

Table S1. *S. cerevisiae* parental strains used in the study

Strain	Genotype	Source	Reference
ADA	MAT α PDR1-3 Δ YOR1::hisG Δ SNQ2::hisG Δ PDR3::hisG Δ PDR10::hisG Δ PDR11::hisG Δ YCF1::hisG Δ PDR55::hisG Δ PDR15::hisG Δ ura3 Δ hisAD124567 Δ PDR5::hisG Δ PDR15::hisG, Δ ura3	Lamping et al, 2007	[1]
Y941	ADA, Δ pdr5::ScCYP51-6xHIS-URA3	Monk et al, 2014	[2]
Y1857	AD2 Δ (ADA, Δ HIS1::dpl200)	Sagatova et al, 2016	[3]
Y2411	AD2 Δ , Δ PDR5::URA3	E. Lamping	
Y2494	AD2 Δ , Δ CYP51pro::GAL1pro	Monk et al, 2019	[4]

Table S2. Oligonucleotides used in the study

A. Construction of CpCYP51-6xHis Y132F cassette by 3-fragment fusion for expression from the PDR5 locus		
1. Amplification of CpCYP51-6xHis Y132F ORF from plasmids received from ATUM		
Forward primer	PDR5us-CpCYP51 Y132F_f (52)	CCGCTCGTTCGAAAGACTTAATTAATAATGG CTTTGGTGGACTTGGCGTTG
Reverse primer	6 HisStop_r (26)	CGAATTTAATGATGATGGTGATGATG
2. Amplification of upstream fragments from gDNA of selected strain		
Forward primer	PDR5Fv3_f (23)	TCGCATTCTGCGCCTTCGAGCAC
Reverse primer	pABC3-PacI_r (31)	CATTTTTTAATTAAGTCTTTCGAACGAGCGG
3. Amplification of downstream fragment from gDNA of selected strain		
Forward primer	6HisStop-f (26)	CATCATCACCATCATCATTAATTCG
Reverse primer	PDR5_186DS_r (25)	TTCGGACATTGAACTTTGATTTATC
B. For wild type strain construct reversion		
Forward primer	CpCyp51_Y132_f (31)	GTAAAGGTGTGATCTACGATTGTCCTAACGC
Reverse primer	CpCyp51_Y132_r (31)	GCGTTAGGACAATCGTAGATCACACCTTTAC
C. Construction of CpCPR-6xHis cassette by 3-fragment fusion for expression from the PDR15 locus		
1. Amplification of CpCPR-6xHis ORF from plasmids received from ATUM		
Forward primer	PDR5us-CpCPR (54)	CCGCTCGTTCGAAAGACTTAATTAATAATGG CTTTAGATAGACTAGATCTTAC
Reverse primer	6 HisStop -r (26)	
2. Amplification of upstream fragments from gDNA of selected strain		
Forward primer	PDR15us_f (25)	GTCACGCCGCCGAACGACGCGCGC
Reverse primer	pABC3-PacI_r (31)	CATTTTTTAATTAAGTCTTTCGAACGAGCGG
3. Amplification of downstream fragment from gDNA of selected strain		
Forward primer	Not1-6xHis (34)	GGCGGCCGCCATCATCACCATCATCATTAAT TC
Reverse primer	PDR15DS_r (25)	GATGGAATAATCCAGTTCGACTCTG
D. Amplification of SchIS1 disruption cassette from gDNA of selected strains		

Forward primer	ScErg11_US-773_f (24)	GCAACAATGGGCGGTTGTTTAGAG
Reverse primer	ScErg11DS346_r (25)	GACTGCTTTATTTCtGCTTGGCCTG

*f and r denotes forward and reverse primer respectively. Number of nucleotides is shown in brackets.

Table S3. MIC₈₀ values for strains overexpressing CpCYP51, CpCYP51 Y132F with/without CpCPR

Strains	MIC ₈₀							
	FLC (μ M)	VCZ (nM)	ITC (nM)	PCZ (nM)	VT-1161 (nM)	VT-1129 (nM)	MCF (nM)	AmpB (μ M)
Y2411	2.4 \pm 0.08	41.7 \pm 1.6	128 \pm 12	157 \pm 3.4	27.1 \pm 0.5	26.7 \pm 0.2	212 \pm 15	2.69 \pm 0.2
Y2718	4.2 \pm 0.4	104 \pm 7.5	187 \pm 16	253 \pm 48	58 \pm 6.1	51.3 \pm 10	288 \pm 31	2.24 \pm 0.13
Y2719	4.65 \pm 0.13	131 \pm 3.6	222 \pm 16	188 \pm 9.6	58 \pm 8	41.6 \pm 3.5	263 \pm 18	2.74 \pm 0.08
Y2720	6.67 \pm 0.09	146 \pm 8.3	213 \pm 18	283 \pm 21	84.7 \pm 8	56.5 \pm 6.5	247 \pm 3.0	2.58 \pm 0.03
Y2721	4.97 \pm 0.03	121 \pm 2.03	291 \pm 3.3	229 \pm 3.5	46.8 \pm 2	47.5 \pm 8.8	237 \pm 1.5	2.77 \pm 0.05
Y2713	38.2 \pm 3.4	1280 \pm 81	202 \pm 1.4	164 \pm 4.7	199 \pm 5.4	264 \pm 24	237 \pm 3.2	2.8 \pm 0.02
Y2714	47.2 \pm 5.8	1820 \pm 18	207 \pm 3.6	242 \pm 17	277 \pm 8.1	367 \pm 16	280 \pm 1.5	2.09 \pm 0.2
Y2715	50 \pm 2.8	1540 \pm 44	201 \pm 0.7	179 \pm 0.5	222 \pm 20	299 \pm 21	184 \pm 16	3.38 \pm 0.3
Y2716	49 \pm 0.01	1410 \pm 20	202 \pm 0.13	228 \pm 25	295 \pm 3.8	293 \pm 5.2	205 \pm 0.9	2.9 \pm 0.2

MIC₈₀s are shown as the mean values \pm SEM for 3 separate clones of each strain using data obtained in triplicate measurements from at least 3 different experiments (a total of 9 measurements per strain).

Table S4. Data collection and refinement statistics for ScCYP51-6×His in complex with VT-1129

Protein	ScCYP51-6×His
Ligand	VT-1129
PDB ID	7RYX
<u>Data collection</u>	
Diffraction source	MX2
Wavelength (Å)	0.9537
Space group	P 1 2 ₁ 1
Cell dimensions	
a, b, c (Å)	78.4, 67.98, 80.3
α , β , γ (°)	90, 99.38, 90
Total reflections	279113
Unique reflections	48767
Resolution (Å)	2.1 - 45.19 (2.16- 2.10)
R _{merge}	0.086 (0.720)
I / σ I	8.6 (1.7)
Completeness (%)	99.8 (99.9)
Redundancy	5.7 (6.0)
CC _{1/2}	0.996 (0.893)
<u>Refinement</u>	
Resolution (Å)	2.1
No. reflections	48681
Rwork / Rfree	0.2336/0.2684
No. of atoms	
Protein	4271
Ligand/ion	79
Water	41
B-factors (Å ²)	
Protein	61.69
Ligand/ion	49.68
Water	52.63
R.m.s. deviations	
Bond lengths (Å)	0.008
Bond angles (°)	0.94
Ramachandran favored (%)	95.44
Ramachandran allowed (%)	3.99
Ramachandran outliers (%)	0.57
Rotamer outliers (%)	0.65
Clashscore	5.79

A. Mass spectrometry result of CpCYP51-6×His

Digestion of CpCYP51-6×His with trypsin

MALVDLALHGNYFM¹TLSTLQQFGLLVFAPFIYNI²IWQLLYSLRKDRVPLVFYWI³PWVGS⁴AVSYGQDPYGF⁵FEQCREKYGD⁶LF
SFVMLGRVMTVYLGPKGHEFVFN⁷AKLSDVSAEDAYQHL⁸TTPVFGKGV⁹IYDCPNARLMEQKKFAKTAL¹⁰TDSFRRYVPLIRGE¹¹I
LDYFTKSKVFNMKKQKSGVVDVLQSQPEITIFTASRSL¹²GEAMRKRFDASFAQLYADLDKGFT¹³PINFVFPHLPLPHYWKRDA¹⁴A
QQKISETYMTEIARRRETGDIDENRDLIDSLLVNSTYKDGVKMTDQEIANLLIGVLMGGQHTSATTSAWFL¹⁵LLHAEKPQLQDE
LYQEVNLALSGKGGNDDLSYEDLQQMPLVNNTIKETLR¹⁶LHMLHSIFRKVVSPLVVPNTKYIVPRGHHVLVSPGYAHTNERF
YKDASDFNPHRWDESASTNDAGEVDYGF¹⁷GKVSKGVSSSYLPF¹⁸GGGRHRCIGE¹⁹QFAYVQLGTIL²⁰TTFVYNLKWKL²¹ANGKVPD²²VD
YTSMTVTL²³PQH²⁴PAEIVWEKRDTCVIGGRHHHHHH

Protein sequence coverage is 67.8%

Digestion of CpCYP51-6×His with chymotrypsin

MALVDLALHGNYFM¹TLSTLQQFGLLVFAPFIYNI²IWQLLYSLRKDRVPLVFYWI³PWVGS⁴AVSYGQDPYGF⁵FEQCREKYGD⁶LF
SFVMLGRVMTVYLGPKGHEFVFN⁷AKLSDVSAEDAYQHL⁸TTPVFGKGV⁹IYDCPNARLMEQKKFAKTAL¹⁰TDSFRRYVPLIRGE¹¹I
LDYFTKSKVFNMKKQKSGVVDVLQSQPEITIFTASRSL¹²GEAMRKRFDASFAQLYADLDKGFT¹³PINFVFPHLPLPHYWKRDA¹⁴A
QQKISETYMTEIARRRETGDIDENRDLIDSLLVNSTYKDGVKMTDQEIANLLIGVLMGGQHTSATTSAWFL¹⁵LLHAEKPQLQDE
LYQEVNLALSGKGGNDDLSYEDLQQMPLVNNTIKETLR¹⁶LHMLHSIFRKVVSPLVVPNTKYIVPRGHHVLVSPGYAHTNERF
YKDASDFNPHRWDESASTNDAGEVDYGF¹⁷GKVSKGVSSSYLPF¹⁸GGGRHRCIGE¹⁹QFAYVQLGTIL²⁰TTFVYNLKWKL²¹ANGKVPD²²VD
YTSMTVTL²³PQH²⁴PAEIVWEKRDTCVIGGRHHHHHH

Protein sequence coverage is 88.32%

B. Mass spectrometry result of CpCYP51-6×His Y132F

Digestion of CpCYP51-6×His Y132F with trypsin. (The 'F' substitution at the 132 position is highlighted in red).

MALVDLALHGNYFM¹TLSTLQQFGLLVFAPFIYNI²IWQLLYSLRKDRVPLVFYWI³PWVGS⁴AVSYGQDPYGF⁵FEQCREKYGD⁶LF
SFVMLGRVMTVYLGPKGHEFVFN⁷AKLSDVSAEDAYQHL⁸TTPVFGKGV⁹I¹⁰DCPNARLMEQKKFAKTAL¹¹TDSFRRYVPLIRGE¹²I
LDYFTKSKVFNMKKQKSGVVDVLQSQPEITIFTASRSL¹³GEAMRKRFDASFAQLYADLDKGFT¹⁴PINFVFPHLPLPHYWKRDA¹⁵A
QQKISETYMTEIARRRETGDIDENRDLIDSLLVNSTYKDGVKMTDQEIANLLIGVLMGGQHTSATTSAWFL¹⁶LLHAEKPQLQDE
LYQEVNLALSGKGGNDDLSYEDLQQMPLVNNTIKETLR¹⁷LHMLHSIFRKVVSPLVVPNTKYIVPRGHHVLVSPGYAHTNERF
YKDASDFNPHRWDESASTNDAGEVDYGF¹⁸GKVSKGVSSSYLPF¹⁹GGGRHRCIGE²⁰QFAYVQLGTIL²¹TTFVYNLKWKL²²ANGKVPD²³VD
YTSMTVTL²⁴PQH²⁵PAEIVWEKRDTCVIGGRHHHHHH

Protein sequence coverage is 63.09%

Digestion of CpCYP51-6×His Y132F with chymotrypsin. (The 'F' substitution at the 132 position is highlighted in red).

MALVDLALHGNYFM¹TLSTLQQFGLLVFAPFIYNI²IWQLLYSLRKDRVPLVFYWI³PWVGS⁴AVSYGQDPYGF⁵FEQCREKYGD⁶LF
SFVMLGRVMTVYLGPKGHEFVFN⁷AKLSDVSAEDAYQHL⁸TTPVFGKGV⁹I¹⁰DCPNARLMEQKKFAKTAL¹¹TDSFRRYVPLIRGE¹²I
LDYFTKSKVFNMKKQKSGVVDVLQSQPEITIFTASRSL¹³GEAMRKRFDASFAQLYADLDKGFT¹⁴PINFVFPHLPLPHYWKRDA¹⁵A
QQKISETYMTEIARRRETGDIDENRDLIDSLLVNSTYKDGVKMTDQEIANLLIGVLMGGQHTSATTSAWFL¹⁶LLHAEKPQLQDE
LYQEVNLALSGKGGNDDLSYEDLQQMPLVNNTIKETLR¹⁷LHMLHSIFRKVVSPLVVPNTKYIVPRGHHVLVSPGYAHTNERF
YKDASDFNPHRWDESASTNDAGEVDYGF¹⁸GKVSKGVSSSYLPF¹⁹GGGRHRCIGE²⁰QFAYVQLGTIL²¹TTFVYNLKWKL²²ANGKVPD²³VD
YTSMTVTL²⁴PQH²⁵PAEIVWEKRDTCVIGGRHHHHHH

Protein sequence coverage is 93.97%

C. Mass spectrometry result of CpCPR-6×His

Digestion of CpCPR-6×His with trypsin.

MALDRDLTLVVIVLAVAVAAAYFIKSQYFSKPESSEGLNTDTAGGNSRDILATLTKNHKNTLLLFSGQTGTAEDYCNKMSRELS
 ARFGLKTMVADFADYDWDNFGDIKEDVLVFFIMATYGEGETDNAIEFVDFLDNEADTLSTLRFTVFGLGNSTYEFFNAIGRK
 INEKLESKGAERFAEYEGEDDGQGTMDDEDFLAWKDGVFDSLNNLNLEEKELKYEPSLKLEIRDDLTIDSSVSLGEPDKSYV
 NTKAGTDLTKGPFHDHSHPYLAPITKIKELFFTKERSCVHVEFDLSNSNLKYTTGDHLAIWPSNANEYVELFLKTFDLTEQRDV
 VFDLKALDSTYQIPFPPTITYEAVVRHHLEISGPVSRQFFLSIAAFAPDEETKTKLTTVANDKQKYAAEVTHKKYNIADGLLY
 FSNGKPWTKVPFEFLIENVQHFTPRYYSISSSSLSEKTHIDITAVVEAETESDGRVVTGVVTNLLKDVEINKNSSDDKPIVS
 DLKGPRNKFQNYKLPVHVRSTFKLPSSSKTPIILVGPGTGVAPLRGFVRERVQQLKNGVNVGPSLLFYGCRNEDEDYLYRDE
 WPQYAKELGESFELITAFSRANPNKKVYVQHKILEQAKKINQLLQDGGIIYVCGDASHMARDVQASFAKVLSQERGIELEKAA
 ELIRSLKVQNRQEDVWGRHHHHH

Protein sequence coverage is 90.14%.

Digestion of CpCPR-6×His with chymotrypsin.

MALDRDLTLVVIVLAVAVAAAYFIKSQYFSKPESSEGLNTDTAGGNSRDILATLTKNHKNTLLLFSGQTGTAEDYCNKMSRELS
 ARFGLKTMVADFADYDWDNFGDIKEDVLVFFIMATYGEGETDNAIEFVDFLDNEADTLSTLRFTVFGLGNSTYEFFNAIGRK
 INEKLESKGAERFAEYEGEDDGQGTMDDEDFLAWKDGVFDSLNNLNLEEKELKYEPSLKLEIRDDLTIDSSVSLGEPDKSYV
 NTKAGTDLTKGPFHDHSHPYLAPITKIKELFFTKERSCVHVEFDLSNSNLKYTTGDHLAIWPSNANEYVELFLKTFDLTEQRDV
 VFDLKALDSTYQIPFPPTITYEAVVRHHLEISGPVSRQFFLSIAAFAPDEETKTKLTTVANDKQKYAAEVTHKKYNIADGLLY
 FSNGKPWTKVPFEFLIENVQHFTPRYYSISSSSLSEKTHIDITAVVEAETESDGRVVTGVVTNLLKDVEINKNSSDDKPIVS
 DLKGPRNKFQNYKLPVHVRSTFKLPSSSKTPIILVGPGTGVAPLRGFVRERVQQLKNGVNVGPSLLFYGCRNEDEDYLYRDE
 WPQYAKELGESFELITAFSRANPNKKVYVQHKILEQAKKINQLLQDGGIIYVCGDASHMARDVQASFAKVLSQERGIELEKAA
 ELIRSLKVQNRQEDVWGRHHHHH

Protein sequence coverage is 94.49%

Figure S1 Identification of the recombinant proteins by mass spectrometry. Sequences identified on excision from SDS-PAGE gel bands are highlighted in grey.

References

1. Lamping E, Monk BC, Niimi K, Holmes AR, Tsao S, Tanabe K, et al. Characterization of three classes of membrane proteins involved in fungal azole resistance by functional hyperexpression in *Saccharomyces cerevisiae*. *Eukaryot Cell*. 2007;6:1150-65.
2. Monk BC, Tomasiak TM, Keniya MV, Huschmann FU, Tyndall JD, O'Connell JD, 3rd, et al. Architecture of a single membrane spanning cytochrome P450 suggests constraints that orient the catalytic domain relative to a bilayer. *Proc Natl Acad Sci U S A*. 2014;111:3865-70.
3. Sagatova AA, Keniya MV, Wilson RK, Sabherwal M, Tyndall JD, Monk BC. Triazole resistance mediated by mutations of a conserved active site tyrosine in fungal lanosterol 14alpha-demethylase. *Sci Rep*. 2016;6:26213.
4. Monk BC, Keniya MV, Sabherwal M, Wilson RK, Graham DO, Hassan HF, et al. Azole Resistance Reduces Susceptibility to the Tetrazole Antifungal VT-1161. *Antimicrob Agents Chemother*. 2019;63:e02114-18.