

Article

Transannular Selenocyclofunctionalization of 1,5-cyclooctadiene: The Antioxidant Properties of 9-selenabicyclo[3.3.1]nonane Derivatives and the Discovery of Increasing Both GPx and GR Activities

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Abstract: Oxidative stress is the cause of various pathologies and disorders of cellular functions. Substances that reduce the pathological effect of oxidative stress on homeostasis include organoselenium compounds of natural and synthetic origin. Depending on the structure, organoselenium compounds can exhibit different biological activities, for example, reducing oxidative stress, participating in the regulation of signaling systems, influencing the synthesis of cytokines, etc. This makes them promising products for the development of new means of metabolic correction and drugs with enzyme-like activity. This study is aimed at developing an effective method for the synthesis of functional organoselenium compounds and studying their antioxidant effect in vivo under stress conditions. A one-pot catalyst-free method of transannular addition-functionalization of cis,cis-1,5-cyclooctadiene with selenium dihalides in the presence of nucleophiles was developed. For the first time, the antioxidant activity of functionalized 9-selenabicyclo[3.3.1]nonanes was studied in vivo. Quantitative characteristics of the effect on the level of lipid peroxidation and the activity of glutathione peroxidase and glutathione reductase under stress conditions were obtained. The effect of substituents in the selenium-containing scaffold on the biological activity of the compounds was studied. The water-soluble 9-selenabicyclo[3.3.1]nonane derivatives, containing hydroxyl and 2-hydroxyethoxy groups, which increased the activity of both glutathione peroxidase and glutathione reductase, were discovered.

Keywords: 1,5-cyclooctadiene; 9-selenabicyclo[3.3.1]nonane; selenium halides; cyclofunctionalization; glutathione peroxidase; glutathione reductase; oxidative stress



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1. Introduction

In modern medicine, attention is increasingly focused on post-therapeutic reactions of the organism, such as allergies, intoxication, drug hepatitis, etc. Among the biochemical processes associated with these reactions is oxidative stress, which is developed due to the increased cell-generated reactive oxygen species (ROS). Peroxide derivatives of unsaturated fatty acids in cell membranes are involved in the biosynthesis of prostaglandins and leukotrienes (mediators of inflammation and allergic reactions). As ROS is accumulated in the macroorganism, many mechanisms mobilize, including enzymes that utilize peroxide compounds. With insufficient capacity for compensatory resources, activation of lipid peroxidation (LPO) can destabilize cell membranes with subsequent complications [1]. Intoxications of various nature activate LPO against the background of inhibition of cell antioxidant protection as well as the accumulation of immunosuppressive factors

in the blood (acylhydroperoxides, diene conjugates of fatty acids, MDA, abnormal lipid metabolites). Disturbances in metabolic status and decreased energy supply to cells elicit immunodeficiency states [2]. In this case, it is advisable to use metabolic correction agents that increase antioxidant potential.

Selenium is recognized as an important micronutrient for humans [3–6]. This element is an important part of glutathione peroxidase, a selenium-containing enzyme whose biological role is to protect the body from oxidative damage. A number of organoselenium compounds exhibit high glutathione peroxidase-like activity [3–10] including ebselen, a well-known selenium heterocycle, which is undergoing evaluation as a therapeutic agent in clinical trials [11,12].

Natural and synthetic organoselenium compounds can reduce the pathological effect of oxidative stress on homeostasis [13,14] and prove to be antioxidants [15–17]. Meanwhile, it becomes clear that an uncontrolled and unreasonable increase in the antioxidant profile of the organism has a negative effect on the immune system and a number of other body functions. Note that enhancement of the catalytic activity of organoselenium compounds is also not a priority, since it leads to lower selectivity and higher toxicity of the studied compounds. This, together with a decrease in biostability, negatively affects the possibilities of using organoselenium compounds as bioactive substances for drug design. For the time being, a standing challenge for medicine remains the development of mild drugs of metabolic correction for use in vaccination processes, including as hepatoprotectors or general protectors of immunogenesis organs, antiviral therapy, and diseases associated with immunodeficiency states.

Important electrophilic reagents, selenium dichloride and dibromide, were recently introduced in organic synthesis [18–21]. The use of these reagents opened up new possibilities for the synthesis of new classes of organoselenium compounds. Novel approaches, including transannular selenocyclization, annulation/selenofunctionalization, and selenobisfunctionalization, were developed [19–27].

Selenofunctionalization reactions have found useful applications in modern organic synthesis, allowing the simultaneous introduction of the selenium atom and a functional group into the double bonds [28]. Seleniranium cations are postulated as intermediates in these reactions. Recently, we developed one-pot bis-alkoxyselenenylation of alkenes with selenium dibromide and alcohols [29]. This approach allows to realize one-pot selenobisfunctionalization reactions under mild conditions and to increase yields of the target products, avoiding the stage of the formation of the intermediate bis(β -halogenorganyl) selenides [29].

Recently, 2,6-dichloro-9-selenabicyclo[3.3.1]nonane and its sulfur analogue [30] were involved in nucleophilic substitution reactions to evaluate a relative anchimeric assistance effect [31]. The anchimeric assistance effect of the selenium atom was found to be considerably higher than that of the sulfur atom [31].

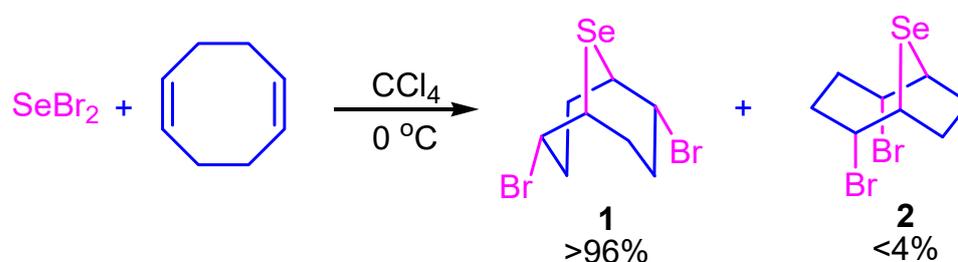
We developed a method for the synthesis of 2,6-dichloro- and 2,6-dibromo-9-selenabicyclo[3.3.1]nonanes based on selenium dihalides and *cis,cis*-1,5-cyclooctadiene (COD) [31,32]. A number of 2,6-bifunctional derivatives were obtained based on 2,6-dibromo-9-selenabicyclo[3.3.1]nonane [33–36]. It was found that the 2,6-dipyridinium salt of 9-selenabicyclo[3.3.1]nonane with high glutathione peroxidase-like activity is a promising drug for metabolic correction during vaccination [35]. The introduction of this compound into the bodies of experimental animals significantly decreases the development of pathological reactions under the influence of the tularemia and brucellosis vaccines [35]. This compound potentiates the proliferative activity of immune cells and shows a high anti-inflammatory effect [36]. It also enhances the immune response against *Yersinia pestis*, thereby increasing resistance to the plague pathogen.

2. Results and Discussion

2.1. Chemistry

This study is aimed at developing an effective method for the synthesis of functional organoselenium compounds and studying their antioxidant effect *in vivo* under stress conditions.

Previously, 2,6-dibromo-9-selenabicyclo[3.3.1]nonane (**1**) was obtained by the reaction of selenium dibromide with COD in carbon tetrachloride (Scheme 1) [32].

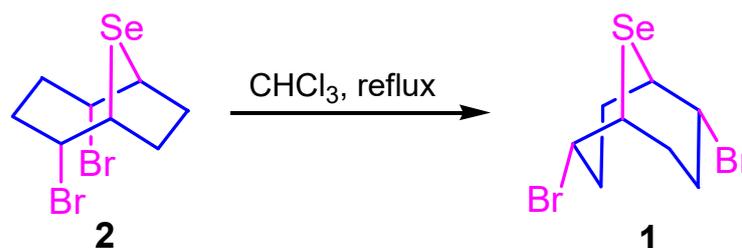


Scheme 1. Reaction selenium dibromide with cis,cis-1,5-cyclooctadiene.

However, the target product contained some amounts of the isomeric compound 2,5-dibromo-9-selenabicyclo[4.2.1]nonane (**2**) as an impurity.

In this work, we propose a new one-pot catalyst-free method of transannular addition-functionalization of COD with selenium dibromide in the presence of nucleophiles. This approach makes it possible to avoid the stage of isolation of the intermediate dibromo adduct **1** with the use of highly toxic and carcinogenic CCl₄ and to increase the yields of target 9-selenabicyclo[3.3.1]nonanes. The proposed reaction conditions make it possible to obtain only 2,6-bifunctional 9-selenabicyclo[3.3.1]nonanes without traces of isomeric 9-selenabicyclo[4.2.1]nonanes and to simplify the preparation of high-purity products.

It was found that carrying out the reaction in more polar solvents (chloroform, methylene chloride, acetonitrile, etc.) decreased the content of compound **2**. Also, the latter can be isomerized into a thermodynamically more stable compound **1** by refluxing the reaction mixture in chloroform (Scheme 2). It was established that the isomerization of compound **2** did not lead to the formation of other products except for nonane **1**.



Scheme 2. Isomerization of 2,5-dibromo-9-selenabicyclo[4.2.1]nonane.

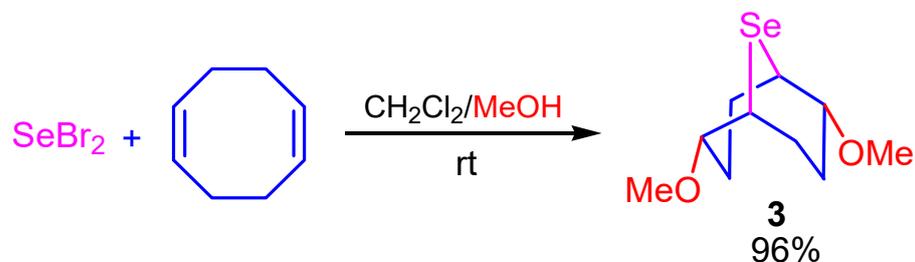
It was shown that the isomeric composition of the substitution products did not depend on the isomeric ratio of compounds **1** and **2**, but was determined by the conditions of the substitution reaction.

For example, methoxylation of compound **1** in a mixture of carbon tetrachloride and methanol (20:1) at 0 °C delivered a mixture of isomeric 2,6-dimethoxy-9-selenabicyclo[3.3.1]nonane (**3**) and 2,5-dimethoxy-9-selenabicyclo[4.2.1]nonane (**4**) in a 93:7 ratio, whereas this reaction in methylene chloride-methanol (5:1) proceeded at room temperature, affording exclusively compound **3**. These trends indicate that the nucleophilic substitution reactions proceed via seleniranium intermediates.

We have recently demonstrated the first examples of selenomethoxylation and annulation-selenomethoxylation reactions of selenium dihalides with alkenes [22,29]. The

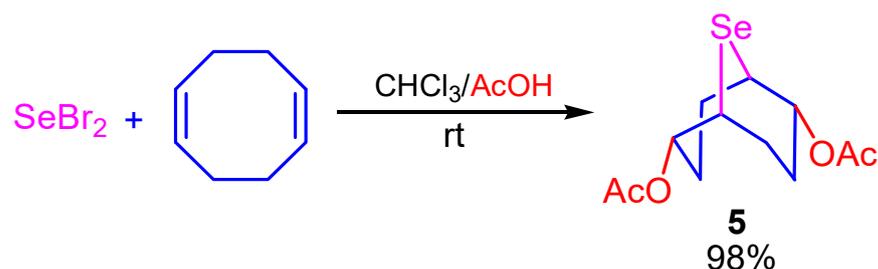
experimental conditions were found to allow the use of selenium dihalides in the methoxysele-
lenation reaction in the presence of alcohols (methanol, ethanol, isopropanol, mercaptoethanol,
etc.) and not involve alcoholysis of the selenium-halogen bond in selenium dihalides.

A one-pot transannular selenomethoxylation of COD was developed. The target
compound **3** was synthesized in high yield (96%) by a one-pot atom-economic reaction of
selenium dibromide with COD in the presence of methanol under mild conditions at room
temperature (Scheme 3).



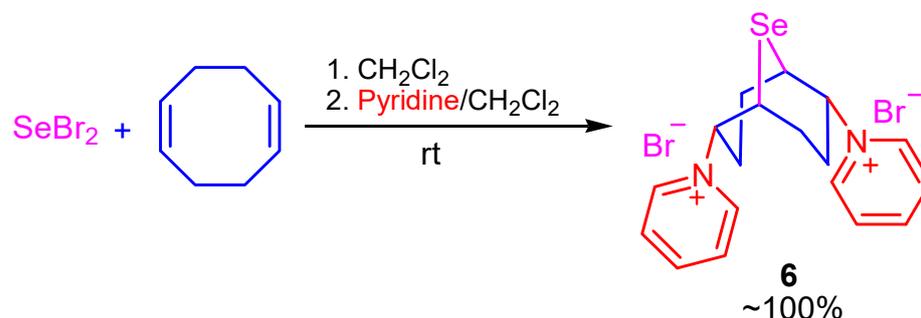
Scheme 3. Selenomethoxylation of COD by SeBr_2 .

The experimental conditions for the acetoxysele-
lenation reaction of selenium dihalides
with COD were found. The target 2,6-diacetoxy-9-selenabicyclo[3.3.1]nonane (**3**) was
synthesized in almost quantitative yield (98%) by a one-pot atom-economic reaction of
selenium dibromide with COD in a chloroform-acetic acid (5:1) mixture (Scheme 4).



Scheme 4. Selenoacetoxylation of COD by SeBr_2 .

Using the reaction of selenium dibromide with COD in chloroform or methylene
chloride in the presence of acetic acid, we showed the possibility of an acetoxysele-
lenation reaction (Scheme 5). The reaction proceeded regio- and stereoselectively and led to the
formation of 2,6-diacetoxy-9-selenabicyclo[3.3.1]nonane (**5**).



Scheme 5. One-pot synthesis of 2,6-dipyridinium 9-selenabicyclo[3.3.1]nonane dibromide (**6**).

The structure of compound **5** was proven by single-crystal X-ray diffraction analysis
(Figure 1). These data indicate that the bis-functionalization reaction proceeds as an anti-
addition to the double bond, giving the product a trans-disposition of the selenium atom
and two functional (acetoxy) groups.

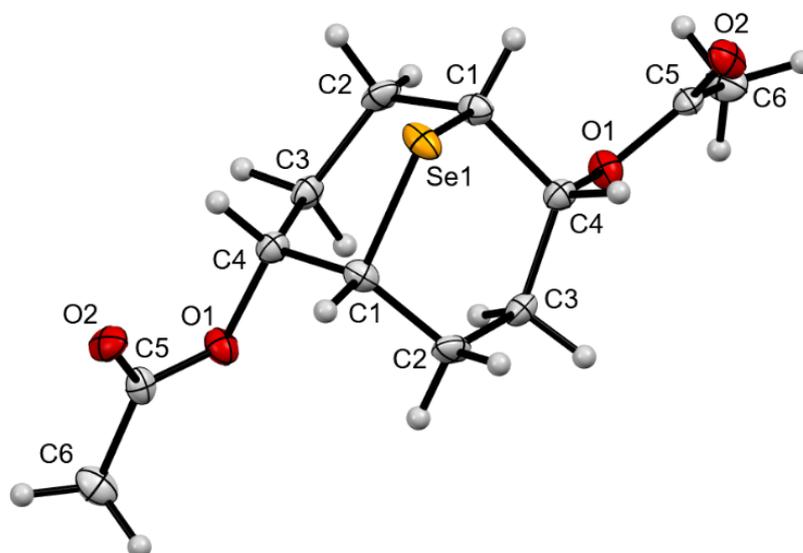
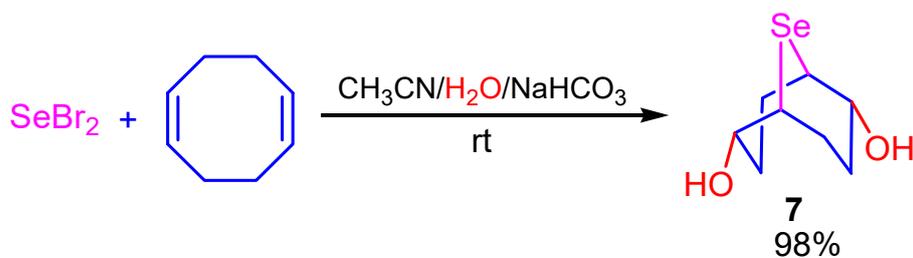


Figure 1. ORTEP molecular structure of compound **5** at 50% thermal ellipsoid probability.

The one-pot, two-stage synthesis of 2,6-dipyridinium 9-selenabicyclo[3.3.1]nonane dibromide (**6**) in quantitative yield was developed based on the addition of selenium dibromide to COD followed by nucleophilic substitution of intermediate compound **1** with pyridine (two equivalents) in methylene chloride (Scheme 5).

When the reaction of selenium dibromide with COD was carried out in the presence of pyridine, mainly halogenated products of double bonds were formed.

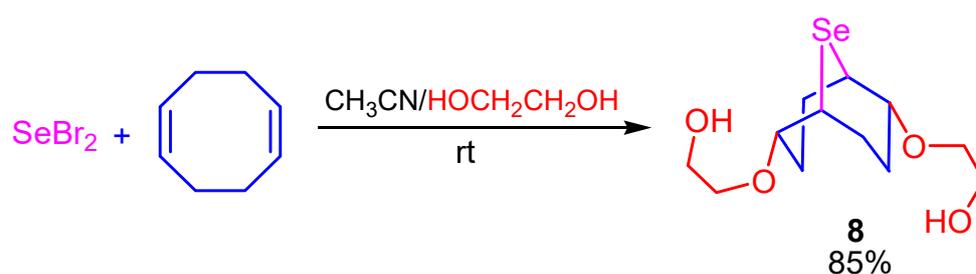
In this case, the reaction was realized as a one-pot, two-stage synthesis, with the addition of selenium dihalide to COD in methylene chloride followed by the addition of two equivalents of pyridine to a solution of intermediate compound **1** (Scheme 6). The reaction proceeds in a chemo- and regioselective fashion to produce 2,6-dipyridinium 9-selenabicyclo[3.3.1]nonane dibromide (**6**) in quantitative yield.



Scheme 6. Hydroxyselenation of COD by SeBr_2 .

We found that the hydroxyselenation reaction of selenium dichloride and selenium dibromide with COD can be realized in the system acetonitrile/water/ NaHCO_3 (Na_2CO_3 , K_2CO_3) at room temperature, affording 2,6-dihydroxy-9-selenabicyclo[3.3.1]nonane (**7**) in almost quantitative yield (98%). The reaction occurred with high chemo- and regioselectivity, and the formation of 2,5-dihydroxy-9-selenabicyclo[4.2.1]nonane as a possible by-product was not observed under these conditions (Scheme 6).

The reaction of selenium dibromide with COD and the excess ethylene glycol in acetonitrile afforded 2,6-bis(2-hydroxyethoxy)-9-selenabicyclo[3.3.1]nonane (**8**) in high yield (Scheme 7).



Scheme 7. Synthesis of 2,6-bis(2-hydroxyethoxy)-9-selenabicyclo[3.3.1]nonane (**8**).

When ethylene glycol was used as a solvent for the reaction of selenium dihalides with COD, the formation of a mixture of oligomeric products was observed.

2.2. Biological Evaluation

In the present work, the effect of functionalized 2,6-disubstituted selenabicyclo nonanes on the main characteristics of the antioxidant profile—the level of lipid peroxidation (LPO), glutathione peroxidase (GPx), and glutathione reductase (GR) activities—was examined. The tested compounds were divided into two groups: hydrophobic compounds **1**, **3**, and **5**, and water-soluble derivatives **6–8** (Table 1).

Table 1. First group—hydrophobic compounds **1**, **3**, and **5** with Ph_2Se_2 as reference. Second group—hydrophilic water-soluble compounds **6**, **7**, and **8** with Na_2SeO_3 as reference.

Compounds	Reference	Compounds	Reference
 1		 6	
 3		 7	Na_2SeO_3
 5		 8	

Increasing salinity was chosen as a method for causing stress in plant cells. The salinity was created using NaCl at a concentration of 200 mM [37]. In fact, such a concentration generates stress since it almost doubles the level of lipid peroxidation as compared with the control. Selenium is known to play a major role in the antioxidant protection of plants [38]. In this line, we commenced our study with the determination of the diene conjugate content.

In the series of water-soluble derivatives **7** and **8**, a significant decrease in lipid peroxidation was observed under stress conditions at both low and high concentrations of compounds (Figure 2). In contrast, derivative **6** at a 1000 μM concentration showed an increase in lipid peroxidation, thus indicating toxicity. However, the increase in lipid peroxidation by compound **6** was about twice lower compared with the reference compound sodium selenite (Figure 1). Taking into account the low lipid peroxidation levels observed

for compounds 7 and 8, the toxicity of compound 6 may be attributed to the blocking of ion channels, which is characteristic for pyridinium and ammonium salts.

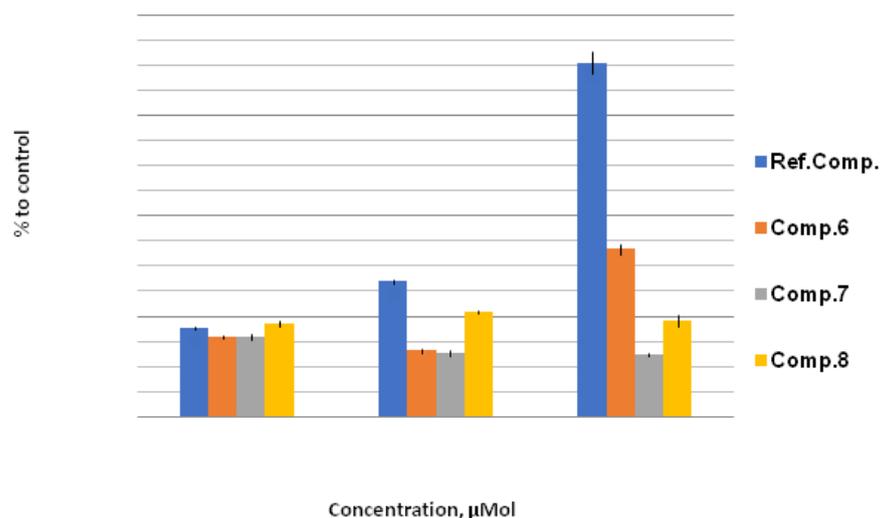


Figure 2. LPO level, % to control. The effect of compounds 6, 7, and 8 on LPO level under stress conditions relative to control ($\text{H}_2\text{O} + 200 \text{ mmol NaCl}$; taken as 100%) at concentrations of 10 μM , 100 μM , and 1000 μM . Quantitative values are given in Electronic Supplementary Information.

Previously, a toxic effect of sulfur analogs of compounds 6 and 7 (2,6-dipyridyl- and 2,6-dihydroxy substituted 9-thiabicyclo[3.3.1]nonanes) was noted when fragmentable oligo- and polycations derived from 2,6-dichloro-9-thiabicyclo[3.3.1]nonane were studied [39].

For lipophilic derivatives 1, 3, and 5, the level of lipid peroxidation at a 1000 μM concentration was significantly decreased with respect to the reference substance (Ph_2Se_2). Compound 1, despite the highly active bromine atom in the β -position relative to the selenium atom, did not exhibit high toxicity (Figure 3).

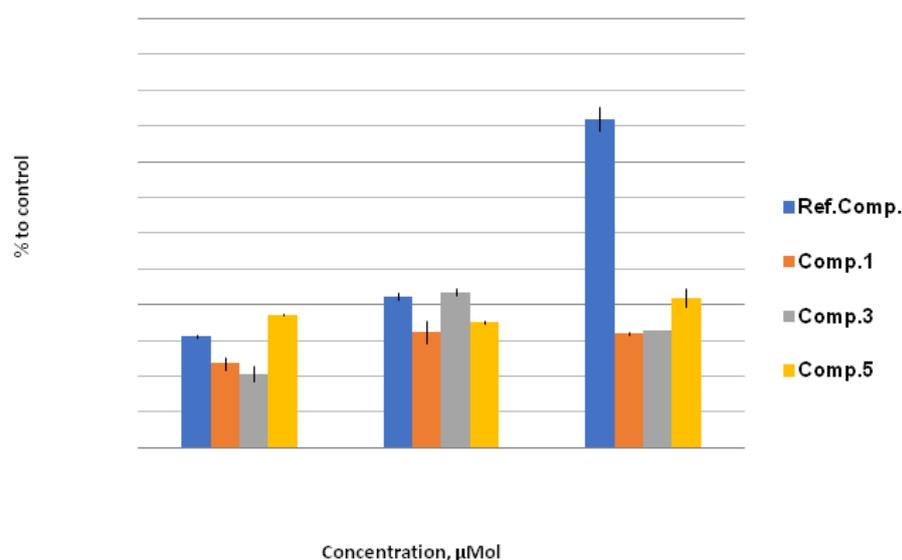


Figure 3. LPO level, % to control. The effect of compounds 1, 3, and 5 on LPO level under stress conditions relative to control ($\text{H}_2\text{O}/1\% \text{ EtOH} + 200 \text{ mmol NaCl}$; taken as 100%) at concentrations of 10 μM , 100 μM , and 1000 μM . Quantitative values are given in Electronic Supplementary Information.

Glutathione peroxidase is a key enzyme in protecting cells from oxidative damage. Glutathione peroxidase activity is a direct characteristic of the antioxidant system's capac-

ity. We have studied the effect of various concentrations of 9-selenabicyclo[3.3.1]nonane derivatives on glutathione peroxidase activity under stress conditions.

In the series of water-soluble derivatives 6–8, a multidirectional tendency was found (Figure 4). The hydroxyethoxy compound 8 at a 10 μM concentration showed a significant increase in GPx activity, with a tendency to decrease, with the elevation of concentration.

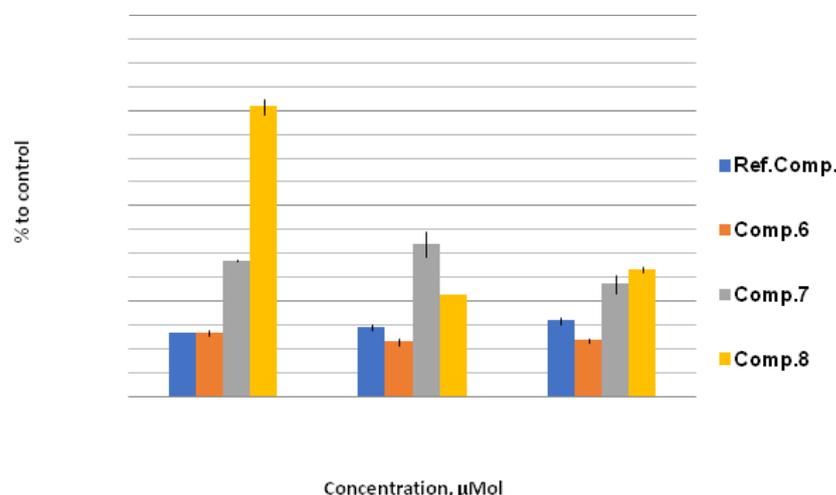


Figure 4. GPx activity, % to control. The effect of compounds 6, 7, and 8 on GPx activity under stress conditions relative to control ($\text{H}_2\text{O} + 200 \text{ mmol NaCl}$; taken as 100%) at concentrations of 10 μM , 100 μM , and 1000 μM . Quantitative values are given in Electronic Supplementary Information.

The pyridine derivative 6 demonstrated a result close to the inorganic source of selenium, sodium selenite, which is characterized by reduced GPx activity in all the concentrations studied, thus contrasting with the previously demonstrated decrease in the lipid peroxidation level (Figure 4).

In the case of lipophilic derivatives 1, 3, and 5, a noticeable effect on the activity of GPx was not observed (Figure 5). A gradual lowering of GPx activity with the growth of the concentration of the acetoxy compound 5 may indicate a weak inhibitory effect. The increase in GPx activity in the case of bromine derivative 1 may be attributed to the accumulation of the metabolites, including hydrolysis products such as hydroxy derivative 7, which enhanced the activity of GPx.

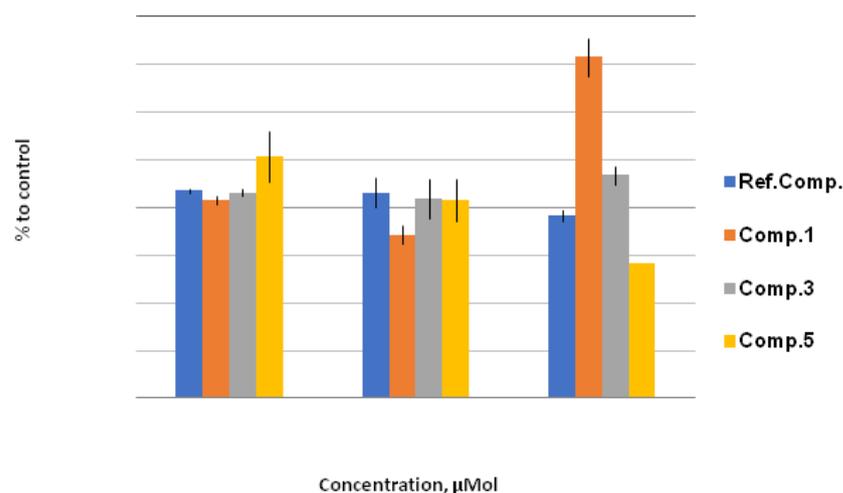


Figure 5. GPx activity, % to control. The effect of compounds 1, 3, and 5 on GPx activity under stress conditions relative to control ($\text{H}_2\text{O}/1\% \text{ EtOH} + 200 \text{ mmol NaCl}$; taken as 100%) at concentrations of 10 μM , 100 μM , and 1000 μM . Quantitative values are given in Electronic Supplementary Information.

The key parameter of the antioxidant protective potential is the redox status of glutathione, which is significantly biased towards the reduced form under normal conditions. The oxidation-reduction status of glutathione is supported by a number of enzymes, the main of which is glutathione reductase (GR). Glutathione reductase is an important component of the ROS detoxification system in plants and catalyzes the conversion of oxidized glutathione (GSSG) to its reduced form (GSH). It is known that selenium affects the activity of enzymes in the glutathione plant protection system [40]. Following this line, we have investigated the effect of compounds **1**, **3**, **5**, and **6–8** on the activity of glutathione reductase (GR) in the present work.

In contrast to the inhibitory effect of compound **6** toward GPx, a significant increase in GR activity with concentration growth was found. These results may indicate the promotion of the GSSG reduction reaction. However, this effect of a multiple increase in GR activity may be a combination of the GPx inhibitory effect of compound **6** and an increase in cellular antioxidant potential under stress, which can also lead to an increase in GR activity.

An increase in GR activity by compounds **7** and **8** was observed, but already against the background of a substantial growth of GPx activity, thus evidencing a drastic augmentation of the antioxidant profile even at concentrations of 10 μM (Figure 6). A sharp increase in the antioxidant profile even at 10 μM concentration is noteworthy, taking into account the significant increase in GPx activity by these compounds.

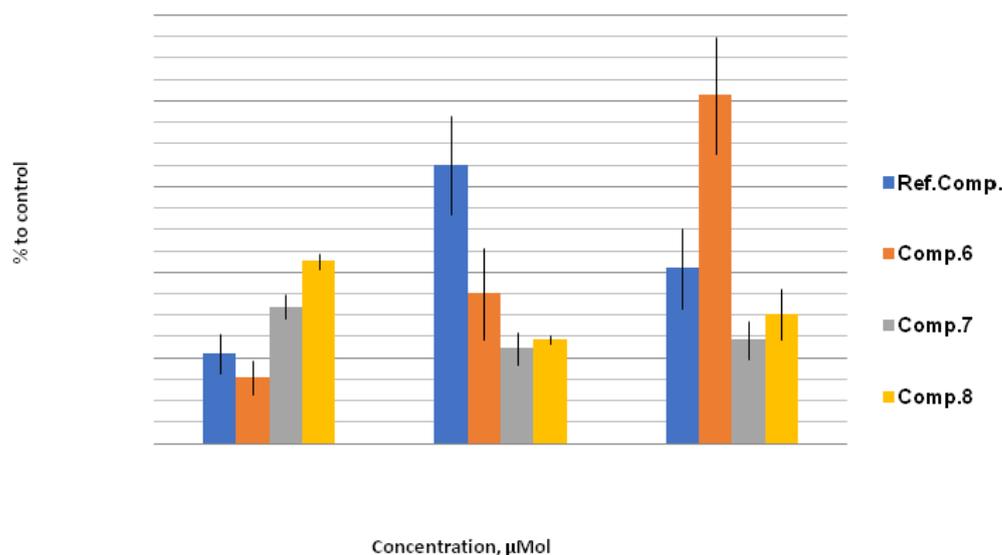


Figure 6. GR activity, % to control. The effect of compounds **6**, **7**, and **8** on GR activity under stress conditions relative to control ($\text{H}_2\text{O} + 200 \text{ mmol NaCl}$; taken as 100%) at concentrations of 10 μM , 100 μM , and 1000 μM . Quantitative values are given in Electronic Supplementary Information.

Lipophilic derivatives of 9-selenabicyclo[3.3.1]nonane demonstrated a considerable drop in GR activity, which, together with a slight increase in GPx activity, allows us to consider these compounds as potential medicines for mild metabolic correction. Furthermore, it is worthwhile to note a significant GR-inhibitory effect of the bromo derivative **1**, which in this case is characterized by a general decrease in lipid peroxidation (Figure 7).

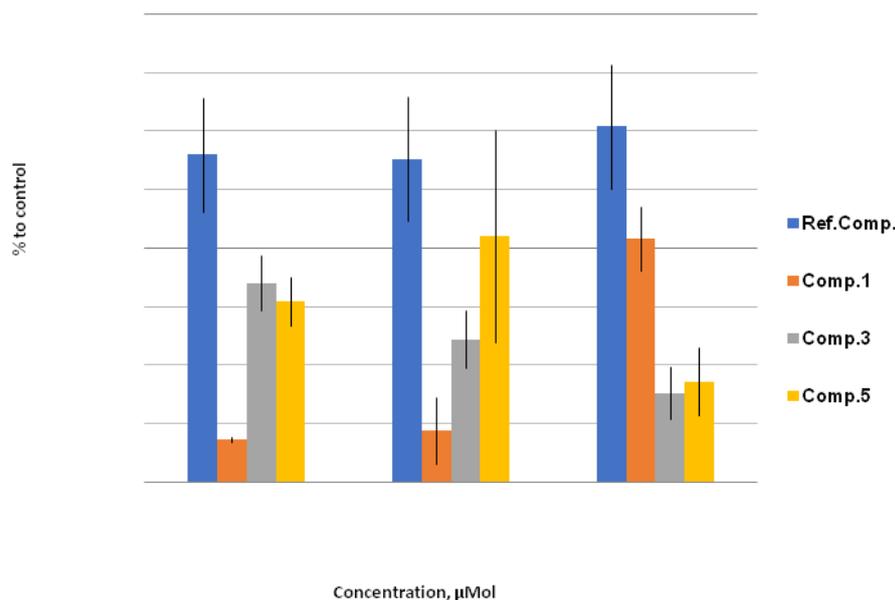


Figure 7. GR activity, % to control. The effect of compounds 1, 3, and 5 on GR activity under stress conditions relative to control ($\text{H}_2\text{O}/1\% \text{ EtOH} + 200 \text{ mmol NaCl}$; taken as 100%) at concentrations of 10 μM , 100 μM , and 1000 μM . Quantitative values are given in Electronic Supplementary Information.

3. Materials and Methods

3.1. General Information

X-ray diffraction experiments were carried out on a Bruker D8 Venture Photon 100 CMOS diffractometer (Bruker BioSpin GmbH, Rheinstetten, Germany) with $\text{Mo-K}\alpha$ radiation ($\lambda = 0.71073 \text{ \AA}$). X-ray crystallographic data for compound 5 (CCDC 2277477) are shown in Electronic Supplementary Information. These data can be obtained free of charge at <http://www.ccdc.cam.ac.uk/conts/retrieving.html> (accessed on 28 June 2023).

^1H (400.1 MHz) and ^{13}C (100.6 MHz) NMR spectra were recorded on a Bruker DPX-400 spectrometer (Bruker BioSpin GmbH, Rheinstetten, Germany) in 5–10% solution in CDCl_3 . ^1H and ^{13}C chemical shifts (δ) are reported in parts per million (ppm), relative to the residual solvent peak of CDCl_3 ($\delta = 7.27$ and 77.16 ppm in ^1H and ^{13}C -NMR, respectively).

The mass spectra were recorded on a Shimadzu GCMS-QP5050A (Shimadzu Corporation, Kyoto, Japan) with electron impact (EI) ionization (70 eV). Elemental analysis was performed on a Thermo Scientific Flash 2000 Elemental Analyzer (Thermo Fisher Scientific Inc., Milan, Italy).

The organic solvents were dried and distilled according to standard procedures.

3.2. The Synthesis of Compound 3

2,6-Dimethoxy-9-selenabicyclo[3.3.1]nonane (3). A solution of SeBr_2 (4.80 g, 20 mmol) in CH_2Cl_2 (50 mL) was added dropwise to a solution of 1,5-cyclooctadiene (2.16 g, 20 mmol) in CH_2Cl_2 (35 mL) and MeOH (15 mL). The reaction mixture was stirred for 15 h at room temperature. The solvent was removed on a rotary evaporator, and the residue was washed with C_6H_{14} (2 mL) at 0°C and dried in vacuum to constant mass. Product compound 3 (4.8 g, 96% yield) was colorless or light-yellow liquid.

^1H NMR (400 MHz, CDCl_3) δ 1.78–1.82 (m, 2H, SeCHCH_2), 2.00–2.04 (m, 2H, SeCHCH), 2.17–2.19 (m, 2H, OCHCH), 2.60–2.64 (m, 2H, OCHCH), 3.02–3.03 (m, 2H, SeCH), 3.36 (s, 6H, CH_3), 3.88–3.92 (m, 2H, MeOCH). ^{13}C NMR (100 MHz, CDCl_3) δ 27.87, 28.05, 28.87, 55.86, 81.04. Found: C, 47.96; H, 7.45; Se, 31.93. Calc. for $\text{C}_{10}\text{H}_{18}\text{O}_2\text{Se}$: C, 48.20; H, 7.28; Se, 31.68. MS (EI): m/z (%) = 250 (30, M^+), 218 (10), 179 (18), 137 (64), 105 (50), 79 (100), 71 (89), 45 (78), 41 (90). IR (film): 1086, 1153, 1186, 2817, 2922, 2977 cm^{-1} .

3.3. The Synthesis of Compound 5

2,6-Diacetoxy-9-selenabicyclo[3.3.1]nonane (5). A solution of SeBr₂ (4.80 g, 20 mmol) in CH₂Cl₂ (30 mL) was added dropwise to a solution of *cis,cis*-1,5-cyclooctadiene (2.16 g, 20 mmol) in a mixture of CH₂Cl₂ (20 mL) and AcOH (10 mL). The reaction mixture was stirred for 16 h at room temperature. Then the mixture was washed with water (3 × 20 mL), dried with CaCl₂, filtrated, and the solvent removed on a rotary evaporator. The residue was washed with C₆H₁₄ (2 mL) and dried in vacuum. Compound 5 (6.0 g, 98% yield) was obtained as colorless crystals, mp 108–109 °C.

¹H NMR (400 MHz, CDCl₃) δ 1.96–2.05 (m, 10H, SeCHCH₂, CH₃), 2.28–2.33 (m, 2H, OCHCH₂), 2.62–2.66 (m, 2H, OCHCH₂), 2.94–2.96 (m, 2H, SeCH), 5.46–5.48 (m, 2H, OCH). ¹³C NMR (100 MHz, CDCl₃) δ 20.97, 27.37, 27.80, 28.00, 74.07, 169.85. Found: C, 46.98; H, 6.09; Se, 26.11. Calc. for C₁₂H₁₈O₄Se: C, 47.22; H, 5.94; Se, 25.87. IR: 2922, 1729, 1364, 1231, 1021, 960, 870 cm⁻¹. MS (EI): *m/z* (%) = 306.00 (9, M⁺) 186.00 (18), 123.15 (10), 105.10 (59), 95.10 (11), 93.10 (11), 79.10 (28), 67.10 (10), 43.05 (100), 41.10 (17), 39.10 (10).

3.4. The Synthesis of Compound 6 (Modified Procedure)

2,6-Dipyridinium-9-selenabicyclo[3.3.1]nonane dibromide (6). A solution of SeBr₂ (2.4 g, 10 mmol) in methylene chloride (20 mL) was added dropwise to a solution of *cis,cis*-1,5-cyclooctadiene (1.08 g, 10 mmol) in methylene chloride (20 mL). The reaction mixture was stirred for 8 h at room temperature. A solution of pyridine (1 g, 1.25 mmol) in methylene chloride (5 mL) was added dropwise to the reaction mixture. The reaction mixture was stirred overnight (16 h) at room temperature. The precipitate was filtered, washed with cold methylene chloride, and dried in vacuum. Compound 6 (5.05 g, ~100% yield) was obtained as a beige powder, mp 177–178 °C.

¹H NMR (400 MHz, D₂O) δ 2.49–2.55 (m, 2H, SeCHCH₂), 2.69–2.73 (m, 4H, NCHCH₂, SeCHCH₂), 3.21–3.33 (m, 2H, NCHCH₂), 3.57–3.59 (m, 2H, SeCH), 5.99–6.04 (m, 2H, NCH), 8.24 (t, 4H, *m*-H_{pyr}), 8.69 (t, 2H, *p*-H_{pyr}), 9.17 (d, 4H, *o*-H_{pyr}). ¹³C NMR (100 MHz, D₂O) δ 28.39, 31.01, 32.19, 77.43, 131.52, 145.99, 149.29. Found: C, 43.08; H, 4.58; Br, 31.48; N, 5.73; Se, 15.76. Calc. for C₁₈H₂₂Br₂N₂Se: C, 42.80; H, 4.39; Br, 31.64; N, 5.55; Se, 15.63. MS (EI): *m/z* (%) = 345.80 (0.36, M⁺-2Br), 187.05 (14.20), 105.15 (19.59), 91.05 (9.20), 80.00 (17.64), 79.05 (100.00), 78.10 (20.72), 77.05 (10.78), 52.05 (56.84), 51.10 (24.21), 39.05 (19.49). IR: 3404, 3044, 1629, 1488, 1134, 769, 690 cm⁻¹.

3.5. The Synthesis of Compound 7

2,6-Dihydroxy-9-selenabicyclo[3.3.1]nonane (7). A solution of SeBr₂ (4.80 g, 20 mmol) in CH₃CN (30 mL) was added dropwise to a mixture of *cis,cis*-1,5-cyclooctadiene (2.16 g, 20 mmol) in CH₃CN (35 mL), H₂O (5 mL), and NaHCO₃ (3.5 g, 42 mmol). The reaction mixture was stirred for 14 h at room temperature. The solvent was removed on a rotary evaporator, and the residue was washed with cold C₆H₁₄ (5 mL) and dried in vacuum. Compound 7 (4.35 g, 98% yield) was obtained as white crystals, mp 249–250 °C.

¹H NMR (400 MHz, DMSO-D₆/CDCl₃) δ 1.73–1.81 (m, 2H, SeCHCH₂), 1.85–1.92 (m, 2H, SeCHCH₂), 2.04–2.14 (m, 2H, OCHCH₂), 2.62–2.70 (m, 4H, OCHCH₂, OH), 3.53–3.57 (m, 2H, SeCH), 4.24–4.27 (m, 2H, OCH). ¹³C NMR (100 MHz, DMSO-D₆/CDCl₃) δ 25.91, 29.50, 30.45, 69.43. Found: C, 43.74; H, 6.56; Se, 35.49. Calc. for C₈H₁₄O₂Se: C, 43.45; H, 6.38; Se, 35.70. MS (EI): *m/z* (%) = 222 (52, M⁺), 205 (5), 178 (20), 149 (15), 133 (17), 123 (27), 95 (58), 79 (69), 71 (49), 67 (41), 57 (31), 55 (58), 41 (100), 39 (51). IR: 873, 982, 1017, 2899, 2932, 3340 cm⁻¹.

3.6. The Synthesis of Compound 8

2,6-Di(2-hydroxyethoxy)-9-selenabicyclo[3.3.1]nonane (8). A solution of SeBr₂ (4.80 g, 20 mmol) in CH₃CN (30 mL) was added dropwise to a solution of *cis,cis*-1,5-cyclooctadiene (2.16 g, 20 mmol) in a mixture of CH₃CN (30 mL) and ethylene glycol (10 mL). The reaction mixture was stirred for 14 h at room temperature. The solvents were removed in vacuum, and water (10 mL) was added to the residue. The product was extracted with CH₂Cl₂ (4 × 20 mL), the combined organic phase was washed with 1M NaHCO₃

solution (3 × 20 mL), dried with CaCl₂, and filtered. The solvent was removed on a rotary evaporator, and the residue was dried in vacuum. Compound 8 (5.27 g, 85% yield) was obtained as a colorless oil.

¹H NMR (400 MHz, D₂O) δ 1.84–1.91 (m, 2H, SeCHCH₂), 2.11–2.16 (m, 2H, SeCHCH₂), 2.27–2.31 (m, 4H, OCHCH₂, OH), 2.63–2.67 (m, 2H, OCHCH₂), 3.20–3.22 (m, 2H, SeCH), 3.63–3.68 (m, 2H, CH₂OCH), 3.70–3.83 (m, 6H, OCH₂CH₂O), 4.19–4.22 (m, 2H, OCH). ¹³C NMR (100 MHz, D₂O) δ 30.28, 30.57, 31.18, 63.45, 71.73, 82.96. Found: C, 46.37; H, 7.03; Se, 25.82. Calc. for C₁₂H₂₂O₄Se: C, 46.60; H, 7.17; Se, 25.53. IR: 3392, 2919, 1649, 1432, 1362, 1089, 888, 653 cm⁻¹. MS (EI): *m/z* (%) = 310.10 (15, M⁺), 186.00 (13), 167.15 (27), 123.05 (15), 105.10 (53), 101.15 (21), 95.10 (20), 91.15 (23), 80.05(29), 79.10 (89), 73.10 (42), 57.05 (53), 45.10 (100), 41.10 (59), 39.10 (30)

3.7. Plant Material

Studies were carried out on oilseed radish seeds (*Raphanus sativus* L. var. *oleiferus* Metzg.) of lines of Irkutsk State Agricultural Academy with laboratory germinability of 80–98% and weighing 1000 seeds 9.5 g. Seeds were germinated on wet filter paper in Petri dishes at a constant 23 °C temperature in the dark for 4 days after wetting with the test solutions. The number of seeds in one cup was 30 pcs. The experiment was repeated 3 times.

3.8. Evaluation of Germinability and Mass of Seedlings

Germinability was analyzed according to the All-Union State Standard 10-14-86, “Oilseed Radish Seeds. Varietal and sowing qualities”. These indicators were determined in accordance with All-Union State Standard 12038-84, “Seeds of agricultural crops. Methods for determining germinability” [41]. The mass of shoots and roots was determined using gravimetric analysis.

3.9. Determination of Protein Content

Protein content was determined by the degree of binding to the Coomassie blue dye (CBB G250 “Sigma”) according to the Bradford method [42].

3.10. Determination of Glutathione Reductase Activity

Glutathione reductase activity (EC 1.6.4.2) was measured according to the method described by Nigmatullina et al. [43]. The activity of glutathione reductase was determined by the change in absorption at 340 nm caused by the oxidation of NADPH in 3.5 min with an interval of 1 s on the spectrophotometer. The enzyme activity was calculated using the extinction coefficient for NADP⁺ at a wavelength of 340 nm, which is equal to 6.22 mmol⁻¹cm⁻¹.

3.11. Evaluation of Diene Conjugates

Analysis of the content of the primary products of lipid peroxidation—diene conjugates (DC)—was carried out according to the method [44] in our modification. The measurement was performed on a spectrophotometer at a wavelength of 203 nm. The obtained optical density (D) was used to calculate the concentration of diene conjugates (re-calculated per 1 g wet mass) using an extinction coefficient equal to 2.2 × 10⁵ mol⁻¹cm⁻¹. Salinization was chosen as a stress, which was created with NaCl, a concentration of 200 mmol was taken from literature data [40]. This concentration causes stress since it significantly increases the level of lipid peroxidation by almost two times compared with the control.

3.12. Statistics

The results obtained are given as relative values in comparison with the control (taken as 100%). Electronic supplementary information contains the data, which are presented as arithmetic mean values and their standard deviations, which were obtained in three

independent experiments and calculated using Microsoft Excel. The statistical significance of the differences between the compared mean values was assessed using the Mann–Whitney U-test.

3.13. X-ray Analysis

Data were collected on a BRUKER D8 VENTURE PHOTON 100 CMOS diffractometer with MoK α radiation ($\alpha = 0.71073 \text{ \AA}$) using the φ and ω scans technique. Using Olex2 [45], the structure was solved with the ShelXS [46] structure solution program using Direct Methods and refined with the XL refinement package using Least Squares minimization. Data were corrected for absorption effects using the multi-scan method (SADABS). All non-hydrogen atoms were refined anisotropically using SHELX [46]. The coordinates of the hydrogen atoms were calculated from geometrical positions.

Crystallographic data for compound **5** and experimental details are given in Tables 1ESM and S1ESM (Electronic Supplementary Information). These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via <http://www.ccdc.cam.ac.uk>.

4. Conclusions

A number of functional selenium-containing compounds were synthesized and isolated. The fundamental possibility of the synthesis of selenium-containing heterocycles via one-pot selenofunctionalization of dienes by selenium dihalides in the presence of nucleophiles was shown and realized for the first time. The synthesized compounds showed promising enzyme-like activity. The effect of 9-selenabicyclo[3.3.1]nonanes on the following major characteristics of the antioxidant profile was first studied: the level of lipid peroxidation, the activity of glutathione peroxidase, and the activity of glutathione reductase. A significant effect of the nature of the substituent in the polycyclic selenium-containing core on the biological activity of the compounds was shown. All studied compounds drastically reduced the level of lipid peroxidation both under normal conditions and under stress at concentrations of up to 100 μm . Unexpectedly, pyridinium compound **6** and hydrophobic derivatives **1**, **3**, and **5** displayed an inhibitory effect towards glutathione reductase activity with a decrease in LPO levels, while hydroxy and hydroxyethoxy derivatives showed an increase in the activity of both glutathione peroxidase and glutathione reductase.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/inorganics11070304/s1>, ^1H and ^{13}C NMR spectra of the obtained compounds, X-ray crystallographic data including bond lengths, bond angles and torsion angles for compound **5** (Tables 1ESM and S1ESM), experimental data of evaluation of diene conjugates, glutathione reductase and glutathione peroxidase activities.

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