

## Article

# Oxidative, Genotoxic and Cytotoxic Damage Potential of Novel Borenium and Borinium Compounds

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**Abstract:** In this study, the biological properties of novel borenium and borinium compounds in terms of their oxidative, genotoxic, and cytotoxic effects were assessed on cultured human peripheral blood cells, as well as several types of cancer cells. Our results revealed that the borinium compounds yielded the best results in terms of supporting total antioxidant capacity (TAC). In fact, borenium 1, borenium 2, borenium 3, borinium 4, and borinium 5 compounds elevated TAC levels of cultured human blood cells at rates of 42.8%, 101.5%, 69.8%, 33.3%, and 49.2%, respectively. There were no statistically significant differences ( $p > 0.05$ ) between the negative control and the groups treated with all borinium and borenium concentrations from the micronucleus (MN) and chromosome aberration (CA) assays, demonstrating the non-genotoxic effects. Moreover, borenium 1 (60.7% and 50.7%), borenium 2 (70.4% and 57.2%), borenium 3 (53.1% and 45.2%), borinium 4 (55.1% and 48.1%), and borinium 5 (51.0% and 36.1%) minimized the mitomycin C (MMC)-induced genotoxic damages at different rates as determined using CA and MN assays, respectively. Again, it was found that the borinium compounds exhibited higher cytotoxic activity on cancer cells when compared to borenium compounds. Consequently, in light of our in vitro findings, it was suggested that the novel borinium and borenium compounds could be used safely in pharmacology, cosmetics, and various medical fields due to their antioxidant and non-genotoxic features, as well as their cytotoxicity potential on cancer cells.

**Keywords:** antioxidant; antigenotoxicity; boron compounds; borenium; borinium; cancer cells; cytotoxicity; human blood cells; genotoxicity



**Citation:** Oguzkan, S.B.; Turkez, H.; Ugras, H.I.; Tatar, A.; Mardinoglu, A. Oxidative, Genotoxic and Cytotoxic Damage Potential of Novel Borenium and Borinium Compounds.

*Inorganics* **2023**, *11*, 324. <https://doi.org/10.3390/inorganics11080324>

Academic Editors: Michael A. Beckett, Marvin Antonio Soriano-Ursúa and Bhaskar C. Das

Received: 5 June 2023

Revised: 20 July 2023

Accepted: 27 July 2023

Published: 31 July 2023



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## 1. Introduction

Around 400 different industrial domains involving the production of ceramics and fertilizers, glass and glass fibers, pharmaceuticals, chemicals, nuclear power, automobiles, and spacecrafts employ boron (B) in different chemical structures. In light of recent scientific findings based on the biological and physicochemical properties of boron-containing compounds (BCCs), B is now regarded as a strategic element whose usage and application domains are expanding constantly [1]. Due to its superior physicochemical properties and its preference in many industrial fields, new boron derivatives are synthesized by scientists and new BCCs are offered for use. B is considered to be a crucial microelement for plants. Despite the vast body of scientific evidence, it has not been conclusively reported that B is necessary for both humans and animals. In fact, several BCCs, such as

borates, boronates, and boronic acids, exhibited interesting biological activities, including antiviral [2], antibacterial [3,4], antifungal and antiparasitic [5,6], antioxidant [7,8], wound healing [9], anti-inflammatory [10,11], antimutagenic [12], anticancerogenic [13,14], radiobiological [15], and neuroprotective properties [16,17]. Interestingly it was found that people living in B-rich regions had less cancer incidence than the people living in B-poor regions [18]. The functions of B in the human body are not known clearly but previously suggested health benefits of boron included protection of the liver, enhancement of fetal development, regulation of enzymatic activity associated with the immune system, and improvement of brain functions in humans and animals [19]. Hence, researchers and industries have recently given extensive effort in studying BCCs with the goal of understanding their physiological process and locating novel health technologies endowed with clinical safety.

A few studies on boron compounds that have been previously synthesized are the ionic liquids based on boronium cations [20]. These cations were considered to be a highly electrophilic species that was elusive and was reported to have a key role in the chemistry of B. A commercially available ionic fluid boron tetra-fluoro borate ( $\text{BF}_4^-$ ) anion has this cation. Compounds with a  $\text{BF}_4^-$  anion have a hypersensitivity to the reactivity of the pipe to air and water [21]. Given this context, the main objective of the current investigation was to synthesize novel cationic B-based compounds that exhibit high resistance properties due to a B cation and introduce novel boron compounds with potential to be used in the biomedical field. In accordance with this goal, we aimed to obtain useful electrolyte compounds indicating higher thermal stability and more resistance to heat treatment.

Understanding the behavior of the novel borenium and borinium compounds can guide researchers or applicators in choosing the relevant biomedical materials for different clinical and anatomical purposes. Along with the proper thermal stability and heat-resistance features of novel biomedical materials, their toxicity potential should be evaluated before safe clinical use. The chemical and physical features of biomedical materials influence their biological and toxic potential [22]. In this regard, the chemical, physical, and biological features of B provide medicinal chemists a unique chance to research and develop brand new fields of biomedical sciences, especially in drug discovery [23–25]. Recent investigations indicated that BCCs could affect the crucial cellular machineries implicating cell survival, tissue regeneration, and immunogenic responses [26]. Although the toxicity potentials by borenium and borinium compounds are not well known, their antibacterial and antiviral properties have been well documented [27–29]. Thus, these limited studies indicate that borenium and borinium compounds can be used safely in medical, cosmetics, and green chemistry domains [28,30]. Hence, the second aim of this investigation was to assess the oxidative, genotoxic, and cytotoxic effects of novel borenium and borinium compounds on human peripheral blood cells, as well as several types of cancer cells. We assessed the *in vitro* effects of these compounds on cytotoxicity using an MTT assay, DNA damage response via MN and CA assays, and antioxidant capacity using a TAC assay.

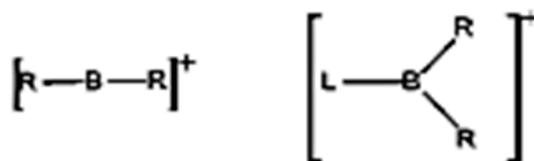
## 2. Materials and Methods

### 2.1. Novel Ionic Liquids

Ionic liquids are salts that exist in liquid form at room temperature, and at lower temperatures, and generally have an organic cationic part. Ionic liquids have high polarity, low vapor pressure, and are resistant to high temperatures. Due to these properties, ionic liquids can be used repeatedly as both solvent and catalyst in many reactions, and they can also be easily removed from the reaction medium without leaving any waste. In this way, ionic liquids are superior to other classic solvents and cause them to be preferred in terms of environment.

In this study, an oxidative, genotoxic, and cytotoxic evaluation of new boron-containing molecules to the family of ionic liquids was performed. In the five novel compounds we synthesized, new ionic liquids with a boron cationic center—not  $\text{BF}_4^-$  anion—were syn-

thesized. Cationic forms of boron are produced in two different forms, borinium and borenium, as shown in Figure 1. The original ionic liquids were synthesized with organic extension derivatives of these structures containing chiral structures.



**Figure 1.** Borinium (L) and borenium (R) forms.

NMR analyses were performed in DMSO,  $CDCl_3$ , and MeOH with an Agilent Premium Contact NMR 600 MHz spectrophotometer, and the chemical shift values ( $\delta$ ) in the spectrum were expressed in ppm. IR measurements were performed with the Agilent Cary 630 FTIR device. The recovery of the silver amino acid salts was confirmed with the Agilent 7700 series ICP-MS instrument. A Heidolph brand rotary evaporator was used for the solvent removal system. Melting point was determined with Stuart SMP40 brand melting point determination device. Analytical TLCs were performed using aluminum plates coated with a layer of silica gel ( $SiO_2$ , Merck 60 F254).

## 2.2. Experimental Design

We used human U87MG, SHSY-5Y, PC-3, and Detroit-562 cell lines for screening of cytotoxicity of boronated compounds. The U87MG, SHSY-5Y, and Detroit-562 cell lines were obtained from Atatürk University, Faculty of Medicine, Erzurum, Turkey. PC-3 cells were provided from the American Type Culture Collection (Manassas, VA, USA). Cells were harvested using 0.25% trypsin–EDTA solution and suspended with RPMI 1640 medium (Sigma-Aldrich, St. Louis, MO, USA) containing 10% FBS, L-glutamine (1%), and penicillin–streptomycin mixture (1%). The different concentrations (from 1.56 to 400 mg/L or from 2.91 to 1316.01  $\mu M$ ) of borenium and borinium compounds were applied to the cultures ( $n = 5$ ). All experiments were performed in accordance with the rules of the World Medical Association.

## 2.3. Cytotoxicity Testing

The cytotoxic potential of the boron compounds was determined by MTT analysis. In brief, compounds were added into the cell culture plates at a wide concentration range from 1.56 to 400 mg/L or from 2.91 to 1316.01  $\mu M$  and incubated for 48 h ( $n = 5$ ). Then, 10  $\mu L$  of MTT solution was added to wells and incubated for an additional 3 h at 37 °C. After discarding cell mediums, DMSO (100  $\mu L$ ) was added to wells for dissolving formazan crystals. Finally, the color intensities were measured via using a microplate reader at 570 nm [31]. A podophyllotoxin derivative, etoposide (Merck), which is a chemotherapy medication, was dissolved in DMSO (<1%) and used as a positive control agent for comparing cytotoxic action of boron compounds.

## 2.4. Genotoxicity Testing

The in vitro genotoxic/antigenotoxic potential of boron compounds on human whole blood cell cultures using chromosomal aberration (CA) and micronucleus (MN) assays was tested. For CA assay, a 0.65 mL aliquot of heparinized blood sample was cultured in 7 mL of culture medium (Chromosome medium B; Biochrom, Berlin, Germany) containing phytohemagglutinin (5 mg/mL, Biochrom). Then, the cultures were incubated for 72 h at 37 °C. Around 2 h before harvesting, 0.1 mL of colchicine (0.2 mg/mL, Sigma; St Louis, Missouri, USA) was added into the culture tubes and hypotonic treatment/fixation steps were performed. The prepared slides were stained using Giemsa solution (3%). For each culture tube ( $n = 5$ ), 30 well-spread metaphases were scored to detect CA frequencies. Chromatid or

chromosome gaps and chromatid or chromosome breaks were scored as recommended by Environmental Health Criteria 46 for environmental monitoring of human populations [32].

For MN assay, cytochalasin B (at a final concentration of 6  $\mu\text{g}/\text{mL}$ , Sigma) was added into the culture tubes (contained  $2 \times 10^6$  cells/mL) after 44 h of culture and incubated for 72 h ( $n = 5$ ). After the incubation period of three days, the lymphocytes were fixed using treatment with ice-cold methanol:acetic acid (1:1) and stained Giemsa (5%) for 12 min. The slides were scored by using a bright-field microscope (at  $400\times$  magnification, Olympus). A total of 2000 binucleated lymphocytes were scored per treatment type for the presence of one, two, or more micronuclei according to previously reported standard criteria [33]. Mitomycin C (MMC,  $5 \times 10^{-6}$  M) was used as the positive control in CA and MN assays [34].

### 2.5. Determining of TAC Levels

The commercially available kit (Rel Assay Diagnostics, Gaziantep, Turkey) was used to determine the antioxidant capabilities of the borenium and borinium compounds on human whole blood cell cultures for 72 h ( $n = 5$ ). The principle of this kit assay is to monitor antioxidant levels of cultures via inhibiting formation of free radical featured 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) compound. Furthermore, the assay is calibrated with Trolox equivalent vitamin E analogue. The cultures without boronated compounds were studied as a negative control group. Ascorbic acid (10  $\mu\text{M}$ ) was also used as a positive control in total antioxidant capacity (TAC) analysis [35].

### 2.6. Statistical Analyses

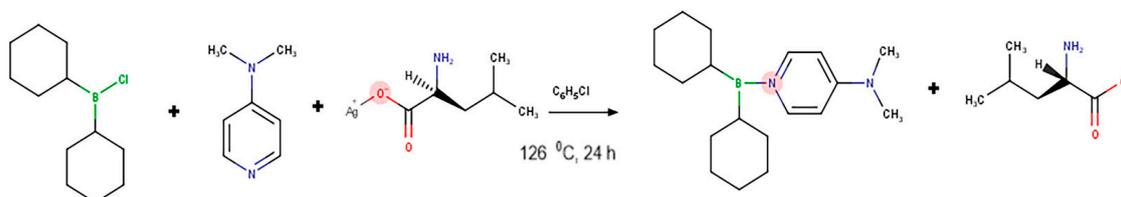
Statistical analysis was performed using IBM program SPSS version 25. All tests were performed for at least five different repeats. The obtained data were analyzed using a variance (ANOVA) test followed by Duncan's test and values with  $p < 0.05$  were accepted as significantly different. Probit regression analyses were performed to estimate the concentrations required to reduce cell viability rates by 50% using SPSS [36].

## 3. Results

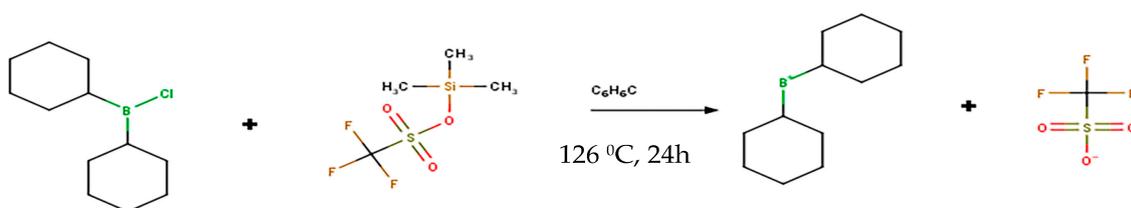
Examples of ionic liquids containing boron cations are very rare in the literature [37]. In previous research, a series of ionic liquids containing N-alkylimidazole-amine BH<sub>2</sub><sup>+</sup> structures were synthesized, and their electronic and spectroscopic properties were examined [38]. Some new boronium cation-based ionic liquids were also synthesized and their potential for use in lithium ion batteries was investigated. In this study, it is of great importance to test the usability of ionic liquids with boron cations in practice [39].

### 3.1. Synthesis of Borenium Ionic Liquids

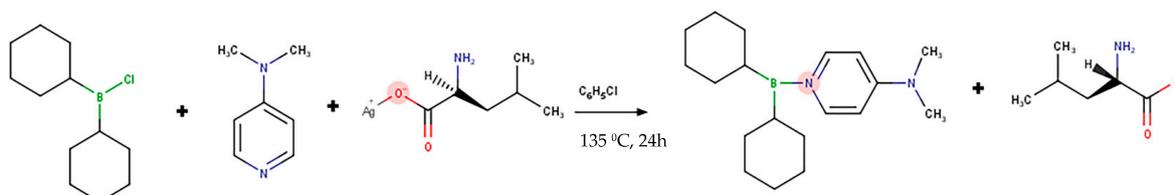
The dialkyl aryl borane and the anion compound, chlorobenzene, were dissolved. After mixing, an aromatic amine compound was added to this reaction solution, and it was allowed to boil at a temperature  $>120$  °C. Centrifugation was applied and the solvent component was removed. The synthesis reaction and code numbers of the synthesized borenium 1, 2, and 3 compounds are presented in Figures 2–4. The IR, <sup>1</sup>H-NMR, and <sup>13</sup>C-NMR (Agilent, Premium Compact, 14.1 tesla, 600 MHz) spectra of three novel borenium compounds are presented in Supplementary Figures S1–S9.



**Figure 2.** Dicyclohexyl borenium dimethyl amino pyridine trifluoro methane sulfonate synthesis reaction (Borenium 1).



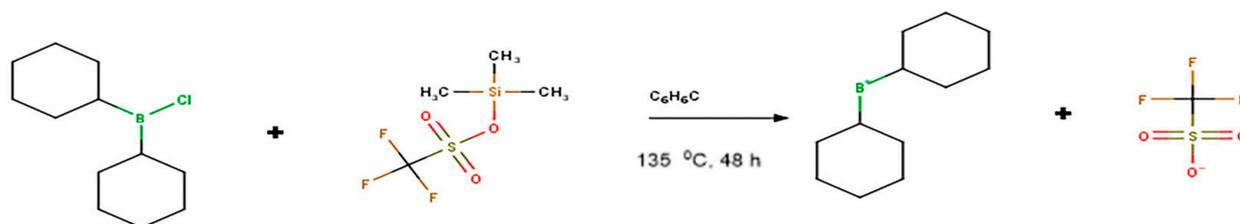
**Figure 3.** Synthesis reaction of dicyclohexylborenyldimethyl amino pyridine 2-amino-4-methylpentanoate (Borenyum 2).



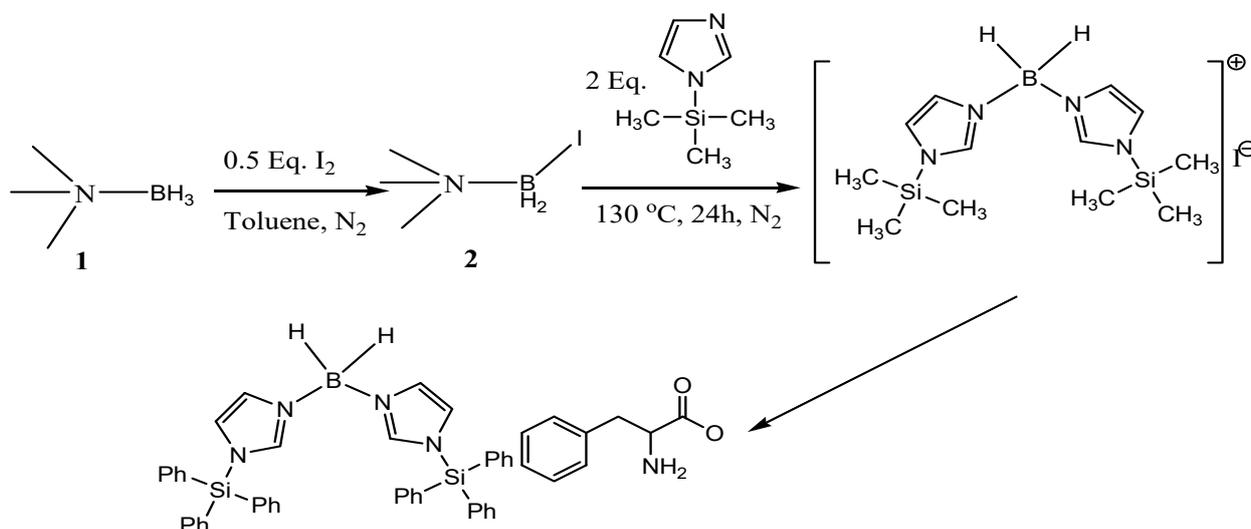
**Figure 4.** Bis dimethyl amino borenyum dimethyl amino pyridine trifluoro acetate synthesis reaction (Borenyum 3).

### 3.2. Synthesis of Borinium Ionic Liquids

The dialkyl aryl borane and the anion compound, chlorobenzene, were dissolved. After mixing, an aromatic amine compound was added to this reaction solution, and it was allowed to boil at a temperature of  $>130\text{ }^\circ\text{C}$ . Centrifugation was applied and the solvent component was removed. After one night, after the reaction was cooled to room temperature, the first centrifugation was applied, and the solvent part was removed. The synthesis reactions of borinium 4 and borinium 5 are presented in Figures 5 and 6. The IR (Agilent Cary 630),  $^1\text{H-NMR}$ , and  $^{13}\text{C-NMR}$  spectra of three novel borinium compounds are presented in Supplementary Figures S10–S15.



**Figure 5.** Synthesis reaction of dicyclohexyl borinium trifluoro methane sulfonate (Borinium 4).



**Figure 6.** Synthesis reaction of bis(1,2-dimethyl-1H-imidazol-3-yl) dihydroboronium iodide (Borinium 5).

The cytotoxicity of the boron compounds on three different cancer cells, including human glioblastoma (U87MG), neuroblastoma (SHSY-5Y), prostate (PC-3) and pharyngeal (Detroit-562) cancer cell lines, as well as human whole blood cells, was determined. The determination of cytotoxicity was evaluated by calculating percent inhibition ( $IC_{50}$ ). Table 1 presents the  $IC_{50}$  values as estimated using the results of the MTT assay and Probit analysis.  $IC_{50}$  values are used to express the concentration corresponding to a survival rate of 50% under in vitro conditions. This value is commonly used for measuring antagonist drug potency using outputs from cell-based cytotoxicity tests; the lower the  $IC_{50}$  value, the more cytotoxic the compound, drug candidate, or drug is. The international authorities, such as the National Cancer Institute, debate  $IC_{50}$  values for classifying cytotoxicity potentials by compounds as high cytotoxic ( $IC_{50} < 20$  mg/L), moderate cytotoxic ( $20$  mg/L  $< IC_{50} < 200$  mg/L), weak cytotoxic ( $200$  mg/L  $< IC_{50} < 500$  mg/L), and non-cytotoxic ( $IC_{50} > 500$  mg/L) [40–43]. No statistical difference ( $p > 0.05$ ) in cell viability was observed between the negative controls and vehicle (DMSO,  $<1\%$ ) controls. Borinium 5 was found to be the most promising compound as an anti-proliferative agent, but it seems necessary to take into consideration that concentration-dependent cytotoxicity on healthy cells might occur at excessive exposure. The PC-3 cells were found to be more sensitive to boronated compounds in comparison to other cancer cells, such as the U87MG, SHSY-5Y, and Detroit-562 lines. The concentration-dependent cell viability alterations are presented in Supplementary Figures S16–S20.

**Table 1.** The determined  $IC_{50}$  values for boronated compounds in several cancer cell lines and human whole blood cells after 48 h of exposure.

Compounds	$IC_{50}$ Value				
	U87MG Cells	SHSY-5Y Cells	PC-3 Cells	Detroit-562 Cells	Human Whole Blood Cells
Positive control (Etoposide)	16.305 mg/L 0.027 $\mu$ M	12.665 mg/L 0.022 $\mu$ M	6.904 mg/L 0.012 $\mu$ M	26.342 mg/L 0.045 $\mu$ M	81.122 mg/L 0.138 $\mu$ M
Borenium 1	117.365 mg/L 317.031 $\mu$ M	86.141 mg/L 232.687 $\mu$ M	67.608 mg/L 182.625 $\mu$ M	106.884 mg/L 288.719 $\mu$ M	235.190 mg/L 635.304 $\mu$ M
Borenium 2	168.410 mg/L 516.304 $\mu$ M	111.361 mg/L 341.406 $\mu$ M	86.773 mg/L 266.025 $\mu$ M	179.662 mg/L 550.800 $\mu$ M	324.655 mg/L 995.314 $\mu$ M
Borenium 3	96.674 mg/L 318.057 $\mu$ M	77.804 mg/L 255.975 $\mu$ M	53.096 mg/L 174.685 $\mu$ M	108.025 mg/L 355.402 $\mu$ M	177.020 mg/L 582.396 $\mu$ M
Borinium 4	88.369 mg/L 197.098 $\mu$ M	59.113 mg/L 131.845 $\mu$ M	60.554 mg/L 135.059 $\mu$ M	92.045 mg/L 205.297 $\mu$ M	145.224 mg/L 323.907 $\mu$ M
Borinium 5	71.436 mg/L 132.783 $\mu$ M	55.238 mg/L 102.804 $\mu$ M	41.941 mg/L 78.057 $\mu$ M	69.786 mg/L 129.880 $\mu$ M	169.208 mg/L 314.916 $\mu$ M

The results of the genotoxicity tests of five different borenium and borinium compounds, which were evaluated in cultured human lymphocytes by CA and MN assays, are shown in Table 2. The rates of chromosomal aberrations (abnormal cell, %) and abnormal cells (CAs/cell) were determined by blindly scoring at least 30 well-spread metaphases for each culture type (a total of at least 150 metaphases for each experimental group). Likewise, the rates of micronuclei (MNs) were monitored by blindly scoring at least 1000 binucleated cells (MN/1000 cells) for each culture type (a total of at least 4000 binucleated cells for each treatment). MMC treatment led to 6.13- and 6.69-fold change increases of CAs and MNs formations, respectively. On the contrary, the frequency of CAs and the rate of MNs in cells treated with different concentrations of the novel boron compounds were similar to those of the untreated control cells ( $p > 0.05$ ). When assessed in terms of genotoxic damage potentials, the rates of CAs/cell and MN/1000 cells did not significantly ( $p > 0.05$ )

increase at all tested concentrations of boron compounds. Hence, our findings reveal the non-genotoxic features of these novel boron compounds.

**Table 2.** The CA and MN frequencies after exposure to boronated compounds in cultured human lymphocytes for 72 h.

Groups		CAs/Cell	MN/1000 Cells
Negative control		0.32 ± 0.04	2.89 ± 0.18
Positive control (MMC, 5 × 10 <sup>-6</sup> M)		1.96 ± 0.22 *	19.35 ± 2.44 *
Borenium 1	1.56 mg/L (4.22 µM)	0.34 ± 0.08	2.81 ± 0.23
	3.12 mg/L (8.44 µM)	0.32 ± 0.05	2.75 ± 0.21
	6.25 mg/L (16.88 µM)	0.35 ± 0.07	2.66 ± 0.27
	12.5 mg/L (33.77 µM)	0.35 ± 0.07	2.71 ± 0.34
	25 mg/L (67.53 µM)	0.37 ± 0.05	2.84 ± 0.32
	50 mg/L (135.06 µM)	0.34 ± 0.08	2.88 ± 0.25
	100 mg/L (270.12 µM)	0.37 ± 0.04	2.93 ± 0.33
	200 mg/L (540.24 µM)	0.39 ± 0.09	2.97 ± 0.37
	400 mg/L (1080.48 µM)	CD	CD
Borenium 2	1.56 mg/L (4.79 µM)	0.24 ± 0.05	2.68 ± 0.25
	3.12 mg/L (9.58 µM)	0.27 ± 0.07	2.61 ± 0.28
	6.25 mg/L (19.16 µM)	0.24 ± 0.09	2.75 ± 0.35
	12.5 mg/L (38.32 µM)	0.26 ± 0.06	2.89 ± 0.23
	25 mg/L (76.64 µM)	0.28 ± 0.08	2.96 ± 0.28
	50 mg/L (153.28 µM)	0.33 ± 0.08	3.08 ± 0.34
	100 mg/L (306.56 µM)	0.37 ± 0.06	2.95 ± 0.37
	200 mg/L (613.12 µM)	0.39 ± 0.09	3.19 ± 0.29
	400 mg/L (1226.24 µM)	0.37 ± 0.07	3.24 ± 0.32
Borenium 3	1.56 mg/L (5.14 µM)	0.30 ± 0.05	2.77 ± 0.27
	3.12 mg/L (10.28 µM)	0.30 ± 0.07	2.73 ± 0.31
	6.25 mg/L (20.56 µM)	0.29 ± 0.08	2.78 ± 0.26
	12.5 mg/L (41.13 µM)	0.34 ± 0.09	2.93 ± 0.37
	25 mg/L (82.25 µM)	0.32 ± 0.09	3.04 ± 0.34
	50 mg/L (164.50 µM)	0.30 ± 0.07	3.08 ± 0.33
	100 mg/L (329 µM)	0.37 ± 0.08	3.11 ± 0.38
	200 mg/L (658 µM)	0.39 ± 0.07	3.16 ± 0.34
	400 mg/L (1316 µM)	CD	CD
Borinium 4	1.56 mg/L (3.49 µM)	0.33 ± 0.05	2.55 ± 0.14
	3.12 mg/L (6.97 µM)	0.34 ± 0.04	2.63 ± 0.22
	6.25 mg/L (13.94 µM)	0.30 ± 0.07	2.94 ± 0.29
	12.5 mg/L (27.88 µM)	0.36 ± 0.06	2.41 ± 0.15
	25 mg/L (55.76 µM)	0.33 ± 0.03	2.77 ± 0.17
	50 mg/L (111.52 µM)	0.30 ± 0.02	2.92 ± 0.22
	100 mg/L (223.04 µM)	0.38 ± 0.06	2.97 ± 0.33
	200 mg/L (446.08 µM)	0.42 ± 0.05	3.12 ± 0.18
	400 mg/L (892.16 µM)	CD	CD
Borinium 5	1.56 mg/L (2.91 µM)	0.30 ± 0.05	2.69 ± 0.21
	3.12 mg/L (5.82 µM)	0.34 ± 0.07	2.66 ± 0.24
	6.25 mg/L (11.63 µM)	0.36 ± 0.09	2.75 ± 0.32
	12.5 mg/L (23.26 µM)	0.36 ± 0.08	2.83 ± 0.31
	25 mg/L (46.53 µM)	0.38 ± 0.05	2.89 ± 0.26
	50 mg/L (93.06 µM)	0.33 ± 0.09	2.93 ± 0.44
	100 mg/L (186.12 µM)	0.47 ± 0.06	2.98 ± 0.38
	200 mg/L (372.24 µM)	0.42 ± 0.05	3.06 ± 0.28
	400 mg/L (744.48 µM)	CD	CD

\* symbol presents statistical difference from the negative control group at the level of  $p < 0.05$ . Positive control: Mitomycin C (MMC, 5 × 10<sup>-6</sup> M), CD: Stimulated cells could not be observed due to cellular death.

The results for the antigenotoxicity assays performed via MMC in combination with three different concentrations (25, 50, and 100 mg/L) of boronated compounds are also reflected in Table 3. MMC alone caused a statistically significant increase in CA and MN frequencies when compared to the control group ( $p < 0.05$ ). On the contrary, co-treatment with boronated compounds significantly reduced the frequencies of CAs and MNs, which were elevated by MMC ( $p < 0.05$ ). In fact, in lymphocytes treated with the combination of boronated compounds plus MMC at 72 h (except for 25 mg/L borenium 1), the frequencies of CAs and MNs were significantly reduced compared to the positive control (MMC) at 25, 50, and 100 mg/L. Borinium 4 (55.1% and 48.1%), borinium 5 (51.0% and 36.1%), borenium 1 (60.7% and 50.7%), borenium 2 (70.4% and 57.2%), and borenium 3 (53.1% and 45.2%) minimized the MMC-induced genotoxic damage at different rates, as determined using CA and MN assays, respectively. Furthermore, borenium 2 was found to be the most potent compound for the prevention of DNA damage induced by MMC.

**Table 3.** Frequencies of CAs and MNs in cultured human peripheral lymphocytes after treatment with different concentrations of boronated compounds plus MMC.

Treatment Type	CAs/Cell	MN/1000 Cells
Negative control	0.32 ± 0.04 <sup>a</sup>	2.89 ± 0.18 <sup>a</sup>
Positive control (MMC, 5 × 10 <sup>-6</sup> M)	1.96 ± 0.22 <sup>f</sup>	19.35 ± 2.44 <sup>e</sup>
MMC + 25 mg/L (67.53 μM) Borenium 1	1.33 ± 0.33 <sup>d</sup>	16.80 ± 2.66 <sup>d</sup>
MMC + 50 mg/L (135.06 μM) Borenium 1	0.92 ± 0.25 <sup>c</sup>	10.85 ± 1.88 <sup>bc</sup>
MMC + 100 mg/L (270.12 μM) Borenium 1	0.77 ± 0.21 <sup>bc</sup>	8.55 ± 1.75 <sup>b</sup>
MMC +25 mg/L (76.64 μM) Borenium 2	1.21 ± 0.33 <sup>d</sup>	14.18 ± 2.80 <sup>d</sup>
MMC +50 mg/L (153.28 μM) Borenium 2	0.86 ± 0.17 <sup>c</sup>	10.19 ± 2.36 <sup>bc</sup>
MMC +100 mg/L (306.56 μM) Borenium 2	0.58 ± 0.19 <sup>b</sup>	7.43 ± 1.55 <sup>b</sup>
MMC +25 mg/L (82.25 μM) Borenium 3	1.46 ± 0.28 <sup>e</sup>	15.22 ± 3.08 <sup>d</sup>
MMC +50 mg/L (164.50 μM) Borenium 3	1.22 ± 0.26 <sup>d</sup>	11.73 ± 2.77 <sup>c</sup>
MMC +100 mg/L (329 μM) Borenium 3	0.92 ± 0.24 <sup>c</sup>	9.66 ± 2.12 <sup>b</sup>
MMC + 25 mg/L (55.76 μM) Borinium 4	1.38 ± 0.26 <sup>de</sup>	14.75 ± 2.61 <sup>d</sup>
MMC + 50 mg/L (111.52 μM) Borinium 4	0.97 ± 0.34 <sup>c</sup>	10.69 ± 2.18 <sup>bc</sup>
MMC + 100 mg/L (223.04 μM) Borinium 4	0.88 ± 0.30 <sup>c</sup>	9.02 ± 1.49 <sup>b</sup>
MMC + 25 mg/L (46.53 μM) Borinium 5	1.68 ± 0.28 <sup>e</sup>	15.32 ± 2.52 <sup>d</sup>
MMC + 50 mg/L (93.06 μM) Borinium 5	1.45 ± 0.13 <sup>e</sup>	13.54 ± 2.48 <sup>d</sup>
MMC + 100 mg/L (186.12 μM) Borinium 5	0.96 ± 0.27 <sup>b</sup>	11.08 ± 2.30 <sup>bc</sup>

Different letters in the same column denote significant differences between treatments at the level of  $p < 0.05$ .

In the TAC assay, the available antioxidants in the cultures reduced the colored free radical to its colorless form, and the alternation in absorbance at 660 nm refers to the total antioxidant level in samples from treated and untreated cultures. The presented values in Table 4 correspond to the mean value of at least four different absorbance readings from each culture type. The most significant contribution to the TAC level was observed after treatment with the borenium 2. In fact, borenium 2 elevated TAC levels at a rate of 10.6, as compared to those levels (6.3) in the negative control group. Furthermore, its highest concentration (400 mg/L) increased the TAC levels at a rate of 7.9%. Moreover, the decreasing order of effectiveness for enhancing TAC levels by the tested compounds was as follows: borenium 2 > borenium 1 > borinium 4 > borenium 3 > borinium 5. In fact, the borinium 4, borinium 5, borenium 1, borenium 2, and borenium 3 compounds elevated TAC levels at rates of 33.3%, 49.2%, 42.8%, 101.5%, and 69.8%, respectively. To the contrary, the increasing concentrations of borinium 4 (at 200 and 400 mg/L), borenium 1 (at 400 mg/L), borenium 3 (at 200 and 400 mg/L), and borinium 5 (at 100, 200 and 400 mg/L) caused statistical ( $p < 0.05$ ) reductions of TAC levels (Table 4).

**Table 4.** The determined TAC levels (as mmoleqv./L) after exposure to novel borinium and borenium compounds for 72 h in cultured human blood cells.

Groups	TAC Level	
Negative control	6.3 ± 0.8 <sup>c</sup>	
Positive control (AA, 10 µM)	15.8 ± 1.2 <sup>f</sup>	
Borenium 1	1.56 mg/L (4.22 µM)	6.3 ± 0.7 <sup>c</sup>
	3.12 mg/L (8.44 µM)	6.5 ± 0.8 <sup>c</sup>
	6.25 mg/L (16.88 µM)	6.6 ± 0.6 <sup>c</sup>
	12.5 mg/L (33.77 µM)	7.5 ± 0.7 <sup>cd</sup>
	25 mg/L (67.53 µM)	7.9 ± 0.9 <sup>cd</sup>
	50 mg/L (135.06 µM)	8.5 ± 0.8 <sup>d</sup>
	100 mg/L (270.12 µM)	9.0 ± 1.1 <sup>d</sup>
	200 mg/L (540.24 µM)	6.1 ± 0.7 <sup>c</sup>
Borenium 2	400 mg/L (1080.48 µM)	5.4 ± 0.5 <sup>b</sup>
	1.56 mg/L (4.79 µM)	6.5 ± 0.7 <sup>c</sup>
	3.12 mg/L (9.58 µM)	6.9 ± 0.7 <sup>c</sup>
	6.25 mg/L (19.16 µM)	7.5 ± 0.9 <sup>cd</sup>
	12.5 mg/L (38.32 µM)	8.2 ± 1.0 <sup>cd</sup>
	25 mg/L (76.64 µM)	9.7 ± 0.9 <sup>d</sup>
	50 mg/L (153.28 µM)	10.6 ± 1.3 <sup>d</sup>
	100 mg/L (306.56 µM)	12.7 ± 1.4 <sup>e</sup>
200 mg/L (613.12 µM)	7.9 ± 0.9 <sup>cd</sup>	
Borenium 3	400 mg/L (1226.24 µM)	6.8 ± 0.7 <sup>c</sup>
	1.56 mg/L (5.14 µM)	6.1 ± 0.8 <sup>c</sup>
	3.12 mg/L (10.28 µM)	6.4 ± 0.7 <sup>c</sup>
	6.25 mg/L (20.56 µM)	6.6 ± 0.8 <sup>c</sup>
	12.5 mg/L (41.13 µM)	6.9 ± 0.7 <sup>c</sup>
	25 mg/L (82.25 µM)	8.9 ± 1.0 <sup>d</sup>
	50 mg/L (164.50 µM)	10.7 ± 1.1 <sup>d</sup>
	100 mg/L (329 µM)	6.5 ± 0.6 <sup>c</sup>
200 mg/L (658 µM)	5.8 ± 0.7 <sup>b</sup>	
Borinium 4	400 mg/L (1316 µM)	5.3 ± 0.5 <sup>b</sup>
	1.56 mg/L (3.49 µM)	6.2 ± 0.8 <sup>c</sup>
	3.12 mg/L (6.97 µM)	6.4 ± 0.6 <sup>c</sup>
	6.25 mg/L (13.94 µM)	6.5 ± 0.5 <sup>c</sup>
	12.5 mg/L (27.88 µM)	7.1 ± 0.7 <sup>c</sup>
	25 mg/L (55.76 µM)	7.7 ± 0.8 <sup>cd</sup>
	50 mg/L (111.52 µM)	8.4 ± 1.0 <sup>d</sup>
	100 mg/L (223.04 µM)	7.8 ± 0.9 <sup>cd</sup>
200 mg/L (446.08 µM)	5.7 ± 0.6 <sup>b</sup>	
Borinium 5	400 mg/L (892.16 µM)	5.1 ± 0.6 <sup>ab</sup>
	1.56 mg/L (2.91 µM)	6.6 ± 0.9 <sup>c</sup>
	3.12 mg/L (5.82 µM)	6.9 ± 0.7 <sup>c</sup>
	6.25 mg/L (11.63 µM)	7.5 ± 0.8 <sup>cd</sup>
	12.5 mg/L (23.26 µM)	8.5 ± 0.9 <sup>d</sup>
	25 mg/L (46.53 µM)	9.4 ± 1.0 <sup>d</sup>
	50 mg/L (93.06 µM)	6.0 ± 0.6 <sup>bc</sup>
	100 mg/L (186.12 µM)	5.8 ± 0.7 <sup>b</sup>
200 mg/L (372.24 µM)	4.2 ± 0.5 <sup>a</sup>	
400 mg/L (744.48 µM)	3.6 ± 0.4 <sup>a</sup>	

Different letters in the same column denote significant differences between treatments at the level of  $p < 0.05$ .

#### 4. Discussion

Previous reports revealed that incorporation of cationic boron centers in organic heterocycles or transition-metal metallocenes could provide opportunities for the development of novel biomedical materials with superior redox activity and optical properties [21].

In our study, new boron-containing molecules were introduced into the family of ionic compounds, and the synthesis and characterization of the compounds were carried out. In the first step, the synthesis of boronium ionic liquids was performed. Briefly, boronium amino acid salts were obtained by anion exchange over the boronium iodide compounds obtained. At this stage, silver salts of various amino acid derivatives, which were dissolved in the appropriate solvent, were slowly added to the solution of boronium iodide salts dissolved in the appropriate solvent at room temperature in the dark, and the targeted compounds were obtained. In the second phase of our study, the synthesis of borenium ionic liquids was performed. When the  $^1\text{H-NMR}$  and  $^{13}\text{C-NMR}$  spectra of the synthesized compounds are examined, the presence of the peaks indicates that the synthesis of the targeted compounds has been successfully achieved.

The cytotoxic, genotoxic, and oxidative damage potentials of the newly synthesized borenium and borinium compounds were investigated in this study. The cytotoxic effects of these new ionic liquids of B have been demonstrated as a result of exposing U87MG, SHSY-5Y, PC-3, and Detroit-562 cancer cells to these liquids. Our findings revealed that the PC-3 cells were found to be more sensitive to boronated compounds in comparison to other cell lines. Furthermore, borinium **5** was found to be the most potent among them (Table 1). Similar to our findings, previous cellular and epidemiological studies revealed that B (as boric acid, BA) did not induce carcinogenicity. Moreover, supplementation with B might decrease the risk for prostate and brain cancers [44,45]. The underlying mechanisms of anticancer properties by BCCs are still unclear. It has been found that a Ca signal has a regulatory role in cell profiling and very little attention has been paid to cancer-preventive therapies. In a previous study, it was determined that supplementation with high amounts of boron (50 mg/L) inhibited the proliferation of human DU-145 prostate cancer cells via decreasing intracellular Ca signals and stores [46]. Another underlying anticancer mechanism due to B supplementation was associated with inhibition of serine proteases, such as prostate specific antigens and the affinity of BA to hydroxyl groups [47]. Again, BA was shown to contribute to proliferative inhibition via dose-dependent reductions in the expression of cyclins A-E and MAPK proteins in DU-145 prostate, HeLa cervical cancer, and DLD-1 colorectal adenocarcinoma cells [48–50]. In addition, ferroptosis (a new type of iron-dependent cell death) was characterized by intracellular iron ion accumulation. Moreover, B was reported to modulate the ferroptosis in HepG2 hepatocellular carcinoma cells, hence it could serve as a sensitizer to anticancer chemotherapeutics [51]. In this context, the induction of ferroptosis might be one of the possible underlying mechanisms for anticancerogenic action by introduced borenium or borinium compounds. In fact, in a recent investigation it was reported that BA (up to 1500 mg/L) was able to trigger both ferroptosis and apoptosis in C6 glioma cells and affected the ephratorin–neuropilin signaling pathway [52]. AKT phosphorylation by B could be proposed as another associated mechanism for explaining the anticancer properties of BCCs in hepatocellular carcinoma and glioma cases [53,54].

The determined  $\text{IC}_{50}$  values after exposure to boronated compounds in several cancer cell lines clearly revealed that the effects of boron compounds on different cancer types might be variable (Table 1). In a recent investigation, BA and borax (BX) enhanced the apoptosis in human DMS-114 lung cancer cells by upregulating pro-apoptotic genes, such as *Bax* and *Casp-3*. In addition, these borates modulated anti-apoptotic genes, such as *BIRC2*, *BIRC5*, and *Bcl-2*, and induced cell cycle arrest at the G2/M phase [55]. Similar to our findings, different concentrations (150–3000 mg/L) of BA exerted cytotoxic action on U87MG and T98G glioblastoma (GBM) cells with high  $\text{IC}_{50}$  values (1050 mg/L) [56]. In this regard, the novel boron compounds, especially borinium **5**, might be novel sources for anti-GBM therapies with their moderate cytotoxic  $\text{IC}_{50}$  values without damaging healthy cells (Table 1). Alongside the cell cycle arresting and apoptosis modulating features, BCCs were found to interfere with other key tumorigenic pathways involving glycolysis, molybdenum Fe–S-containing flavin hydroxylases, and intratumoral IGF-I levels—as well as transient receptor potential in glioma cases [57].

In the present investigation, it was revealed that the tested borenium and borinium compounds exerted non-genotoxic features. All of the concentrations of these compounds did not induce the formations of CAs or MNs as compared to untreated cells (Table 2). In accordance with the present findings, the World Health Organization propounded that genotoxic damage cannot be associated with exposure to B in both animals and humans [58]. Likewise, previous multiplexed in vitro studies proved the non-genotoxic features of certain commercially important BCCs, including potassium tetraborate, BX, ulexite, colemanite, and B-ionic liquids [30]. In addition to these in vitro studies, the three orally administered BA doses (5, 10, and 20 mg/kg) did not cause DNA damage in the mononuclear leukocytes of rats [59]. Relatively high doses of BX (100 mg/kg) also did not induce DNA strand breaks in rat lymphocyte cells; hence, BX was reported to have a non-genotoxic nature [60]. Our findings also revealed that the genotoxic damages in the co-treated with MMC and borenium or borinium compounds monitored via CA and MN frequencies were lower than the solely MMC-applied group (Table 3).

Our findings indicated that both the borenium and borinium compounds also have in vitro antigenotoxic action potential. Cross-linking to DNA occurred in MMC-treated human cells, thus MMC application induced persistent DNA double-strand breaks [61]. BCCs, such as boric acid (BA), reduced the formation of DNA double-strand breaks and prevented chromosome loss of cells [62]. Moreover, ataxia–telangiectasia-mutated (ATM) protein kinase was reported to initiate DNA repair after formation of double-strand breaks by mutagens [63]. Furthermore, BA was able to lead to ATM activation and a DNA damage response of the cells [62]. The observed antigenotoxicity action by the borenium and borinium compounds (Table 3) could also be attributed to their direct chemical interaction before MMC caused genotoxic damage [64]. Previous evidence exerted that antioxidant-featured substances can eliminate ROS before these reactive chemicals interact with DNA and change in a DNA sequence [64]. Consistent with our findings, previous reports suggested that BCCs have antigenotoxic effects against several genotoxic agents, such as titanium, aluminum, aflatoxins, lead, bismuth, arsenic, and cadmium [65]. The observed antigenotoxic action by several boron compounds on animal or cell culture models could be primarily linked to their antioxidant properties. In fact, the antioxidative features of several BCCs were associated with their antigenotoxic action [66,67].

The results of our study put forward that the borinium compounds yielded the best results in terms of TAC values when compared to the untreated control treatment. Moreover, concentrations below 100 mg/L supported the TAC levels of human blood cultures (Table 4). Various experimental studies indicated that erythrocytes were especially susceptible to oxidative stress. Furthermore, B regulated the activity of cellular antioxidant enzymes, such as oxidoreductases, aldehyde dehydrogenase, xanthine oxidase, and cytochrome b5 oxidoreductase, and affected coagulation factors, such as glyceraldehyde-3-phosphate dehydrogenase and lactate dehydrogenase, by interacting with enzymes, such as serine proteases [61,68]. It was also observed that BA could alter the oxidative metabolism in animals. However, the eventual mechanisms of this change are still unclear [69]. The primer findings in this field manifested that several BCCs at relatively low doses (<80 mg/L) supported antioxidant enzyme activities in human whole blood cultures. Indeed, the activities of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPX), glutathione-S-transferase (GST), glucose-6-phosphate dehydrogenase (G6PD), and the levels of total glutathione (TGSH), as well as TAC levels, were strengthened by application with certain BCCs, including BA, BX, and calcium borates [8,50,70].

In conclusion, the five novel borenium and borinium compounds exerted key biological functions involving (I) antioxidant (supporting TAC levels up to 101.5%), (II) non-genotoxic (having no clastogenic and eugenic effects), (III) antigenotoxic (minimizing MMC induced genotoxic damages in different rates up to 70.4%), and (IV) moderate cytotoxic ( $IC_{50}$  values < 200 mg/L on glioblastoma, neuroblastoma, prostate, and pharyngeal cancer cell lines and ranging from 41.941 to 179.662) properties. Our findings will contribute to further investigations on the biomedical evaluation of these borenium and borinium

derivatives. They have great potential to be employed for nutritional, pharmacological, and medicinal purposes. These boron compounds deserve to be studied further in cancer treatment and nutrition due to their multi-biological functions. In the meantime, concentration adjustment should be undertaken while using these new boron compounds to promote health benefits.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/inorganics11080324/s1>. Figure S1: IR spectrum of dicyclohexyl borenium dimethyl amino pyridine trifluoro methane sulphonate (Borenium 1); Figure S2: <sup>1</sup>H-NMR spectrum of dicyclohexyl borenium dimethyl amino pyridine trifluoro methane sulphonate (Borenium 1); Figure S3: <sup>13</sup>C-NMR spectrum of dicyclohexyl borenium dimethyl amino pyridine trifluoro methane sulphonate (Borenium 1); Figure S4: IR spectrum of dicyclohexyl borenium dimethyl amino pyridine 2-amino-4-methylpentanoate (Borenium 2); Figure S5: <sup>1</sup>H-NMR spectrum of dicyclohexyl borenium dimethyl amino pyridine 2-amino-4-methylpentanoate (Borenium 2); Figure S6: <sup>13</sup>C-NMR spectrum of dicyclohexyl borenium dimethyl amino pyridine 2-amino-4-methylpentanoate (Borenium 2); Figure S7: IR spectrum of bisdimethyl amino borenium dimethyl amino pyridine trifluoro acetate (Borenium 3); Figure S8: <sup>1</sup>H-NMR spectrum of bisdimethyl amino borenium dimethyl amino pyridine trifluoro acetate (Borenium 3); Figure S9: <sup>13</sup>C-NMR spectrum of bisdimethyl amino borenium dimethyl amino pyridine trifluoro acetate (Borenium 3); Figure S10: IR spectrum of dicyclohexyl borinium trifluoro methane sulphonate (Borinium 4); Figure S11: <sup>1</sup>H-NMR spectrum of dicyclohexyl borinium trifluoro methane sulphonate (Borinium 4); Figure S12: <sup>13</sup>C-NMR spectrum of dicyclohexyl borinium trifluoro methane sulphonate (Borinium 4); Figure S13: IR spectrum of Bis (2-methyl-1H-imidazol-3-yl) dihydroboronium iodide (Borinium 5); Figure S14: <sup>1</sup>H-NMR spectrum of Bis (2-methyl-1H-imidazol-3-yl) dihydroboronium iodide (Borinium 5); Figure S15: <sup>13</sup>C-NMR spectrum of bis(1,2-dimethyl-1H-imidazol-3-yl) dihydroboronium iodide (Borinium 5); Figure S16: Cytotoxic activity of compounds in SHSY-5Y cells; Figure S17: Cytotoxic activity of compounds in U87MG cells; Figure S18: Cytotoxic activity of compounds in PC-3 cells; Figure S19: Cytotoxic activity of compounds in Detroit-562 cells; Figure S20: Cytotoxic activity of compounds in cultured human whole blood cells.

**Author Contributions:** Conceptualization, S.B.O., A.M. and H.T.; methodology, S.B.O., A.T., H.I.U. and H.T.; software, S.B.O.; investigation, S.B.O., H.I.U. and H.T.; writing-original draft preparation, S.B.O. and H.T.; writing-review and editing, S.B.O., H.I.U., A.T., A.M. and H.T. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding author.

**Acknowledgments:** The synthesis of the compounds was obtained with the results of the TUBITAK project with the code number 114M933.

**Conflicts of Interest:** The authors declare no conflict of interest.

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