

# Chili Pepper Scent: Study and Recognition with Chemiresistors Array <sup>†</sup>

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**Abstract:** Chili peppers are spices worldwide appreciated and represent a real culinary tradition and cultural identity for several populations. Here we present an effort to evaluate whether an array of chemiresistor gas sensors is capable to distinguish different chili pepper samples by the analysis of the emitted aroma. Our approach is based on two methods. A classical method based on GC-MS-SPME to characterize intensity and composition of chemical compounds emitted by the fruits and a novel approach on Multivariate sensors response that produces an online graphic representation of the sensor features.

**Keywords:** scent; chemiresistor; e-nose; artificial nose; volatile organic compounds; GC-MS-SPME; food security; food safety

## 1. Introduction

Food authenticity and adulteration is a nowadays topic trend. Sensor Systems in food quality and security control assessment are strongly requested [1,2]. The wide range of chili peppers cultivars is a suitable and poor explored test [3] for embedded system sensors.

The most important parameter of food from the sensory point of view it's the aroma. It is composed by the set of volatile organic compounds (VOC) that form the so called fingerprint. The characterization of the volatile fingerprint may offer information about safety and quality and could be use as an indicator resource when it varies somehow from the standard. The set of VOC could provide also information about geographical origins and cultivars [4].

## 2. Materials and Methods

Chili peppers are fruit of plants from the genus *Capsicum*, members of the nightshade family, Solanaceae. Today in the world are cultivated five Species of *Capsicum* that have generated more than 50.000 cultivars. Measurements starts from six *Capsicum* cultivars belonging from three of the five major species of cultivated chili peppers: from the Species *Annuum* we test "tondo Calabrese" (TCCA) with a taste similar to sweet bell peppers and "Goat's Weed" (GWCA); from the Species *Baccatum* we test "Jamy" (JBC) often very sweet; from the Species *Chinense* we test "Trinidad Perfume" (TPCC), "Fatali", "Habanero Orange" and "Habanero red" very aromatic and often fruity,

with a bitter, soapy aftertaste in some cultivars, mostly the superhot ones. This variability was needed so as to highlight the aromatic differences between both the species and the single cultivars [5].

## 2.1. Analytical Method

To extract and identify the set of VOCs fraction the pepper fruits were divided in parts placed in a 20 mL chromatographic vial. Vials were closed hermetically with silicon-PTFE septum, crimped with an aluminum crimp. To perform the HS-SPME analysis a triphasic fiber DVB/CAR/PDMS—50/30  $\mu\text{m}$  (Supelco Co. Bellefonte, PA, USA) and a HT280T auto-sampler (HTA S.r.l., Brescia, Italy). The vials were incubated at 50 °C for 15 to reach the equilibrium, subsequently, the fiber was exposed at the same conditions. Desorption of volatiles took place in the injector of the GC-MS for 6 min at 250 °C. For the analysis of the volatile set of peppers was used a Shimadzu Gas Chromatograph GC2010 PLUS (Kyoto, KYT, Japan) interfaced with a Shimadzu single quadrupole Mass Spectrometer MS-QP2010 Ultra (Kyoto, KYT, Japan). GC separations were performed on a DB-WAX capillary column (30m  $\times$  0.25mm  $\times$  0.25 $\mu\text{m}$ , Agilent Technologies, Santa Clara, CA, USA). Ultrapure helium (99.99%) was used as the carrier gas at the constant flow rate of 1.0 mL/min. The GC oven temperature was programmed as follows: initially the column was held at 50 °C for 1 min followed by a rise from 50 to 80 °C at 5 °C/min and kept for 6 min; finally a second rise from 80 °C to 250 °C at a rate of 7 °C/min was performed and the temperature was maintained at 250°C for 1 min. The identification of the volatile compounds was carried out using the NIST11 and the FFNSC2 libraries of mass spectra [5,6].

## 2.2. E-Nose Method

Samples for the analysis are prepared cutting single fresh or unfreezed Chili pepper into pieces and placed in a 0.50cl PET bottle at room temperature. These samples are analyzed, within a few hours, sucking the air from the bottle by the mean of an HAPSITE H1552 Portable Gas Chromatograph with a Mass Spectroscopy detector (GC-MS). The same Sample is inhaled by the e-nose platform based on a max number of twelve chemiresistors [7]. Chemiresistor matrix can be divided in two typologies: 6 commercial heated sensors (TGS family) and 4 not heated nanostructured sensing films dropcasted onto transducer with an USB connector for an easily change on the sensor board (USB family). TGS sensors are FIGARO sensors of the 260, 261 and 262 families. USB sensors are affected by low dynamics, low stability and high humidity interference but, in spite of all, their low-temperature chemical kinetic can have a key role for odour discrimination.

Analysis is conducted without the use of a gas concentrator. For every Bottle-head-space of a cut chili pepper we produce a GC-MS chromatogram illustrating the principal chemical compounds emitted and a matrix 10  $\times$  150  $R_{ij}$  (Odour Pattern) based on 150 sampling of the ten sensors resistance ( $j = 1:4$  USB sensors;  $J = 5:10$  TGS sensors) measured every second during the e-nose analysis. Solid state sensors are affected by drift and it is a challenge to extract features useful for odour discrimination. We use the following steps to extract from the e-nose Odour Pattern parameters that can be used to compare different Odours:

- for the Sensor  $j$  baseline  $R0\_j$  is calculated taking into account the mean value of 30 sampling before odour injection;
- percentage relative response:  $Rsp\_j = \max |R_{ij} - R0\_j| / R0\_j * 100$ ;
- relative response velocity (signal differential with sampling at 1 Hz):  $Vrsp\_ij = (R_{ij} - R_{(i-1)j}) / R0\_j$ ;
- max relative response velocity:  $MVrsp\_j = \max |R_{ij} - R_{(i-1)j}| / R0\_j$ . This parameter can be calculated both during the odour inhaling phase (ads) that during the odour exhaling phase (des);
- a new homogenized matrix  $S_{ij}$  (20  $\times$  150) composed by  $(Rsp\_j - R_{ij}) / Rsp\_j$  for  $j = 1:10$  and  $Vrsp\_ij / MVrsp\_j$  for  $j = 11:20$ ;
- Taking into account the USB Sensor that give the best  $MVrsp$  and the TGS Sensor that give the Best  $MVrsp$  we calculate  $RMVrsp\_j$  as the ratio of the  $MVrsp$  to the best one for each Sensor family.

In this manner became possible to compare different Odour Pattern simply by subtracting their RMVrsp and their  $S_{ij}$ . We choose max relative response velocity instead of percentage relative response because we have reason to believe [8] that this differential parameter is less affected by sensors drifting, and saturation although it contains the same discrimination feature of the Sensor Response.

### 3. Results and Discussion

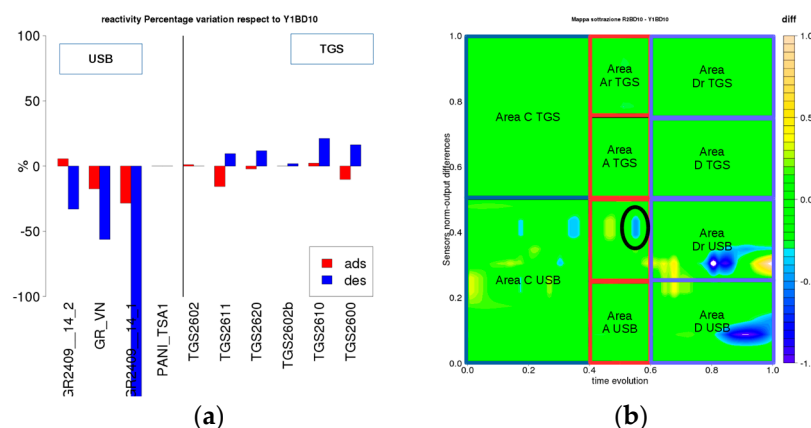
#### 3.1. Chili Pepper Scent Chemical Composition

Analytical measurements performed through GC-MS-SPME describe a chemical pattern emitted by the chilli pepper samples composed by more than 160 chemical compounds. GC-MS data have been recorded for all the four different cultivars producing 12 chromatograms (3 for each cultivar). Data have been normalized and projected on the 3D principal component (PC) subspace. The patterns of the four cultivars appear to be significantly separated in the PC space supporting the occurrence of a specific olfactive fingerprint to be exploited by an e-nose. JCB data express a significant correlation with Caryophyllene, Acetoin, 1,6-Octadien-3-ol,3,7-dimethyl, GWCA and TCCA with Cyclopentane, 3-hexyl-1,1-dimethyl, Tetradecane,2-methyl-, 2,3-Butanediol,[R-(R\*,R\*)]-, finally TPCC cultivar is correlated with Hexyl n-valerate, Cyclohexanol,2,4-dimethyl-, Cyclohexanol, 3,3-dimethyl-. Ethanol, 1-Butanol-3 methyl, Phosphonic acid (p-dydroxyphenil) and Phenylethyl Alcohol appear to be present in all the cultivar always eliciting a significant response with low inter-cultivar variance inducing a common mode signal that should be rejected by an e-nose. Apart from the above mentioned highly class-correlated analytes, a K-NN based wrapper analyte selection scheme (r-relief -Kira) [9] reported that Decanoic acid,ethyl ester, Oxacyclotetralon-2-one-, 14 methyl and Citronellic Acid provides for the most discriminant information while 1-Butanol-3methyl, cis-2-methyl-7-octadecene, Decanoic acid-silver provide a little amount of information in order to discriminate among the four different cultivar in a supervised scheme. Both unsupervised and supervised schemes agree that 1-Butanol-3methyl do not provide useful discriminant information.

#### 3.2. E-Nose Odour Comparison between Habanero and Fatalii

These two cultivars were tested 3 times: first time with two fresh fruits, the other two time with six unfreezed fruits. GC-MS analysis shows the abundance of hexyl valerate and ethanol for all the samples. For the GC-MS habanero orange and red emitted similar chemical compounds while in Fatalii is not present the TrimethylDihydrofuran 2-1. E-nose odour pattern are acquired taking into account odour intensity and avoiding e-nose chamber contamination. Perhaps we use an odour dilution too high for the USB sensors causing a lack of stability in the USB odour Pattern.

In the all comparison TGS sensors shows minimal differences both in the histogram and in the comparison map (in Figure 1 an example of comparison). USB sensors seem to discriminate habanero from fatalii with the third sensor that show always strong differences in all the comparison between habanero and fatalii. Also in the comparison map is possible to isolate a distortion region (black circle in figure) correlated to the difference between fatalii and habanero that disappear in the comparison between two fatalii. However, the instability of the USB family can affect the reliability of such a result.



**Figure 1.** example of e-nose odor pattern comparison between Fatalii and Habanero for freezeed samples: (a) percentage relative change in the sensors adsorbing and desorbing response max velocity; (b) Signal and Signal rate differences among sensors—comparison map. The black circle indicates an area that show discrimination capability.

## 4. Conclusions

This study try to answer to a question: if and how is possible to smell the chili peppers with an e-nose based on chemiresistive solid state sensors. We encountered several difficulties for the correct discrimination of cultivars with the e-nose: cultivar discrimination is based on second order variation of chemical compounds. So already the intensity variation of odours between the samples or e-nose chamber contamination can affect the discrimination reliability. Heated sensors TGS seems to go in crisis for cultivars discrimination so USB not heated sensing films became crucials if is possible to demonstrate enough reliability in the measurement. Work in progress deals with measuring a statistical substantial number of samples based on fresh fruits of different cultivars to confirm the e-nose cultivars discrimination capability.

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**Conflicts of Interest:** The authors declare no conflict of interest.

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