable veins to cure wounds, ulcers and toothache [66]. *P. guajava* is widely dispersed in the provinces of KwaZulu-Natal, Eastern Cape, Mpumalanga and Limpopo [31]. In the Eastern Cape, the warm water extracted from the dried leaves of the plant is taken orally to treat diabetes [67]. The leaves of the plant are also used for other ailments such as boils, wounds, and coughs [68]. It has been reported that *P. guajava* ethanol extract demonstrates a considerable decrease in blood glucose levels in alloxan-induced diabetic rats at an oral dose of 250 mg/kg [69]. Another study reported by Shukla and Dubey [70] revealed that the aqueous and ethanolic extracts of *P. guajava* produced blood glucose homeostasis but also reversed metabolic and pathologic changes in pancreatic islets. Tella et al. [71] also reported that the treatment of diabetic animals with the *P. guajava* extract ameliorated damage to the pancreatic islets and enhanced the lowering of blood glucose. This antidiabetic activity of the plant has also been confirmed in in vitro studies. Another report by van de venter et al. [43] revealed that the leaves of dichloromethane/methanol (1:1) extract of *P. guajava* increased glucose uptake in C2C12 and 3T3-L1 cells. The acetone extract of the plant also showed good inhibition against alpha-amylase and alpha-glucosidase activities [41]. The aqueous leaf extract of *P. guajava* was also reported to exhibit excellent inhibitory activity against alpha-glucosidase with an IC₅₀

value of 5.6 mg/mL [72]. The major active antidiabetic compound isolated from *P. guajava* includes arachidic acid (1), β -sitosterolxylopyranoside (2), lanost-7-en-3 β -ol-26-oic acid (3) lanost-7-en-3 β , 12 β -diol-26-oic acid (4), lanost-7-en-3 β , 12 β , 29-triol-26-oic acid (5), lanost-cis-1,7,23-trien-3 β , 12 β , 18, 22 α -tetraol-26-oic acid (6), lanosteryl-3 β -O-D-xylopyranosyl-2'-*p*-benzaldehyde (7) and lanost-7-en-3 β -ol-26-oic acid-3 β -D-glucopyranoside (8) [66]. The isolated compounds 3,4,5 and 8 have been reported to show strong antidiabetic activity against streptozotocin-induced diabetic rats [66].

3.14. *Solanum aculeastrum* Dunal subsp. *aculeastrum*

Solanum aculeastrum (Solanaceae family) is a small tree of approximately 1–5 m high. The leaves of the plant are shortly petiolate, ovate, up to 150 × 130 mm. The plant is found naturally in grassland, woodland and in forest margins across some South African provinces, particularly Eastern Cape and Western Cape provinces. Traditionally, the decoction from the root of the plant is taken orally to treat diabetes in the Eastern Cape [30]. However, there is still no scientific report of its antidiabetic activity. The literature survey also showed no reported antidiabetic isolated compounds from *S. aculeastrum*.

3.15. *Sutherlandia frutescens* (L.) R.Br.

Sutherlandia frutescens (Fabaceae family) is a small plant around 1.2 m in height. It is a fast-growing and drought-tolerant plant that prefers full sun. The compound leaves of the plant are greyish green in colour. It is broadly dispersed in the dry areas of the Western, Eastern, Northern Cape provinces, frequently in distressed places [31]. The plant is used by indigenous communities throughout South Africa. In the Eastern Cape province, the leaf infusion of the plant is used to treat diabetes [50]. It is also used for the treatment of gastric ailments, cancer, and gynaecological complications [31]. The literature survey showed various in vitro antidiabetic studies of the plant. William et al. [73] stated that the aqueous extract of *S. frutescens* prevented insulin resistance (a precursor of type II diabetes) in Chang liver cells. Another report by Elliot, [74] also revealed that the aqueous extract of the plant stimulated insulin secretion from INS-1 cells. MacKenzie et al. [75] also showed that the plant extract enhanced glucose uptake in 3T3-L1 adipocytes. To confirm its antidiabetic activity in the animal model, Chadwink et al. [33] revealed that the *S. frutescens* extract exhibited a strong capacity to normalize insulin levels, increased glucose uptake in peripheral tissues, and also subdue intestinal glucose uptake with no significant weight gain in pre-diabetic rats. The active antidiabetic compounds isolated from *S. frutescens* include flavonoids, pinitol, saponins and triterpenoid [76]. Among the antidiabetic compounds, pinitol, a well-known antidiabetic agent, has been reported to have the same properties as insulin [77]. Therefore, the presence of pinitol explains the antidiabetic use of *S. frutescens*.

3.16. *Tarchonanthus camphoratus* L.

Tarchonanthus camphoratus is a semi-deciduous small tree with a height of 2–9 m. *T. camphoratus* belongs to the Asteraceae family. The leaves of the plant are thin, with entire finely toothed margins. *T. camphoratus* is widely spread in a range of habitats, such as thickets of bushveld, grassland and forests across Southern African countries. In South Africa, the plant is widely dispersed in the Eastern Cape, Gauteng, Free State and Northern Cape provinces. In traditional medicine, the fresh leaves of this plant are taken orally as an infusion to treat diabetes in the Eastern Cape [30]. The literature report indicated that the aqueous and ethanol extracts of *T. camphoratus* exhibited high glucose uptake in C2C12 muscle cells [47]. However, there are no scientific records of its antidiabetic activity in an animal model. Additionally, no isolated antidiabetic molecule has been reported from *T. camphoratus*.

3.17. *Vernonia oligocephala* Sch. Bip.

Vernonia oligocephala (Table 1) is an upright, perennial, herbaceous plant that belongs to the Asteraceae family. It measures up to 1 m in height and its stems grow from a

woody rootstock. It is widely spread throughout the grassland regions in several South African provinces, including the Eastern Cape [31,57]. The infusion of the leaves of this plant is taken orally to treat diabetes [34]. The infusion of the leaves is also used to cure rheumatism and dysentery [68]. The methanol extract of *V. oligocephala* has been reported to show weak inhibition against alpha amylase inhibition [78]. Additionally, oligocephalate, a compound isolated from *V. oligocephala*, was reported to exhibit strong inhibition against alpha glucosidase enzymes, with an IC₅₀ value of 18.5 µM. [79].

Table 1. List of medicinal plants used to treat diabetes in the Eastern Cape province.

S/N	Scientific Name	Local Name (Xhosa)	Family	Part Used	Method of Preparation	Reference
1	<i>Albuca setosa</i> Jacq.	Ingwe beba	Hyacinthaceae	Corms	Fresh corms are milled, boiled and taken orally.	[30]
2	<i>Allium sativum</i> L. fam.	Ikoronofile	Alliaceae	Whole plants	The fresh plant is crushed, boiled and the infusion is taken orally.	[30]
3	<i>Artemisia afra</i> Jacq. ex Willd.	Umhlonyane	Asteraceae	Leaves, roots	The leaves or roots are boiled, then the infusion is mixed with sugar to reduce the bitterness.	[34]
4	<i>Aloe ferox</i> Mill	Ikhala- lasekoloni	Aloaceae	Leaves	The liquid from the leaves is boiled to powder, submerge in water and taken orally.	[30]
5	<i>Anacampseros ustulata</i> E.Mey. ex Sond.	Igwele	Portulacaceae	Corms	Fresh corms are crushed, boiled and taken orally.	[30]
6	<i>Bridelia micrantha</i> (Hochst.) Baill.	umhlalamakwaba,	Phyllanthaceae	Stem bark.	Unspecified	[79]
7	<i>Brachylaena discolor</i> DC.	UmPhahla	Asteraceae	Leaves	Leaves of the plant are boiled, and the infusion is taken orally	[80]
8	<i>Brachylaena elliptica</i> (Thunb.) DC.	isiduti	Asteraceae	Leaves	An infusion is made from fresh leaves serves as a gargle and mouthwash	[31,81]
9	<i>Brachylaena ilicifolia</i> (Lam.) E. Phillips & Schweick.	Umgqeba.	Asteraceae	Leaves	An infusion is prepared from fresh leaves serves as a gargle	[31]
10	<i>Bulbine abyssinica</i> A.Rich	Uyakayakana	Asphodelaceae	whole plant	The whole plant parts are crushed, boiled and the infusion is taken orally.	[30]
11	<i>Bulbine frutescens</i> L. (Willd.)	Ibhucu	Xanthorrhoeaceae	Roots	The infusion is prepared from freshly boiled roots and taken orally.	[34]
12	<i>Bulbine natalensis</i> Baker	Ibhucu	Asphodelaceae	Roots	An infusion is prepared from boiled fresh roots and taken orally.	[30]
13	<i>Cannabis sativa</i> L.	Umya	Cannabaceae	Whole plants	Unspecified	[82]
14	<i>Carpobrotus edulis</i> (L.) L. Bolus	unomatyumtyum,	Mesembryanthemaceae	Leaves	The leaf juice and leaf palp are taken orally.	[31]
15	<i>Catha edulis</i> (Vahl) Forrsk. ex Endl.	iqgwaka	Celastraceae	Leaves	The leaves are chewed	[48]
16	<i>Catharanthus roseus</i> (L.) G. Don.	Isisushlungu	Apocynaceae	Leaves	The infusion is prepared from boiled leaves and is taken orally.	[34]
17	<i>Chilianthus olearaceus</i> Burch.	Umgeba	Buddlejaceae	Leaves and twigs	The fresh leaves or twigs are taken orally as an infusion.	[34]

Table 1. Cont.

S/N	Scientific Name	Local Name (Xhosa)	Family	Part Used	Method of Preparation	Reference
18	<i>Chironia baccifera</i> L.	NA	Gentianaceae	Rhizomes	The fresh or dry rhizomes are chewed.	[83]
19	<i>Cissampelos capensis</i> L.f.	Umayisake	Menispermaceae	Roots	The fresh corms are milled, boiled and taken orally.	[30]
20	<i>Conyza scabrida</i> DC.	Isavu	Asteraceae	Leaves	The fresh leaves are taken orally as an infusion.	[30]
21	<i>Dianthus thunbergii</i> Hooper	Ungcana,	Caryophyllaceae	Roots	The extract from the freshly crushed roots is taken orally.	[30]
22	<i>Euclea natalensis</i> A.DC.	umKhasa	Ebenaceae	Roots	The roots are used to make a decoction and then taken orally	[84]
23	<i>Euclea undulata</i> Thunb.	Umgwali	Ebenaceae	Roots, bark	An aqueous infusion from the ground root bark is usually drunk as tea.	[54]
24	<i>Helichrysum gymnocomum</i>	Impepho	Asteraceae	leaves	A decoction is prepared from boiled leaves and taken orally.	[30]
25	<i>Helichrysum nudifolium</i> (L.) Less.	Ichocholo	Asteraceae	Leaves and roots	Fresh leaves or roots are boiled, then taken orally.	[34]
26	<i>Helichrysum odoratissimum</i> L.	Imphepho	Asteraceae	Whole plant	An infusion from the whole plant is milled, boiled and then taken orally.	[34]
27	<i>Helichrysum petiolare</i> Hilliard & B.L.	Imphepho	Asteraceae	Whole plant	The whole plant is crushed, boiled and the concentrated solution is taken orally.	[34]
28	<i>Heteromorpha arborescens</i> (Spreng.) Cham.	Umbangandlala	Apiaceae	Leaves and roots	The infusion is prepared from boiled leaves or roots and taken orally	[34]
29	<i>Hypoxis argentea</i> Harv Ex Baker	Ixalanxa,	Hypoxidaceae	Corms	The corms are boiled in water and then taken orally	[30]
30	<i>Hypoxis colchicifolia</i> Bak.	Inongwe	Hypoxidaceae	Corms	Fresh corms are crushed, boiled in water and then taken orally.	[34]
31	<i>Hypoxis hemerocallidea</i> Fisch. and C. A	Inongwe	Hypoxidaceae	Corms	Fresh corms are crushed, boiled in water and then taken orally.	[34]
32	<i>Lauridia tetragonia</i> (L.f.) R.H. Archer	Umdlavuzza	Celastraceae	Barks	The Infusion from the powdered bark is taken orally	[30]
33	<i>Leonotis leonorus</i> (L.) R.Br.	Unfincafincane	Lamiaceae	Whole plants	A whole plant is milled, boiled, mixed with coke and half a cup of decoction taken orally.	[30]
34	<i>Momordica balsamina</i> L.	NA	Cucurbitaceae	Fruits, Stem and flowers	The Infusion from fresh young fruit and taken orally	[31,83]
35	<i>Momordica foetida</i> Schumach.	NA	Cucurbitaceae	Whole plants	Unspecified	[82]
36	<i>Ornithogalum longibracteatum</i> (Jacq)	Ingwe beba	Hyacinthaceae	Bulb	The fresh bulb soaked in water and the concoction taken orally	[37]
37	<i>Psidium guajava</i> L.	NA	Myrtaceae	Leaves	The warm water extract from the dried leaves is taken orally	[67]
38	<i>Ruta graveolens</i> L.	iyeza lomoya	Rutaceae	Leaves	Unspecified	[43]
39	<i>Sclerocarya birrea</i> (A. Rich.) Hochst. subsp. <i>caffra</i> (Sond.) Kokwaro	NA	Anacardiaceae	Leaves	The decoctions or infusions from leaves taken orally	[85]

Table 1. Cont.

S/N	Scientific Name	Local Name (Xhosa)	Family	Part Used	Method of Preparation	Reference
40	<i>Solanum aculeastrum</i> Dunal subsp. <i>aculeastrum</i>	umthuma, itunga	Solanaceae	Roots	The decoction is prepared from fresh crushed roots and then taken orally	[30]
41	<i>Strychnos henningsii</i> Gilg	Umnonono	Loganiaceae	Barks	The barks are crushed to powder and half of a cup of the decoction is taken orally	[30]
42	<i>Sutherlandia frutescens</i> L.	umnwele	Fabaceae	leaves	An infusion of the leaves is taken orally	[50]
43	<i>Tarchonanthus camphoratus</i> L.	Umgqemba	Asteraceae	leaves	The fresh leaves are taken orally as an infusion	[30]
44	<i>Tulbaghia alliacea</i> L.	Umwelala	Alliaceae	Roots	The fresh roots are crushed, boiled and three quarters of a cup of decoction is taken orally	[47]
45	<i>Tulbaghia violacea</i> . Harv.	utswelane	Alliaceae	leaves	Unspecified	[47]
46	<i>Vernonia amygdalina</i> DeL.	Umlunguhlungu	Asteraceae	Leaves	Powdered from the fresh leaves are soaked in water and the solution is taken orally.	[34]
47	<i>Vernonia oligocephala</i> Sch. Bip.	Umlunguhlungu	Asteraceae	leaves, roots or twigs	The infusion prepared from fresh leaves, roots or twigs is taken orally.	[34]
48	<i>Vinca major</i> L.	Iflawa	Apocynaceae	Leaves, roots, stem	Unspecified	[43]

Table 2. Plant used traditionally in the Eastern Cape with reported antidiabetic activity.

S/N	Scientific Name	Plant Part Used	Solvent/Extract Used	Antidiabetic Isolated Compounds	Antidiabetic Mechanism of Action	References
1	<i>A. setosa</i>	Bulb	Aqueous and acetone	*	High glucose utilization in cell lines and weak inhibition against carbohydrate digesting enzymes	[37]
2	<i>A. sativum</i>	Corms	Ethanol	*	Inhibition against alpha-amylase	[86]
3	<i>A. afra</i>	Leaves	Acetone	*	Weak Inhibition of carbohydrate digesting enzymes	[41]
4	<i>A. ferox</i>	Leaves	Aqueous	*	Stimulate adiponectin secretion from 3T3-L1 cells	[42]
5	<i>A. ustulata</i>	*	*	*	*	*
6	<i>B. micrantha</i>	Bark, stem, roots	Methanol	Quercetin-3-O-β-d-glucoside (1→4)-α-L-rhamnoside (1)	The methanol extract exhibited high inhibition against glucosidase and moderate inhibition against alpha-amylase. The isolated compound exhibited (Quercetin-3-o-β-d-glucopyranoside (1→4)-α-L-rhamnoside) strong inhibition against alpha glucosidase	[87,88]
7	<i>B. discolor</i>	Leaves, roots and stems	Aqueous	*	High glucose utilisation in Chang liver, C2C12 muscle and 3T3-L1 fat cells	[43]
8	<i>B. elliptica</i>	Leaves	Aqueous	*	Moderate glucose utilization in HepG2 liver cells	[89]

Table 2. Cont.

S/N	Scientific Name	Plant Part Used	Solvent/Extract Used	Antidiabetic Isolated Compounds	Antidiabetic Mechanism of Action	References
9	<i>B. ilicifolia</i>	Leaves	Aqueous	*	Moderate glucose utilisation in HepG2 liver cells	[90]
10	<i>B. abyssinica</i>	Leaves, flowers, stems and roots	Aqueous	*	Strong Inhibition against alpha amylase	[91]
11	<i>B. frutescens</i>	Whole plant	Aqueous	*	High glucose utilisation in Chang liver and C2C12 muscle cells	[47]
12	<i>B. natalensis</i>	*	*	*	*	*
13	<i>C. sativa</i>	Leaves, roots	Aqueous	*	Stimulate glucose uptake in 3T3-L1 fat cells and strong inhibition of alpha-amylase	[43,92]
14	<i>C. edulis</i>	Leaves	Aqueous	*	Strong inhibition against alpha glucosidase.	[93]
15	<i>C. edulis</i>	Leaves	Dichloromethane/methanol (1:1)	*	Moderate glucose utilisation in C2C12 muscle and 3T3-L1 fat cells	[43]
16	<i>C. roseus</i>	Leaves and twig	Dichloromethane/methanol (1:1)	*	Moderate glucose utilisation in 3T3-L1 cells	[43]
17	<i>C. olearaceus</i>	*	*	*	*	*
18	<i>C. baccifera</i>	Whole plant	Aqueous	*	Moderate glucose utilisation in Chang liver, C2C12 and 3T3-L1 cells	[43]
19	<i>C. capensis</i>	Leaves	Aqueous	*	Increase glucose utilisation in 3T3-L1 cells	[43]
20	<i>C. scabrida</i>	*	*	*	*	*
21	<i>D. thunbergii</i>	Roots	Ethanol and Aqueous	*	Moderate glucose utilisation in L6 muscle cells	[53]
22	<i>E. natalensis</i>	Root-bark	Acetone	*	Strong inhibition against alpha amylase	[41]
23	<i>E. undulata</i>	Stem-bark and root-bark	Acetone	Epicatechin (2), α -amyrin-3O- β -(5-hydroxy) ferulic acid (3)	High glucose utilisation in Chang liver, C2C12 and 3T3-L1 cells. The isolated compound (α -amyrin-3O- β -(5-hydroxy) ferulic acid) exhibited strong inhibition against carbohydrate digesting enzymes	[7,56]
24	<i>H. gymnocomum</i>	*	*	*	*	*
25	<i>H. nudifolium</i>	*	*	*	*	*
26	<i>H. odoratissimum</i>	leaf and stem	Aqueous	*	Hypoglycaemic effect	[94]
27	<i>H. petiolare</i>	Whole plant	Aqueous	*	High glucose utilisation in L6 cells Strong inhibition against carbohydrate digesting enzymes	[95]
28	<i>H. arborescens</i>	Leaf	Aqueous and Ethanol	*	Moderate glucose utilization in C3A and L6 cells and strong inhibition against alpha amylase activity	[96]
29	<i>H. argentae</i>	Corms	Aqueous	*	Moderate glucose utilisation in HepG2 liver and L6 muscle cells, stimulate INS-1 cells proliferation and prevent fat accumulation in 3T3-L1 adipocytes.	[53]
30	<i>H. colchicifolia</i>	Corms	Methanol	Hypoxoside (4), bisphenol A diglycidyl ether (5)	Moderate glucose utilisation in C2C12 muscle cells and Chang liver cells	[58]

Table 2. Cont.

S/N	Scientific Name	Plant Part Used	Solvent/Extract Used	Antidiabetic Isolated Compounds	Antidiabetic Mechanism of Action	References
31	<i>H. hemerocallidea</i>	Leaves and bark	Acetone and Hexane	β -Sitosterol (6)	The extract exhibited moderate Inhibition against alpha-amylase and strong inhibition against alpha glucosidase. The isolated compound (β -sitosterol) stimulates insulin release	[97,98]
32	<i>L. tetragonia</i>	*	*	*	*	*
33	<i>L. leonorus</i>	Leaves	Aqueous	Marrubin (7)	Hypoglycaemic effect	[30,63]
34	<i>M. balsamina</i>	Stems and flowers	Dichloromethane/methanol (1:1)	Betulinic acid (8)	High glucose utilisation in 3T3-L1 cells, Chang liver and C2C12. The isolated compound (betulinic acid) exhibited antihyperglycemic effect	[43,99]
35	<i>M. foetida</i>	Whole plant	Dichloromethane/methanol (1:1)	Foetidin (9)	Moderate glucose utilisation in C2C12 muscle cells	[43]
36	<i>O. longibracteatum</i>	Bulbs	Aqueous and ethanol	*	High glucose utilisation in Chang liver and C2C12 muscle cells	[47]
37	<i>P. guajava</i>	Leaves and roots	Dichloromethane/methanol (1:1)	lanost-7-en-3 β -ol-26-oic acid (10) lanost-7-en-3 β , 12 β -diol-26-oic acid (11), lanost-7-en-3 β , 12 β , 29-triol-26-oic acid (12), lanost-7-en-3 β -ol-26-oic acid-3 β -glucoside (13), gallic acid (14), quercetin (15), kaempferol (16), myricetin (17), catechin (18)	Moderate glucose uptake in C2C12 and 3T3-L1 cells. Exhibited strong inhibition against alpha amylase	[41,43]
38	<i>R. graveolens</i>	Arial parts	Aqueous and ethanol	*	High glucose utilisation in Chang liver cells	[47]
39	<i>S. birrea</i>	Bark	Aqueous and methanol	*	High glucose utilisation in C2C12, 3T3-L1 and HepG2 cells. Strong Inhibition of carbohydrate digestive enzymes	[7]
40	<i>S. aculeastrum</i>	*	*	*	*	*
41	<i>S. henningsii</i>	Stem bark	Aqueous	*	High glucose uptake in differentiated 3T3-L1 cells and moderate inhibition against alpha glucosidase and weak inhibition against alpha amylase	[100]
42	<i>S. frutescens</i>	Leaves and shoots	Aqueous	Pinitol (19)	Prevention of insulin resistance in Chang liver cells, stimulate secretion of insulin from INS-1 cells and high glucose uptake in C2C12 muscle and 3T3-L1 fat cells	[73–75]
43	<i>T. camphoratus</i>	Leaves and twig	Aqueous and ethanol	*	High glucose uptake in C2C12 cells	[47]
44	<i>T. alliaacea</i>	*	*	*	*	*
45	<i>T. violacea</i>	Whole plant	Aqueous and ethanol	*	High glucose utilisation in Chang liver and C2C12 cells	[47]
46	<i>V. amygdalina</i>	Leaves	Aqueous, methanol, acetone	*	High glucose uptake in C2C12 and Chang liver cells and Strong inhibition of carbohydrate digesting enzyme	[7,34]

Table 2. Cont.

S/N	Scientific Name	Plant Part Used	Solvent/Extract Used	Antidiabetic Isolated Compounds	Antidiabetic Mechanism of Action	References
47	<i>Vernonia oligocephala</i> Sch. Bip.	Leaves	Methanol	Oligocephalate (20)	The extract exhibited weak inhibition against alpha amylase. The isolated compound showed strong inhibition against alpha glucosidase	[78,79]
48	<i>V. major</i>	Leaves, roots and stems	Dichloromethane/methanol (1:1)	*	High glucose uptake in Chang liver, C2C12 and 3T3-L1 cells	[43]

*: No reported antidiabetic activities available in the literature.

*A. afra.**A. Setosa**B. discolor**A. ferox**B. ilicifolia**B. micrantha*

Figure 3. Cont.



B. frutescens



B. elliptica



C. sativa



B. abyssinica



C. edulis



B. natalensis



C. olearaceus



C. edulis

Figure 3. *Cont.*



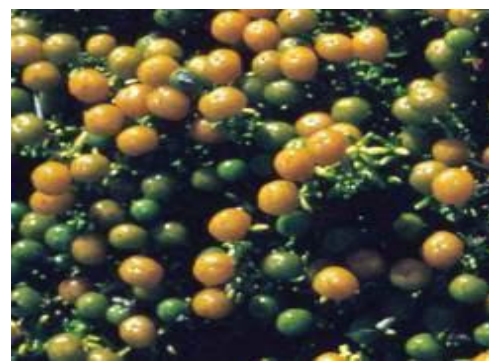
C. capensis



C. roseus



D. thunbergia



C. baccifera



E. undulata



C. scabrida



H. nudifolium



E. natalensis

Figure 3. *Cont.*



H. petiolare



H. gymnocomum



H. argentea



H. odoratissimum



H. hemerocallidea



H. arborescens



L. leonorus



H. colchicifolia

Figure 3. *Cont.*



L. tetragonia



M. balsamina



P. guajava



O. longibracteatum



S. birrea



R. graveolens



S. frutescens



S. aculeastrum

Figure 3. *Cont.*



Figure 3. Selected medicinal plants used by the people of Eastern Cape for the treatment of diabetes [101–143].

4. Conclusions and Future Direction

The present trend in the management of diabetes involves the use of medicinal plants, since oral antidiabetic drugs are known to be expensive with unwanted side effects. This high prevalence justifies the special attention toward the use of these plants for the management of diabetes. In this review, it is evident that the majority of these medicinal plants exert their antidiabetic mechanism of action through the enhancement of glucose uptake in cells (hepatic, skeletal and fat cells), stimulation of the release of insulin by pancreatic beta-cells or through the alteration of certain hepatic enzymes involved in glucose metabolism and reducing intestinal glucose absorption. Out of the 48 medicinal plants used in the Eastern Cape for the treatment of diabetes, only eight plants have not been scientifically studied. Furthermore, it is imperative to note that a large number of active antidiabetic molecules of these plants are yet to be isolated and clinically studied. Thus, an effort needs to be devoted

to the isolation and purification of these antidiabetic molecules and the determination of their mechanism of action, both in in vitro and in vivo experimental animal models.

Author Contributions: Conceptualization: I.J.S. and A.A.H.; Writing of the original draft: I.J.S.; Editing: I.J.S. and A.A.H.; Supervision: A.A.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: Not applicable.

Acknowledgments: The authors are thankful to the Cape Peninsula University of Technology, for publication support.

Conflicts of Interest: The authors state that there is no conflict of interest in publishing this article.

References

- Genuth, S.; Alberti, K.G.; Bennet, P.; Buse, J.; DeFronzo, R.; Kahn, R.R.; Kitzmiller, J.; Knowler, W.C.; Lebovitz, L.; Lerman, A.; et al. Follow-up report on the diagnosis of diabetes mellitus. *Diabetes Care* **2003**, *26*, 3160–3167. [PubMed]
- American Diabetes Association (ADA). Diagnosis and classification of diabetes mellitus. *Diabetes Care* **2009**, *32*, s62–s67. [CrossRef] [PubMed]
- Ozougwu, J.C.; Obimba, K.C.; Belonwu, C.D.; Unakalamba, C. The pathogenesis and pathophysiology of type 1 and type 2 diabetes mellitus. *J. Physiol. Pathophysiol.* **2013**, *4*, 46–57. [CrossRef]
- Cnop, M.; Welsh, N.; Jonas, J.; Jorns, A.; Lenzen, S.; Eizirik, D. Mechanisms of pancreatic β cell death in type 1 and type 2 diabetes. *Diabetes* **2005**, *54* (Suppl. 2), s97–s107. [CrossRef]
- International Diabetes Federation (IDF). *Diabetes Atlas*, 9th ed.; International Diabetes Federation: Brussels, Belgium, 2019; Available online: www.idf.org/diabetesatlas (accessed on 28 February 2022).
- World Health Organization (WHO). *Global Report on Diabetes*; WHO: Geneva, Switzerland, 2016; Available online: https://www.who.int/gho/publications/world_health_statistics/2016/EN_WHS2016_TOC.pdf (accessed on 24 February 2021).
- Mousinho, N.M.H.D.C.; van Tonder, J.J.; Steenkamp, V. In vitro anti-diabetic activity of *Sclerocarya birrea* and *Ziziphus mucronata*. *Nat. Prod. Commun.* **2013**, *8*, 1279–1284. [PubMed]
- Kokil, G.R.; Rewatkar, P.V.; Verma, A.; Thareja, S.; Naik, S.R. Pharmacology and chemistry of diabetes mellitus and antidiabetic drugs: A critical review chemistry. *Curr. Med. Chem.* **2010**, *17*, 4405–4423. [CrossRef]
- Pleuvry, B.J. Pharmacological control of blood sugar. *Anaesth Intensive Care Med.* **2005**, *6*, 344–346. [CrossRef]
- Dornhost, A. Insulinotropic meglitinide analogues. *Lancet* **2001**, *358*, 1709–1716. [CrossRef]
- Nolte, M.S.; Karam, J. Pancreatic hormones and anti-diabetic drugs. In *Basic and Clinical Pharmacology*, 8th ed.; Katzung, B.G., Ed.; Lange Medical Books; Mc Graw-Hill: San Francisco, CA, USA, 2001; pp. 711–734.
- Iheanyi, E.I. The Antidiabetic Activities of the Methanolic Leaf Extract of *Hymenocardia acida* (Tul). Ph.D. Thesis, University of Nigeria, Nsukka, Nigeria, 2010.
- Maideen, N.M.P.; Manavalan, G.; Balasubramanian, K. Drug interactions of meglitinide antidiabetics involving CYP enzymes and OATP1B1 transporter. *Ther. Adv. Endocrinol. Metab.* **2018**, *9*, 259–268. [CrossRef]
- Shaw, R.J.; Lamia, K.A.; Vasquez, D.; Koo, S.H.; Bardeesy, N.; Depinho, R.A.; Montminy, M.; Cantley, L.C. The kinase LKB1 mediates glucose homeostasis in liver and therapeutic effects of metformin. *Science* **2005**, *310*, 1642–1646. [CrossRef]
- Hawley, S.A.; Gadalla, A.E.; Olsen, G.S.; Hardie, D. The antidiabetic drug metformin activates the AMP-activated protein kinase cascade via an adenine nucleotide-independent mechanism. *Diabetes* **2002**, *51*, 2420–2425. [CrossRef] [PubMed]
- Hanefeld, M. The role of acarbose in the treatment of non-insulin-dependent diabetes mellitus. *J. Diabetes Complicat.* **1998**, *12*, 228–237. [CrossRef]
- Stafylas, P.C.; Sarafidis, P.A.; Lasaridis, A. The controversial effects of thiazolidinediones on cardiovascular morbidity and mortality. *Int. J. Cardiol.* **2008**, *131*, 298–304. [CrossRef] [PubMed]
- Semple, R.K.; Krishna, K. PPAR α and human metabolic disease. *J. Clin. Investig.* **2006**, *116*, 581–586. [CrossRef]
- Chiaasson, J. Acarbose for the prevention of diabetes, hypertension and cardiovascular disease in subjects with impaired glucose tolerance: The study to prevent non-insulin-dependent diabetes mellitus (STOP-NIDDM) trial. *Endocr. Pract.* **2006**, *12*, 25–30. [CrossRef] [PubMed]
- Ahren, B. DPP-4 inhibitors. *Best Pract. Res. Clin. Endocrinol. Metab.* **2007**, *21*, 517–533. [CrossRef]
- Hasani-Ranjbar, S.; Larijani, B.; Abdollahi, M. Systematic review of the potential herbal sources of future drugs effective in oxidant-related diseases. *Inflamm. Allergy Drug Targets* **2009**, *2*, 2–10. [CrossRef]
- Salehi, B.; Ata, A.; Kumar, N.V.A.; Sharopov, F.; Ram, K.; Ruiz-ortega, A.; Ayatollahi, S.A.; Fokou, P.V.T.; Kobarfard, F.; Zakaria, Z.A.; et al. Antidiabetic potential of medicinal plants and their active components. *Biomolecules* **2019**, *9*, 551. [CrossRef]
- Mishra, A.P.; Sharifi-Rad, M.; Shariati, M.A.; Mabkhot, Y.N.; Al-Showiman, S.S.; Rauf, A.; Salehi, B.; Župunski, M.; Sharifi-Rad, M.; Gusain, P.; et al. Bioactive compounds and health benefits of edible *Rumex* species—A review. *Cell Mol. Biol.* **2018**, *64*, 27–34. [CrossRef]

24. Mishra, A.P.; Saklani, S.; Salehi, B.; Parcha, V.; Sharifi-Rad, M.; Milella, L.; Iriti, M.; Sharifi-Rad, J.; Srivastava, M. *Satyrium nepalense*, a high altitude medicinal orchid of Indian Himalayan region: Chemical profile and biological activities of tuber extract. *Cell Mol. Biol.* **2018**, *64*, 35–43. [CrossRef]
25. Abdolshahi, A.; Naybandi-Atashi, S.; Heydari-Majd, M.; Salehi, B.; Kobarfard, F.; Ayatollahi, S.A.; Athar, A.; Tabanelli, G.; Sharifi-Rad, M.; Montanari, C.; et al. Antibacterial activity of some lamiaceae species against *Staphylococcus aureus* in yoghurt-based drink (Doogh). *Cell Mol. Biol.* **2018**, *64*, 71–77. [CrossRef] [PubMed]
26. Bailey, C.J. Metformin: Historical overview. *Diabetologia* **2017**, *60*, 1566–1576. [CrossRef] [PubMed]
27. Germano, M.P.; D'Angelo, V.D.; Biasini, T.; Sanogo, R.; De Pasquale, R.; Catania, S. Evaluation of the antioxidant properties and bioavailability of free and bound phenolic acids from *Trichilia emetica* Vahl. *J. Ethnopharmacol.* **2006**, *105*, 368–373. [CrossRef] [PubMed]
28. Chauhan, A.; Sharma, P.K.; Srivastava, P.; Kumar, N.; Dudhe, R. Plants having potential antidiabetic activity: A review. *Der Pharmacia Lettre* **2010**, *2*, 369–387.
29. Mamum-or-Rashid, A.N.M.; Hossain, M.S.; Hassan, N.; Dash, B.K.; Sapon, M.A.; Sen, M. A review on medicinal plants with anti-diabetic activity. *J. Pharmacogn. Phytochem.* **2014**, *3*, 149–159.
30. Oyedemi, S.O.; Bradley, G.; Afolayan, A. Ethnobotanical survey of medicinal plants used for the management of diabetes mellitus in the Nkonkobe municipality of South Africa. *J. Med. Plants Res.* **2009**, *3*, 1040–1044.
31. Deutschlandler, M.S.; Lall, N.; Van de venter, M. Plant species used in the treatment of diabetes by South Africa traditional healer: An Inventory. *Pharm Biol.* **2009**, *47*, 348–365. [CrossRef]
32. Balogun, F.O.; Tshabalala, N.T.; Ashafa, A.O.T. Antidiabetic medicinal plants used by the Basotho Tribe of Eastern Free State: A Review. *J. Diabetes Res.* **2016**, *2016*, 4602820. [CrossRef]
33. Chadwick, W.A.; Roux, S.; Van de Venter, M.; Low, J.; Oelofsen, W. Anti-diabetic effects of *Sutherlandia frutescens* in Wistar rats fed on a diabetogenic diet. *J. Ethnopharmacol.* **2007**, *109*, 121–127. [CrossRef]
34. Erasto, P. Phytochemical Investigation of *Vernonia amygdalina*: A medicinal Plant Used for the Treatment of Diabetes. Ph.D. Thesis, University of Fort Hare, Alice, South Africa, 2006.
35. Sahadew, N.; Podiatry, B.; Singaram, V.S. A diabetes profile of the eight districts in the public health sector Eastern Cape Province, South Africa. *S. Afr. Med. J.* **2019**, *109*, 957–962. [CrossRef]
36. Musara, C.; Silas, M.; Mudyiwa, S.M.; Maroyi, A. Pharmacological potential, botany, biological and chemical properties of *Albuca setosa* (Asparagaceae) endemic to Southern Africa. *J. Pharma. Nutr. Sci.* **2019**, *9*, 195–199. [CrossRef]
37. Odeyemi, S. A Comparative Study of the In Vitro Antidiabetic Properties, Mechanism of Action and Cytotoxicity of *Albuca setosa* and *Albuca bracteata* bulb Extracts. Ph.D. Thesis, University of Fort Hare, Alice, South Africa, 2016.
38. Umaphathy, E.; Ndebia, E.J.; Meeme, A.; Adam, B.; Nkeh-chungag, B.; Iputo, J.E. An experimental evaluation of *Albuca setosa* aqueous extract on membrane stabilization protein denaturation and white blood cell migration during acute inflammation. *J. Med. Plants Res.* **2010**, *4*, 789–795.
39. van der Walt, L. Kirstenbosch National Botanical Garden. 2004. Available online: <http://www.plantzafrica.com/plantab/artemisafra.html> (accessed on 26 December 2020).
40. Afolayan, A.J.; Sunmuno, O.T. *Artemisia afra* Jacq. Ameliorates oxidative stress in the pancreas of streptozotocin-Induced diabetic wistar rats. *Biosci. Biotechnol. Biochem.* **2011**, *75*, 2083–2086. [CrossRef] [PubMed]
41. Nkobile, N.K. Antidiabetic Activity of Pentacyclic Triterpenes and Flavonoids Isolated from Stem Bark of *Terminalia sericea* Burch Ex DC. Master's Thesis, University of Pretoria, Pretoria, South Africa, 2009.
42. Patil, G.V.; Dass, S.K.; Chandra, R. *Artemisia afra* and modern diseases. *J. Pharmacogenom. Pharm.* **2011**, *2*, 1000105. [CrossRef]
43. van de Venter, M.; Roux, S.; Bungu, L.C.; Louw, J.; Crouch, N.R.; Grace, O.M.; Maharaj, V.; Pillay, P.; Sewnarian, P.; Bhagwandin, N.; et al. Antidiabetic screening and scoring of 11 plants traditionally used in South Africa. *J. Ethnopharmacol.* **2008**, *119*, 81–86. [CrossRef]
44. Mellem, J. Antidiabetic Potential of *Brachylaena discolor*. *Afr. J. Tradit. Complement. Altern. Med.* **2015**, *12*, 38–44. [CrossRef]
45. Mellem, J.J. Isolation and Characterization of the Leaves of *Brachylaena discolor* Extract as an Anti-Diabetic Agent. Ph.D. Thesis, Durban University of Technology, Durban, South Africa, 2013.
46. Harris, S. *Bulbine frutescens*. Free State National Botanical Garden. 2003. Available online: <http://pza.sanbi.org/bulbine-frutescens> (accessed on 27 December 2020).
47. van Huyssteen, M.; Milne, P.J.; Campbell, E.E.; van der Venter, M. Antidiabetic and cytotoxicity screening of 5 medicinal plants used by traditional African Health practitioners in the Nelson Mandela Metropole South Africa. *Afr. J. Tradit. Complement. Altern. Med.* **2011**, *8*, 150–158.
48. Debecho, D.A.; Ababa, A. Effect of fresh juice of Khat (*Catha edulis*) on blood glucose levels of normoglycemic and streptozotocin-induced diabetic rats. *Int. J. Pharm. Sci. Res.* **2018**, *9*, 784–789.
49. Piero, N.M.; Joan, M.N.; Cromwell, K.M.; Joseph, N.J.; Wilson, N.M.; Daniel, M.; Peter, G.K.; Eliud, N. Hypoglycemic activity of some kenyan plants traditionally used to manage diabetes mellitus in Eastern province. *J. Diabetes Metab.* **2011**, *2*, 2–7. [CrossRef]
50. Thring, T.S.A.; Weitz, F.M. Medicinal plant use in the Bredasdorp/Elim region of the Southern Overberg in the Western Cape Province of South Africa. *J. Ethnopharmacol.* **2006**, *103*, 261–275. [CrossRef]
51. Scott, G.; Springfield, E.P.; Coldrey, N. A pharmacognostical study of 26 South African plant species used as traditional medicines. *Pharm Biol.* **2004**, *42*, 186–213. [CrossRef]

52. Pooley, E. *A Field Guide to Wild Flowers KwaZulu-Natal and the Eastern Region*; Natal Flora Publications Trust, Natal Herbarium: Durban, South Africa, 1998.
53. Akinrinde, A. Antidiabetic and Toxicological Properties of *Dianthus thunbergii* (Caryophyllaceae) Roots and *Hypoxis argentea* (Hypoxidaceae) Corms. Ph.D. Thesis, University of Fort Hare, Alice, South Africa, 2017.
54. Maroyi, A. *Euclea undulata* Thunb.: Review of its botany, ethnomedicinal uses, phytochemistry and biological activities. *Asian J. Pac. Trop. Biomed.* **2017**, *10*, 1030–1036. [[CrossRef](#)] [[PubMed](#)]
55. Deutschländer, M.S.; Lall, N.; Van de Venter, M.; Dewanjee, S. The hypoglycemic activity of *Euclea undulata* Thunb var myrtina (Ebenaceae) root bark evaluated in a streptozotocin—nicotinamide induced type 2 diabetes rat model. *S. Afr. J. Bot.* **2012**, *80*, 9–12. [[CrossRef](#)]
56. Deutschländer, M.S.; Lall, N.; van de Venter, M.; Hussein, A. Hypoglycemic evaluation of a new triterpene and other compounds isolated from *Euclea undulata* Thunb. var. myrtina (Ebenaceae) root bark. *J. Ethnopharmacol.* **2011**, *133*, 1091–1095. [[CrossRef](#)] [[PubMed](#)]
57. Van Wyk, B.-E.; Van Oudtshoorn, B.; Gericke, N. *Medicinal Plants of South Africa*; Briza Publications: Pretoria, South Africa, 2005.
58. Cumbe, J. Antidiabetic Compounds from *Hypoxis colchicifolia* and *Terminalia sericea*. Master's Thesis, University of KwaZulu-Natal, Durban, South Africa, 2015.
59. Afolayan, A.J.; Sunmonu, T.O. In vivo studies on antidiabetic plants used in South African herbal medicine. *J. Clin. Biochem. Nutr.* **2010**, *47*, 98–106. [[CrossRef](#)]
60. Oyedemi, S.O.; Yakubu, M.T.; Afolayan, A.J. Antidiabetic activities of aqueous leaves extract of *Leonotis leonurus* in streptozotocin induced diabetic rats. *J. Med. Plants Res.* **2011**, *5*, 119–125.
61. Odei-Addo, F.; Shegokar, R.; Müller, R.H.; Levendal, R.-A.; Frost, C. Nanoformulation of *Leonotis leonurus* to improve its bioavailability as a potential antidiabetic drug. *Biotechnology* **2017**, *7*, 344. [[CrossRef](#)]
62. Ojewole, J.A. Antinociceptive, antiinflammatory and antidiabetic effects of *Leonotis leonurus* (L.) R. BR. [Lamiaceae] leaf aqueous extract in mice and rats. *Methods Find Exp. Clin. Pharmacol.* **2005**, *27*, 257–264. [[CrossRef](#)]
63. Mnonopi, N.; Ruby-Ann, R.; Davies-Coleman, M.T.; Frost, C.L. The cardioprotective effects of marrubiin, a diterpenoid found in *Leonotis leonurus* extracts. *J. Ethnopharmacol.* **2011**, *138*, 67–75. [[CrossRef](#)]
64. Akinwunmi, O. In vitro antioxidant, α -amylase and α -glucosidase activities of methanol extracts from three *Momordica* species. *Int. J. Phytomed.* **2019**, *11*, 8–14. [[CrossRef](#)]
65. Marquis, V.O.; Adanlawo, T.A.; Olaniyi, A.A. The effect of foetidin from *Momordica foetida* on blood glucose level of albino rats. *Planta Med.* **1977**, *31*, 367–374. [[CrossRef](#)] [[PubMed](#)]
66. Bagri, P.; Ali, M.; Aeri, V.; Bhowmik, M. Isolation and antidiabetic activity of new lanostenoids from the leaves of *Psidium guajava* L. *Int. J. Pharm. Pharm. Sci.* **2016**, *8*, 14–18. [[CrossRef](#)]
67. Mushtaq, A.; Khan, M.A.; Arshad, M.; Zafar, M. Ethnophytotherapical Approaches for the Treatment of Diabetes by the Local Inhabitants of District Attock (Pakistan). Available online: www.siu.edu/~|ebl/leaflets/phyto.htm (accessed on 23 December 2020).
68. Hutchings, A.; Scott, A.H.; Lewis, G.; Cunningham, A. *Zulu Medicinal Plants*; University Natal Press: Pietermaritzburg, South Africa, 1996.
69. Mukhtar, H.M.; Ansari, S.H.; Naved, T.; Bhat, Z.A. Hypoglycemic activity of *Psidium guajava* Linn. leaf extract. *J. Nat. Remed.* **2004**, *4*, 186–189.
70. Shukla, K.; Dubey, P.K. Antidiabetic activity of *Psidium guajava* (Guava) leaves extract. *Res. J. Sci. Tech.* **2009**, *1*, 13–15.
71. Tella, T.; Masola, B.; Mukaratirwa, S. Anti-diabetic potential of *Psidium guajava* leaf in streptozotocin induced diabetic rat. *Phytomed. Plus.* **2022**, *2*, 100254. [[CrossRef](#)]
72. Simamora, A.; Paramita, L.; Azreen, N.; Santoso, A.W.; Timotius, K.H. In vitro antidiabetic and antioxidant activities of aqueous extract from the leaf and fruit of *Psidium guajava* L. *Indones. Biomed. J.* **2018**, *10*, 154–156. [[CrossRef](#)]
73. Williams, S.; Roux, S.; Koekemoer, T.; van de Venter, M.; Dealtry, G. *Sutherlandia frutescens* prevents changes in diabetes-related gene expression in a fructose-induced insulin resistant cell model. *J. Ethnopharmacol.* **2013**, *146*, 482–489. [[CrossRef](#)] [[PubMed](#)]
74. Elliott, G.P. Implementation of Novel Flow Cytometric Methods to Assess the In Vitro Antidiabetic Mechanism of a *Sutherlandia frutescens* Extract. Ph.D. Thesis, Nelson Mandela Metropolitan University, Port Elizabeth, South Africa, 2011.
75. MacKenzie, J.; Koekemoer, T.C.; Roux, S.; Van de Venter, M.; Dealtry, G.B. Effect of *Sutherlandia frutescens* on the lipid metabolism in an insulin resistant rat model and 3T3-L1 adipocytes. *Phytother. Res.* **2012**, *26*, 1830–1837. [[CrossRef](#)]
76. Van Wyk, B.E. A broad review of commercially important south Africa medicinal plants. *J. Ethnopharmacol.* **2008**, *119*, 342–355. [[CrossRef](#)]
77. Jawla, S.; Kumar, Y.; Khan, M. Isolation of antidiabetic principle from *Bougainvillea spectabilis* Willd (Nyctaginaceae) stem bark. *Trop. J. Pharm. Res.* **2013**, *12*, 761–765. [[CrossRef](#)]
78. Nkala, B.A. The Cytotoxic Effects, Anti-Inflammatory, Antioxidant, Antibacterial, and Antidiabetic Properties of Eight Selected South African Plants for Medicinal Purposes. Ph.D. Thesis, University of KwaZulu-Natal, Durban, South Africa, 2009.
79. Riaz, N.; Feroze, K.; Saleem, M.; Musaddiq, S.; Ashraf, M.; Alam, U.; Mustafa, R.; Jabeen, B.; Irshad Ahmad, I.; Jabbar, A. Oligocephlate, a new α -glucosidase inhibitory neohopane triterpene from *Vernonia oligocephala*. *J. Chem. Soc. Pak.* **2013**, *35*, 972–975.

80. Naidoo, A. Pretoria National Botanical Garden. Available online: <http://pza.sanbi.org/bridelia-micrantha> (accessed on 27 December 2020).
81. Coates Palgrave, K. *Trees of Southern Africa*; Struik: Cape Town, South Africa, 1984; pp. 1–959.
82. Odeyemi, S. Medicinal plants used for the traditional management of diabetes in the Eastern Cape, South Africa: Pharmacology and toxicology. *Molecules* **2018**, *23*, 2759. [CrossRef] [PubMed]
83. Van Wyk, B.E.; Gericke, N. *People's Plants, A Guide to Useful Plants of Southern Africa*; Briza: Pretoria, South Africa, 2000; pp. 218–219.
84. Maroyi, A. Review of ethnomedicinal uses, phytochemistry and pharmacological properties of *Euclea natalensis* A.DC. *Molecules* **2017**, *22*, 2128. [CrossRef] [PubMed]
85. Iwu, M.M. *Handbook of African Medicinal Plants*; CRC Press: Boca Raton, FL, USA, 1993; pp. 1–435.
86. Nickavar, B.; Yousefian, N. Inhibitory effects of six *Allium* species on α -amylase enzyme activity. *Iran J. Pharm. Res.* **2009**, *8*, 53–57.
87. Maluleke, K.A. Isolation and Characterization of Antidiabetic Constituents of *Bridelia Micrantha*. Master's Thesis, University of Venda, Thohoyandou, South Africa, 2009.
88. Ezekiel, A.; Zubai, S.; Abiodun, O.; Iqbal, J. GC-MS analysis and antidiabetic potentials of *Bridelia micrantha* crude extracts. *Integr. Mol. Med.* **2018**, *5*, 1–5. [CrossRef]
89. Sagbo, I.J.; Van de venter, M.; Koekemoer, T.; Bradley, G. In vitro antidiabetic activity and mechanism of action of *Brachylaena elliptica* (Thunb) DC. *Evid. Based Complement. Alternat. Med.* **2018**, *2018*, 4170372. [CrossRef]
90. Sagbo, I.J. Antidiabetic Activity and Mechanism of Action of Extracts of *Brachylaena elliptica* (Thurb.) DC. and *Brachylaena ilicifolia* (Lam) Phill & Schweick. Ph.D. Thesis, University of Fort Hare, Alice, South Africa, 2017.
91. Kibiti, C. Evaluation of the Medicinal Potentials of *Bulbine abyssinica* A. Rich in the Management of Diabetes Mellitus in the Eastern Cape, South Africa. Ph.D. Thesis, University of Fort Hare, Alice, South Africa, 2015.
92. Shah, S.B.; Sartaj, L.; Hussain, S.; Ullah, N.; Idrees, M.; Shaheen, A.; Javed, M.S.; Aslam, M.K. In-vitro evaluation of antimicrobial, antioxidant, alpha-amylase inhibition and cytotoxicity properties of *Cannabis sativa*. *Adv. Tradit. Med.* **2019**, *20*, 181–187. [CrossRef]
93. Mulaudzia, R.O.; Aremu, A.O.; Rengasamy, K.R.R.; Adebayo, S.A.; McGaw, L.J.; Amoo, S.A.; Van Staden, J.; Du Plooy, C.P. Antidiabetic, anti-inflammatory, anticholinesterase and cytotoxicity determination of two *Carpobrotus* species. *S. Afr. J. Bot.* **2019**, *125*, 142–148. [CrossRef]
94. Njagi, J.M.; Ngugi, P.M.; Kibiti, C.M.; Ngeranwa, J.; Njue, W.; Gathumbi, P.; Njagi, E. Hypoglycemic effect of *Helichrysum odoratissimum* in alloxan induced diabetic mice. *J. Phytopharm.* **2015**, *4*, 30–33. [CrossRef]
95. Aladejana, A.E.; Bradley, G.; Afolayan, A.J. In vitro evaluation of the anti-diabetic potential of *Helichrysum petiolare* Hilliard & B.L. Burt using HepG2 (C3A) and L6 cell lines. *F1000Research* **2021**, *9*, 1240.
96. Abifar, T.O.; Otunola, G.A.; Afolayan, A.J. Cytotoxicity, anti-obesity and anti-diabetic activities of *Heteromorpha arborescens* (Spreng.) Cham Leaves. *Processes* **2021**, *9*, 1671. [CrossRef]
97. Mkolo, N.M. Antidiabetic Activity of Extracts and a Bioactive Compound Isolated from *Hypoxis Hemerocallidea* (Hypoxidaceae) in a Murine Model of Spontaneous Diabetes. Ph.D. Thesis, University of Pretoria, Pretoria, South Africa, 2019.
98. Boaduo, N.K.K.; Katerere, D.; Eloff, J.N.; Naidoo, V. Evaluation of six plant species used traditionally in the treatment and control of diabetes mellitus in South Africa using in vitro methods. *Pharm Biol.* **2014**, *52*, 756–761. [CrossRef] [PubMed]
99. Kabir, N.; Umarb, I.A.; Damac, H.A.; James, D.B.; Inuwa, H.M. Isolation and structural elucidation of novel antidiabetic compounds from Leaves of *Momordica balsamina* Linn and *Leptadenia hastata* (Pers) Decne. *Iran J. Pharm. Res.* **2021**, *20*, 390–402. [PubMed]
100. Oyedemi, S.; Koekemoer, T.; Bradley, G.; Van de Venter, M.; Afolayan, A.J. In vitro anti-hyperglycemia properties of the aqueous stem bark extract from *Strychnos henningsii* (Gilg). *Int. J. Diab. Dev. Ctries* **2013**, *33*, 120. [CrossRef]
101. SANBI (South African National Biodiversity Institute). Available online: <http://redlist.sanbi.org/species.php?species=3795-71> (accessed on 15 August 2022).
102. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/artemisia-afra> (accessed on 16 August 2022).
103. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/aloe-ferox> (accessed on 16 August 2022).
104. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/bridelia-micrantha> (accessed on 16 August 2022).
105. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/brachylaena-discolor> (accessed on 16 August 2022).
106. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/brachylaena-elliptica> (accessed on 14 August 2022).
107. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/bulbine-abyssinica> (accessed on 14 August 2022).
108. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/bulbinefrutescens#:~:text=Bulbine%20frutescens%20is%20often%20used,flowering%20at%20the%20same%20time> (accessed on 13 August 2022).
109. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/bulbine-latifolia> (accessed on 15 August 2022).

110. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/cannabis-sativa> (accessed on 16 August 2022).
111. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/carpobrotusedulis#:~:text=Carpobrotus%20edulis%20is%20an%20easy,the%20herb%20or%20kitchen%20garden> (accessed on 15 August 2022).
112. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/catha-edulis> (accessed on 15 August 2022).
113. Dladla, T.P. Effects of Catharanthus Roseus and *Centella asiatica* Leaf Extracts on Enzymes of Glutamine Catabolism in Human Colon Carcinoma (Caco-2) Cell Line and in Enterocytes from Male Sprague-Dawley Rats. Master's Thesis, University of KwaZulu-Natal, Durban, South Africa, 2015.
114. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/buddleja-saligna> (accessed on 16 August 2022).
115. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/chironia-baccifera> (accessed on 16 August 2022).
116. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/cissampelos-capensis> (accessed on 16 August 2022).
117. SANBI (South African National Biodiversity Institute). Available online: <http://redlist.sanbi.org/species.php?species=3058-45> (accessed on 16 August 2022).
118. SANBI (South African National Biodiversity Institute). Available online: <http://redlist.sanbi.org/species.php?species=3958-4002> (accessed on 16 August 2022).
119. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/euclea-natalensis#:~:text=Euclea%20natalensis%20is%20a%20shrub,grey%2C%20thin%20and%20finely%20cracked> (accessed on 16 August 2022).
120. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/euclea-undulata#:~:text=Euclea%20undulata%20is%20one%20of,different%20climatic%20and%20habitat%20conditions> (accessed on 16 August 2022).
121. FOSTER (The Friends of the St Francis Nature Areas). Available online: <https://foster.org.za/plant-gallery-2/> (accessed on 28 February 2022).
122. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/Helichrysum-nudifolium> (accessed on 28 February 2022).
123. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/Helichrysum-odoratissimum> (accessed on 28 February 2022).
124. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/Helichrysum-petiolare> (accessed on 28 February 2022).
125. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/heteromorpha-arborescens> (accessed on 28 February 2022).
126. Wildflower nursery. Available online: <https://wildflownursery.co.za/indigenous-plant-database/hypoxis-argentea/> (accessed on 28 February 2022).
127. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/hypoxis-colchicifolia> (accessed on 28 February 2022).
128. SANBI (South African National Biodiversity Institute). Available online: <http://redlist.sanbi.org/species.php?species=1603-21> (accessed on 28 February 2022).
129. Inaturalist. Available online: <https://inaturalist.mma.gob.cl/taxa/588764-Lauridia-tetragona> (accessed on 16 August 2022).
130. SANBI (South African National Biodiversity Institute). Available online: <http://redlist.sanbi.org/species.php?species=1702-15> (accessed on 16 August 2022).
131. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/momordica-balsamica> (accessed on 16 August 2022).
132. CJM Grower. Available online: <https://cjmgrowers.co.za/ornithogalum-longibracteatum/> (accessed on 16 August 2022).
133. CABI. Available online: <https://www.cabi.org/isc/datasheet/45141> (accessed on 16 August 2022).
134. Kew Royal Botanical. Available online: <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:775099-1> (accessed on 16 August 2022).
135. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/sclerocarya-birrea> (accessed on 16 August 2022).
136. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/solanum-aculeastrum> (accessed on 16 August 2022).
137. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/lessertia-frutescens> (accessed on 16 August 2022).
138. Wildflower Nursery. Available online: <https://wildflownursery.co.za/indigenous-plant-database/tarchonanthus-camphoratus/> (accessed on 16 August 2022).
139. kumbula Nursery. Available online: <https://kumbulanursery.co.za/plants/tulbaghia-alliacea> (accessed on 16 August 2022).
140. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/tulbaghia-violacea> (accessed on 16 August 2022).

-
141. Wikipedia. Available online: https://en.wikipedia.org/wiki/Vernonia_amygdalina (accessed on 16 August 2022).
 142. SANBI (South African National Biodiversity Institute). Available online: <http://pza.sanbi.org/hilliardiella-oligocephala-0> (accessed on 16 August 2022).
 143. Biodiversity Explorer. Available online: <http://www.biodiversityexplorer.info/plants/apocynaceae/vinca.htm> (accessed on 16 August 2022).