

## Supplementary Materials

**Table S1:** Tips and tricks of successful MCAs analysis by our experience.

Problem	Recommendations
Cells do not form MCAs	<ul style="list-style-type: none"> <li>- Increase cell density.</li> <li>- Centrifugation step is crucial to gather the cells.</li> </ul>
MCAs are too small, easily lost during their manipulation	<ul style="list-style-type: none"> <li>- Increase cell density.</li> </ul>
Irregular MCAs formation due to the presence of fibres into the wells	<ul style="list-style-type: none"> <li>- Use a laminar hood with good filters.</li> <li>- Use synthetic lab coats.</li> <li>- Keep the plate uncovered the least time possible.</li> <li>- Plate the cells using multichannel pipettes to be faster.</li> </ul>
MCAs lost or damaged during medium change	<ul style="list-style-type: none"> <li>- Use a 100 <math>\mu</math>l micropipette and place the tip against the wall of the well to aspirate and replace the medium.</li> <li>- Both aspiration and medium replacement should be done very slowly and carefully, as too much pressure can cause MCAs' damage.</li> <li>- Lean the plate around 30°, since this helps seeing the MCAs and prevent their aspiration.</li> </ul>
MCAs lost or damaged during harvesting for optical and electron microscopy	<ul style="list-style-type: none"> <li>- Carefully remove the medium and replace it with the proper fixative, in the well. Incubate for 10 min before harvesting. Be careful not to leak the fixative</li> </ul>

	<p>into other wells destined for other types of proxies. In this step leave the lid open because fixatives release vapours than can interfere with other techniques.</p> <ul style="list-style-type: none"> <li>- For harvesting use a 1000 µl micropipette with a sectioned tip that offers a larger circumference and prevents the MCAs' damage.</li> <li>- The harvesting procedure should be performed slowly and carefully.</li> <li>- Transfer to an Eppendorf tube containing the appropriate fixative.</li> </ul>
<p>MCAs lost or damaged during processing for optical and electron microscopy</p>	<ul style="list-style-type: none"> <li>- After the fixation step, embed the MCAs in histogel according to the supplier's instructions.</li> </ul>
<p>Difficult in finding MCAs during paraffin block sectioning</p>	<ul style="list-style-type: none"> <li>- Histogel containing the MCAs should be oriented during the embedding procedure. Sometimes it is necessary to remove the excess of histogel to facilitate the sectioning step.</li> <li>- When sectioning, it is extremely important not to remove a large thickness of paraffin, as MCAs are not easy to see and therefore can be easily lost.</li> <li>- Sections should be checked under the microscope to assess the presence of the MCA. Its observation is easier if the sections have been recently collected from the water.</li> </ul> <p>Note: It is important to mention that to check the presence of an apoptotic core, it is necessary to section nearly all the MCA to assure that we have passed through the core</p>