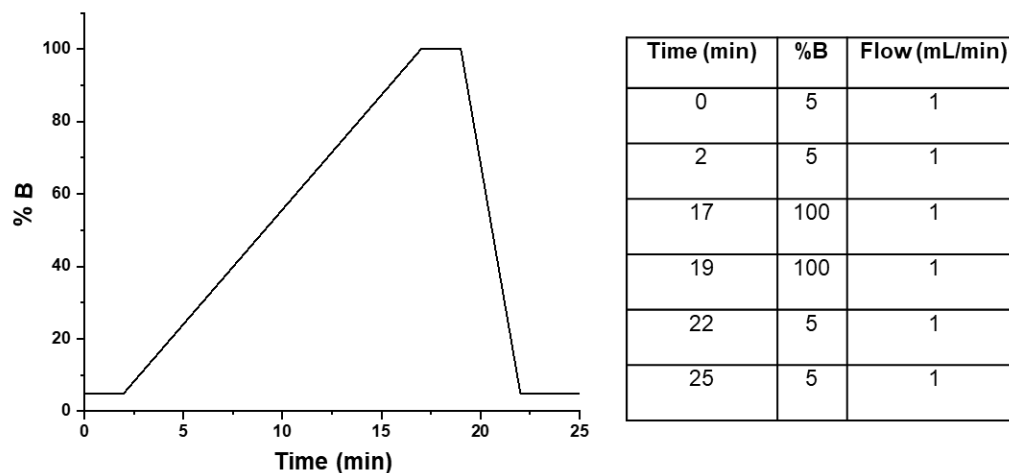
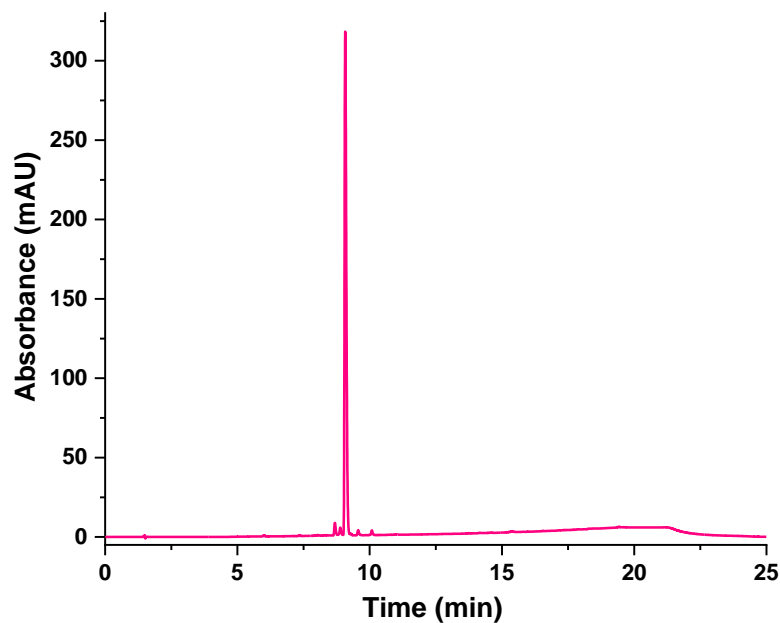


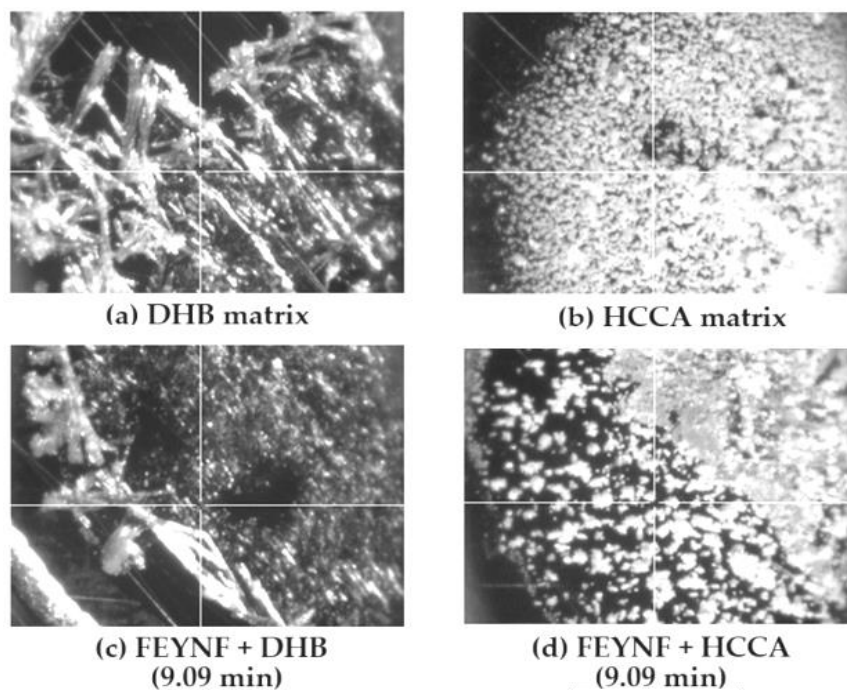
## Supplementary materials



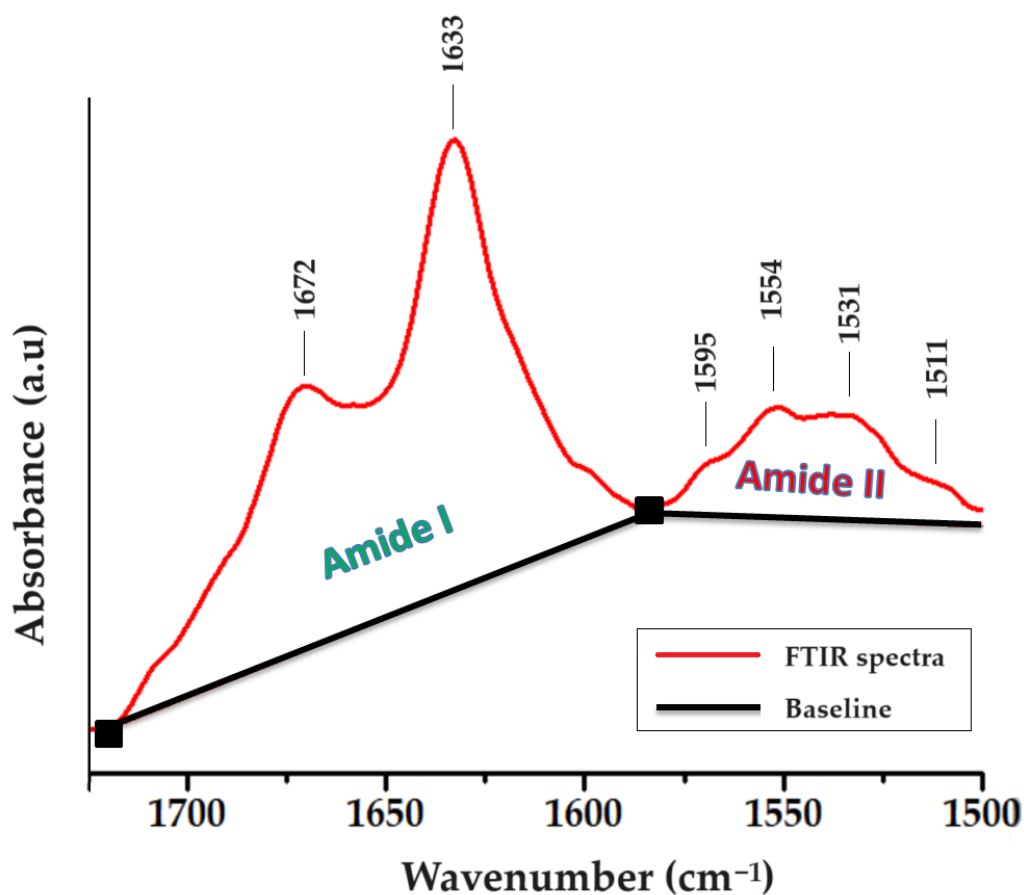
**Figure S1.** The gradient protocol used in HPLC analysis.



**Figure S2.** HPLC chromatogram of the FEYNF-NH<sub>2</sub> peptide following purification through solid-phase extraction, revealing a calculated purity of 99.54%. The chromatogram was generated using an injection volume of 5  $\mu$ L, and the data were recorded at a detection wavelength of 215 nm. The observed peak patterns and their intensities affirm the successful purification, highlighting the peptide's exceptional purity achieved through the solid-phase extraction process.



**Figure S3.** Visualization of the crystals formed by the HCCA and DHB matrices (a,b) and the co-crystals with the FEYNF samples on the target (c,d). Images obtained by means of the camera equipped with the MALDI-ToF mass spectrometer. Retention times are reported based on HPLC analysis.

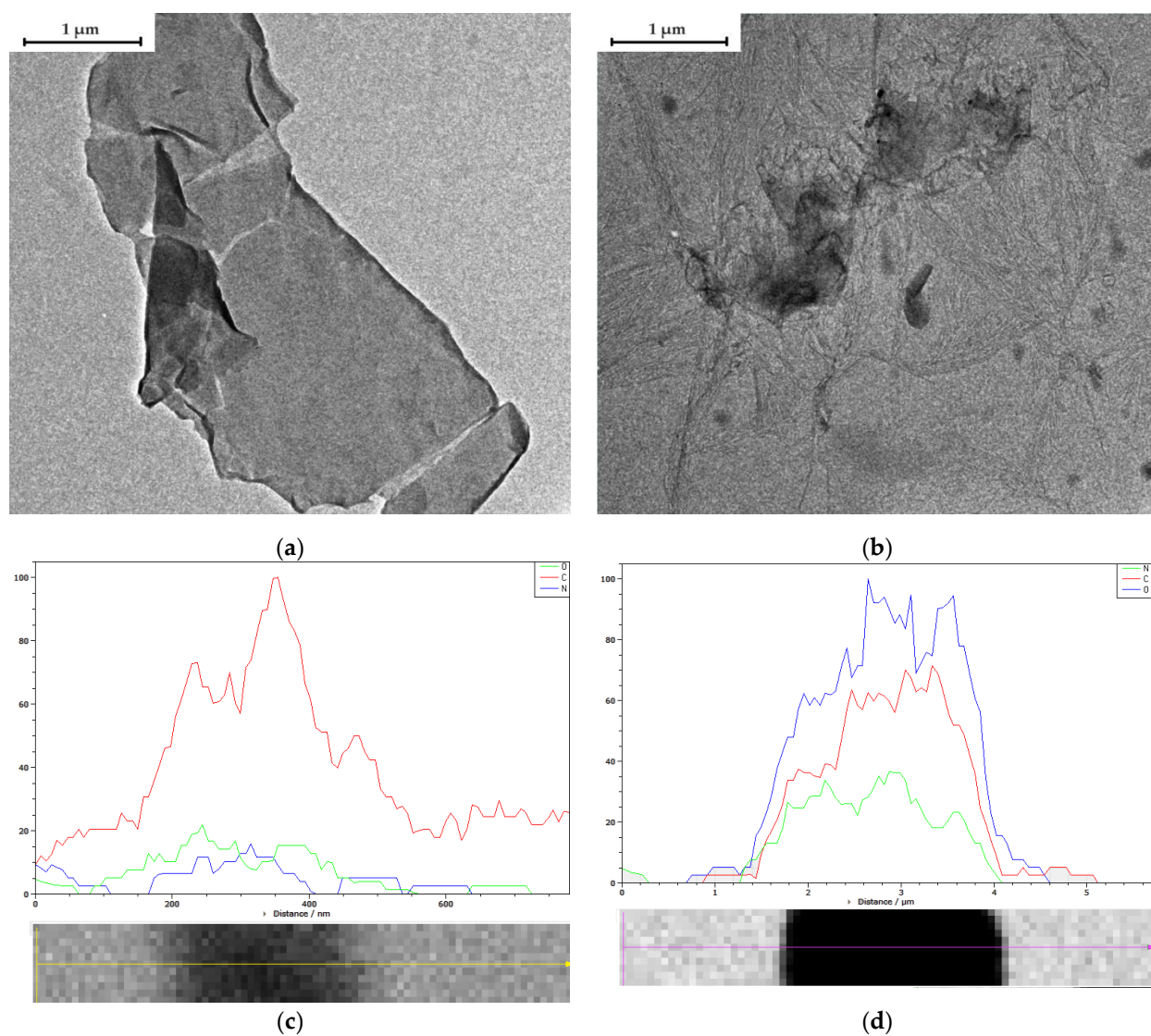


**Figure S4.** FTIR spectrum of FEYNF-NH<sub>2</sub> peptide evidencing the amide I-amide II spectral range and the baseline applied for data processing.

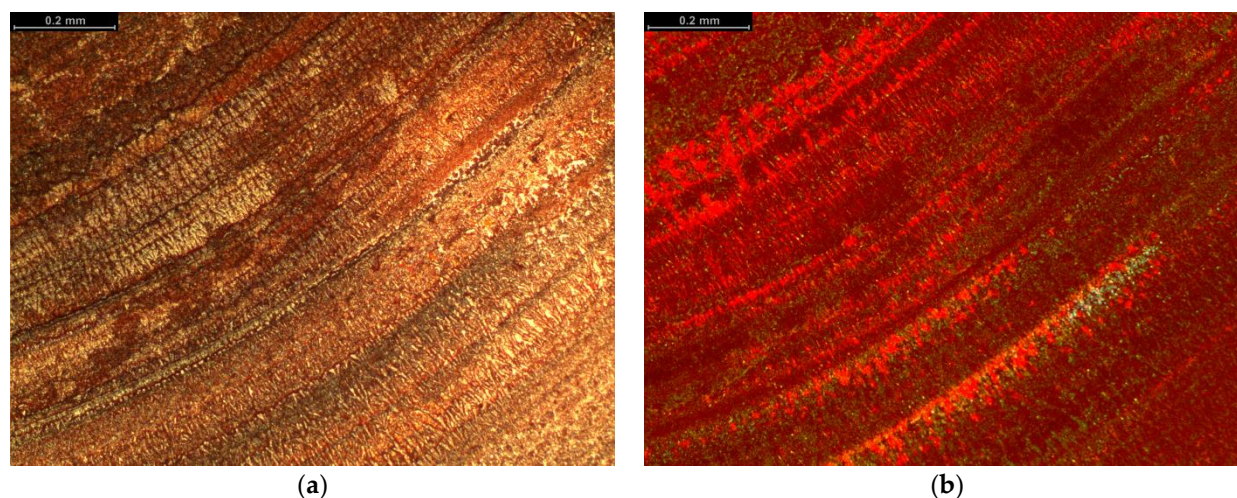
**Table S1.** The content of secondary structures (%) of the FEYNF-NH<sub>2</sub> revealed from FTIR spectrum analysis.

Peak no.	Structure	Wavenumber (cm <sup>-1</sup> )	Relative intensity (%) <sup>*</sup>
1	Antiparallel $\beta$ -sheet	1694	3.12
2	$\beta$ -turns	1672	16.12
3	$\alpha$ -helix	1657	28.01
4	$\beta$ -sheet	1633	34.53
5	$\beta$ -sheet aggregate strands	1613	10.56
6	Side chain	1596	7.65

<sup>\*</sup> The relative intensity indicates the proportion of a component band in the total integrated area of the Amide I region.



**Figure S5.** TEM images of FEYNF-NH<sub>2</sub> before (a) and after 20 h of incubation at physiological pH of 7.4 and a temperature of 37 °C; EDX spectra for FEYNF peptide sample before (c) and after incubation (d);



**Figure S6.** Microscopic images of FEYNF-NH<sub>2</sub> sample after Congo red staining captured under two polarization angles: 0 degrees (a), and 90 degrees (b).

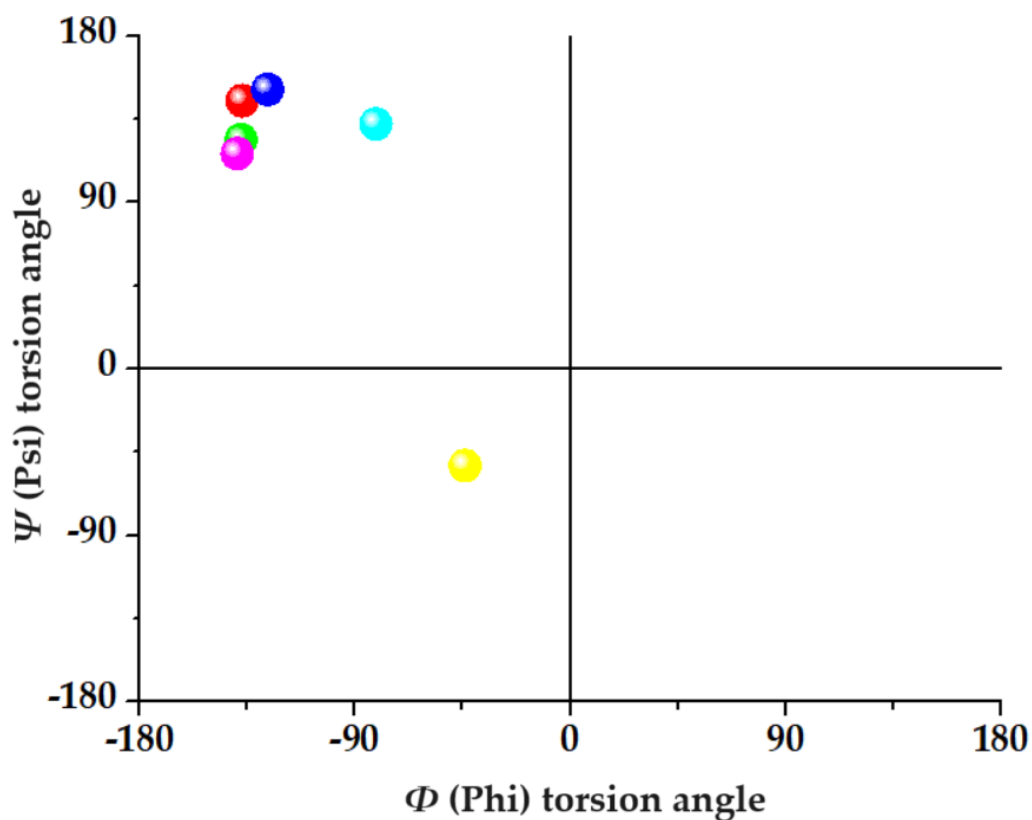
**Table S2.** Representative clusters of the docking simulation for the FEYNF@FEYNF dimer complex, highlighting the amino acid residues involved in the peptide–peptide interaction, binding energy ( $E_b$ ) and dissociation constant ( $K_d$ ) values

Cluster	Binding energy, ( $E_b$ , kcal/mol)	Dissociation constant ( $K_d$ , $\mu$ m)	Contacting amino acid residue
1	−5.38	113.87	PHE1 GLU2 TYR3 ASN4 PHE5
2	−5.21	150.69	PHE1 GLU2 TYR3 ASN4 PHE5
3	−5.02	208.01	PHE1 GLU2 TYR3 ASN4 PHE5
4	−5.00	216.98	PHE1 GLU2 TYR3 ASN4 PHE5
5	−4.94	238.89	PHE1 GLU2 TYR3 ASN4 PHE5
6	−4.94	239.70	PHE1 GLU2 TYR3 ASN4 PHE5
7	−4.94	240.92	PHE1 GLU2 TYR3 ASN4 PHE5
8	−4.91	250.45	PHE1 GLU2 TYR3 ASN4 PHE5
9	−4.90	257.31	PHE1 GLU2 TYR3 ASN4
10	−4.87	271.59	PHE1 GLU2 TYR3 ASN4 PHE5
11	−4.81	300.53	PHE1 GLU2 TYR3 ASN4 PHE5

**Table S3.** Intermolecular and intramolecular interactions occurring in the FEYNF@FEYNF dimer complex, with corresponding amino acid residues involved.

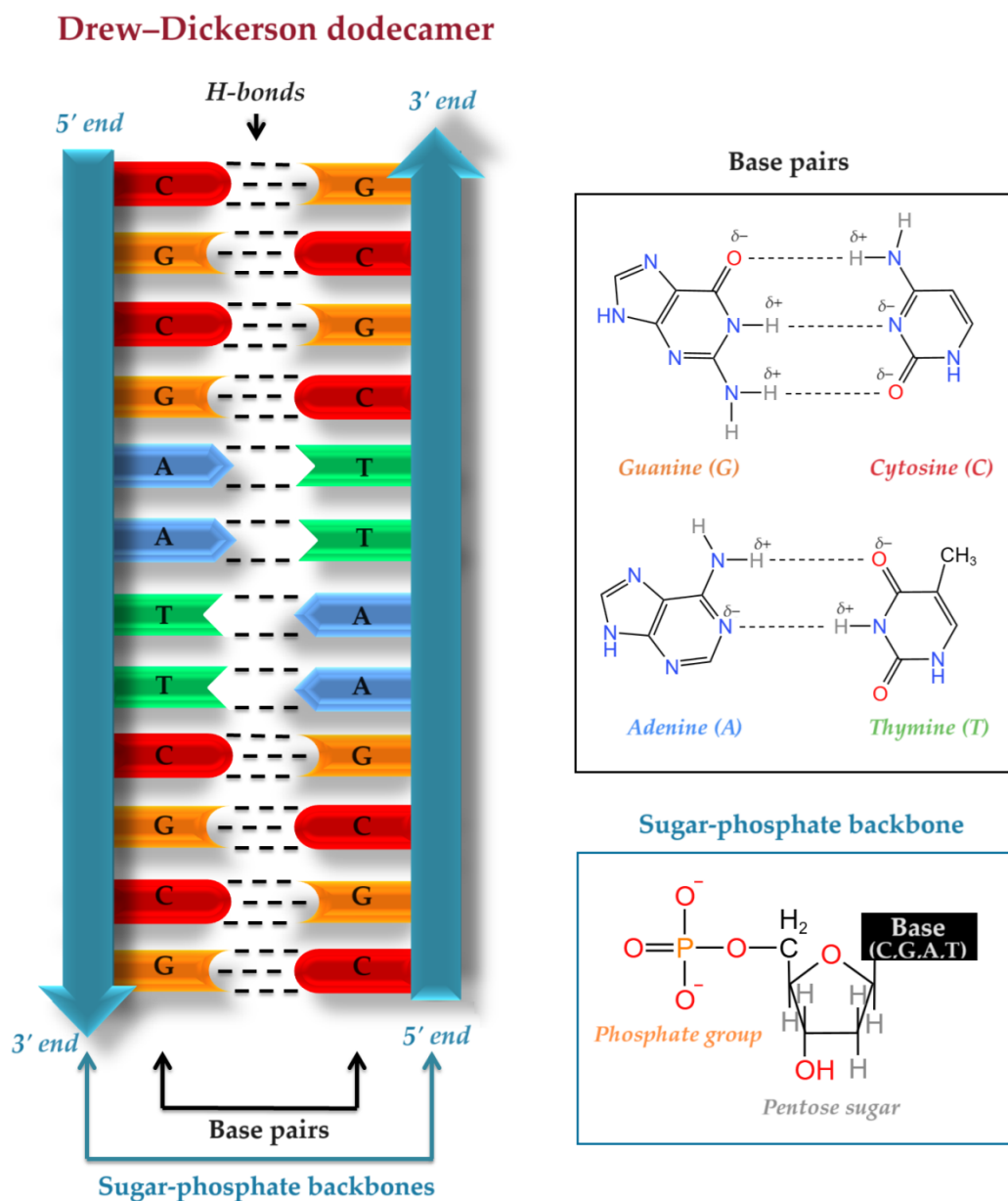
Interaction type	Ligand amino acid residue	Receptor amino acid residue
H-bonds	PHE1 (O)	GLU2 (N-H)

	N4 (O)( -	GLU2 (O-H) TYR3 (O)...ASN4 (N-H) (intramolecular)
Hydrophobic interactions	PHE1	PHE1
	PHE1	TYR3
	GLU2	GLU2
	TYR3	TYR3
	TYR3	PHE5
	PHE5	ASN4
	PHE5	PHE5
$\pi$ - $\pi$ stacking	PHE1	PHE1
	TYR3	TYR3
	TYR3	PHE5
	PHE5	PHE5



**Figure S7.** Graphical representation of  $(\Phi_i, \Psi_i)$  plot (Chain 1:  $(\Phi_1, \Psi_1)$  – red,  $(\Phi_2, \Psi_2)$  – green,  $(\Phi_3, \Psi_3)$  – blue; Chain 2:  $(\Phi_4, \Psi_4)$  –cyan,  $(\Phi_5, \Psi_5)$  – magenta,  $(\Phi_6, \Psi_6)$  – yellow) based on determined torsion angles.





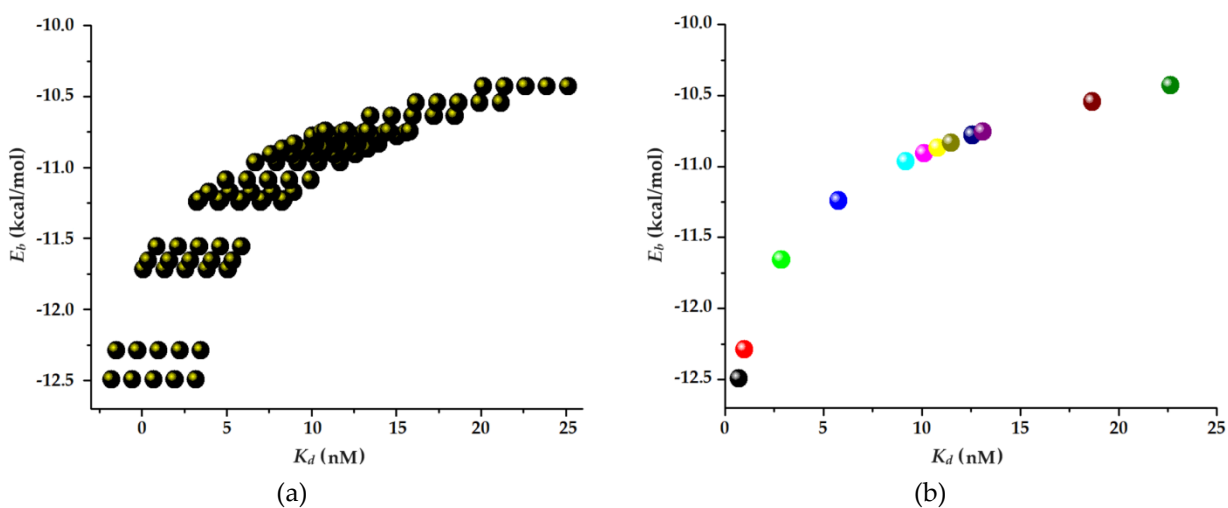
**Figure S8.** Graphical representation of Drew–Dickerson dodecamer sequence (d(CGCGAATTCGCG)<sub>2</sub>) with molecular structures of corresponding nitrogenous bases (C-cytosine, G-guanine, A-adenine, T-thymine) and sugar-phosphate backbone (KingDraw V3.0.2.).

**Table S4.** Representative clusters of the docking simulation for the investigated polyplex (binding energy ( $E_b$ ), dissociation constant ( $K_d$ ) and contacting residues)

Cluster	Binding energy, ( $E_b$ , kcal/mol)	Dissociation constant ( $K_d$ , nm)	Contacting receptor residue (nitrogenous base) <sup>1</sup>
1	−12.494	0.695	C3(a) G4(a) A5(a) A6(a) T7(a) C3(b) G4(b) A5(b) A6(b) T7(b)
2	−12.289	0.982	C3(a) G4(a) A5(a) A6(a) T7(a) C3(b) G4(b) A5(b) A6(b) T7(b)
3	−11.656	2.858	C3(a) G4(a) A5(a) A6(a) A5(b) A6(b) T7(b) T8(b) C9(b)
4	−11.241	5.758	A5(a) A6(a) T7(a) T8(a) C9(a) C3(b) G4(b) A5(b) A6(b) T7(b)
5	−10.964	9.189	C1(a) G2(a) C3(a) G4(a) A5(a) T7(b) T8(b) C9(b) G10(b) C11(b)
6	−10.908	10.100	C1(a) G2(a) C3(a) G4(a) A5(a) T7(b) T8(b) C9(b) G10(b) C11(b)
7	−10.868	10.806	T7(a) T8(a) C9(a) G10(a) C1(b) G2(b) C3(b) G4(b) A5(b)
8	−10.832	11.483	A6(a) T7(a) T8(a) C3(b) G4(b) A5(b) A6(b)
9	−10.779	12.557	C3(a) G4(a) A5(a) A6(a) T7(a) A5(b) A6(b) T7(b) T8(b)
10	−10.754	13.098	A6(a) T7(a) G2(b) C3(b) G4(b) A5(b) A6(b)
11	−10.544	18.670	G4(a) A5(a) A6(a) G4(b) A5(b) A6(b) T7(b)
12	−10.43	22.631	G4(a) A5(a) A6(a) T7(a) T8(a) C9(a) C3(b) G4(b) A5(b) A6(b)

<sup>1</sup> The contact residues are the nitrogenous bases cytosine (C), guanine (G), adenine (A) and Thymine (T); the base residues are found on both sugar-phosphate backbones, referred to as (a) and (b), respectively.





**Figure S9.** Molecular docking outcomes for the polyplex formation: (a) data for all 100-docked poses of polyplex conformation (binding energy vs. dissociation constant); (b) data for the 12 representative clusters (obtained from clustering analysis).