

Optimization of Extraction Parameters for Enhanced Recovery of Bioactive Compounds from Quince Peels Using Response Surface Methodology

Vassilis Athanasiadis, Theodoros Chatzimitakos, Eleni Bozinou, Konstantina Kotsou, Dimitrios Palaogiannis and Stavros I. Lalas *

Department of Food Science & Nutrition, University of Thessaly, Terma N. Temponera Str.,
43100 Karditsa, Greece

* Correspondence: slalas@uth.gr; Tel.: +30-24410-64783

2.1 Chemicals and reagents

All of the solvents were purchased from Carlo Erba in Val de Reuil, France, and were at least of HPLC grade. Penta (Prague, Czech Republic) provided the gallic acid, anhydrous sodium carbonate, 2,4,6-tri-2-pyridinyl-1,3,5-triazine (TPTZ), and Folin-Ciocalteu reagent. Chemical standards for the HPLC-based determination of polyphenols [i.e., chlorogenic acid (3-O-caffeoylquinic acid), neochlorogenic acid (5-O-caffeoylquinic acid), cryptochlorogenic acid (4-O-caffeoylquinic acid), rutin, quercetin 3-O-galactoside, and kaempferol 3-O- β -rutinoside], aluminum chloride, iron (III) chloride, hydrochloric acid, ascorbic acid, and trichloroacetic acid were purchased from Sigma-Aldrich (Steinheim, Germany). Deionized water was used for all conducted experiments.

2.4. Determination of total polyphenol content (TPC)

Briefly, 100 μ L of quince peel extracts were mixed with 100 μ L of Folin-Ciocalteu reagent in a 1.5 mL Eppendorf tube. After 2 min, 800 μ L of Na₂CO₃ solution (5% *w/v*) was added and the solutions were incubated at 40 °C for 20 min. Eventually, the absorbance at 740 nm was determined using a Shimadzu spectrophotometer (UV-1700, Shimadzu Europa GmbH, Duisburg, Germany). Gallic acid was used to create a calibration curve (10–80 mg/L) using a standard compound. The TPC (C_{TP}) was expressed as mg gallic acid equivalents (GAE) per L. Equation (S1) was used to express the extraction yield in terms of total polyphenols (Y_{TP}) as mg GAE per g of dry weight (dw):

$$Y_{TP} \text{ (mg GAE/g dw)} = \frac{C_{TP} \times V}{w} \quad (S1)$$

where V is the volume of the extraction medium (in L) and w is the dry weight of the sample (in g).

2.5. Determination of total flavonoid content (TFC)

100 μ L of the extract was mixed with 860 μ L of aqueous ethanol (35% *v/v*) and 0.04 μ L of a reagent containing 0.5 M sodium acetate and 5% *w/v* aluminum chloride in an Eppendorf tube. The resulting mixture was vortexed and reacted at room temperature for 30 minutes. The absorbance at 415 nm was then measured against a suitable blank. The concentration of total flavonoids (TFC), expressed in mg/L, was estimated by calibration curve (20–100 mg/L), which was constructed by plotting known rutin solutions against absorbance at 415 nm. The TFC was expressed as mg rutin equivalents (RtE) per g dw, using the following Equation (S2):

$$\text{TFC (mg RtE/g dw)} = \frac{C_{TFC} \times V}{w} \quad (S2)$$

where, V is the volume of the extraction medium (in L), and w is the dry weight of the sample (in g).

2.6. Ferric reducing antioxidant power (FRAP) assay

FeCl_3 solution (4 mM in 0.05 M HCl) was combined with 0.05 mL of the extract in an Eppendorf tube, and the mixture was then incubated at 37 °C for 30 min. Following the addition of 0.90 mL of the TPTZ solution (1 mM in 0.05 M HCl), the absorbance at 620 nm was measured after 5 minutes. Ascorbic acid was used as a standard compound to produce a calibration curve (C_{AA} , 50–500 $\mu\text{mol/L}$ in 0.05 M HCl). Equation (S3) was used to calculate the ferric reducing antioxidant power (P_R) as μmol ascorbic acid equivalents (AAE) per g of dw:

$$P_R (\mu\text{mol AAE/g dw}) = \frac{C_{AA} \times V}{w} \quad (\text{S3})$$

where V is the volume of the extraction medium (in L) and w is the dry weight of the sample (in g).

2.7. DPPH Radical Scavenging Activity

Briefly, 25 μL of the prepared extract was added to 975 μL of 100 μM DPPH solution, thoroughly mixed, and the solution's absorbance was measured at 515 nm ($A_{515(i)}$) and after 30 min of incubation in the dark ($A_{515(f)}$). Equation (S4) was utilized to compute the antiradical activity (A_{AR}):

$$A_{AR} (\mu\text{mol DPPH/g dw}) = \frac{\Delta A}{\varepsilon \times l \times C} \times Y_{TP} \quad (\text{S4})$$

where $\Delta A = A_{515(i)} - A_{515(f)}$; ε (DPPH) = $11,126 \times 10^{-6} \mu\text{M}^{-1} \text{cm}^{-1}$; $C = C_{TP} \times 0.025$; Y_{TP} is the total polyphenol yield of the extract (mg/g), and l is the path length (1 cm).

2.9. HPLC-based determination of the chlorogenic acid content and other phenolic compounds

The analysis was conducted using a Shimadzu CBM-20A liquid chromatograph and a Shimadzu SPD-M20A diode array detector (both supplied by Shimadzu Europa GmbH, Duisburg, Germany). The compounds were separated using a Phenomenex Luna C18(2) column from Phenomenex Inc. in Torrance, California, maintained at 40 °C (100 Å, 5 μm , 4.6 \times 250 mm). The mobile phase consisted of 0.5% aqueous formic acid (A) and a mixture of 0.5% formic acid in acetonitrile/water (6:4) (B). The gradient program used was as follows: 0% B to 40% B, then to 50% B in 10 min, to 70% B in another 10 min and then held constant for 10 min. The flow rate of the mobile phase was 1 mL/min. The retention time and absorbance spectrum were compared to those of pure chemical standards to identify the compounds and then quantified using calibration curves (0–50 $\mu\text{g/mL}$).

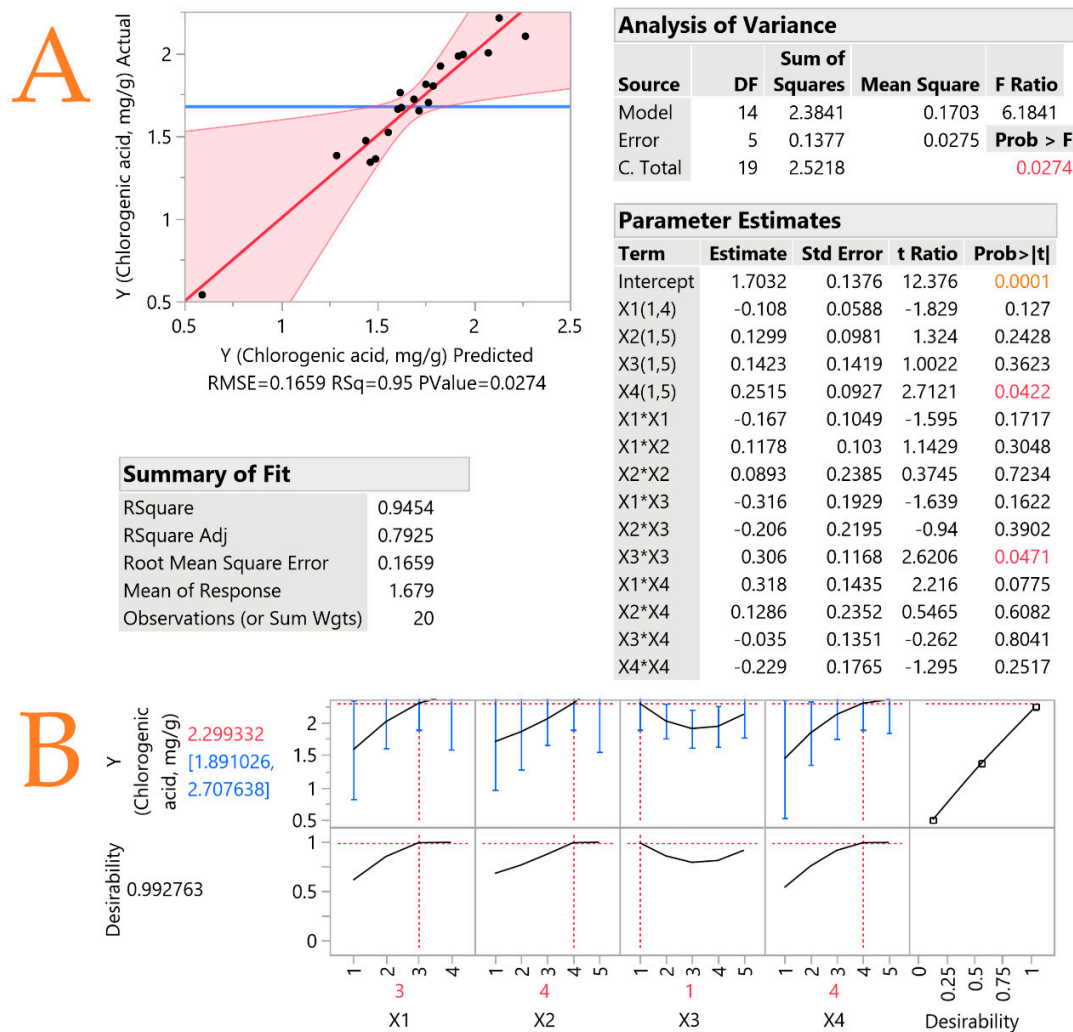


Figure S1. Plots A and B show plots of the actual response versus the predicted response (Chlorogenic acid, mg/g) and the desirability function for the optimization of quince peel extracts performed with hydroethanolic solutions, respectively. Inset tables provide statistics related to the evaluation of the resulting model, and colored values indicate statistically significant values.

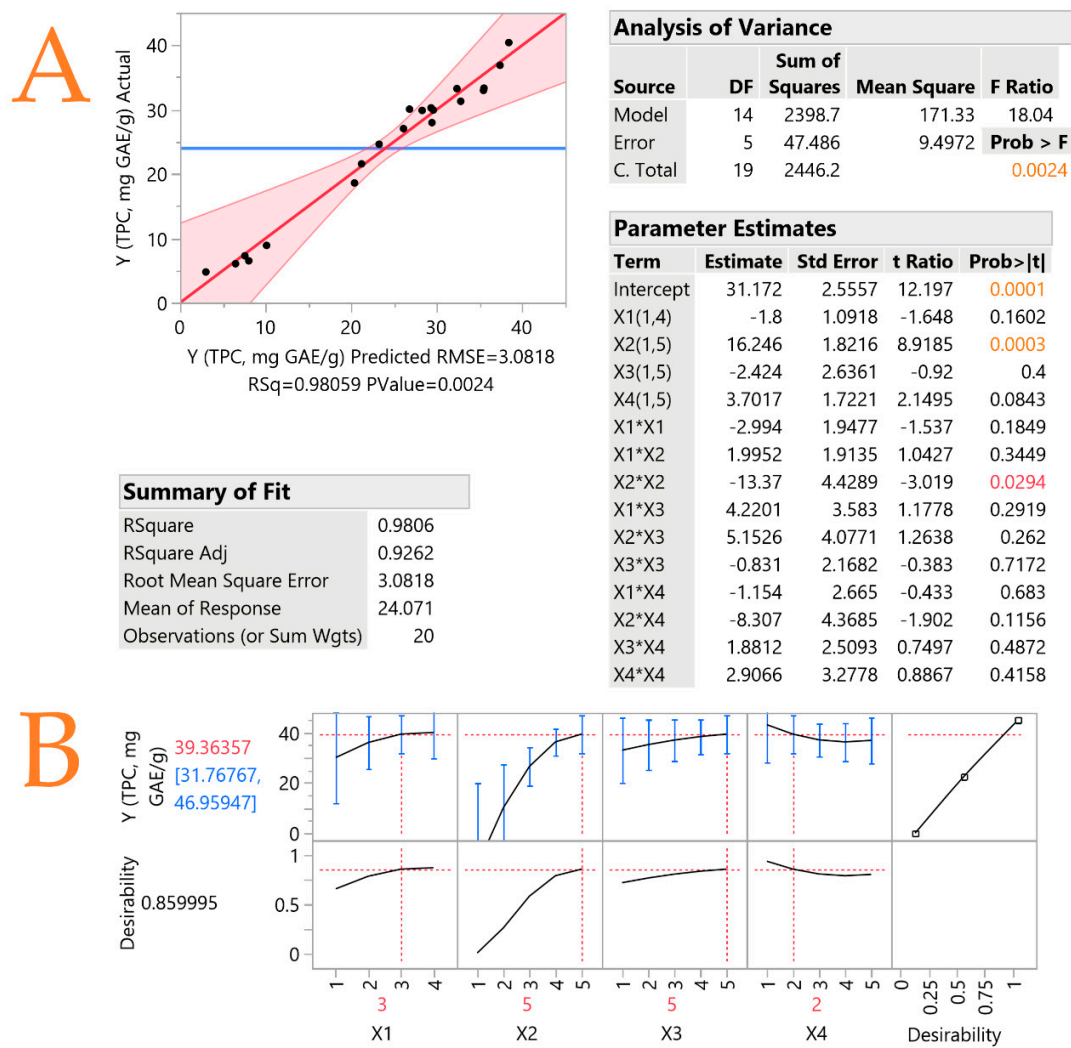


Figure S2. Plots A and B show plots of the actual response versus the predicted response (Total Polyphenol Content-TPC, mg GAE/g) and the desirability function for the optimization of quince peel extracts performed with hydroethanolic solutions, respectively. Inset tables provide statistics related to the evaluation of the resulting model, and colored values indicate statistically significant values.

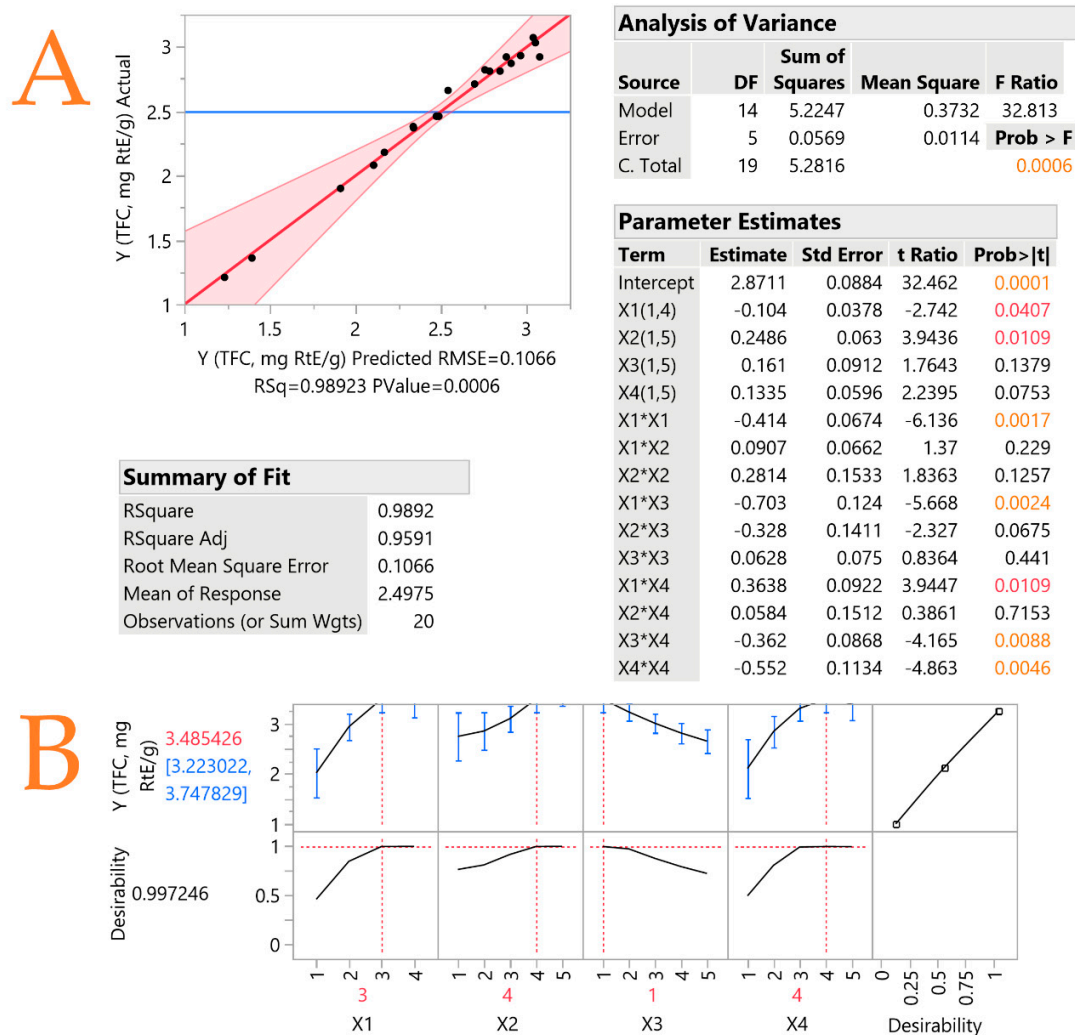


Figure S3. Plots A and B show plots of the actual response versus the predicted response (Total Flavonoid Content-TFC, mg RtE/g) and the desirability function for the optimization of quince peel extracts performed with hydroethanolic solutions, respectively. Inset tables provide statistics related to the evaluation of the resulting model, and colored values indicate statistically significant values.

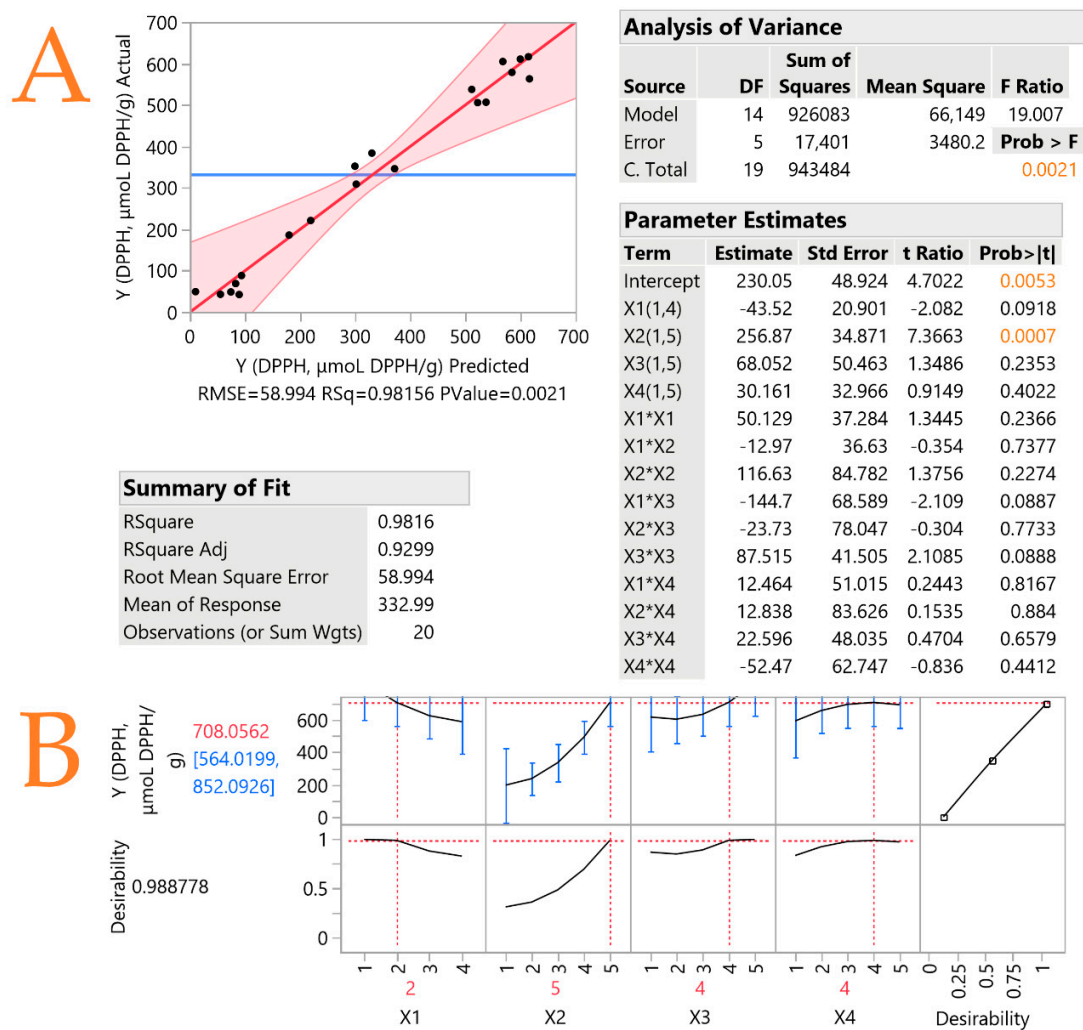


Figure S5. Plots A and B show plots of the actual response versus the predicted response (DPPH, $\mu\text{mol DPPH/g}$) and the desirability function for the optimization of quince peel extracts performed with hydroethanolic solutions, respectively. Inset tables provide statistics related to the evaluation of the resulting model, and colored values indicate statistically significant values.

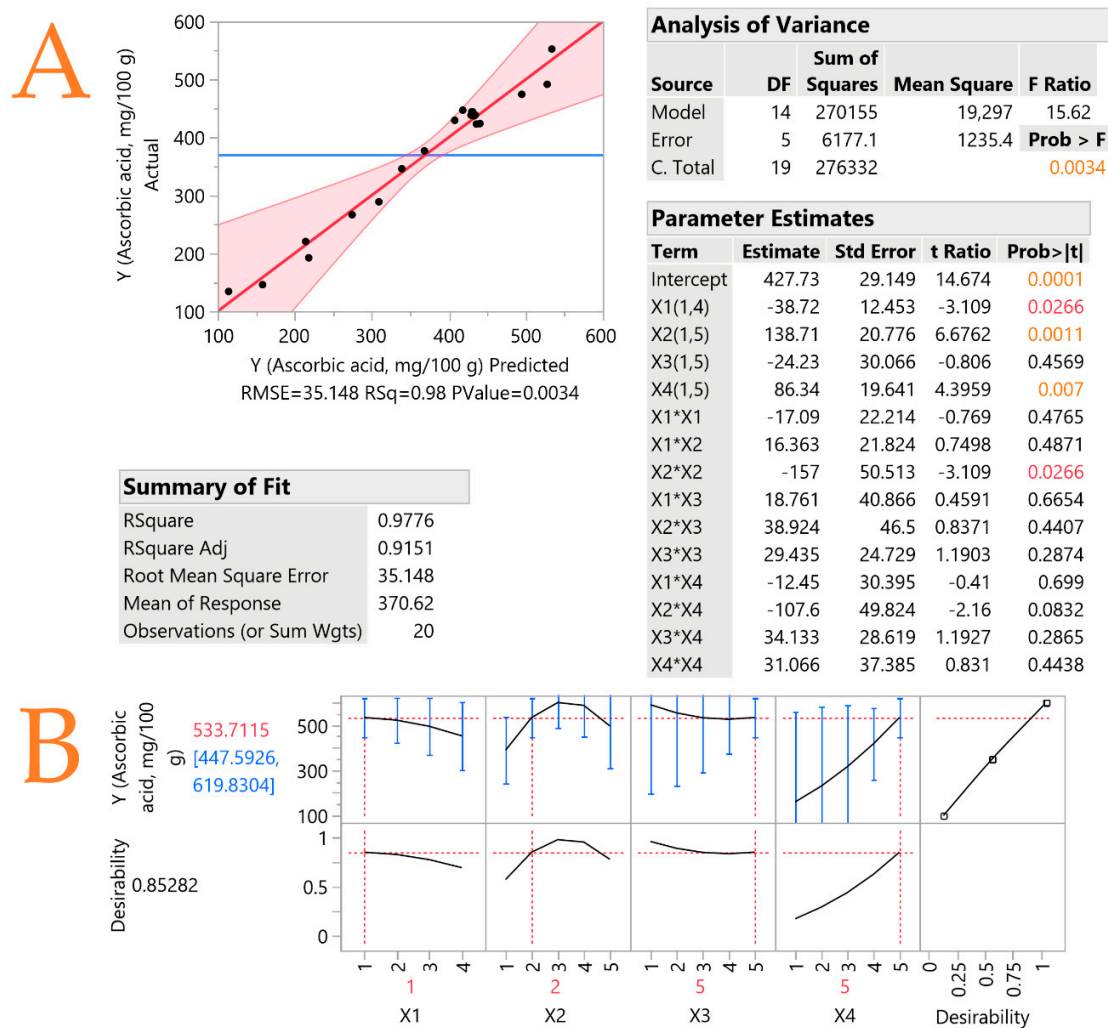


Figure S6. Plots A and B show plots of the actual response versus the predicted response (Ascorbic acid, mg/100 g) and the desirability function for the optimization of quince peel extracts performed with hydroethanolic solutions, respectively. Inset tables provide statistics related to the evaluation of the resulting model, and colored values indicate statistically significant values.

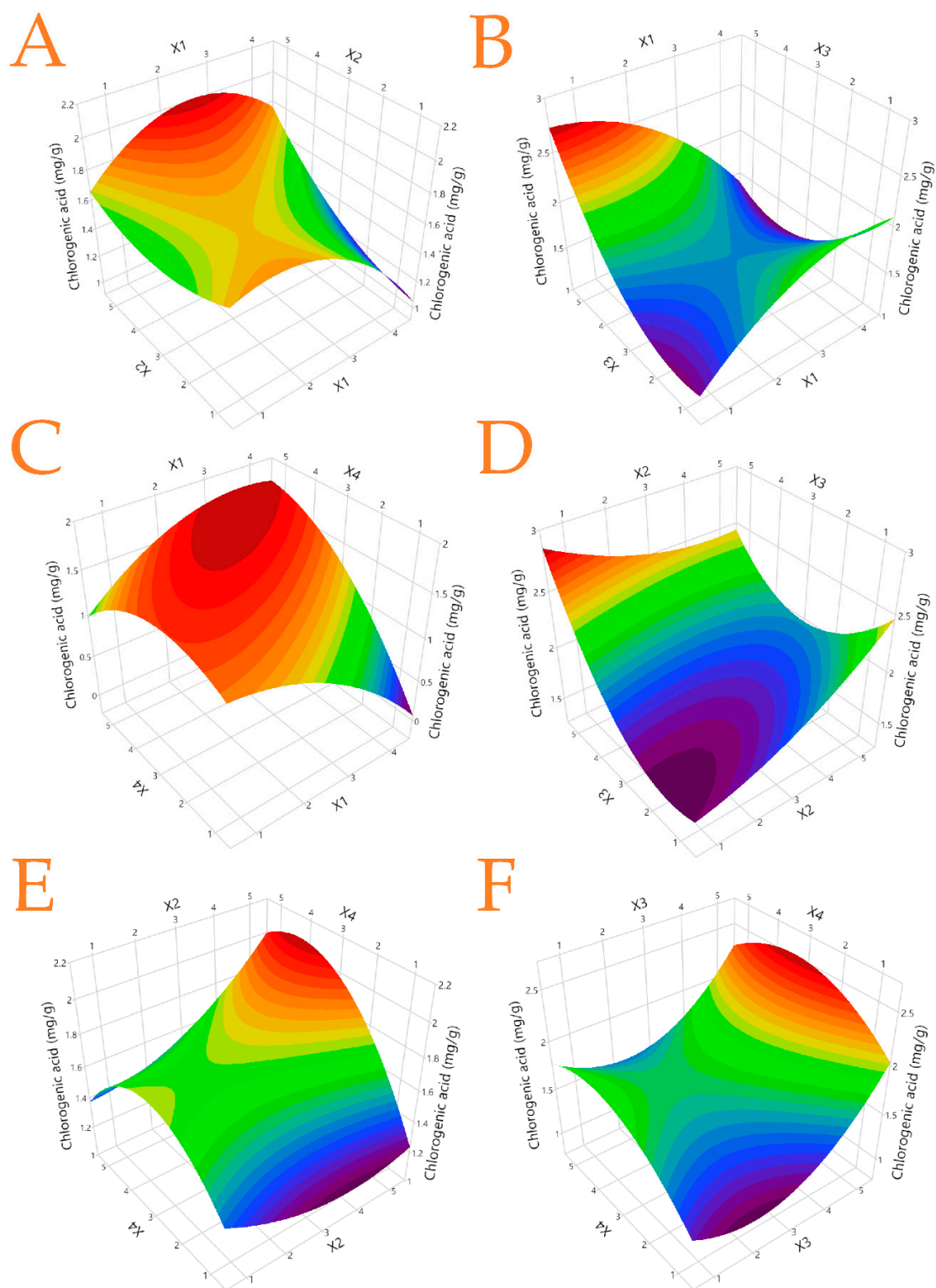


Figure S7. 3D graphs depicting the effect of the process variables considered in the response (Chlorogenic acid, mg/g), to optimize the extraction of the quince peel, using various extraction techniques and hydroethanolic solutions. Plot (A), covariation of X₁ and X₂; plot (B), covariation of X₁ and X₃; plot (C), covariation of X₁ and X₄; plot (D), covariation of X₂ and X₃; plot (E), covariation of X₂ and X₄; plot (F), covariation of X₃ and X₄.

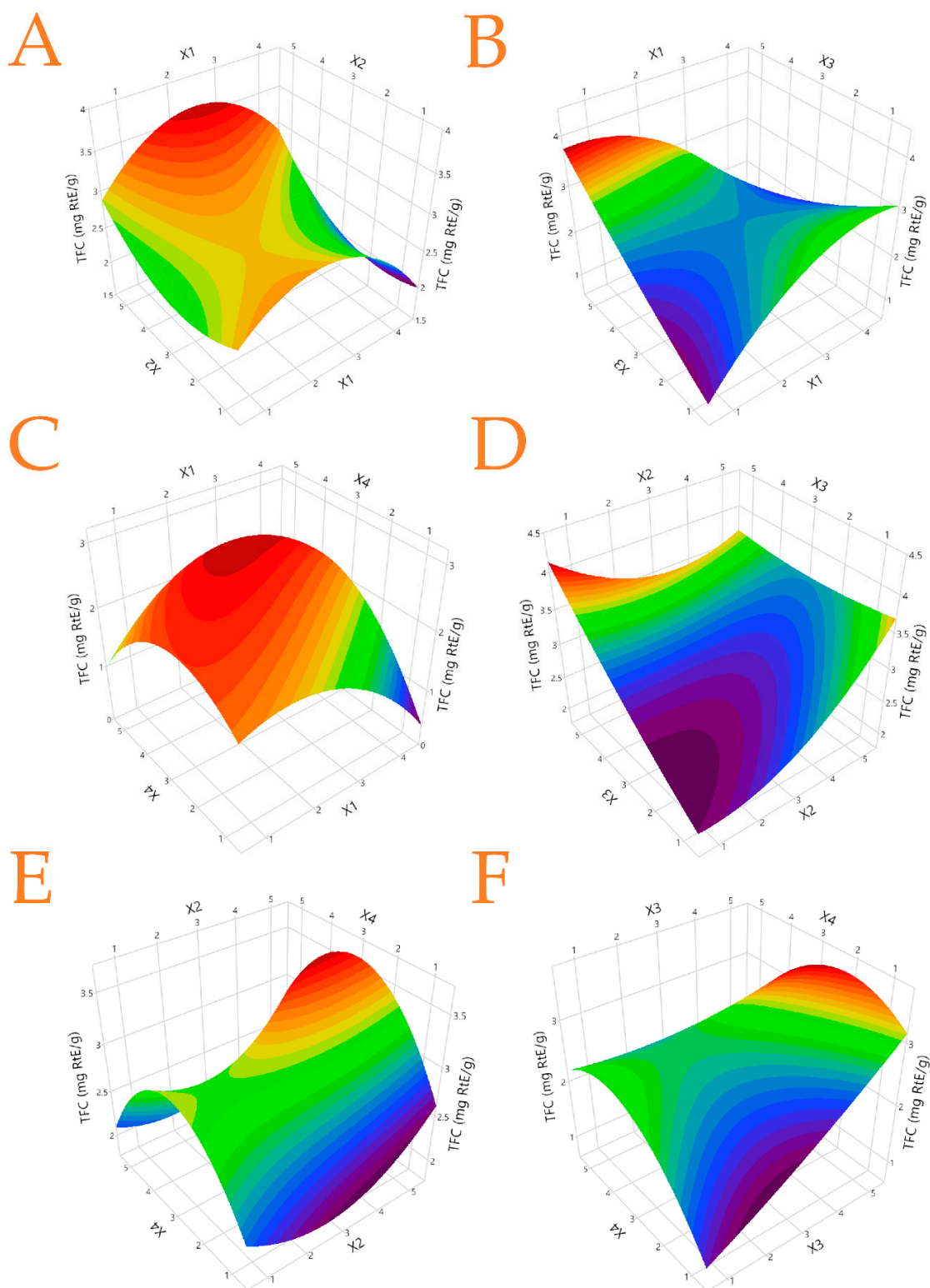


Figure S8. 3D graphs depicting the effect of the process variables considered in the response (Total Flavonoid Content-TFC, mg RtE/g), to optimize the extraction of the quince peel, using various extraction techniques and hydroethanolic solutions. Plot (A), covariation of X_1 and X_2 ; plot (B), covariation of X_1 and X_3 ; plot (C), covariation of X_1 and X_4 ; plot (D), covariation of X_2 and X_3 ; plot (E), covariation of X_2 and X_4 ; plot (F), covariation of X_3 and X_4 .

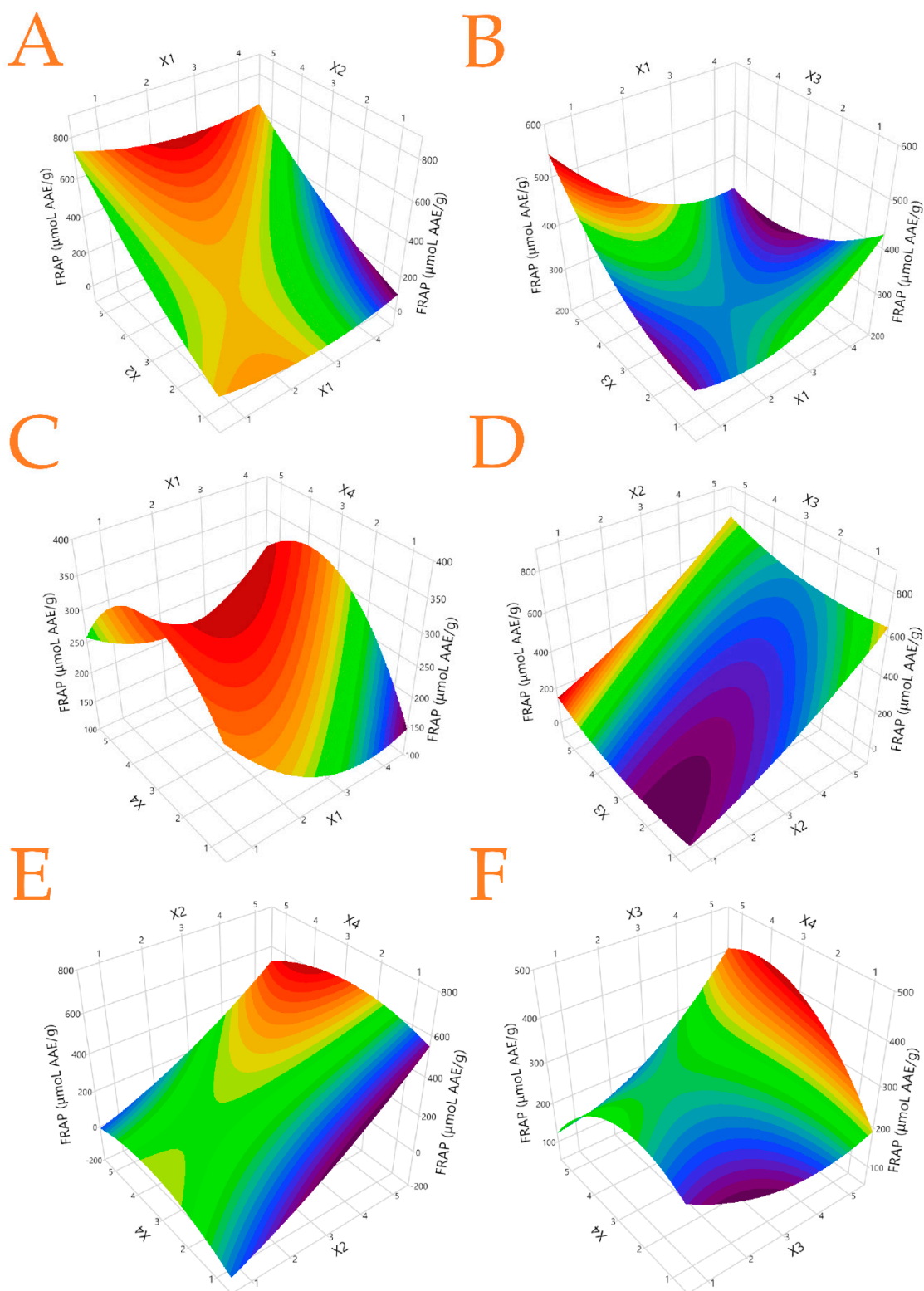


Figure S9. 3D graphs depicting the effect of the process variables considered in the response (FRAP, $\mu\text{mol AAE/g}$), to optimize the extraction of the quince peel, using various extraction techniques and hydroethanolic solutions. Plot (A), covariation of X_1 and X_2 ; plot (B), covariation of X_1 and X_3 ; plot (C), covariation of X_1 and X_4 ; plot (D), covariation of X_2 and X_3 ; plot (E), covariation of X_2 and X_4 ; plot (F), covariation of X_3 and X_4 .

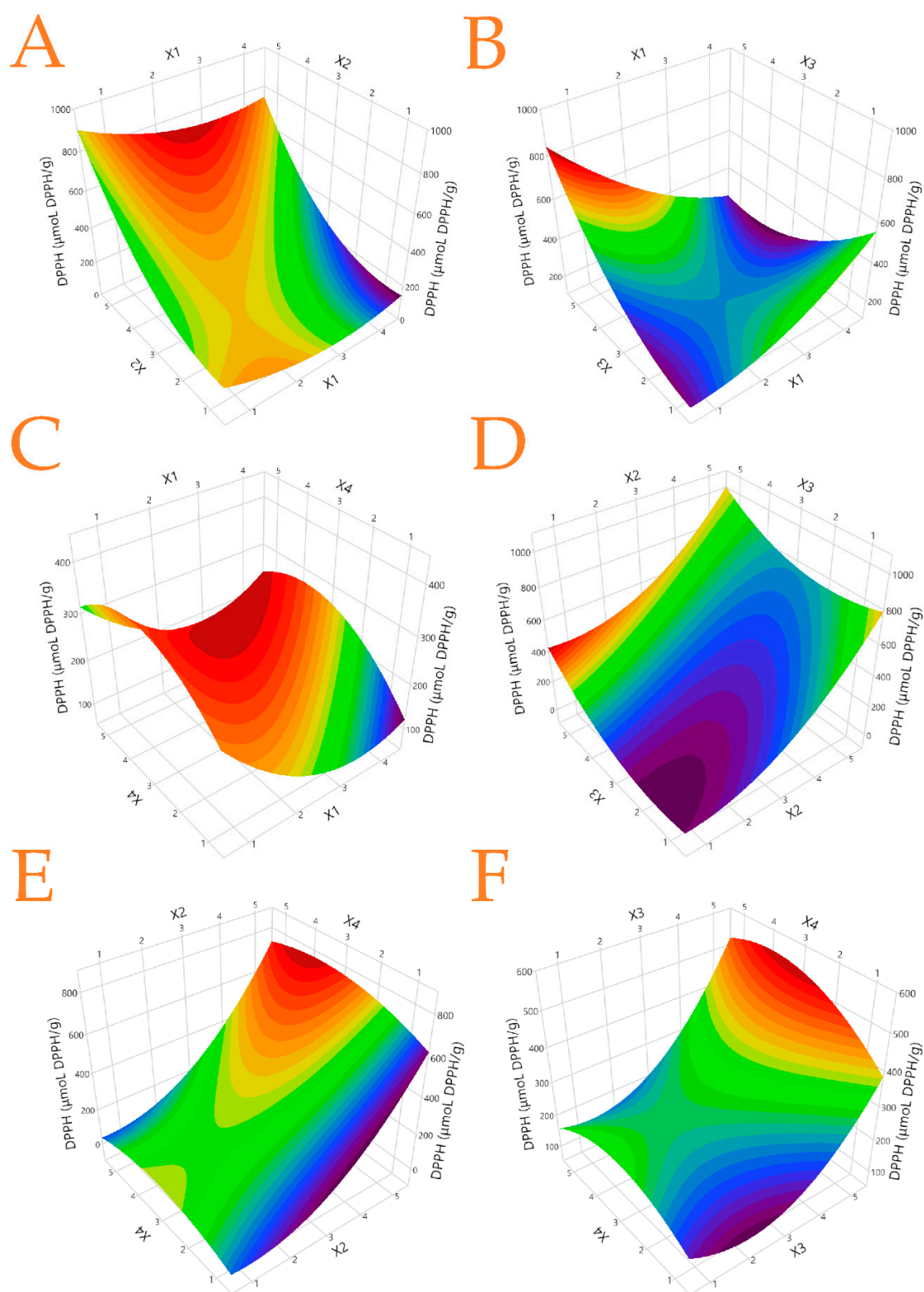


Figure S10. 3D graphs depicting the effect of the process variables considered in the response ($DPPH$, $\mu\text{mol DPPH/g}$), to optimize the extraction of the quince peel, using various extraction techniques and hydroethanolic solutions. Plot (A), covariation of X_1 and X_2 ; plot (B), covariation of X_1 and X_3 ; plot (C), covariation of X_1 and X_4 ; plot (D), covariation of X_2 and X_3 ; plot (E), covariation of X_2 and X_4 ; plot (F), covariation of X_3 and X_4 .