

Proceeding Paper

Bioaugmentation Performance of a Bacterial Consortium for Moving Bed Biofilm Reactor (MBBR) Treating Municipal Wastewater [†]

Eliza-Gabriela Brettfeld ^{1,2}, Oana-Andreea Cheoafa ¹, Diana Constantinescu-Aruxandei ¹ 
and Florin Oancea ^{1,3,*} 

¹ Bioresources Department, National Institute for Research & Development in Chemistry and Petrochemistry—ICECHIM, Splaiul Independentei nr. 202, 060021 Bucharest, Romania; eliza.brettfeld@icechim.ro (E.-G.B.); oana.cheoafa@icechim.ro (O.-A.C.); diana.constantinescu@icechim.ro (D.C.-A.)

² Faculty of Chemical Engineering and Biotechnology, National University of Science and Technology Politehnica Bucharest, Splaiul Independentei nr. 313, 060042 Bucharest, Romania

³ Faculty of Biotechnologies, University of Agronomic Sciences and Veterinary Medicine of Bucharest, Bd. Marasti nr. 59, 011464 Bucharest, Romania

* Correspondence: florin.oancea@icechim.ro; Tel.: +40-213-16307

[†] Presented at the Exploratory Workshop “Innovative Cross-Sectoral Technologies”, Vth Edition, Bucharest, Romania, 22–23 May 2023, Secvent project meeting.

Abstract: A compatible microbial consortium with a high organic pollutant degradation ability, which includes a gram-positive *Brevibacillus parabrevis* B50 NCAIM B 001413 bacterial strain and a gram-negative *Pseudoxanthomonas mexicana* P32 NCAIM (P) B 001414 bacterial strain, was selected using high-throughput screening techniques. The compatible microbial consortium, encapsulated in alginate beds, was used to inoculate moving bed biofilm reactors from a small municipal wastewater treatment plant. The bioaugmentation performance of the inoculated consortium was evaluated by determining the water quality parameters before inoculation and one month after bioaugmentation treatment. The removal of organic matter was enhanced after treatment with the selected microbial consortium.

Keywords: organic matter removal; gram-positive—gram-negative bacterial consortium; alginate beads encapsulation; inoculation efficiency



Citation: Brettfeld, E.-G.; Cheoafa, O.-A.; Constantinescu-Aruxandei, D.; Oancea, F. Bioaugmentation Performance of a Bacterial

Consortium for Moving Bed Biofilm Reactor (MBBR) Treating Municipal Wastewater. *Chem. Proc.* **2023**, *13*, 29.

<https://doi.org/10.3390/chemproc2023013029>

Academic Editors: Mihaela Doni and Radu Claudiu Fierăscu

Published: 20 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Bioaugmentation of wastewater treatment intends to enhance efficiency in removing organic matter and pollutants by inoculation with selected microbial strains [1]. The inoculation efficiency also depends on the ability of the introduced bacteria to form protective biofilms [2]. The moving bed biofilm reactor (MBBR) is a wastewater treatment process that uses plastic carriers with a buoyancy close to water to support the development of the bioactive microbial biofilm [3,4]. This MMBR technology benefits by inoculation with bacterial strains with organic pollutant degradation ability and promoting biofilm formation [5–7].

This study describes the effect of applying a compatible microbial consortium with high organic pollutant degradation ability, which promotes biofilm formation, selected with a combination of high-throughput screening techniques [8–10], to enhance the efficiency of a small municipal wastewater treatment plant using MMBR technology. The bioaugmentation performance of the inoculated consortium was evaluated by determining the water quality parameters before inoculation and two weeks after inoculation. The collected water samples were analyzed following the guidelines specified in the Romanian “Normative regarding the establishment of pollutant load limits for industrial and urban wastewater when discharged into natural receivers—the NTPA-001:2002”, that fulfills the European

Union Directive 91/271/EEC concerning urban waste-water treatment. This normative document sets essential regulatory limits for pollutant concentrations in wastewater discharges, ensuring the protection of natural water bodies. The analysis of the treated water demonstrated that the inoculation with the bioaugmentation consortium determined the compliance of wastewater discharges with legal requirements.

2. Materials and Methods

2.1. Biological Material

The biocompatible consortium was selected using a set of high-throughput screening techniques developed by our group [8–10] from the microbial strain collection of the National Institute for Research & Development in Chemistry and Petrochemistry—ICECHIM Bucharest. The gram-positive spore-forming bacteria *Brevibacillus parabrevis* B50 NCAIM B 001413 bacterial strain was isolated from soil and demonstrated a high organic matter degradation ability and capacity to produce manganese oxide, promoting the degradation of recalcitrant pollutants and quorum sensing (QS) signal autoinducer-2, AI-2 [11]. The gram-negative *Pseudoxanthomonas mexicana* P32 NCAIM (P) B 001414 bacterial strain was isolated by selective enrichment from a soil polluted with crude oil and demonstrated a high ability to produce biosurfactant and Acyl-homoserine lactones (AHL) QS signals.

2.2. Bioproduct Preparation and Bioaugmentation Treatment

The bacterial strains were separately cultivated on potato-dextrose broth supplemented by yeast extract from wine lees for 48 h, and biomass was harvested by centrifugation [12]. The biomass pellets were resuspended in phosphate saline buffer to reach 10^{10} ufc per mL and mixed in a 1:1 ratio. In the resulting suspension, 3 g of sodium alginate, with an average molecular mass of 40 kDa (Sigma-Aldrich, Merck Group, Darmstadt, Germany) per each 97 g of bacterial suspension, were added and dissolved by gentle stirring. The alginate suspension was prilled into calcium alginate beads using Encapsulator B-390 (Büchler Flawil, Switzerland), with a flow vibration nozzle, according to manufacturer instructions. The resulting beads were used to inoculate the first aeration tank of a small municipal wastewater treatment plant (Filipeștii-de-Pădure, Romania) that uses the MMBR technology according to the design of the DFR System (Bucharest, Romania). The used dose was 100 g beads per 1 m^3 . The treatment was performed at the beginning of September 2021. Samples of discharged water from the wastewater treatment plant were taken before inoculation and one month after bioaugmentation treatment.

2.3. Analysis of the Water Samples

The water samples were analyzed following NTPA-001:2002, the European Union Directive 91/271/EEC, and corresponding standards [13,14]. pH measurements were performed using a calibrated pH meter equipped with a glass electrode (Seven Compact with InLab[®] Viscous Pro-ISM electrode, Mettler Toledo, Columbus, OH, USA), according to ISO 10523:2008 [13]. Electrical conductivity was determined using a calibrated conductivity meter (model E-45, Phoenix Instrument, Garbsen, Germany). The sample was directly measured without dilution, in conformity with ISO 7888:1985 [14]. Chloride concentration was determined using titration with silver nitrate (AgNO_3) and sodium chromate, with ISO 9297:2001 [15] as a guide. The chemical oxygen demand (COD) analysis was conducted using the sealed tube method. The sample was treated with a dichromate solution in the presence of a silver catalyst, as in the ISO 6060:1996 standard [16]. The biochemical oxygen demand (BOD₅) was determined by incubating the sample for five days at 20 °C in an incubator (Algaetron A230, Photon Systems Instruments, Drásov, Czech Republic). Dissolved oxygen levels were measured before and after the incubation with an oxygen meter HI98193 (Hanna Instruments, Nufalau, Romania), and the BOD₅ value was calculated according to SR EN 1899:2003-2 [17]. Total suspended solids (TSS) content was determined by filtering a known sample volume through a pre-weighed glass fiber filter. The filter was then dried in a laboratory oven (UE200 Memmert, Buechenbach, Germany). The glass filter

increase in weight corresponded to the TSS concentration, according to ISO 11923:1997 [18]. The residue at 105 °C was determined by evaporating a determined sample volume at 105 °C until constant weight (using an analytical balance, model MS105DU, Mettler Toledo, Columbus, OH, USA). The residue remaining after evaporation represented the non-volatile solids in the sample -STAS 9187-84 [19]. Extractable organic substances were determined by extracting the sample with an appropriate solvent (petroleum ether or hexane), followed by gravimetric analysis SR 7587:1996 [20]. Total nitrogen analysis was performed according to SR EN ISO 11905:2003 [21] as the total nitrogen from the forms of free ammonia, ammonium, nitrites, nitrates, and nitrogenous organic compounds that can be converted into nitrates under oxidative conditions achieved with sodium peroxodisulfate. Quantification was performed with a spectrometer (Ocean FX[®] UV-Vis spectrometer Ocean Insights, Orlando, FL, USA). The biodegradable synthetic detergents were analyzed using the spectrophotometric methods specified in SR EN 903:2003 [22], as the index of active substances on methylene blue (MBAS). Total phosphorus was determined by the ammonium molybdate spectrometric method, involving the reaction of phosphorus with ammonium molybdate to form a blue complex, following ISO 6878:2008 [23].

3. Results and Discussion

The determined water quality parameters are illustrated in the following figures, compared with the thresholds established by the NTPA-001:2002 and the European Union Directive 91/271/EEC. Figure 1 represents pH, electrical conductivity, and chloride values, and Figure 2 illustrates COD, BOD 5, and total suspended solids values.

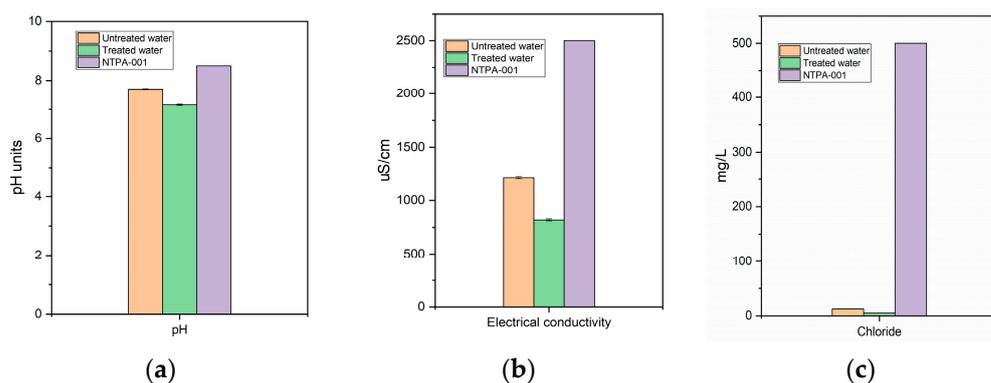


Figure 1. pH (a), electrical conductivity (b), and chloride values (c), determined for initial samples from untreated water, samples after bioaugmentation treatment, and maximum threshold values.

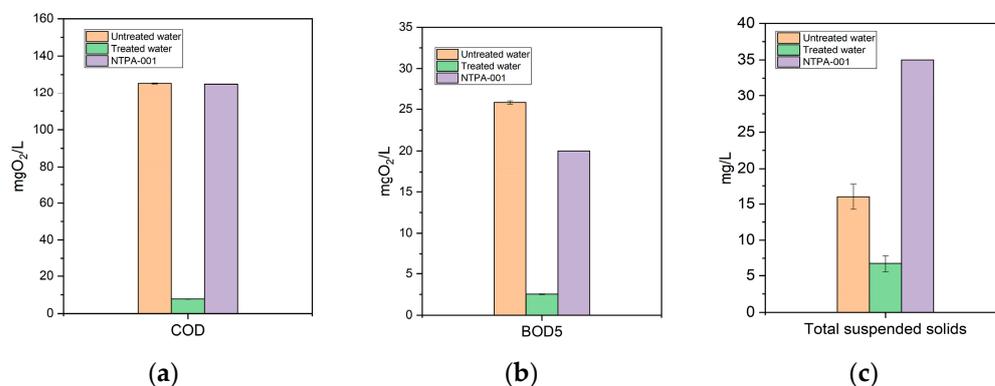


Figure 2. Chemical oxygen demand (COD, a), biochemical oxygen demand (BOD5, b), total suspended solid (c), determined for initial samples from untreated water, samples after bioaugmentation treatment, and maximum threshold values.

The pH values for both untreated (7.71) and treated (7.15) samples were within the acceptable range of 6.5 to 8.5, meeting the NTPA-001 criteria. Similarly, the electrical

conductivity values (1210, respectively, $820 \pm 10 \mu\text{S}/\text{cm}$) remained below the specified maximum threshold ($2500 \mu\text{S}/\text{cm}$), indicating compliance with the standard. Chlorides are a significant water quality parameter that measures the concentration of chloride ions (Cl^-) in water. The measured values in samples before treatment (12.8 mg/L) and after treatment (5.3 mg/L) were below the permissible limits set by the NTPA-001 standard [24].

COD is an essential parameter used to assess the organic content and pollution level in water. On the water sample before the bioaugmentation treatment, the COD measured was 125.33 mg/L, being at the superior limit of the NTPA-001, while after the treatment, the water COD value was reduced to 7.7 mg/L.

BOD5 indicates the level of organic matter pollution in the water and helps assess its overall health and the potential for self-purification. The substantial decrease in BOD5 between the water samples taken before the inoculation ($25.8 \text{ mg O}_2/\text{L}$) and after the inoculation ($2.5 \text{ mg O}_2/\text{L}$) suggests a successful bioaugmentation. The high level of BOD5 is most probably due to the COVID-19 pandemic conditions, which affected the functionality of wastewater treatment plant, due to high level of disinfectants and other organic pollutants [25,26]. Bioaugmentation with robust bacterial strains, with high organic degradative ability, seems to re-balance the functionality of the wastewater treatment plant.

Total suspended solids (TSS) represent the concentration of particles and solid matter that remain in the water after filtration and drying at 105°C . After the application of the treatment, the TSS value decreased by 57% from the initial value (16 mg/L), down to 7 mg/L. The values were both under the NTPA-001.

Figure 3 illustrates residues at 105°C , extractible organic substances, total nitrogen, determined for initial samples from untreated water, samples after bioaugmentation treatment, and maximum thresh- old values.

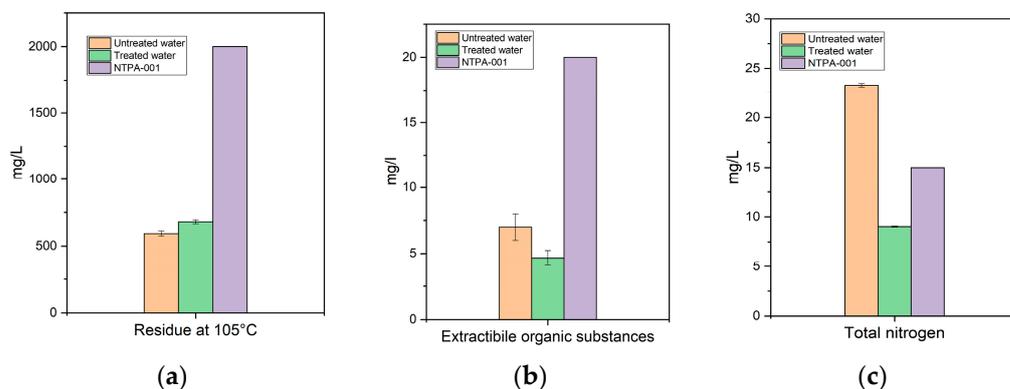


Figure 3. Residues at 105°C (a), extractible organic substances (b), total nitrogen (c), determined for initial samples from untreated water, samples after bioaugmentation treatment, and maximum threshold values.

Residue at 105°C represents the sum of all the organic and inorganic substances dissolved in the water. After the treatment, a slight increment occurred ($680 \text{ mg}/\text{L}$ vs. $593 \text{ mg}/\text{L}$), but remained within the acceptable limit. Most probably, the increase residues at 105°C is due to enhanced mineralization of organic matter—and the enhancement of the mineralization process is supported by the finding related to BOD5.

The threshold for extractible organic substances, as indicated by the NTPA-001 standard [24], is $20 \text{ mg}/\text{L}$. Both water samples were well below this limit, and after the treatment, a 33% lower value was obtained ($4.8 \text{ mg}/\text{L}$), indicating that the treatment process effectively reduced the concentration of extractible organic substances in the water.

On the first water sample (untreated), the total nitrogen concentration in the water samples taken before bioaugmentation treatment was $23.3 \text{ mg}/\text{L}$, higher than the $15 \text{ mg}/\text{L}$ NTPA-001 threshold. This value correlates with the higher level of BOD and probably also resulted from the overloading of wastewater treatment plants with organic pollutants

during the COVID-19 pandemic, as already mentioned [25,26]. After the treatment, this parameter decreased to 9.04 mg/L.

Figure 4 represents the biodegradable synthetic detergents and total phosphorus, determined for initial samples from untreated water, samples after bioaugmentation treatment, and maximum threshold values according to regulations in force.

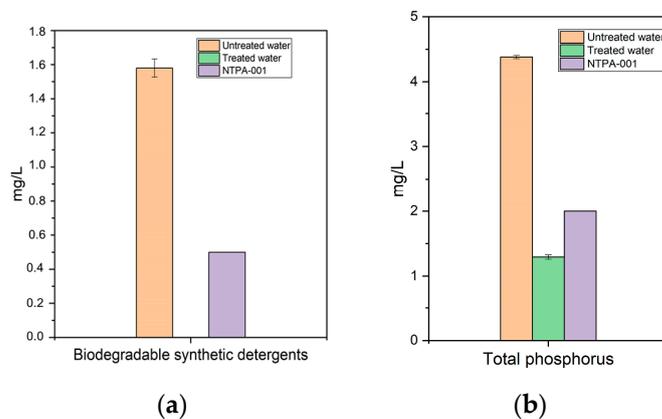


Figure 4. Biodegradable synthetic detergents (a), and total phosphorus (b), determined for initial samples from untreated water, samples after bioaugmentation treatment, and maximum threshold values.

Biodegradable synthetic detergents negatively impact the environment due to their foaming effect, contribution to eutrophication, and altered salinity and pH [27]. In the water sample taken before bioaugmentation treatment, the value was 1.58 mg/L, more than three times over the NTPA-001 limit [24]. Synthetic detergent consumption (including soaps) was much higher during the COVID-19 pandemic and significantly influenced wastewater treatment plants [18]. Notably, after the treatment, the concentration of biodegradable synthetic detergents was below the detection limit (<0.2 mg/L) in the treated samples, indicating effective treatment to remove these substances. The application of bacteria from *Bacillus* and *Pseudomonas* genera (*sensu lato*) was recently proposed as a sustainable approach for synthetic detergent degradation and environmental protection [28].

Total phosphorus represents the sum of all forms of phosphorus present in the water, including dissolved and particulate phosphorus compounds. The concentration of total phosphorus in the water sample taken before bioaugmentation treatment was found to be 4.38 mg/L, 2.4 times over the NTPA-001 limit (2 mg/L). Such an increase in phosphorus concentration is probably also related to the increased detergent utilization [29]. In the treated water sample, the total of phosphorus decreased to 1.29 mg/L, indicating that the treatment process effectively reduced the total phosphorus concentration in the water.

Overall, the results indicate the performance of the bacterial consortium used as inoculant for the bioaugmentation of a small wastewater treatment plant using MBBR technology.

4. Conclusions

The bioaugmentation treatment with the bacterial consortia, which includes a gram-positive *Brevibacillus parabrevis* B50 NCAIM B 001413 bacterial strain and a gram-negative *Pseudoxanthomonas mexicana* P32 NCAIM (P) B 001414 bacterial strain, proved to be effective in the re-equilibration of the main functions of wastewater treatment plant unbalanced due to higher level of organic contaminants resulting from specific conditions of the COVID-19 pandemic. The data presented in this work assess the efficiency of the bioaugmentation treatment that ensures compliance with environmental regulations to safeguard water resources.

Author Contributions: Conceptualization, E.-G.B. and F.O.; methodology, O.-A.C.; validation, D.C.-A. and F.O.; formal analysis, E.-G.B.; investigation, E.-G.B. and D.C.-A.; resources, F.O.; data curation, D.C.-A.; writing—original draft preparation, E.-G.B.; writing—review and editing, F.O.; visualization, D.C.-A.; supervision, F.O.; project administration, F.O.; funding acquisition, F.O. All authors have read and agreed to the published version of the manuscript.

Funding: The research leading to these results has received funding from European Regional Development Fund (ERDF), the Competitiveness Operational Program (POC), Axis 1, project POC-A1-A1.2.3-G-2015-P_40_352, My_SMIS 105684, “Sequential processes of closing the side streams from bioeconomy and innovative (bio)products resulting from it—SECVENT”, subsidiary projects 1518/2019 and 1612/2018 BioDeg.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data are included in the present work.

Acknowledgments: The authors thank Vasile Surugiu, manager of Apă-Canal Srl Filipești-de-Pădure, for his support.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

1. Herrero, M.; Stuckey, D.C. Bioaugmentation and its application in wastewater treatment: A review. *Chemosphere* **2015**, *140*, 119–128. [[CrossRef](#)] [[PubMed](#)]
2. Ma, H.; Zhao, Y.; Yang, K.; Wang, Y.; Zhang, C.; Ji, M. Application oriented bioaugmentation processes: Mechanism, performance improvement and scale-up. *Bioresour. Technol.* **2022**, *344*, 126192. [[CrossRef](#)] [[PubMed](#)]
3. Morgan-Sagastume, F. Biofilm development, activity and the modification of carrier material surface properties in moving-bed biofilm reactors (MBBRs) for wastewater treatment. *Crit. Rev. Environ. Sci. Technol.* **2018**, *48*, 439–470. [[CrossRef](#)]
4. McQuarrie, J.P.; Boltz, J.P. Moving Bed Biofilm Reactor Technology: Process Applications, Design, and Performance. *Water Environ. Res.* **2011**, *83*, 560–575. [[CrossRef](#)] [[PubMed](#)]
5. Fu, H.-m.; Wang, J.; Ren, H.; Ding, L. Acceleration of start-up of moving bed biofilm reactor at low temperature by adding specialized quorum sensing bacteria. *Bioresour. Technol.* **2022**, *358*, 127249. [[CrossRef](#)] [[PubMed](#)]
6. Irankehah, S.; Ali, A.A.; Soudi, M.R.; Gharavi, S.; Ayati, B. Highly efficient phenol degradation in a batch moving bed biofilm reactor: Benefiting from biofilm-enhancing bacteria. *World J. Microbiol. Biotechnol.* **2018**, *34*, 164. [[CrossRef](#)] [[PubMed](#)]
7. Chen, X.; Yuan, C.B.; Zhu, Y.A.; Liu, H.; Chen, W.; Zhang, Q. Bioaugmentation with *Acinetobacter* sp. TAC-1 to enhance nitrogen removal in swine wastewater by moving bed biofilm reactor inoculated with bacteria. *Bioresour. Technol.* **2022**, *359*, 127506. [[CrossRef](#)] [[PubMed](#)]
8. Cheoafa, O.A.; Constantinescu-Aruxandei, D.; Popa, D.G.; Dimitriu, L.; Oancea, F.; Cornea, C.P. Screening of bacterial consortia for a bioaugmented bioassay of flushable wipes biodegradation. *Agrolife Sci. J.* **2022**, *11*, 27–36. [[CrossRef](#)]
9. Cornea, P.C.; Oancea, F.; Israel, R.; Raut, I.; Voaides, C.M.; Burlacu, A. Selecting Microbial Consortium with High Organic Matter Degradation Ability. RO132587-B1, 2020.
10. Margarit, E.; Oancea, F.; Constantinescu-Aruxandei, D.; Cheoafa, O.A. Selecting Consortia of Microorganism that Involves Selecting Potential Consortia of Mutualistic Microorganisms, Identifying Compatible Combinations, and Chemical Mapping. RO135838-A2, 2022.
11. Calin, M.; Doni, M.; Jecu, M.L.; Oancea, F.; Pairault, A.L.; Raut, I.; Jecu, M.; Pairault, A. Brevibacillus Parabrevis Strain and Controlled Release Composition Based on the Same. EP2765185-A3, 2014.
12. Oancea, F.; Raut, I.; Şesan, T.E.; Cornea, P.C.; Badea-Doni, M.; Popescu, M.; Jecu, M.L. Hydro-gelified and film forming formulation of microbial plant biostimulants for crop residues treatment on conservation agriculture systems. *Stud. Univ. Vasile Goldis Ser. Stiintele Vietii (Life Sci. Ser.)* **2016**, *26*, 251–260.
13. ISO 10523:2008; Water Quality—Determination of pH. International Organization for Standardization (ISO): Geneva, Switzerland, 2008.
14. ISO 7888:1985; Water Quality—Determination of Electrical Conductivity. International Organization for Standardization (ISO): Geneva, Switzerland, 1985.
15. ISO 6060:1996; Water Quality—Determination of the Chemical Oxygen Demand. International Organization for Standardization (ISO): Geneva, Switzerland, 1996.
16. SR ISO 9297:2001; Water Quality—Determination of Chloride Content. Titration with Silver Nitrate Using Chromate as Indicator (Mohr Method). ISO: Geneva, Switzerland, 2001.
17. SR EN 1899-2:2002; Water Quality—Determination of Biochemical Oxygen Demand after N Days (CBO_n). Part 2: Method for Undiluted Samples. CEN: Brussels, Belgium, 2002.

18. ISO 11923:1997; Water Quality—Determination of Suspended Solids by Filtration through Glass-Fibre Filters. ISO: Geneva, Switzerland, 1997.
19. STAS 9187-84; Surface water, groundwater and waste water, Determination of the residue. ASRO: Bucharest, Romania, 1984.
20. SR 7587:1996; Water quality—Determination of solvent-extractable substances, Gravimetric method. ASRO: Bucharest, Romania, 1996.
21. SR EN ISO 11905-1:2003; Water Quality—Determination of Nitrogen Content. Part 1: Oxidative Mineralization Method with Peroxodisulphate. ISO: Geneva, Switzerland, 2003.
22. NTPA 001; Pollutant Load Limit Values for Industrial and Urban Waste Water Discharged into Natural Receptors. Romanian Government Decision 188/2002-Annex 3. Available online: <http://legislatie.just.ro/Public/DetaliuDocumentAfis/98311> (accessed on 14 November 2023).
23. Grimeaud, D. The EC Water Framework Directive—An Instrument for Integrating Water Policy. *Rev. Eur. Community Int. Environ. Law* **2004**, *13*, 27–39. [[CrossRef](#)]
24. Hernández-Chover, V.; Castellet-Viciano, L.; Fuentes, R.; Hernández-Sancho, F. Circular economy and efficiency to ensure the sustainability in the wastewater treatment plants. *J. Clean. Prod.* **2023**, *384*, 135563. [[CrossRef](#)]
25. Parveen, N.; Chowdhury, S.; Goel, S. Environmental impacts of the widespread use of chlorine-based disinfectants during the COVID-19 pandemic. *Environ. Sci. Pollut. Res.* **2022**, *29*, 85742–85760. [[CrossRef](#)]
26. Di Marcantonio, C.; Chiavola, A.; Gioia, V.; Frugis, A.; Cecchini, G.; Ceci, C.; Spizzirri, M.; Boni, M.R. Impact of COVID19 restrictions on organic micropollutants in wastewater treatment plants and human consumption rates. *Sci. Total Environ.* **2022**, *811*, 152327. [[CrossRef](#)] [[PubMed](#)]
27. Mousavi, S.A.; Khodadoost, F. Effects of detergents on natural ecosystems and wastewater treatment processes: A review. *Environ. Sci. Pollut. Res.* **2019**, *26*, 26439–26448. [[CrossRef](#)] [[PubMed](#)]
28. Chirani, M.R.; Kowsari, E.; Teymourian, T.; Ramakrishna, S. Environmental impact of increased soap consumption during COVID-19 pandemic: Biodegradable soap production and sustainable packaging. *Sci. Total Environ.* **2021**, *796*, 149013. [[CrossRef](#)]
29. Arora, J.; Ranjan, A.; Chauhan, A.; Biswas, R.; Rajput, V.D.; Sushkova, S.; Mandzhieva, S.; Minkina, T.; Jindal, T. Surfactant pollution, an emerging threat to ecosystem: Approaches for effective bacterial degradation. *J. Appl. Microbiol.* **2022**, *133*, 1229–1244. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.